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1           **SYMPTOMS AND FUNGI ASSOCIATED WITH DECLINING**  
2           **MATURE GRAPEVINE PLANTS IN NORTHEAST SPAIN**

3

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20   Running title: Grapevine decline and associated pathogenic fungi

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26 *Key words:* black dead arm, esca, eutypiose, grapevine decline, phytopathogenic fungi,  
27 *Vitis vinifera*

28

29 **Summary**

30

31 A field survey was carried out in the Catalonia region of northeast Spain to  
32 characterize the decline of mature grapevines. The relationships of both external and  
33 internal symptoms of diseased plants and their associated mycoflora were studied.  
34 Co-occurrence of different internal disease symptoms was frequent in Catalonia, since  
35 44% of sampled plants had wood lesions commonly associated with at least two of the  
36 following decline diseases: eutypiose, black dead arm or esca. The results obtained also  
37 suggest that apoplexy might not be associated only with esca-affected plants, since 60%  
38 of surveyed plants showing apoplexy were also affected by V-shaped necroses which  
39 are commonly associated with eutypiose and black dead arm, and 20% were exclusively  
40 affected by V-shaped necroses. An experiment was conducted to establish the  
41 pathogenicity of most representative fungi isolated from the diseased tissues of  
42 declining plants, by artificially inoculating 1-year-old plants of 'Macabeo' and  
43 'Tempranillo' varieties. Pathogenicity of fungi, as indicated by vascular lesion  
44 extension, was confirmed for most of the species tested, namely *Botryosphaeria*  
45 *dothidea*, *Diplodia seriata*, *Eutypa lata*, *Neofusicoccum luteum*, *N. parvum* and  
46 *Phaeomoniella chlamydospora*.

47

48 **Introduction**

49

50 The area of vineyards in Spain consists of about 1,2 Mha, thus making  
51 Spain the leading country in grapevine culture in the world, the third in the  
52 world in wine production, and the second in raisin production (data from OIV,  
53 year 2005 [retrieved from Internet Oct. 15, 2008 at <http://www.oiv.org>];  
54 Anonymous, 2006). Grapevines are widespread in Spain, where about 97% of  
55 the total grapevine area is managed for wine production (Anonymous, 2006).  
56 Although viticulture is an essential component of the agriculture sector in Spain,  
57 grapevine declines and their associated pathogenic fungi are poorly known in  
58 this country. In the last decade, studies on diseases and pathogenic mycoflora  
59 associated to rootstocks (Aroca *et al.*, 2006), young vines (Armengol *et al.*, 2002;  
60 Giménez-Jaime *et al.*, 2006), and mature vines (Armengol *et al.*, 2001a, b; Úrbez-  
61 Torres *et al.*, 2006a) have been carried out in Spain, but further studies are  
62 needed to increase the knowledge regarding these complex diseases.  
63 Unfortunately, no quantitative data about the economic impact of these  
64 declines on Spanish grapevine production have been calculated. Main decline  
65 diseases of mature grapevine observed in Spain include esca, eutypiose, and  
66 black dead arm (BDA), as reported by Armengol *et al.* (2001a, b) and Úrbez-  
67 Torres *et al.* (2006a).

68

69 Esca is a complex disease where symptoms and their expression over  
70 time are highly variable (Mugnai *et al.*, 1999; Surico *et al.*, 2006). Two main types  
71 of esca episodes can be defined: the chronic esca and the acute syndrome, the  
72 latter also known as apoplexy (Mugnai *et al.*, 1999). Briefly, foliar symptoms of  
73 chronic esca are characterized by interveinal chlorosis or discolorations  
74 (yellowish in white cultivars and reddish in red cultivars) that later coalesce in  
75 large necrotic areas during summer. Vine apoplexy usually occurs in mid  
76 summer, when leaves of affected plants wither rapidly in a few days (Mugnai *et*  
77 *al.*, 1999). Despite of the external foliar symptoms of both esca types, several  
78 types of wood degradation have been described for esca, mainly including i)  
79 longitudinal brown streakings that appear as necrotic black spots in cross  
80 sections, ii) pink-brown or dark red-brown necrotic areas, and iii) wood decay.

81 Many fungi have been reported to be involved in the esca syndrome; several  
82 Basidiomycetes species are responsible for the wood decay, with species in the  
83 genera *Fomitiporia*, *Fomitiporella*, and *Inocutis* (Fischer, 2006), while vascular  
84 necroses are caused mainly by *Phaeomoniella chlamydospora* and several  
85 *Phaeoacremonium* species (Surico *et al.*, 2006). Eutypiose, also known as Eutypa  
86 dieback, is caused by the fungus *Eutypa lata* Carter (1988). The most recognized  
87 symptom of this disease is the stunted appearance of shoots at the early growth  
88 season, with small, cupped, and chlorotic leaves, and short internodes. Wood  
89 internal symptoms include characteristic V-shaped necroses when cross sections

90 of affected arms and trunks are made. Additionally, external cankers developing  
91 from old pruning wounds can be observed. BDA was first described by Lehoczky  
92 (1974) who associated this disease with *Botryosphaeria stevensii*, but later  
93 several other species of Botryosphaeriaceae have been associated with the  
94 disease, including *B. dothidea*, *Diplodia seriata* and *Lasiodiplodia theobromae* as  
95 the most frequently related fungi (Larignon *et al.*, 2001; van Niekerk *et al.*, 2006).  
96 Wood symptoms of BDA include V-shaped necroses, similar to those caused by  
97 *E. lata*, and longitudinal brown streakings along the affected tissues. Stunted  
98 growth in early season has also been described occasionally for diseases caused  
99 by Botryosphaeriaceae species (Castillo-Pando *et al.*, 2001; Taylor *et al.*, 2005),  
100 thus resembling the symptoms caused by *E. lata*. BDA foliar symptoms are also a  
101 matter of controversy; while Lehoczky (1974) reported a slight diffuse chlorosis  
102 and an eventual later leaf wilting, Larignon and Dubos (2001) reported an early  
103 red or yellow-orange patching (in red and white grape varieties, respectively)  
104 that later develop large marginal and interveinal necroses. However, Lecomte *et*  
105 *al.* (2005) and Surico *et al.* (2006) have shown and discussed the similarity  
106 between these late BDA foliar symptoms and those typical of esca. Additional  
107 more comprehensive information on the above diseases can be found in Carter  
108 (1988; 1991), Larignon *et al.* (2001), Lecomte *et al.* (2005), Mugnai *et al.* (1999),  
109 Surico *et al.* (2006) and van Niekerk *et al.* (2006).  
110

111        The present study aims to characterize the grapevine decline of mature  
112        grapevines in the northeast Spanish region of Catalonia, by 1) determining the  
113        relationship of both external and internal symptoms of diseased plants with the  
114        existing mycoflora, and 2) establishing the pathogenicity in grapevine of fungi  
115        isolated from the diseased tissues of declining plants.

116

117        **Materials and methods**

118

119        *Field survey*

120

121        Seventy-nine vineyards known to be affected by decline diseases from previous  
122        field surveys were visited between 2003 and 2005 in Catalonia, NE Spain. Field data  
123        and plant samples were collected each year from May to August. Eighteen grapevine  
124        varieties and three rootstocks were surveyed: the white varieties included Chardonnay,  
125        Chenin Blanc, Garnatxa Peluda, Macabeo, Parellada, Sauvignon Blanc, White  
126        Grenache, and Xarel·lo; the red varieties included Cabernet Sauvignon, Carignane,  
127        Merlot, Pinot Noir, Red Grenache, Syrah, and Tempranillo; three unknown varieties and  
128        three rootstocks (110R, 140Ru and SO4). A total of 192 vines showing decline  
129        symptoms were surveyed from over 1500 total inspected plants (about 20 decline-  
130        affected plants being examined per vineyard). Two to four affected plants per vineyard  
131        were chosen for a careful symptom examination and wood sampling. Declining vines  
132        were examined visually and the nature of the external symptoms was annotated and  
133        attributed to known diseases: eutypiose, BDA or esca. Vines which showed stunted

134 shoot growth in late spring, and V-shaped wood necroses were classified as affected by  
135 eutypiose/BDA during the field survey, since both diseases show similar symptoms  
136 (Castillo-Pando *et al.*, 2001; Taylor *et al.*, 2005). BDA foliar symptoms occurring in  
137 summer, as described by Larignon and Dubos (2001), were not considered as these  
138 symptoms could be confused with those of esca (Lecomte *et al.*, 2005; Surico *et al.*,  
139 2006). Vines with characteristic interveinal chloroses and necroses, wood decay and  
140 vascular necroses different from V-shaped ones were classified as esca-affected plants.  
141 Vines with sectorial necrosis and either one of the esca-associated wood necroses were  
142 classified as affected by eutypiose/BDA and esca. Plants affected by apoplexy whether  
143 partial (1 to several arms) or total (whole plant) were considered as a separate class  
144 from the above diseases. Sections of trunks and arms, as well as the whole plant when  
145 appropriate, were taken to the laboratory for further examination and to conduct fungal  
146 isolations.

147

148 *Fungal isolation and identification*

149

150 Cross and longitudinal sections of diseased arms and trunks were carefully  
151 examined and the type of wood necrosis was recorded. Four types of wood alteration  
152 were considered: V-shaped necroses, irregular central necroses, black spots seen  
153 in cross sections, and wood decay (see Figure 1, c to h). Pieces of each type of  
154 necrosis (approximate size 10 cm in length) were obtained every 20 cm of affected arms  
155 and trunks, and processed separately to isolate the fungi. Wood chips (about 5x5x5 mm;  
156 minimum n=15 pieces per sample and necrosis type) were surface-sterilized (3-4 min in  
157 70% ethanol), blotted on sterile filter paper to remove excessive ethanol, and plated

158 onto Potato Dextrose Agar (PDA, Difco Laboratories, Detroit, MI, USA) amended with  
159 sulphate streptomycin (Sigma-Aldrich Co., St. Louis, MO, USA) at 100 units per ml  
160 (Johnston and Booth, 1983). Plates were incubated at 25 °C in darkness to get pure  
161 cultures. When necessary, sporulation was induced by incubating the fungal colonies in  
162 water agar with sterilized grapevine wood chips at 25 °C and under near-UV  
163 light/darkness for 12/12 hours. Representative isolates were maintained at 4 °C in sterile  
164 distilled water tubes with mycelial plugs.

165

166 Isolated fungi were identified on the basis of morphological characters of  
167 colonies and reproductive structures. Eventually, identification of isolates was  
168 confirmed by analysing the DNA sequences from selected regions: the internal  
169 transcribed spacers ITS1 and ITS2 flanking the 5.8s rRNA gene (ITS), and parts of the  
170 translation elongation factor 1-alfa (EF1- $\alpha$ ) and the  $\beta$ -tubulin genes when applicable.  
171 Procedures of DNA extraction were done as described in Alves *et al.* (2004), and PCR  
172 amplifications were done according to the methods described by Alves *et al.* (2004)  
173 [ITS; for most of fungi], Phillips *et al.* (2005) [EF1- $\alpha$ ; for Botryosphaeriaceae species]  
174 and Mostert *et al.* (2006) [ $\beta$ -tubulin; for *Phaeoacremonium* spp. and  
175 Botryosphaeriaceae species]. DNA sequencing was done as described by Alves *et al.*  
176 (2004). All regions were sequenced in both strands to clarify any nucleotide ambiguous  
177 position. BLAST searches at GenBank showing high identites with reference sequences  
178 (>97%) were used to confirm the identifications.

179

180 *Pathogenicity test*

181

182        Twenty-eight isolates representing 11 fungal taxa were chosen for the  
183        pathogenicity trial, where several isolates (2 to 5) were selected for species which  
184        showed to be more frequent after the field survey (Table 4). Artificial inoculations were  
185        conducted in May 2004 on 1-year-old grapevine plants of 'Macabeo' (white) and  
186        'Tempranillo' (red) cultivars grafted onto Richter 110 rootstocks. Plants were  
187        maintained in 3 liter pots filled with a sand:peat mixture (6:1, v:v; Floratorf peat, from  
188        Floragard, Oldenburg, Germany) and watered regularly in a greenhouse. Plants were  
189        fertilised every two weeks with 10 ml of double-strength Hoagland-Aron's solution  
190        (Hoagland and Arnon, 1950). The pathogenicity test was performed in a completely  
191        randomized experimental design, with 18 inoculated plants per cultivar and isolate. A  
192        superficial wound (15 x 5 mm, reaching into the xylem) was made on the stem of each  
193        plant with a sterilized scalpel, 10 cm above the graft union. A mycelial plug (5 mm  
194        diameter) obtained from the margin of a fungal colony was placed in the wound with  
195        the mycelium facing the stem, and the wound was wrapped with Parafilm® (Pechiney  
196        Plastic Packaging, Menasha, WI, USA). Control plants were inoculated with sterile  
197        PDA plugs instead of the fungal inoculum.

198

199        Nine months after inoculation, the length of the internal vascular lesions was  
200        recorded, by removing the bark from the stem and measuring the necrotic lesions  
201        upwards and downwards from the site of inoculation. Surface sterilized wood pieces  
202        taken from the stem necrotic tissues were plated on PDA to reisolate the inoculated  
203        fungi and thus fulfill Koch's postulates. The length of necroses was used as an indicator  
204        of the pathogenicity of fungi. Necrosis lengths were analyzed using ANOVA with the  
205        aid of the SPSS v.10 statistical package (SPSS Inc., Chicago, IL, USA), with 'grapevine

206 variety' and 'isolate' as independent factors. After ANOVA, mean values of each  
207 treatment (isolate) were compared against their respective controls with the Dunnett  
208 two-tailed test. Additional ANOVA followed by Tukey's test were used to detect  
209 differences among isolates within a given species.

210

## 211 **Results**

212

### 213 *Field survey*

214

215 A total of 192 diseased plants belonging to 18 different grapevine varieties and  
216 three different rootstocks were visually analysed and sampled for laboratory analyses.  
217 The most surveyed white varieties were the local cvs. 'Macabeo' (56 vines), 'Xarel·lo'  
218 (24) and 'Parellada' (11), whereas the red varieties included 'Tempranillo' (30), 'Red  
219 Grenache' (17), 'Cabernet Sauvignon' (14), and 'Carignane' (11). Frequencies for the  
220 remaining varieties and rootstocks were never over 5 plants. According to the external  
221 symptoms observed in the field, 58 % of the surveyed plants were diagnosed as affected  
222 by eutypiose/BDA, 19 % of plants were affected by esca, and 14 % by apoplexy. The  
223 remaining cases included dead plants (5 %), uncertain diagnoses (1 %), and plants  
224 combining both eutypiose/BDA and esca symptoms on the same individual (3 %).

225

226 Internal symptoms appeared to be the result of multiple diseases and were  
227 frequently co-occurrent, as shown in Table 1 and Figure 1.g, h. Forty-four percent of all  
228 sampled plants (n=84) showed internal symptoms characteristic of each of the three  
229 main grapevine diseases occurring in the same plant. Moreover, 38 % of plants with

230 typical external symptoms of eutypiose/BDA (n=42) also showed internal symptoms of  
231 esca, 56 % of plants (n=21) with external symptoms of esca showed the typical V-  
232 shaped necrosis of eutypiose/BDA as well as internal esca symptoms, and 60 % of the  
233 apoplectic plants (n=16) showed internal symptoms of both eutypiose/BDA and esca  
234 (Table 1). In 19 plants (10 % of total sampled plants), the internal symptoms did not  
235 match with the external visual diagnosis of the disease. Two of these plants showing  
236 external esca symptoms were free from any internal wood lesion, whereas the remaining  
237 17 plants showed stunted growth externally but only esca symptoms internally.

238

239 Forty-five percent of the plants observed with external symptoms of  
240 eutypiose/BDA showed at least two different kinds of internal lesions (Table 2).  
241 Percentages of two or more concomitant internal symptoms for esca,  
242 eutypiose/BDA+esca, and apoplectic plants were even higher, namely 80%, 100% and  
243 81%, respectively (Table 2). In all, an average of 63% of surveyed plants (n=121)  
244 showed at least two different types of internal lesions.

245

#### 246 *Fungal isolation and identification*

247

248 Isolations were attempted from 657 samples with V-shaped necroses, 314 with  
249 black spots, 297 with other necroses and 187 with wood decay. Isolations yielded 502  
250 fungal isolates: 236 isolates from V-shaped necroses, 104 from black spots, 97 from  
251 other necroses and 65 from wood decay (Table 3). Fungi were more frequently isolated  
252 in arms than in trunks, as shown by the number of taxa isolated from each plant part and  
253 the number of isolates per taxon.

254

255 *Diplodia seriata* and *E. lata* were predominantly isolated from the V-shaped  
256 necroses, with 44.9 % and 23.3 % of the isolations made from arms and trunks,  
257 respectively (Table 3). However, the remaining species of Botryosphaeriaceae  
258 (*Neofusicoccum luteum*, *N. parvum*, *N. vitifusiforme*, *Dothiorella viticola*, and other  
259 unidentified *Botryosphaeria* species) accounted for an additional 10.6 % of isolations  
260 from arms and trunks. A small number of isolates of *E. lata* and *Botryosphaeriaceae*  
261 species originated from other lesion types. *Phaeomoniella chlamydospora* was  
262 predominantly isolated from the black spots, and was identified in 73.1 % of all the  
263 isolations (Table 3). Several fungal species were associated with central necroses, with  
264 no single species clearly predominant. The most frequently associated fungi to these  
265 lesions were *Pa. chlamydospora* (24.7 %), *D. seriata* (20.6 %), *Pm. aleophilum*  
266 (12.4 %), and *E. lata* (10.3 %). Additionally, about 10 more taxa were isolated from  
267 central necroses, but with low frequencies (Table 3). *Fomitiporia mediterranea* was  
268 predominant in the decayed wood, as was identified in 53.8 % of the isolations (Table  
269 3). Low frequencies of other fungi (e.g. *D. seriata*, *E. lata* and *Pa. chlamydospora*)  
270 were isolated from decayed wood.

271

272 *Pathogenicity test*

273

274 Control plants of both grapevine cultivars grew normally during the  
275 experimental period. Wounds of control plants healed successfully although some  
276 vascular discolourations were noticed (Table 4). Isolations from control plants yielded  
277 no fungus. Only *N. luteum* and *N. parvum* caused the wilting of the plant in a variable

278 number of inoculated plants (Table 4). While *N. luteum* caused wilting of ten  
279 'Tempranillo' plants and five 'Macabeo' plants, *N. parvum* caused a higher proportion  
280 of wilted plants in 'Macabeo' than in 'Tempranillo'. No additional foliar symptoms  
281 were observed among the remaining inoculated plants that could be related to a  
282 potential pathogenic effect of the tested isolates.

283

284 ANOVA showed the significance of the factors 'isolate' and 'grapevine variety'  
285 and their interaction (all  $P<0.01$ ) on the necrosis lengths. In general, necrosis lengths  
286 recorded on 'Tempranillo' for each inoculated isolate were longer than for their  
287 equivalents in 'Macabeo' ( $P<0.05$ ), but the differences between varieties were not  
288 significant for isolates *Botryosphaeria dothidea* 353, *Cryptovalsa ampelina* 476, *D.*  
289 *viticola* 412, *E. lata* 481, *F. mediterranea* I-62, and *N. parvum* 434 and 444,. However,  
290 significant differences between isolates and their respective controls were more frequent  
291 among the 'Macabeo' plants. This was probably due to the longer necrosis ( $P<0.01$ )  
292 observed in the 'Tempranillo' control (1.4 cm), twice as long as those on 'Macabeo'.  
293 The most severe lesions were caused by *N. parvum* (necroses up to 12.7 cm long in  
294 'Macabeo' and 13.8 cm in 'Tempranillo') and *N. luteum* (8.6 cm and 8.2 cm,  
295 respectively). Other fungi that caused significant lesions in plants of both grapevine  
296 cultivars were *Pa. chlamydospora* (2.5 to 5.5 cm), *E. lata* (1.3 to 4.0 cm), *D. seriata*  
297 (0.8 to 3.6 cm), and *B. dothidea* (3.2 cm in both cvs.) (Table 4). *Fomitiporia*  
298 *mediterranea*, *Pm. aleophilum* and *C. ampelina* only caused significant necroses in  
299 'Macabeo'; in general, mean lesion lengths for these fungi were never longer than 2 cm  
300 in 'Macabeo' and 2.5 cm in 'Tempranillo' (Table 4). The isolates *D. seriata* 421, *F.*

301 *mediterranea* 356 and *Phomopsis* sp. 459 did not cause any significant lesion on any  
302 cultivar.

303

304 Fungal species with multiple isolates being tested for pathogenicity showed  
305 some variability on the lesion lengths they caused. Thus, a wide range was observed for  
306 *N. parvum* (maximum mean values about 2-3 times greater than the minimum ones), *D.*  
307 *seriata* (2-3 times), and *E. lata* (1.5-2 times), whereas less variation was observed for *C.*  
308 *ampelina*, *F. mediterranea*, *Pa. chlamydospora* and *Pm. aleophilum* (Table 4). Only  
309 significant differences among isolates in the necrosis lengths for *D. seriata* and *N.*  
310 *parvum* were detected (Table 4).

311

312 Reisolations from the inoculated plants were successful for all of the inoculated  
313 fungi, although percentages of positive reisolations were variable among the fungal  
314 species (Table 4). In general, reisolations from 'Macabeo' plants were higher than those  
315 from 'Tempranillo'. Additionally, reisolation percentages were generally higher for the  
316 most virulent fungi (those causing longer necrosis, e.g. *Botryosphaeriaceae* species, and  
317 *Pa. chlamydospora*) than those from weak pathogens (e.g. *C. ampelina* and *F.*  
318 *mediterranea*).

319

## 320 **Discussion**

321

322 The results obtained during the field survey confirmed the occurrence of the  
323 three main decline diseases of adult grapevine plants in Catalonia: eutypiose, BDA and  
324 esca. External and internal symptoms of both eutypiose and BDA, recorded in May and

325 June, looked very similar and thus were not reliable to distinguish between these two  
326 diseases, as it has been reported previously (Castillo-Pando *et al.*, 2001; Taylor *et al.*,  
327 2005; Urbez-Torres *et al.*, 2006b). Eutypiose and BDA were only differentiated after  
328 isolating the respective pathogens, *E. lata* and Botryosphaeriaceae spp, from the  
329 diseased tissues. Additionally, observations of late symptoms of BDA as described by  
330 Larignon *et al.* (2001) (including leaf chlorosis and necrosis, leaf fall, cluster wilting,  
331 and the occurrence of a brown streaking of the wood under the bark) were occasionally  
332 associated with plants affected by esca in this study. Thus, external symptom expression  
333 of BDA-affected plants would need further investigation to clearly identify this disease.  
334 Since BDA could be related with several species of Botryosphaeriaceae (Larignon *et al.*,  
335 2001; Surico *et al.*, 2006; van Niekerk *et al.*, 2006), it would be also interesting to know  
336 the pathogenic role of each botryosphaeriaceous fungus and the particular symptoms it  
337 causes on adult plants.

338

339 Survey results also showed the high co-occurrence of internal symptoms  
340 associated with eutypiose/BDA and esca in the same plant. This has been reported for  
341 esca and eutypiose (Mugnai *et al.*, 1999), although we were not able to find any  
342 quantitative example in previous literature. The occurrence of multiple lesion types in  
343 the same plant in NE Spain, which were especially frequent in arms, may reflect  
344 different events of infections through the pruning wounds. It is widely accepted that  
345 most fungal pathogens associated with grapevine declines are airborne pathogens and  
346 penetrate into the plant through the annual pruning of shoots (Carter, 1988; Mugnai *et*  
347 *al.*, 1999; Surico *et al.*, 2006; van Niekerk *et al.*, 2006).

348

349 Apoplexy is characterized by the sudden wilting and death of vines or vine-parts  
350 including clusters in midsummer. Apparent healthy leaves rapidly wilt and dry  
351 basipetally in a few days (Mugnai *et al.*, 1999). Weather conditions are thought to  
352 influence this phenomenon, since the apoplectic events often occur in hot summers,  
353 when rainfall is followed by dry, hot weather (Mugnai *et al.*, 1999). Apoplexy has been  
354 frequently described as a severe form of esca or specifically as an “acute esca  
355 syndrome” (Larignon and Dubos, 1997; Mugnai *et al.*, 1999; Graniti *et al.*, 2000;  
356 Surico, 2001; Surico *et al.*, 2006). However, the results obtained in this study suggest  
357 that apoplectic events might not be restricted only to esca-affected plants, since a  
358 significant percentage of surveyed plants showing apoplexy (60 %; n=15) were also  
359 affected by V-shaped necroses, which are commonly associated with eutypiose and  
360 BDA (Carter, 1988; van Niekerk *et al.*, 2006). Moreover, 20 % of apoplectic plants  
361 (n=5) had V-shaped necroses but no black spots, central necroses or wood decay, which  
362 are usually associated with esca. Mugnai *et al.* (1999) reported that both *D. seriata* and  
363 *E. lata*, often isolated from V-shaped necroses, are frequently isolated from esca-  
364 affected plants, which supports our field observations. In this study, cross sections of  
365 arms and trunks of apoplectic vines showed a great percentage of dead, non-functional  
366 tissues. No quantitative data were recorded on the type and extension of these internal  
367 lesions. Further research is then needed to establish whether wood deterioration is  
368 related with apoplexy. Additionally, water relationships of apoplectic plants should be  
369 studied to establish whether water stress could be related to apoplexy.

370

371 Fungal isolations of diseased wood showing a particular symptom indicated a  
372 general relationship between the lesion type and the isolated fungi. Thus, *D. seriata* and

373 *E. lata* were mainly isolated from V-shaped necroses, *Pa. chlamydospora* from black  
374 spots, and *F. mediterranea* from decayed wood. Fungi isolated from central necroses  
375 included several species, such as *D. seriata*, *Pa. chlamydospora* and *Pm. aleophilum*. In  
376 general, these results are in accordance with previous reports (Mugnai *et al.*, 1996;  
377 Larignon and Dubos, 1997; Mugnai *et al.*, 1999; Serra, 1999). However, some regional  
378 differences are observed in the distribution of some of these pathogens when comparing  
379 our data with those obtained in neighbouring regions. In France, Larignon and Dubos  
380 (1997) isolated *E. lata* more frequently than any Botryosphaeriaceous fungi from  
381 V-shaped necrosis, whereas our study and a previous one (Armengol *et al.*, 2001a)  
382 showed a greater incidence of *D. seriata* than *E. lata* in Spanish vineyards. Úrbez-  
383 Torres *et al.* (2006b) also showed this for California. It is suggested that *E. lata* is  
384 probably less abundant in dryer Mediterranean climate countries as compared to other  
385 cooler and rainy regions, since *E. lata* dispersion is enhanced when mean annual rainfall  
386 exceeds 350 mm (Carter, 1991; Mugnai *et al.*, 1999).

387

388 Most of the species tested for pathogenicity showed significant longer necrotic  
389 lesions than those in the 'Macabeo' and 'Tempranillo' controls. Only *N. huteum* and *N.*  
390 *parvum* caused the wilting of inoculated plants but no external disease symptoms were  
391 recorded for any other fungus-plant combination during the experimental period. Some  
392 influencing factors have been suggested to explain this phenomenon, which include a  
393 short experimental period, and other unsuitable experimental conditions such as the use  
394 of young, potted plants, and the inoculation of fungi into green, non-lignified plant  
395 tissues. Moreover, pathogenicity tests in this study were done using mycelium instead  
396 of spores as inoculum sources, which does not correspond to natural conditions for

397 fungal infection. Additionally, it has been reported that some fungi (e.g. *F. mediterranea*) are only able to colonize grapevine tissues previously damaged by other  
398 fungi (Larignon and Dubos, 1997). An unsuitable combination of the above factors  
400 could lead to the unsuccessful fungal colonization of the inoculated plant tissues, as seen  
401 on the low recovery of some fungi (e.g. *C. ampelina*, *F. mediterranea*) and the short,  
402 non-significant necroses recorded occasionally in the pathogenicity test. Lack of foliar  
403 symptom expression in artificially inoculated plants with known grapevine pathogens  
404 has been reported occasionally (Larignon and Dubos, 1997; Mugnai *et al.*, 1999). Since  
405 foliar symptom expression often fails to occur in artificial inoculations, pathogenicity  
406 and virulence of fungi have been often concluded from the analysis of the necrotic  
407 lesions caused by fungi in the plant vascular tissues, as reported in previous works  
408 (Mugnai *et al.*, 1999; Van Niekerk *et al.*, 2004; Surico *et al.*, 2006).

409

410 Pathogenicity has been previously reported for several Botryosphaeriaceae  
411 species (van Niekerk *et al.*, 2004; Taylor *et al.*, 2005; van Niekerk *et al.*, 2006), *E. lata*  
412 (Carter *et al.*, 1985; Carter, 1991; Pérès *et al.*, 1999; Sosnowski *et al.*, 2007), *F. mediterranea* (Sparapano *et al.*, 2001), and *Pm. aleophilum* and *Pa. chlamydospora*  
413 (Adalat *et al.*, 2000; Eskalen *et al.*, 2001; Sparapano *et al.*, 2001; Halleen *et al.*, 2007).  
414 *Neofusicoccum luteum* and *N. parvum* were the most virulent pathogens tested in our  
415 study. While *N. parvum* was proven to be a virulent pathogen by van Niekerk *et al.*  
416 (2004), pathogenicity of *N. luteum* seems controversial. Van Niekerk *et al.* (2004)  
417 considered this species as a low virulent pathogen since it caused no significant  
418 necroses on inoculated mature canes of 'Chardonnay' and 'Cabernet Sauvignon'  
419 varieties in South Africa. However, in our study *N. luteum* was clearly pathogenic.

421 Pathogenicity of *D. seriata* has been also disputed, as summarized by Úrbez-Torres *et*  
422 *al.* (2006b); it has been considered weakly pathogenic in Portugal (Phillips, 2002), but a  
423 virulent pathogen in Chile (Auger *et al.*, 2004), Australia (Castillo-Pando *et al.*, 2001)  
424 and South Africa (van Niekerk *et al.*, 2004). Additionally, Taylor *et al.* (2005) observed  
425 no significant vascular lesions caused by this fungus on inoculated grapevine cuttings in  
426 Australia. In our study, field observations of diseased tissues and the results obtained in  
427 the pathogenicity test would suggest that *D. seriata* is pathogenic. Additionally, the  
428 range observed in necrosis lengths also suggests variability in pathogen  
429 virulence. This is in accordance with previous reports (Larignon *et al.*, 2001; van  
430 Niekerk *et al.*, 2004).

431

432 *Eutypa lata* is a widely-known pathogen of grapevine (Carter, 1988; Carter,  
433 1991; Dubos, 1996). In our study, all five isolates tested for pathogenicity on 'Macabeo'  
434 plants caused significant necrosis while those in 'Tempranillo' were significant only in  
435 three isolates (401, 411 and 427). These findings may indicate variability in pathogen  
436 virulence but more isolates should be tested to confirm this hypothesis. Variability in  
437 virulence of *E. lata* has been shown previously (Péros *et al.*, 1999; Sosnowski *et al.*,  
438 2007). Pathogenicity of *Pa. chlamydospora* was also confirmed in this study, although  
439 no foliar symptoms were observed during the experimental period. Foliar symptom  
440 expression due to *Pa. chlamydospora* only have been observed after long inoculation  
441 periods (2-3 years) in mature plants artificially inoculated with this pathogen  
442 (Sparapano *et al.*, 2001). Foliar symptoms of esca have been reproduced on Thompson  
443 Seedless vines 6 months after inoculation with *Pm. aleophilum* and *Pa. chlamydospora*  
444 (W. D. Gubler, *pers. comm.*). The remaining species tested in our study were considered

445 non-pathogenic (*Phomopsis* sp. taxon 1) or weakly pathogenic (*C. ampelina*, *F.*  
446 *mediterranea* and *Pm. aleophilum*) from the necroses they caused. However, pathogenic  
447 effects have been observed for the latter two species (Adalat *et al.*, 2000; Eskalen *et al.*,  
448 2001; Sparapano *et al.*, 2001).

449

450 Concomitant fungal pathogens occur in the same grapevine, each one causing a  
451 particular wood lesion. This may lead to complex relationships among these pathogens  
452 and the host plant. This study has shown that co-occurrence of internal disease  
453 symptoms and their associated fungi are frequent in NE Spain, and that the relationships  
454 between visual external symptoms and inferred internal lesions often are misleading.  
455 This makes field diagnosis of the diseases difficult when only the external symptoms  
456 are considered.

457

458

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460

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468

469

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577

578 *Table 1.* Percentages of declining grapevine plants showing different internal symptoms for a  
579 given external symptomatology.

580

External symptoms	Internal symptoms
<i>Number of plants</i>	
Eutypiose/BDA	40 % V-shaped necrosis
111 plants	22 % Black spots, central necroses, wood decay
	38 % V-shaped necrosis, black spots, central necroses, wood decay
Esca	7 % V-shaped necrosis
37 plants	37 % Black spots, central necroses, wood decay
	56 % V-shaped necrosis, black spots, central necroses, wood decay
Eutypiose/BDA + Esca	100 % V-shaped necrosis, black spots, central necroses, wood decay
5 plants	
Apoplexy	20 % V-shaped necrosis
26 plants	20 % Black spots, central necroses, wood decay
	60 % V-shaped necrosis, black spots, wood decay

581

582

583 *Table 2.* Percentages of declining grapevine plants showing different lesion types.

584

External symptoms <sup>2</sup>	Number of different internal lesion types <sup>1</sup>				
	0	1	2	3	4
Eutypiose / BDA	2	53	30	12	3
Esca	3	16	52	24	5
Eutypiose / BDA + Esca	0	0	20	80	0
Apoplexy	0	19	46	19	16

585 <sup>1</sup>: Sum of row percentages = 100. Lesion types may include one or more of the following  
 586 internal symptoms in the same plant: V-shaped necrosis, irregularly-shaped necrosis around the  
 587 pith, black spots, wood decay.

588 <sup>2</sup>: Summary of observed external symptoms: *Eutypiose*: stunted appearance of shoots at the  
 589 early growth season, with small, cupped, and chlorotic leaves, and short internodes.  
 590 *Esca*: interveinal chlorosis or discolorations (yellowish in white cultivars and reddish in  
 591 red cultivars) eventually with leaf marginal or interveinal necrosis. *Apoplexy*: drastic leaf  
 592 withering, whether partial (1 to several arms) or total (whole plant).

593

594 Table 3. Number of fungi isolated from wood lesions of declining grapevines.

Plant part	Species	Lesion type <sup>1</sup>				Total
		V-shaped necroses	Black spots	Central necroses	Wood decay	
Arms	<i>Acremonium</i> sp.	0	0	1	1	2
	<i>Botryosphaeria dothidea</i>	0	0	1	0	1
	<i>Cryptovalsa ampelina</i>	0	0	1	0	1
	<i>Cylindrocarpon liriodendri</i>	0	0	1	0	1
	<i>Diplodia seriata</i>	91	8	17	5	121
	<i>Dothiorella viticola</i>	1	0	0	0	1
	<i>Eutypa lata</i>	47	1	8	3	59
	<i>Eutypa leptoplaca</i>	1	0	0	0	1
	<i>Eutypella vitis</i>	1	0	0	0	1
	<i>Fomitiporia mediterranea</i>	3	1	2	19	25
	<i>Fusarium</i> spp.	3	0	3	1	7
	<i>Neofusicoccum parvum</i>	11	1	3	0	15
	<i>Phaeoacremonium aleophilum</i>	2	4	11	1	18
	<i>Phaeoacremonium viticola</i>	0	0	1	0	1
	<i>Phaeoacremonium</i> sp.	1	0	0	0	1
	<i>Phaeomoniella chlamydospora</i>	11	61	19	4	95
	<i>Phoma</i> -like sp.	1	2	0	0	3
Trunk	<i>Phomopsis</i> spp.	7	0	0	0	7
	<i>Stereum hirsutum</i>	0	1	0	0	1
	Unidentified <i>Botryosphaeriaceae</i>	6	0	1	0	7
	Unidentified <i>Diatrypaceae</i>	1	0	1	1	3
	Unidentified species	8	6	6	2	22
	<i>Acremonium</i> sp.	0	1	0	0	1
	<i>Botryosphaeria dothidea</i>	0	1	0	0	1
	<i>Cryptovalsa ampelina</i>	3	0	0	1	4
	<i>Cylindrocarpon liriodendri</i>	0	0	1	0	1
	<i>Diplodia seriata</i>	15	0	3	3	21
	<i>Eutypa lata</i>	8	1	2	2	13
	<i>Fomitiporia mediterranea</i>	0	0	3	16	19
	<i>Fusarium</i> spp.	1	0	3	0	4
	<i>Neofusicoccum luteum</i>	1	0	0	0	1
	<i>Neofusicoccum parvum</i>	5	1	0	0	6
	<i>Neofusicoccum vitifusiforme</i>	1	0	0	0	1
	<i>Phaeoacremonium aleophilum</i>	1	0	1	0	2
	<i>Phaeoacremonium</i> sp.	1	0	0	0	1
	<i>Phaeomoniella chlamydospora</i>	4	15	5	4	28
	<i>Phomopsis</i> spp.	0	0	1	0	1
	Unidentified <i>Botryosphaeriaceae</i>	1	0	1	0	2
	Unidentified species	0	0	1	2	3
Totals		236	104	97	65	502

595 1: Numbers of lesions examined for isolations: V-shaped necroses: n=657; black spots: n=314;

596 central necroses: n=297; wood decay: 187.

597

598 *Table 4.* Number of wilted plants, length of vascular necroses and percentage of mycelium  
 599 recovery in grapevine plants (n=18) cvs. 'Macabeo' and 'Tempranillo' inoculated with selected  
 600 fungi isolated from declining grapevines.

601

Species	Isolate	Macabeo			Tempranillo		
		Wilted plants	Necrosis (cm) <sup>1</sup>	% Myc. recovery	Wilted plants	Necrosis (cm) <sup>1</sup>	% Myc. recovery
<i>Botryosphaeria dothidea</i>	353	0	<b>3.2</b>	100	0	<b>3.2</b>	100
<i>Diplodia seriata</i> <sup>2</sup>	398	0	<b>2.4</b> a	100	0	<b>3.5</b> a	67
<i>D. seriata</i>	421	0	0.8 c	83	0	1.9 b	28
<i>D. seriata</i>	I-29	0	<b>1.6</b> b	94	0	<b>3.0</b> ab	78
<i>D. seriata</i>	I-50	0	<b>1.5</b> b	100	0	<b>3.6</b> a	83
<i>Dothiorella viticola</i>	412	0	<b>1.7</b>	61	0	2.1	39
<i>Cryptovalsa ampelina</i>	413	0	<b>1.5</b>	50	0	2.5	39
<i>C. ampelina</i>	476	0	<b>1.7</b>	28	0	2.0	17
<i>Eutypa lata</i>	401	0	<b>1.7</b>	100	0	<b>2.9</b>	67
<i>E. lata</i>	411	0	<b>1.9</b>	94	0	<b>4.0</b>	50
<i>E. lata</i>	427	0	<b>1.3</b>	83	0	<b>3.6</b>	61
<i>E. lata</i>	438	0	<b>1.4</b>	100	0	2.3	50
<i>E. lata</i>	481	0	<b>2.0</b>	88	0	2.2	39
<i>Fomitiporia mediterranea</i>	356	0	0.7	33	0	1.5	11
<i>F. mediterranea</i>	452	0	<b>1.4</b>	33	0	2.5	22
<i>F. mediterranea</i>	I-62	0	<b>1.1</b>	18	0	1.4	17
<i>Neofusicoccum luteum</i>	519	5	<b>8.6</b>	100	10	<b>8.2</b>	100
<i>Neofusicoccum parvum</i> <sup>2</sup>	387	0	<b>4.0</b> c	94	0	<b>5.6</b> b	100
<i>N. parvum</i>	396	5	<b>10.8</b> a	100	1	<b>13.8</b> a	78
<i>N. parvum</i>	434	5	<b>12.7</b> a	100	0	<b>11.6</b> a	100
<i>N. parvum</i>	444	1	<b>6.7</b> b	100	0	<b>6.9</b> b	94
<i>Phaeoacremonium aleophilum</i>	449	0	<b>1.1</b>	67	0	2.4	83
<i>Pm. aleophilum</i>	477	0	<b>1.2</b>	83	0	2.2	100
<i>Pm. aleophilum</i>	I-10	0	<b>1.4</b>	89	0	1.9	94
<i>Phaeomoniella chlamydospora</i>	454	0	<b>2.5</b>	89	0	<b>4.5</b>	67
<i>Pa. chlamydospora</i>	I-8	0	<b>2.5</b>	78	0	<b>5.3</b>	89
<i>Pa. chlamydospora</i>	I-64	0	<b>2.5</b>	83	0	<b>5.5</b>	89
<i>Phomopsis taxon 1</i>	459	0	1.0	89	0	1.8	89
Control		0	0.7	0	0	1.4	0

602 <sup>1</sup>: Values in bold characters are significantly different from the corresponding control mean  
 603 value according to the two-tailed Dunnett's test.

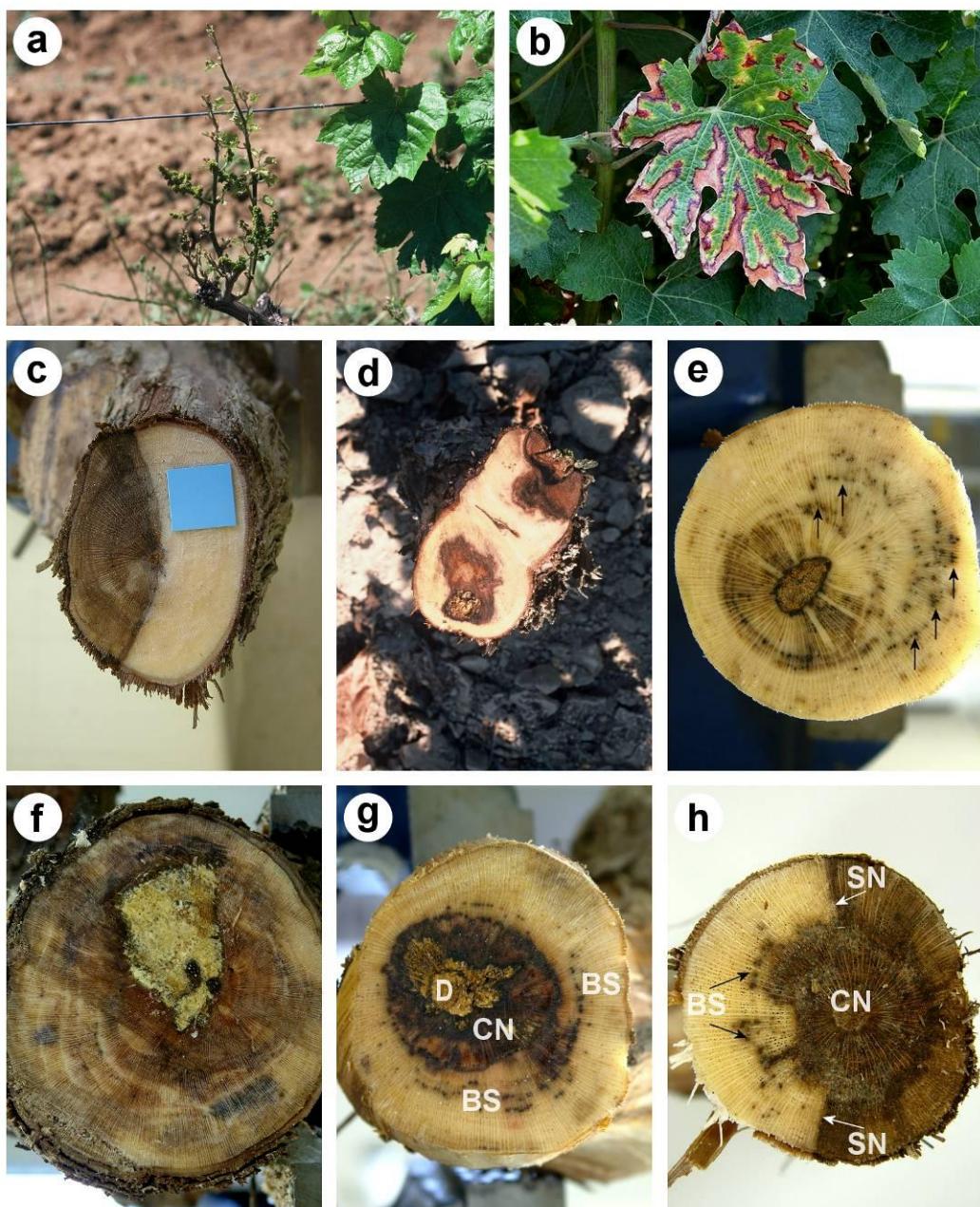
604 <sup>2</sup>: Mean values of necrosis lengths for *D. seriata* and *N. parvum* followed by different letters are  
 605 different according to the Tukey's test (P<0.05) within each species.

606 *Figure 1.* Most habitual symptoms associated with grapevine declines: a) Reduced growth, and  
607 small, chlorotic leaves. b) Foliar symptoms of esca in a red grape cultivar, characterised by the  
608 interveinal necroses and chloroses. c) V-shaped necrosis. d) Irregularly-shaped necrosis around  
609 the pith (central necrosis). e) Black spots (shown by arrows), corresponding to vascular  
610 necroses. f) Wood decay, characterised by a yellowish soft tissue. g and h) Co-occurrence of  
611 internal symptoms; BS, black spots, CN, central necrosis, D, wood decay, SN, V-shaped  
612 necrosis. Symptoms a) and c) are usually associated with eutypiose and black dead arm,  
613 whereas symptoms b) and d) to f) are associated to esca.

614

615 Figure 1

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(low resolution image, for reviewing purposes only)