

Life in Submarine Hell

Lidia Montiel Fontanet

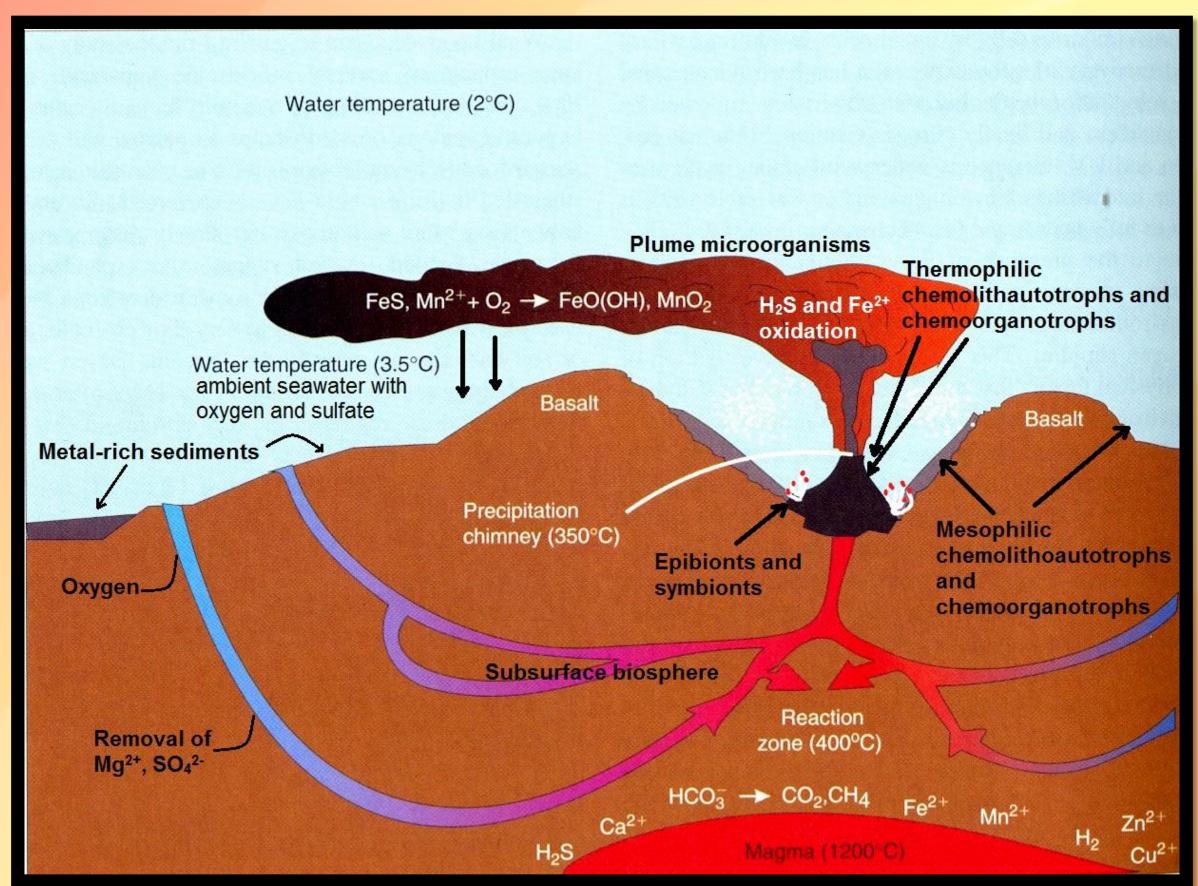
Treball Fi de Grau 2013 – Grau de Microbiologia





INTRODUCTION

Deep-sea hydrothermal vents were first discovered in 1977. They are geophysical and tectonic processes that occur at seafloor spreading centers (Fig.1). The vent fluid are hot (350°C), anoxic and contain high concentrations of hydrogen sulfide and other compounds. An extrem environment given by the pressure (260 atm), high temperature, chemical toxicity of the fluids and total lack of photosynthetic production for animal nutrition. Its communities characterized by large clams, mussels, shrimps and vestimentiferian worms thrive on chemosynthetic microbial production. Microorganisms form the base of the ecosystem's food chain which are primary producers and also live symbiotically with larger fauna.



Locations of the submarine hydrothermal vents.

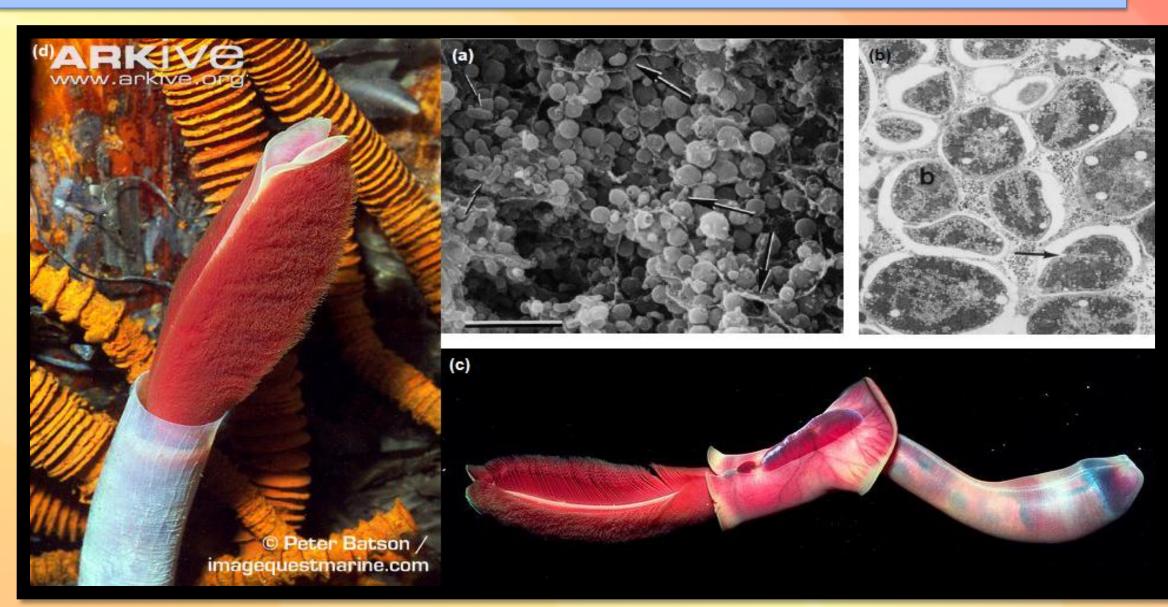


Fig.3 Riftia pachyptila and chemosynthetic endosymbionts. (a) Symbionts within trophosome, symbionts occur as both spherical and rod-shaped cells. (b) Portion of trophosome lobule. (c) Giant tube worm specimen. (d) Tevnia jerichonana in background.

Fig.1 Formation and a sketch of a hydrothermal vent.

RESULTS

Most of vent fauna depends on H₂S to survive. Symbiotic relationships:

- Riftia pachyptila -> giant tube worm without mouth and digestive system. Chemoautotrophic bacterial symbionts are inside his trophosome (Fig.3).
- Large clams (Calyptogena) and mussels (Bathymodiolus) have bacterial endosymbionts too \rightarrow majority are gammaproteobacteria.
- The pompeii worms (Alvinella pompejana) and shrimps (Rimicaris) have epibiotic bacteria \rightarrow majority are epsilonproteobacteria.

There is a considerable diversity of bacteria in which epsilonproteobacteria are the most abundant (Fig.4).

Large part of found archaea are methanogenics or marine groups Crenarchaeota total 16S rRNA gene pool. and Euryarchaeota representatives.

The most significant microbial process taking place is chemosynthesis: biosynthesis of organic carbon compounds from CO2 with the source of energy being chemical oxidations.

Dominant autotrophic metabolisms according to the groups of microorganisms:

- **Mesophiles** (2-40°C) as gamma- and epsilonproteobacteria \rightarrow H₂S/S° and H₂ oxidation with O_2 or NO_3^{2-} .
- (40-80°C) Thermophiles epsilonproteobacteria, Aquificales, Desulfurobacteriaceae and methanogenic archaea \rightarrow same as above but H₂ oxidation coupled to SO_4^{2-} and S^0 reduction and methanogenesis.
- Hyperthermophiles (80-125°C) as Desulfurobacteriaceae, methanogenic archaea, Thermococcales and crenarchaeota \rightarrow methanogenesis, S^0 reduction (Fig.5) and acetogenesis?.

Taxonomic level Phylum	Taxonomic level Class	% of prokaryotic SSU 454 sequence reads
Proteobacteria	Gammaproteobacteria	0.2
Proteobacteria	Betaproteobacteria	0.1
Proteobacteria	Epsilonproteobacteria	36.1
Proteobacteria	Deltaproteobacteria	0.1
Firmicutes	Clostridia	0.1
Deferribacteres	Deferribacterales	0.1
Thermotogae	Thermotogae	0.6
Aquificae	Aquificae	26.1
Thermodesulfobacteria	Thermodesulfobacteria	0.8
Candidate division SR1	_	0.8
Chloroflexi	Dehalococcoides	0.1
Crenarchaeota	Thermoprotei	0.9
Euryarchaeota	Thermoplasmata	0.3
Euryarchaeota	Methanococci	2.8
Euryarchaeota	Archaeoglobi	1.8
Euryarchaeota	Thermococci	28.4

Bold font indicates taxonomic groups represented by more than 1% of t

Fig.4 Taxonomic affiliation, abundances and 16S rRNA gene numbers of microbial populations hydrothermal vent at the Arctic Mid-Ocean Ridge.

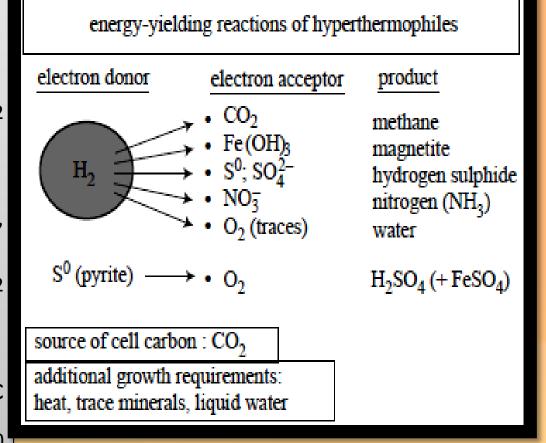


Fig.5 Schematic of main energy-yielding reactions in chemolithoautotrophic hyperthermophiles.

Hyperthermophiles

- Bacteria
 - Thermotoga 80ºC
 - Aquifex 85°C (most thermophile bacteria)
- Archaea
 - Phylum Euryarchaeota
 - Thermococcales: *Thermococcus* 90°C; *Pyrococcus* 100°C
 - Methanococcales: Methanocaldococcus 85°C
 - Methanopyrales: *Methanopyrus* 100°C
 - Nanoarchaeum 90°C: obligate symbiotic of Ignicoccus
 - Arqueogloblales: Archaeoglobus 83°C; Ferroglobus 85°C
 - Phylum Crenarchaeota
 - Thermoproteales: *Pyrobaculum* 100°C
 - Desulfurococcales: *Pyrodictium* 105°C; *Pyrolobus* 105°C; Strain 121 106°C; Desulfurococcus 85°C; Ignicoccus 90°C; Staphylothermus 92°C

Nanoarchaeum equitans is a member of a novel group of hyperthermophilic virus-sized archaea. It has the smallest and the most compact microbial genome known to date. Cells grow attached to the surface of a specific crenarchaeal host, a member of the genus *Ignicoccus*.⁴

Archaeal strain 121 has upper temperature limit of 121°C, which is the highest upper temperature limit reported.

The membrane lipids of thermophiles contain more saturated and straight chain fatty acids than mesophiles. This allows to grow at higher temperatures by providing the right degree of fluidity needed for membrane function. Many archaeal species contain a paracrystalline surface layer (S-layer) with protein or glycoprotein and this is likely as an external protective barrier.

Histone-like proteins identified in hyperthermophiles protect their DNA (Fig.7). Reverse gyrase, a type 1 DNA topoisomerase causes positive supercoiling and stabilize the DNA. Heat shock proteins, chaperones, are likely to play a role in stabilizing and refolding proteins as they begin to denature.4

STATE OF THE ART

- Biotechnology interest^{4,3}. Wide industrial and medical applications:
 - Useful to combat industrial pollution (H₂S) and clean up sites contaminated with Cu, Cd, Hg,...
 - Source of new heat resistant industrial chemicals and new drugs to combat germs now resistant to plant and soil based drugs.
 - Renewable source of natural gas.
 - Proteins and enzymes of hyperthermophiles with high thermal stabilities: longer shelf life, produce less waste and lower risk of contamination.
 - Paper and detergent industries.
 - DNA polymerases for DNA amplifications by PCR (*Thermococcus littoralis*)
- Inactive sulfide chimneys represent a biogeochemically active microbial ecosystem. Communities on inactive sulfides are different from those on active chimneys.
- Interest in origin of life studies.

Fig.6 Cells of *N. equitans* (small) attached to Ignicoccus.

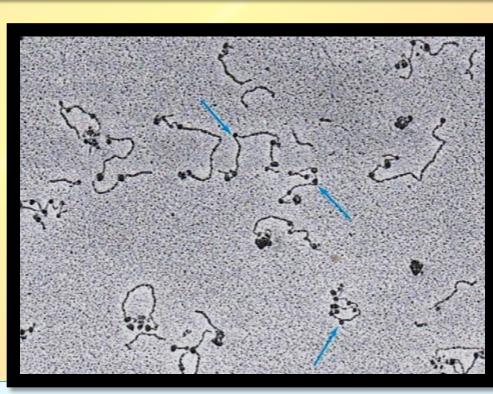


Fig.7 Histone like proteins protecting DNA.

CONCLUSIONS

- Microbial primary producers have an important role to keep alive these ecosystems.
- The findings of recent years revealed significant metabolic and phylogenetic diversity of cultivated thermophilic microorganisms.
- The discovery of Nanoarchaeota suggests that further major groups of microbes may still be unrecognized, and are waiting for their isolation to tell us more about the origin and evolution of life.
- The potential biotechnological applications associated with these microbes and their products has driven extensive and intensive research efforts on deep-sea hydrothermal vent microorganisms.

REFERENCES

- Grassle, J.F. Hydrothermal vent animals: distribution and biology. Science, 1985, 229, 713–717.
- Jannasch, H.W.; Mottl, M.J. Geomicrobiology of deep-sea hydrothermal vents. Science, 1985, 229, 717–725.
- Satyanarayana, T.; et al. Extremophilic microbes: diversity and perspectives. Current Science, 2005, 89, 78–90. Stetter, K.O. Hyperthermophiles in the history of life. Phil. Trans. R. Soc. B, 2006, 361, 1837–1843.

Miroshnichenko, M.L.; et al. Recent developments in the thermophilic microbiology of deep-sea hydrothermal vents. Extremophiles, 2006, 10, 85–96.

Suárez Arriaga, M.C. Evaluación del potencial, biogénesis y características esenciales de los sistemas geotérmicos submarinos en México. Geotermia, 2004, 17, 31–43. Sylvan, J.B; et al. Life and death of deep-sea vents: bacterial diversity and ecosystem succession on inactive hydrothermal sulfides. mBio, 2012, 3, e00279-11.