Additional file 12: Oligonucleotides used in this work

name	Sequence (5'→3')	Application
P1_sulA	TTAATGATACAAATTAGAGTGAATTTTTAGCCCGGAAAGTTGTCTCGTGG CGTGAGAGGAGTGTAGGCTGGAGCTGCTTC	P1 primer for the construction of UA6189 strain
P2_sulA	ATGTACACTTCAGGCTATGCACATCGTTCTTCGTCGTTCTCATCCGCAGC AAGTAAAATTATGGGAATTAGCCATGGTCC	P2 primer for the construction of UA6189 strain
P1_lexA	ATGAAAGCGTTAACGGCCAGGCAACAAGAGGTGTTTGATCTCATCCGTGA TCACATCAGCGTGTAGGCTGGAGCTGCTTC	P2 primer for the construction of UA6189 strain
P2_lexA	TTACAGCCAGTCGCCGTTGCGAATAACCCCAACCGCCAGCCCTTCAATGG TGAAGCTCTGATGGGAATTAGCCATGGTCC	P2 primer for the construction of UA6189 strain
NdelexAVpa	*CATATG*AAGCCGTTAACGCCACGCC[a]	Upper primer for cloning the *V.parahaemolyticus lexA* gen in pET15b overexpression vector
XholexAVpa	*CTCGAG*TTACATCCAATCGGTATTG[a]	Lower primer for cloning the *V.parahaemolyticus lexA* gen in pET15b overexpression vector
NdelexAEco	*CATATGA*AGCGTTAACGGCCAGGC[a]	Upper primer for cloning the *E. coli lexA* gen in pET15b overexpression vector
XholexAEco	*CTCGAG*TTACAGCCAGTCGCCGTTGC[a]	Lower primer for cloning the *E. coli lexA* gen in pET15b overexpression vector
wtintVpaF	AAAAAGCATGATAACTGGGCCAGTATTGATAAATTACAACACCTGTATAA ATAAACAGACTTATAATATTATGAAAAGTCAATTTCTGCTAAGTGTAAAA	Synthetic oligo to obtain the Pint1$^-$ EMSA probe
wtpintVpaR	ATTTTACACTTAGCAGAAATTGACTTTTCATAATATTATAAGTCTGTTTA TTTATACAGGTGTTGTAATTTATCAATACTGGCCCAGTTATCATGCTTTT	Synthetic oligo to obtain the Pint1$^-$ EMSA probe
wtpint1-pMURF	AGTAACGGCGCAGTGGCGGTTTTCATGGCTTGTTATGACTGTTTTTTTGT ACAGTCTATGCCTCGGGCATCCAAGCAGCAAGCGCGTTACGCCGTGGGTC	Synthetic oligo to obtain the P_{int1-} EMSA probe
wtpint1-pMURR	AGACCCACGGCGTAACGCGCTTGCTGCTTGGATGCCCGAGGCATAGACTG TACAAAAAAACAGTCATAACAAGCCATGAAAACCGCCACTGCGCCGTTAC	Synthetic oligo to obtain the P_{int1-} EMSA probe
wtpint1+pMURF	AACGGCGCAGTGGCGGTTTTCATGGCTTGTTATGACTGTTTTTTTGGGGT ACAGTCTATGCCTCGGGCATCCAAGCAGCAAGCGCGTTACGCCGTGGGT	Synthetic oligo to obtain P_{int1+} EMSA probe
wtpint1+pMURR	AACCCACGGCGTAACGCGCTTGCTGCTTGGATGCCCGAGGCATAGACTGT ACCCCAAAAAAACAGTCATAACAAGCCATGAAAACCGCCACTGCGCCGT	Synthetic oligo to obtain P_{int1+} EMSA probe
dxs_upVpa	AGTGCTTTCCGGTAGTCTTTA	Upper primer of *V. parahaemolyticus dxs* gene for quantitative real time RT-PCR assays
dxs_dwVpa	AACCTTTGCCTTTCTTAGTCA	Lower primer of *V. parahaemolyticus dxs* gene for quantitative real time RT-PCR assays

dxs_upEco	GACGAACTGCGCCGCTATT	Upper primer of *E. coli dxs* gene for quantitative real time RT-PCR assays
dxs_dwEco	CCGGCACTGATGGAGGTTGAT	Lower primer of *E. coli dxs* gene for quantitative real time RT-PCR assays
EcoInt_up	AAACCGAGGATGCGAACCACTT	Upper primer of pMUR050 *int* gene for quantitative real time RT-PCR assays
EcoInt_dw	TTACCAACCGAACAGGCTTATG	Lower primer of pMUR050 *int* gene for quantitative real time RT-PCR assays
VpaInt_up	CTGAATGCTATTTCGTTTTTAT	Upper primer of *V. parahaemolyticus int* gene for quantitative real time RT-PCR assays
VpaInt_dw	CACCTTTCCCTTGCCAGACT	Lower primer of *V. parahaemolyticus int* gene for quantitative real time RT-PCR assays
RecAEco_up	CCTTGCGGCACGTATGATGA	Upper primer of *E. coli recA* gene for quantitative real time RT-PCR assays
RecAEco_dw	CACCACGTTTTCGCCCTCTTT	Lower primer of *E. coli recA* gene for quantitative real time RT-PCR assays
RecAVpa_up	AAGTAGCATTTCACGCAGTTTG	Upper primer of *V. parahaemolyticus recA* gene for quantitative real time RT-PCR assays
RecAVpa_dw	AGAAGGCGATGAAGTTGTAGGT	Lower primer of *V. parahaemolyticus recA* gene for quantitative real time RT-PCR assays

[a] *Nde*I or *Xho*I endonuclease restriction sites included in the oligonucleotide sequences are shown in italics.