Multifocal Granulomatous Panniculitis with Ceroid Pigment in Two Mediterranean Striped Dolphins (*Stenella coeruleoalba*)

Sara Soto,^{1,5} Dolors Fondevila,¹ Beatriz González,² Encarna Gómez-Campos,³ and Mariano Domingo^{1,4}
¹ Departament de Sanitat i d'Anatomia Animals, Facultat de Veterinária, Universitat Autònoma de Barcelona (UAB), 08193 Bellaterra, Spain; ² CRAM (Fundació per la Conservació y Recuperació d'Animals Marins), Camí Ral 239, 08330 Premià de Mar, Spain; ³ Department de Biologia Animal–Vertebrats, Facultat de Biologia, Universitat de Barcelona (UB), Avda. Diagonal 645, 1ª planta, 08028 Barcelona, Spain; ⁴ CRESA (Centre de Reserca en Sanitat Animal), Campus Universitat Autònoma de Barcelona (UAB), 08193 Bellaterra, Spain; ⁵ Corresponding author (email: sara.soto@uab.es)

ABSTRACT: Two striped dolphins (Stenella coeruleoalba) were found stranded on the Catalonian Spanish coast. The main pathologic finding in both animals was the existence of multiple granulomatous lesions in the blubber, microscopically composed of macrophages and multinucleated cells containing vacuolar material. This material was identified as ceroid pigment due to its ultrastructural morphology, autofluorescence, and positive staining with periodic acid-Schiff and Ziehl-Neelsen techniques. The special stains and electron microscopy did not reveal any microorganisms associated with the lesions. These findings are very suggestive of "nutritional panniculitis," a welldefined entity associated with vitamin E deficiency that has been rarely described in free-living species.

Key words: Blubber, ceroid pigment, granulomatous panniculitis, nutritional panniculitis, Stenella coeruleoalba, striped dolphin, vitamin E deficiency.

Several processes have been described affecting the panniculus of free-ranging cetaceans (Cowan, 1993; Schulman and Lipscomb, 1999; Bowenkamp et al., 2001; Bonar and Wagner, 2003). Here we describe macroscopic, microscopic, and ultrastructural findings in two free-ranging Mediterranean striped dolphins (*Stenella coeruleoalba*) affected by a granulomatous panniculitis with intralesional ceroid pigment resembling nutritional panniculitis.

A 62 kg, 1.89 m adult female striped dolphin found dead on the coast of Roses-Girona (Catalonia, Spain, 42°15′N, 3°09′E) in April 2008, and a 98.5 kg, 2.24 m adult male striped dolphin that died shortly after being found stranded on the coast of Platja d'Aro-Girona (Catalo-

nia, Spain, 41°48′N, 3°03′E) in February 2009, were transported to the Veterinary School in Barcelona, Spain. Complete necropsies were performed following the protocol of the Catalonian Mediterranean Morbillivirus Surveillance Program.

Samples of pharynx, lung, heart, liver, intestine, stomach, spleen, lymph node, pancreas, adrenal, kidney, urinary bladder, tongue, brain, skeletal muscle, and skin were fixed in 10% neutral-buffered formalin and embedded in paraffin. Sections were cut at 5 µm and stained with hematoxylin and eosin (H&E). Skin sections were also examined under polarized light and stained with periodic acid-Schiff (PAS), Ziehl-Neelsen (ZN), Fite, Gram's, and Grocott's methenamine silver (GMS) techniques. Formalin-fixed blubber samples were processed for electron microscopy. Several brain sections and the prescapular, thoracic, and mesenteric lymph nodes were tested for morbillivirus by immunohistochemistry (IHC) using the avidin-biotin-peroxidase method (Thermo Fisher Scientific, Rockford, Illinois, USA). A mouse monoclonal antibody against canine distemper virus nucleoprotein (VMRD Laboratories, Pullman, Washington, USA), diluted to 1:200, and a goat antimouse IgG (Dako, Glostrup, Denmark) as secondary antibody, were used.

The most relevant macroscopic lesions were several irregular, well-defined, non-raised yellow-orange lesions, approximately 0.5–2.0 cm in diameter in the middle-to-deep blubber (Fig. 1). The lesions were distributed ventrally and dorsally in the



FIGURE 1. Three skin fragments containing well-defined, non-raised, irregular focal lesions in the deep blubber characterized by an intense yellow-orange color. Bar=1 cm.

female and mainly in the abdominal area in the male. In both cases, up to 10% of the blubber layer was affected. Both had a normal external skin surface except for numerous conspecific bite marks and two tattoo lesions in the male. Both had a medium-to-high number of merocercoids of *Phyllobothrium delfini* in the blubber of the genital area. A few copepods (*Penella* spp.) in the fluke of the female were also found. The keratinized stomach was empty in both animals, with low quantities of sand in the male dolphin. The second stomach of the male contained one prominent parasitic nodule.

Microscopically, the blubber lesions corresponded to multifocal to coalescing, well-defined granulomas (Fig. 2) composed of numerous macrophages and multinucleated giant cells. The cytoplasm of these cells contained abundant roundto-oval, nonrefringent, colorless or pale yellow vacuolar material, varying from approximately 2-30 µm in diameter (Fig. 3). Scant foci with adipocyte necrosis and fat saponification were also observed, surrounded by macrophages and scattered karyorrhectic neutrophiles (Fig. 3, inset). An increased number of collagen bundles (fibrosis) was associated with the inflammatory infiltrate. No foreign material was visible under polarized light. The vacuolar material in the blubber lesions was intensely autofluorescent under blue light

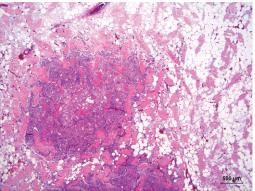


FIGURE 2. Skin blubber showing well-defined, coalescing granulomatous lesions. H&E stain. Bar=500 µm.

and stained with PAS and ZN (Fig. 4). The PAS, ZN, Fite, Gram's, and GMS stains did not reveal microorganisms associated with the granulomatous skin lesions. Ultrastructurally, in the blubber lesions, numerous osmiophilic rounded or granular structures of variable size were observed in the cytoplasm of macrophages (Fig. 5). Additional microscopic findings in both dolphins included mild, multifocal lymphoplasmacytic nephritis with tubular mineralization and moderate neuronal lipopigment deposit. The male also had chronic nodular parasitic gastritis with intralesional trematode eggs. Results of IHC for morbillivirus were negative in both dolphins.

Several processes associated with granulomatous panniculitis have been described in cetaceans, including nocardiosis (Leger et al., 2009), mycobacteriosis (Bowenkamp et al., 2001), lobomycosis (Cowan, 1993), and fusariosis (Frasca et al., 1996). The absence of acid-fast bacteria (ZN and Fite stains) associated with the granulomas ruled out Mycobacterium and *Nocardia* as possible etiologies of the lesions. Mycotic infections are also an unlikely cause, as these organisms are usually easily detected in affected tissues using PAS and GMS stains. If these lesions were the normal result of a response to fat breakdown in marine

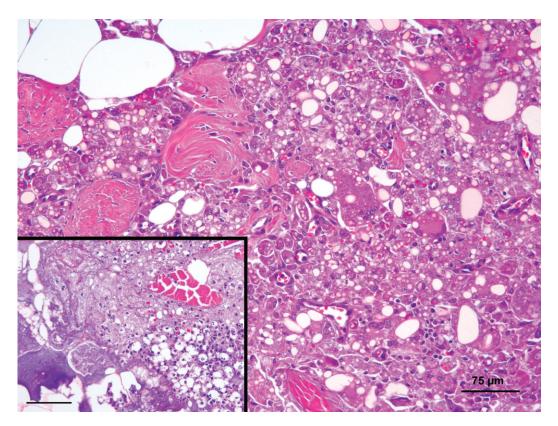


Figure 3. Locally extensive granulomatous panniculitis composed of abundant macrophages and multinucleated giant cells, showing abundant, cytoplasmatic, colorless or pale yellow vacuolar material. H&E stain. Bar=75 μ m. Inset: Focal blubber fat necrosis with degenerated neutrophils and macrophages. Bar=75 μ m.

mammals, they would have been described more often in routine necropsies. Conditions associated with granulomatous panniculitis in dogs and cats have included foreign-body reactions, trauma, pancreatic panniculitis, nutritional panniculitis, and idiopathic-nodular panniculitis (Gross et al., 2005). However, the absence of macroscopic and microscopic pancreatic lesions in these dolphins ruled out a pancreatic origin. The high number of lesions affecting the blubber, their scattered distribution, and the absence of associated foreign material made a foreign-body reaction unlikely. No lesions were evident in the epidermis associated with the granulomas, and fat necrosis was not a significant finding, making trauma an unlikely cause. Although idiopathic inflammation can still be considered, the

granulomas in both dolphins were characterized by the presence of abundant intralesional material with morphologic, physical, and staining properties consistent with ceroid pigment, including ultrastructural electrodense vacuolar structures with marked autofluorescence under blue light and positive staining with PAS and ZN techniques (Seehafer and Pearce, 2006; Ginn et al., 2007). This finding strongly suggested nutritional panniculitis, as observation of ceroid pigment is the characteristic histopathologic finding of this condition and, to our knowledge, has not been described in association with other causes of panniculitis (Gross et al., 2005).

Nutritional panniculitis in several animal species (Ginn et al., 2007) is associated with deficiency of vitamin E due to

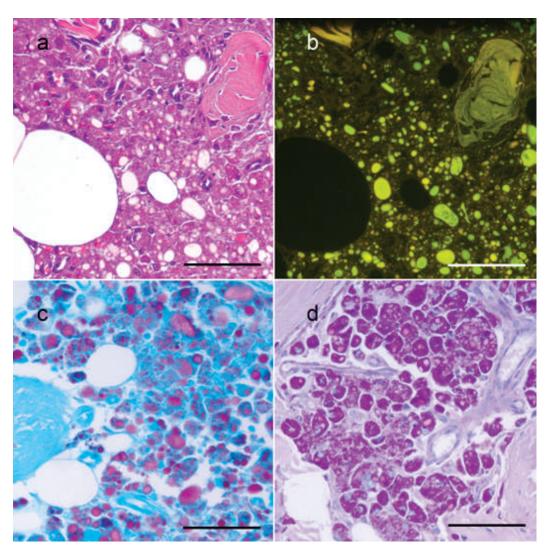


FIGURE 4. Photomicrographs showing characteristic staining and physical properties of ceroid pigment in the granulomatous lesion of the dolphin blubber: (a) Colorless or pale yellow vacuolar material with H&E stain. (b) Intense yellow-green autofluorescence under blue light. (c) Acid-fast red staining with Ziehl-Neelsen method. (d) Intense magenta staining with periodic acid-Schiff method. Bar in the four figures= $75 \mu m$.

inadequate intake of the vitamin, consumption of high levels of unsaturated fatty acids that deplete vitamin E, or both (Niza et al., 2003). The negative antioxidant status leads to peroxidation of susceptible lipids, with formation of free radicals that provoke the characteristic inflammatory reaction in response to formation of irritant soaps and ceroid pigment (Brown et al., 2007). The yellow color of the lesions is a consequence of the deposition of ceroid (Ginn et al., 2007).

Contrary to the systemic presentation of nutritional panniculitis in cats (Ginn et al., 2007), fat deposits in any organs other than the blubber, which in dolphins are confined to small quantities of adipose tissue surrounding organs such as lymph nodes and heart, were not affected in these two cases. One explanation could be that the process targets sites with high biochemical activity. The blubber is the primary site for energy storage, and the middle and deep blubber layers where the

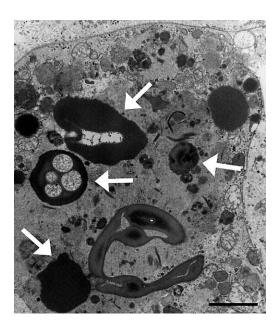


FIGURE 5. Electron micrograph illustrating intracytoplasmic, heterogenous black, osmiophilic rounded and granular structures compatible with lipopigments (arrow) within a macrophage. Bar= $2~\mu m.$

granulomas were localized appear to be the most metabolically dynamic layers (Struntz et al., 2004). Skeletal muscle lesions, which could be expected with vitamin E deficiency, were not observed in either case. Dolphins may respond similarly to other carnivores in which nutritional myopathy due to vitamin E imbalance is unusual (Van Vleet and Valentine, 2007). Another adult striped dolphin, found on the Catalonian coast and affected by fatal sarcocystosis, had blubber granulomas similar to those we describe. The authors associated the lesion to a supposed vitamin E deficiency due to fat malabsorption associated with chronic parasitic pancreatitis (Resendes et al., 2002). In our cases, where no apparent pancreatic lesion was present, the cause of vitamin E deficiency could have been consumption of an abnormal quantity of oily fish with a high concentration of unsaturated fatty acids. Sardines and anchovies are normal prey for Mediterranean dolphins (Hammond et al., 2008).

The same cause has been implicated in nutritional panniculitis in domestic cats and mink farming (Niza et al., 2003). Because no serum or liver samples were available for the measurement of vitamin E in our dolphins, a nutritional origin for the granulomatous panniculitis could not be definitively shown.

Other processes affecting the panniculus have also been described in cetaceans, including bacterial infections (e.g., Streptococcus spp. [Bonar and Wagner, 2003], Erysipelothrix rhusiopathiae [Thurman et al., 1983], Pseudomonas aeruginosa [Diamond et al., 1979], and Aeromonas hydrophila [Cusick and Bullock, 1973]), as well as parasitic infections (e.g., ciliated protozoa [Schulman and Lipscomb, 1999], tetraphyllidean cysticerci [Norman, 1997], and nematodes [Dailey and Stroud, 1978]). The histopathologic findings described in association with these processes are not compatible with those observed in the blubber lesions we describe. In addition, no bacteria or parasites were detected during the histologic (routine examination and staining) and ultrastructural procedures, making these infections an unlikely cause of the lesions.

Finally, the differential diagnosis could include a long list of viruses such as coxsackie, influenza, cytomegalovirus, rabies, vaccinia, Ross River, and hantaviruses, all of which have been described as causes of inflammation or necrosis of fat deposits in rodents (Nisimura et al., 2000). Those viruses are an unlikely cause of the process affecting our dolphins due to the absence of a granulomatous response associated with these agents (Nisimura et al., 2000) and to the absence of viral structures in the electron microscopic examination of the blubber lesions.

In summary, generalized granulomatous panniculitis with intralesional ceroid pigment was the only pathologically relevant finding in the two striped dolphins reported on in this paper. These findings suggest nutritional panniculitis due to vitamin E deficiency. However, further work to

confirm the etiology of the lesions is needed.

LITERATURE CITED

- BONAR, C. J., AND R. A. WAGNER. 2003. A third report of "golf ball disease" in an Amazon River dolphin (*Inia geoffrensis*) associated with *Strep*tococcus iniae. Journal of Zoo and Wildlife Medicine 34: 296–301.
- Bowenkamp, K. E., S. Frasca, A. Draghi, II, G. J. Tsongalis, C. Koerting, L. Hinckley, S. De Guise, R. J. Montali, C. E. C. Goertz, D. J. St. Aubin, and J. L. Dunn. 2001. *Mycobacterium marinum* dermatitis and panniculitis with chronic pleuritis in a captive white whale (*Delphinapterus leuca*) with aortic rupture. Journal of Veterinary Diagnostic Investigation 13: 524–530.
- Brown, C. C., D. C. Baker, and I. K. Barker. 2007. Alimentary system. In Jubb, Kennedy and Palmer's pathology of domestic animals. Vol. 2, 5th Edition, M. G. Maxie (ed.). Elsevier Saunders, Edinburgh, UK, pp. 1–296.
- COWAN, D. F. 1993. Lobo's disease in a bottlenose dolphin (*Tursiups truncatus*) from Matagorda Bay, Texas. Journal of Wildlife Diseases 29: 488– 489.
- Cusick, P. K., and B. C. Bullock. 1973. Ulcerative dermatitis and pneumonia associated with *Aeromonas hydrophila* infection in the bottle-nosed dolphin. Journal of the American Veterinary Medical Association 163: 578–579.
- DAILEY, M., AND R. STROUD. 1978. Parasites and associated pathology observed in cetaceans stranded along the Oregon coast. Journal of Wildlife Diseases 14: 503–511.
- DIAMOND, S. S., D. E. EWING, AND G. A. CADWELL. 1979. Fatal bronchopneumonia and dermatitis caused by *Pseudomonas aeruginosa* in an Atlantic bottle-nosed dolphin. Journal of the American Veterinary Medical Association 175: 984– 987
- FRASCA, JR., S., J. L. DUNN, J. C. COOKE, AND J. D. BUCK. 1996. Mycotic dermatitis in an Atlantic white-sided dolphin, a pygmy sperm whale, and two harbor seals. Journal of the American Veterinary Medical Association 208: 727–729.
- GINN, P. E., J. E. K. L. MANSELL, AND P. M. RAKICH. 2007. Skin and appendages. In Jubb, Kennedy and Palmer's pathology of domestic animals. Vol. 1, 5th Edition, M. G. Maxie (ed.). Elsevier Saunders, Edinburgh, UK, pp. 553–781.
- GROSS, T. L., P. J. IHRKE, E. J. WALDER, AND V. K. AFFOLTER. 2005. Diseases of the panniculus. In Skin diseases of the dog and cat. Clinical and histopathological diagnosis, 2nd Edition. Blackwell Publishing, Oxford, UK, pp. 537–560.

- HAMMOND, P. S., G. BEARZI, A. BJØRGE, K. FORNEY, L. KARCZMARSKI, T. KASUYA, W. F. PERRIN, M. D. SCOTT, J. Y. WANG, R. S. WELLS, AND B. WILSON. 2008. Stenella coeruleoalba. In IUCN 2008. 2008 IUCN red list of threatened species. http://www.iucnredlist.org. Accessed June 2009.
- NISIMURA, H., S. ITAMURA, T. IWASAKI, T. KURATA, AND M. TASHIRO. 2000. Characterization of human influenza A (H5N1) virus infection in mice: Neuro-, pneumo- and adipotropic infection. Journal of General Virology 81: 2503–2510.
- NIZA, M. M. R. E., C. L. VILELA, AND L. M. A. FERREIRA. 2003. Feline pansteatitis revisited: Hazards of unbalanced home-made diets. Journal of Feline Medicine and Surgery 5: 271–277.
- NORMAN, R. J. DE B. 1997. Tetraphyllidian cysticerci in the peritoneal cavity of the common dolphin. Journal of Wildlife Diseases 33: 891–895.
- RESENDES, A. R., C. JUAN-SALLÉS, S. ALMERIA, N. MAJO, M. DOMINGO, AND J. P. DUBEY. 2002. Hepatic sarcocystosis in a striped dolphin (Stenella coeruleoalba) from the Spanish Mediterranean coast. Journal of Parasitology 88: 206–209.
- Seehafer, S. S., and D. A. Pearce. 2006. You say lipofucsin, we say ceroid: Defining autofluorescent storage material. Neurobiology of Aging 27: 576–588.
- Schulman, F. Y., and T. P. Lipscomb. 1999. Dermatitis with invasive ciliated protozoa in dolphins that died during the 1987–1988 Atlantic bottlenose dolphin morbilliviral epizootic. Veterinary Pathology 36: 171–174.
- St. Leger, , J. A. L. Begeman, M. Fleetwood, S. Frasca, Jr., M. M. Garner, S. Lair, S. Trembley, M. J. Linn, and K. A. Terio. 2009. Comparative pathology of nocardiosis in marine mammals. Veterinary Pathology 46: 299–308.
- STRUNTZ, D. J., W. A. McLellan, R. M. Dillaman, J. E. Blum, J. R. Kucklick, and D. A. Pabst. 2004. Blubber development in bottlenose dolphins (*Tursiops truncatus*). Journal of Morphology 259: 7–20.
- THURMAN, G. D., S. J. DOWNES, M. B. FOTHERGILL, N. M. GOODWIN, AND M. M. HEGARTY. 1983. Diagnosis and successful treatment of subacute erysipelas in a captive dolphin. Journal of the South African Veterinary Association 54: 193–200.
- Van Vleet, J. F., and B. A. Valentine. 2007. Muscle and tendon. *In* Jubb, Kennedy and Palmer's pathology of domestic animals. Vol. 1, 5th Edition, M. G. Maxie (ed.). Elsevier Saunders, Edinburgh, UK, pp. 185–280.

Received for publication 20 July 2009.