



Straightforward measurement of individual $^1J(\text{CH})$ and $^2J(\text{HH})$ in diastereotopic CH_2 groups



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ABSTRACT

The C-H_A cross-peak corresponding to a diastereotopic CH_AH_B methylene spin system exhibits a characteristic 1:0:1 multiplet pattern along the indirect dimension of a ω_1 -coupled HSQC spectrum. It is shown here that the use of the initial ^{13}C Boltzmann polarization instead of the regular INEPT-based ^1H Boltzmann polarization makes visible the central lines of this multiplet pattern. A spin-state-selective method is proposed for the efficient measurement of both $^1J(\text{CH}_A)$ and $^1J(\text{CH}_B)$ along the indirect dimension of a 2D spectrum as well as to the magnitude and the sign of the geminal $^2J(\text{H}_A\text{H}_B)$ coupling constant from the straightforward analysis of a single four-component E.COSY cross-peak. Additionally, the extraction of $^1J(\text{CH})$ values for CH and CH_3 multiplicities can be also performed from the same spectrum. The success of the method is also illustrated for the determination of residual dipolar $^1D(\text{CH})$ and $^2D(\text{HH})$ coupling constants in a small molecule weakly aligned in a PMMA swollen gel.

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1. Introduction

In recent years, it has appeared an enormous interest for the measurement of scalar and residual dipolar (RDC) one-bond proton–carbon coupling constants ($^1J(\text{CH})$ and $^1D(\text{CH})$, respectively) in small molecules dissolved in weakly aligned anisotropic media [1–3]. HSQC-based pulse schemes have been generally chosen for this purpose and the accuracy and the simplicity on the experimental measurement of $^1J(\text{CH})$ are subjects of discussion. Some topics of recent interest have been (i) the design of general and robust NMR methods that works efficiently for all multiplicities, (ii) the discussion about whether the $^1J(\text{CH})$ splitting should be measured from the direct ω_2 (^1H) or the indirect ω_1 (^{13}C) dimension, (iii) the accurate measurement of $^1J(\text{CH})$ for individual protons in diastereotopic CH_2 or NH_2 groups, or (iv) the simultaneous determination of additional coupling constants from the analysis of the same cross-peak, being the maximum interest the sign-sensitive determination of geminal $^2J(\text{HH})$ values.

The measurement of $^1J(\text{CH})$ from the detected dimension is relatively easy and high levels of digital resolution are readily available. For instance, the CLIP-HSQC experiment prove to be an efficient tool to determine the $^1J(\text{CH})$ value from the resulting clean in-phase doublets [4]. However, strong $J(\text{HH})$ coupling effects can

generate a high degree of asymmetry between the high- and low-field multiplet lines in ω_2 -coupled HSQC spectra, which can preclude reliable determination of $^1J(\text{CH})$ coupling constants values. In addition, broad signals and/or the large contributions of RDCs can generate poorly defined multiplets that make difficult accurate measurements. These drawbacks have already been described, particularly for CH spin systems in carbohydrates or on the typical strong geminal interaction found in diastereotopic CH_2 spin systems, and some practical solutions have been proposed [5–9]. To avoid such inconveniences, the measurement of $^1J(\text{CH})$ along the ω_1 dimension have been advisable [9,10] although this requires the need for a large number of t_1 increments, and therefore longer acquisition times. The successful use of non-linear uniform sampling, J scaling factors or spectral folding can speed up data acquisition and/or increase the digital resolution in the ω_1 dimension [10].

The accurate measurement of $^1J(\text{CH})$ for individual protons in diastereotopic CH_AH_B (or NH_AH_B) groups is one of the most challenging tasks in this field. Several methods have been proposed that measure them from the ω_1 or ω_2 dimension, but they all present some drawback that can prevent their general use [11–25]. For instance, the passive $^1J(\text{C-H}_B)$ value can be separately measured into the active H_A cross-peak, and vice versa, along the ω_1 dimension of a J-resolved HMQC experiment [11]. In addition, the large doublet is further split by the $^2J(\text{H}_A\text{H}_B)$ coupling yielding a double-doublet. The disadvantage is that additional experiments can

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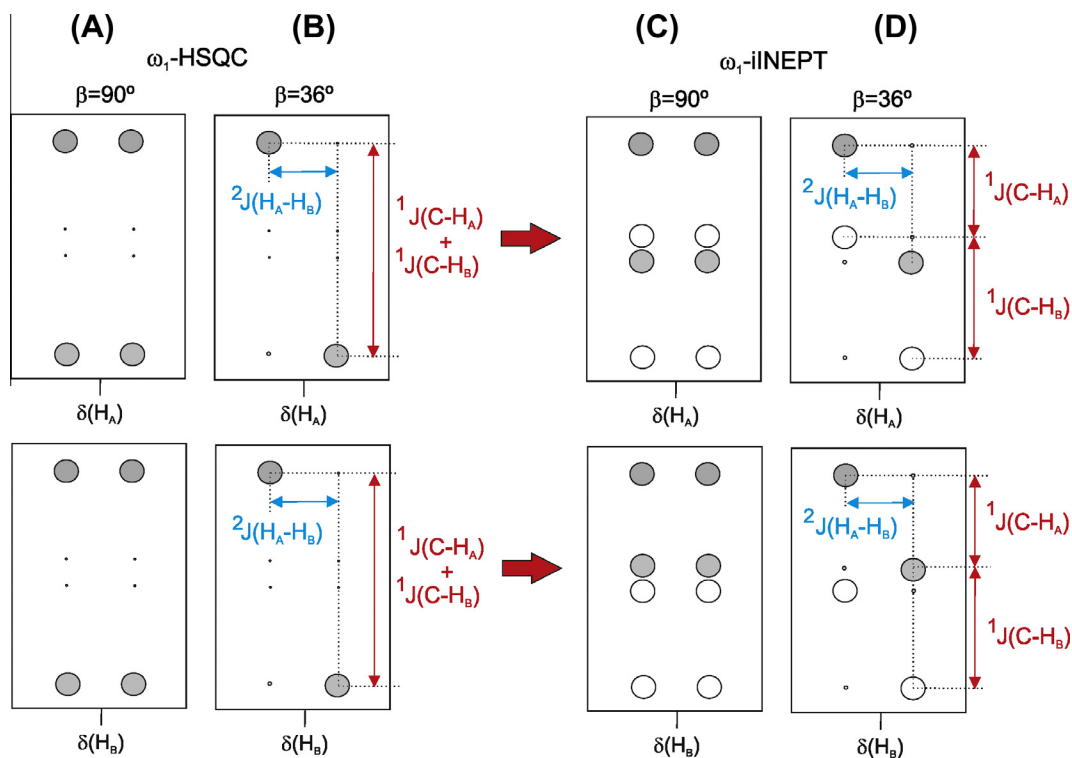


Fig. 2. Schematic representation of the 2D multiplet pattern of each individual H_A and H_B proton belonging to a methylene CH_AH_B group. (A and B) 1H -Boltzmann polarization driven (ω_1 -HSQC) experiments using $\beta = 90^\circ$ and 36° , respectively, and (C and D) ^{13}C -Boltzmann polarization driven experiments (ω_1 -iINEPT) using $\beta = 90^\circ$ and 36° , respectively. In (D), the magnitude and the sign of all involved couplings (defined as $^2J(H_AH_B)$ and assuming $^1J(CH_A) < ^1J(CH_B)$) can be readily extracted. Open and dotted circles represent peaks with opposite phase.

heteronuclear NOE enhanced pre-scan period by means of a 1H WALTZ-16 pulse train saturation, generates in-phase $-C_y$ magnetization (Fig. 1B) which evolves under the effect of $^1J(CH)$ during the variable t_1 BIRD-based period:

$$\begin{aligned}
 & -2H_{Az}C_x[\sin(\pi^1J(CH_A)t_1)\cos(\pi^1J(CH_B)t_1)] \\
 & -2H_{Bz}C_x[\cos(\pi^1J(CH_A)t_1)\sin(\pi^1J(CH_B)t_1)]
 \end{aligned}
 \quad (3)$$

The result is a pure absorptive 2D ω_1 -iINEPT spectra displaying double-doublet coupling patterns along the ω_1 dimension for each individual H_A or H_B cross-peaks, that initially would consist of eight different components as shown in Fig. 2C. Analyzing only the H_A spin, it will show an anti-phase doublet pattern with respect to $^1J(CH_A)$ (sine modulated) and an additional in-phase doublet pattern with respect to $^1J(CH_B)$ (cosine modulated) along the ω_1 dimension. As discussed before, the effect to apply a small flip angle ($\beta = 36^\circ$) will generate a simplified four-component cross-peak with a characteristic E.COSY multiplet pattern due to the mutual $^2J(H_AH_B)$ (Fig. 2D), which facilitates both the multiplet interpretation and analysis (see Fig. S1; supporting information). Thus, the active $^1J(CH_A)$ coupling is directly extracted from the anti-phase 1:–1 pattern along the same column in ω_1 , whereas the passive $^1J(CH_B)$ coupling can be also extracted directly by measuring the in-phase components in each part of the E.COSY pattern. Otherwise, the sign and the magnitude of $^2J(HH)$ is easily extracted from the frequency separation between tilted peaks along the w_2 dimension. Fig. 2 summarizes the expected cross-peak pattern for a single diastereotopic CH_AH_B proton using the different ω_1 -HSQC and ω_1 -iINEPT approaches with $\beta = 90^\circ$ and 36° . For CH groups, a doublet with relative 1:–1 intensities is obtained whereas a 1:1:–1:–1 coupling pattern will be displayed for a CH_3 group.

As a first example, Fig. 3A and B shows the equivalent ω_1 -HSQC and ω_1 -iINEPT correlation spectra, respectively, of strychnine (**1** in Scheme 1) acquired with the pulse schemes of Fig. 1, under the same experimental conditions and using a scaling factor of $k = 8$. The general coupling pattern for individual CH and CH_2 groups are marked with highlighted boxes. CH cross-peaks present the same doublet structure in both approaches. On the other hand, whereas the central lines for each individual CH_2 cross-peak are absent in the ω_1 -HSQC spectrum, they are clearly distinguished in the ω_1 -iINEPT version. A close inspection of the multiplet patterns for CH_2 cross-peaks in both spectra reveals the simplified E.COSY multiplet structure as described in other related experiments [8,19]. Also note the different in-phase vs anti-phase pattern behavior along the indirect dimension, although this is not relevant for the measurement. In cases where chemical shift assignment is already known and/or signal overlapping is not severe, the proposed method can be recorded in a J-resolved mode by simple omission of the ^{13}C chemical shift evolution period ($t_{1/2} - 180^\circ(^1H) - t_{1/2}$). In this way the spectral width in the indirect dimension could be reduced by a factor of about 40, from 20,000 Hz (160 ppm at 500 MHz) to 500 Hz, and therefore the spectral resolution in ω_1 should be improved by a similar factor if all other experimental conditions remain the same. Fig. 3C and D shows the equivalent ω_1 -HSQC-J and ω_1 -iINEPT-J spectra, respectively, acquired with the same number of t_1 increments and using a scaling factor of $k = 1$. It can be observed that the absence of ^{13}C chemical shift signal dispersion does not introduce a serious problem on severe signal overlapping in a small molecule like **1**, where all cross-peaks can be successfully analyzed. All the discussion and conclusions described in Ref. [10] about the application of non-linear sampling to accelerate data acquisition and/or to increase digital resolution in the indirect dimension could

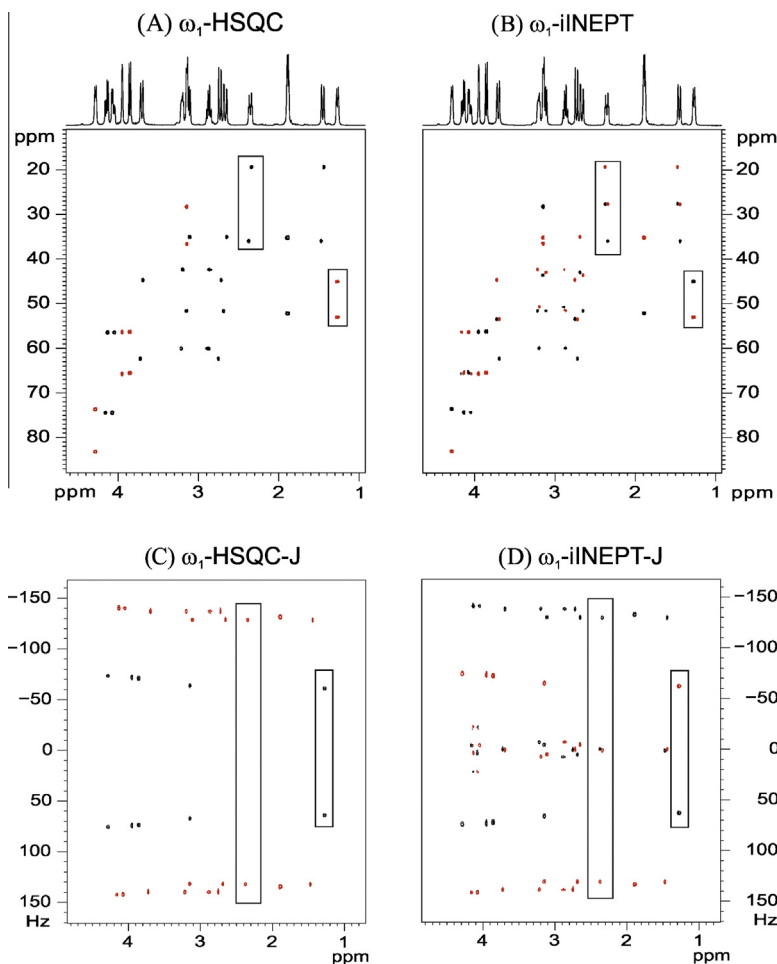


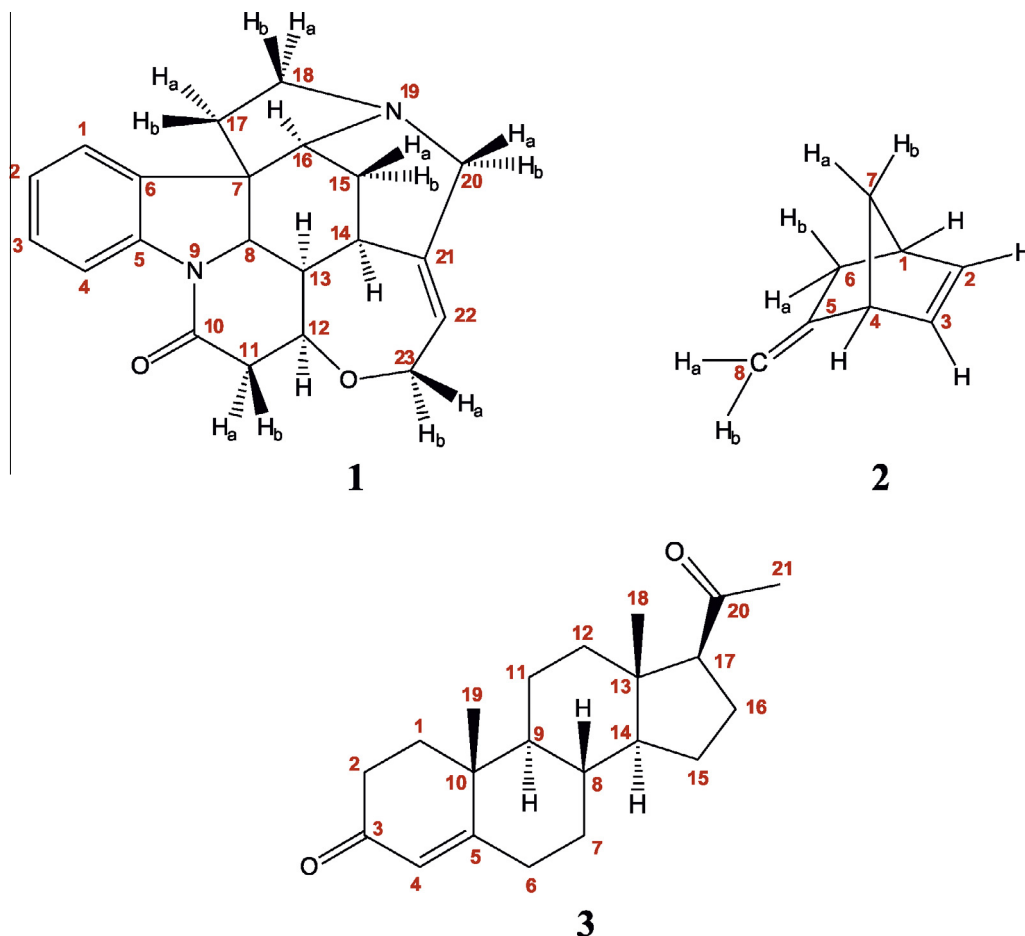
Fig. 3. 2D ^1H - ^{13}C spectra of **1** acquired with the pulse sequences of Fig. 1. (A) ω_1 -HSQC and (C) ω_1 -HSQC-J spectra were obtained starting from ^1H Boltzmann magnetization, and (B) ω_1 -iINEPT and (D) ω_1 -iINEPT-J spectra were achieved starting from ^{13}C Boltzmann magnetization. (A) and (B) are heteronuclear correlation maps (acquired with a scaling factor of $k = 8$) whereas (C) and (D) are the corresponding J-resolved versions (scaling factor $k = 1$ and omission of the ^{13}C chemical shift t_1 evolution period). All spectra were acquired and processed under the same experimental conditions. 2 scans were collected for each one of the 256 t_1 increments using a pre-scan delay of 3 s. Squared boxes mark specific CH and diastereotopic CH_2 cross-peak as examples in each spectrum.

be extrapolated here for the proposed methods. Further increase of resolution by a factor of 3 can be additionally achieved by allowing signal folding in the indirect dimension (see Fig. S2 in the supporting information for an example provided using a reduced spectral width of 180 Hz).

Fig. 4 shows an expanded area of the 2D ω_1 -iINEPT-J spectrum, where the clean tilt, the straightforward analysis and the excellent resolution of the resulting peak patterns can be quickly observed. Note the perfect equivalence between the cross peaks of diastereotopic protons (for instance H-18a vs H-18b or H-11a vs H-11b) which permit the easy and direct measurement of the same three different involved couplings ($^1\text{J}(\text{CH}_A)$ and $^1\text{J}(\text{CH}_B)$) as well as the geminal $^2\text{J}(\text{HH})$ from two independent cross-peaks. The comparison of the experimental J values extracted from these two different measurements evaluates the accuracy of the measurement, and also ensures the measurement of all couplings even in the case of accidental signal overlapping of one of the two diastereotopic proton prevents its analysis. For well resolved $^2\text{J}(\text{HH})$ values, the difference between diastereotopic $^1\text{J}(\text{CH})$ values is quickly ascertained from the relative displacement between the two central lines and accurate $^1\text{J}(\text{CH})$ values can be easily measured even in the case of minor differences between $^1\text{J}(\text{CH}_A)$ and $^1\text{J}(\text{CH}_B)$. For instance, whereas a small difference smaller than 2 Hz is measured for the H-15a/H-15b pair, a big difference about 14.5 Hz is found

for H-18a/H-18b. Experimental $^1\text{J}(\text{CH})$ and $^2\text{J}(\text{HH})$ data extracted from these spectra for compound **1** are in agreement with those reported previously in other works (see Table S1 in the supporting information) [5,36–39]. Even in the case of signal overlapping, CH cross-peaks can be easily distinguished from those of CH_2 groups by their different doublet or double-doublet coupling patterns and also from the relative opposite phase of the anti-phase components for CH/ CH_3 and CH_2 groups because the BIRD element acts as a multiplicity-editing element. For instance, note the clear distinction and straightforward measurement that can be performed for the three different protons resonating close to 3.1–3.2 ppm.

When two diastereotopic protons have similar chemical shift and $^1\text{J}(\text{CH})$ values, the central lines can be partially or completely cancelled, as shown for the H-17a/H-17b protons resonating at 1.9 ppm in Fig. 3D. Another special case is when the geminal $^2\text{J}(\text{HH})$ is near to 0 Hz, where the distinction of the four E.COSY components will depend of the different $^1\text{J}(\text{CH})$ sizes. One illustrative example is the H-8a and H-8b olefinic protons belonging to the exocyclic CH_2 group in 5-methylene-2-norbornene (**2**) (Fig. 5A) which present unresolved signals in the conventional ^1H spectrum, and where the mutual $^2\text{J}(\text{H8a-H8b})$ coupling can not be directly measured. The well differentiated four components observed in the ω_1 -iINEPT-J spectra allow a measurement of $^2\text{J}(\text{HH}) = +1.1$ Hz,



Scheme 1. Molecules used in this work.

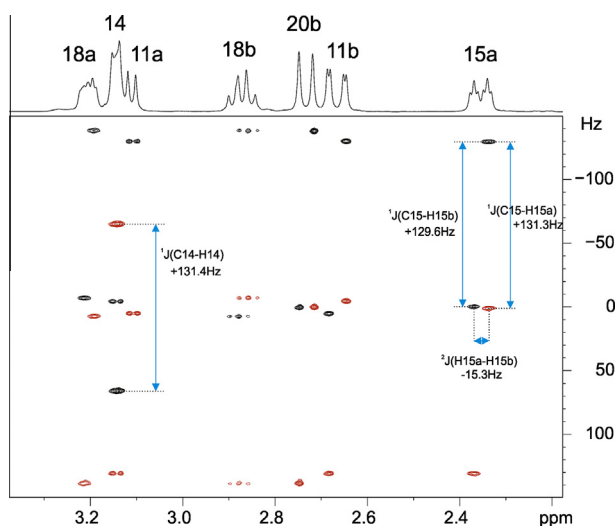


Fig. 4. Expanded area corresponding to the ω_1 -iINEPT-J spectrum of Fig. 3D, where the different multiplet patterns for a CH group and several diastereotopic CH_2 spin systems can be clearly visualized and analyzed, and all $^1J(\text{CH})$ and $^2J(\text{HH})$ can be measured with simplicity and accuracy.

where the positive sign can be determined by comparison with the opposite E.COSY tilt presented by other diastereotopic H-6 and H-7 methylene protons, which have large negative $^2J(\text{HH})$ values of -15.3 Hz and -8.1 Hz, respectively. Although there are small dif-

ferences between the two central lines, the measurement of each individual $^1J(\text{CH})$ (155.1 vs 157.1 Hz) can be performed twice, independently from each cross-peak and with a minimal deviation of ± 0.1 Hz (Table S2; supporting information).

The simplicity of the proposed ω_1 -iINEPT methods make them highly interesting for the measurement of small $^1D(\text{CH})$ and $^2D(\text{HH})$ RDCs, by comparison the difference between experimental measurements performed in isotropic vs anisotropic conditions ($D = T_{(\text{aniso})} - J_{(\text{is})}$). Compound **2** was weakly aligned in a poly(methyl methacrylate) (PMMA) gel swollen in CDCl_3 using the reversible compression/relaxation method [40], and $^1D(\text{CH})$ and $^2D(\text{HH})$ RDCs magnitudes and signs could be easily determined for all signals (see Fig. S3C; supporting information). Fig. 5 compares some cross-peaks obtained in both isotropic and anisotropic conditions. It can be seen how the relative orientation of each diastereotopic HH pair is clearly differentiated from their $^2D(\text{HH})$ values: -0.2 Hz for H-8a/H-8b protons, -4.1 Hz for H-6a/H-6b protons and $+3.1$ Hz for H-7a/H-7b protons. A list of all measured scalar and residual dipolar coupling constants can be found in Table S2 of the supporting information.

The last example corresponds to a molecule having a more complex ^1H spectrum, progesterone (**3**), with high levels of signal overlapping in its aliphatic region. Fig. 6 shows an expanded area of the ω_1 -iINEPT spectrum, where cross-peaks for all multiplicities can be distinguished and the corresponding $^1J(\text{CH})$ and $^2J(\text{HH})$ values conveniently measured (see Table S3 in the supporting information). For instance, note the excellent signal dispersion and multiplet editing for the five resonances fully overlapped in the 1.5–1.65 ppm area. Accidental overlapping of multiplet components

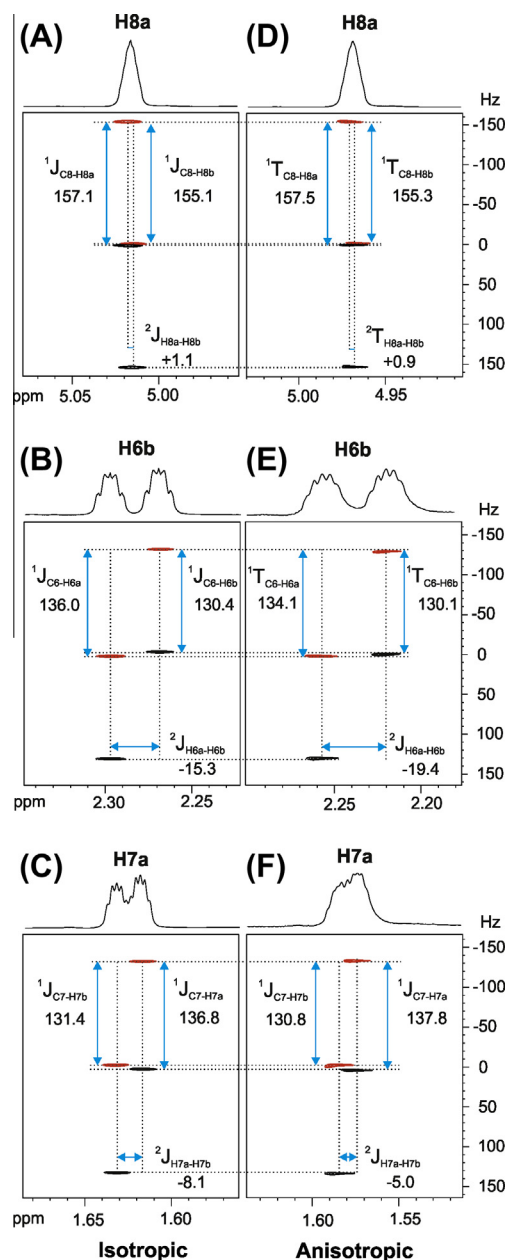


Fig. 5. Some illustrative 2D cross-peaks extracted from the ω_1 -iINEPT-J spectra of **2** showing the easy measurement of the experimental $^1J(\text{CH})/^1T(\text{CH})$ and $^2J(\text{HH})/^2T(\text{HH})$ values measured in (A–C) isotropic (CDCl_3) and (D–F) anisotropic (weakly aligned in PMMA gel swollen in CDCl_3) conditions. Similar values for the same couplings are also measured from the cross-peaks of the other diastereotopic protons (see a complete set of coupling values in Table S2 in the Supporting information).

can be overcome by using the J-resolved version or by changing the scaling factor. The diastereotopicity in the three protons belonging to a methyl groups is not observed and they usually appear as a singlet due to their free rotation under isotropic conditions. However, in analogy with the discussion presented here for diastereotopic CH_2 spin systems, the same conclusions could be extracted from the analysis of a hypothetical non-equivalent protons in a CH_3 group [8,41]. Whereas isotropic CH_3 cross-peaks with no distinction between equivalent protons present a typical 3:1:1:3 multiplet pattern in ω_1 -HSQC experiments [34], they display a symmetrical 1:1:–1:–1 coupling pattern in the ω_1 -iINEPT, as seen for the Me-21 in Fig. 6. A modified HSQC experiment has been

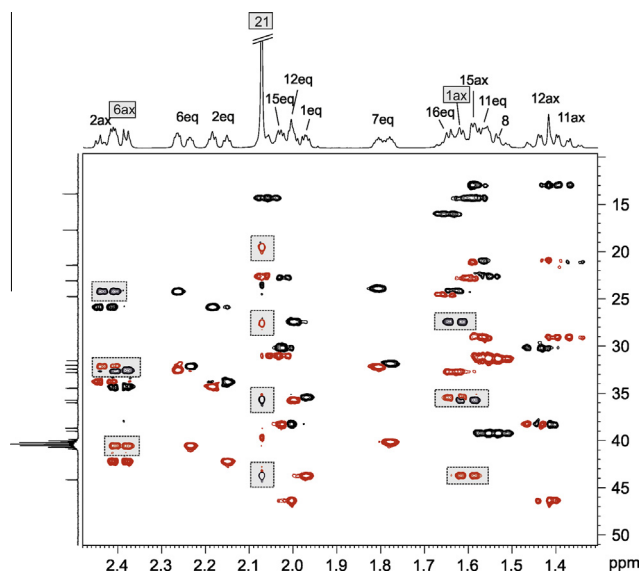


Fig. 6. Expanded area of the 2D ω_1 -iINEPT spectrum of **3** acquired with 4 scans for each one of the 256 t_1 increments, and using a pre-scan delay of 3 s. Boxes enhance the different components corresponding to the H-6ax and H-1ax protons. In addition, the 1:1:–1:–1 multiplet corresponding to the methyl group 21 (at 2.08 ppm) is also highlighted.

reported to recover the 1:2:1 and 1:3:3:1 pattern in NH_2 and NH_3^+ groups, respectively, and spin-state selected methods to study differential relaxation of the different line multiplets of methyl cross-peaks in proteins [41].

In terms of sensitivity, the ω_1 -iINEPT experiment present a sensitivity decrease when compared to the analog ω_1 -HSQC experiment, because of the differential signal enhancement achieved by heteronuclear polarization transfer via INEPT or by heteronuclear NOE effects. In addition, the pre-scan delay must be optimized as a function to the longer ^{13}C T_1 values, although that protonated carbons relax relatively fast. Our experimental data confirms such theoretical prediction and signal-to-noise enhancements by a factor of about 3 and 4 can be achieved for ω_1 -iINEPT and ω_1 -HSQC experiments, respectively, when compared with a reference non signal-enhanced ω_1 -iINEPT experiment acquired without proton saturation and a pre-scan delay of 3 s (see Fig. S4 in the supporting information). Although the proposed methodology could distinguish diastereotopic protons in NH_2 groups, the large difference in sensitivity enhancement achieved by polarization transfer when compared with those obtained by direct ^{15}N Boltzmann magnetization without NOE enhancement (a theoretical factor about 10) makes the experiment of limited practical use due to its very low sensitivity. In addition, the two central lines are likely to be quite broad for large molecules.

In summary, a general and simple NMR method to obtain a characteristic spin-state-selected multiplet pattern for diastereotopic CH_AH_B methylene systems has been described. The magnitude and the sign of the three involved coupling values ($^1J(\text{CH}_A)$, $^1J(\text{CH}_B)$ and $^2J(\text{H}_A\text{H}_B)$) can be measured simultaneously from the analysis of a single and clean four-component E.COSY cross-peak. The method also measures $^1J(\text{CH})$ for all other carbon multiplicities, and it is easily adapted for a J-resolved representation that allows the use of a more reduced spectral width in the carbon dimension, obtaining higher levels of resolution within the same experimental time. We have also shown that small $^1D(\text{CH})$ and $^2D(\text{HH})$ RDCs can be measured for small molecules weakly aligned in anisotropic media. The proposed techniques are appropriate for routine use because require minimum set-up and afford simple data analysis and interpretation.

3. Methods and materials

The isotropic samples used in this work were 0.12 M strychnine dissolved in CDCl_3 (**1**), 0.14 M 5-methylene-2-norbornene dissolved in CDCl_3 (**2**) and 0.13 M progesterone dissolved in DMSO (**3**) (see chemical structures in Scheme 1). For the measurement of RDCs, 10 mg of **2** was aligned in a poly(methyl methacrylate) (PMMA) gel swollen in CDCl_3 using the reversible compression/relaxation method [40]. The ^2H quadrupolar splitting ($\Delta\nu_Q$) for the CDCl_3 signal was of 24 Hz. NMR experiments on **1** and **3** were recorded on a BRUKER DRX-500 spectrometer equipped with a 3-channel 5-mm cryoprobe incorporating a z-gradient coil. NMR experiments on **2** were carried out in a Bruker Avance 600 spectrometer equipped with a TXI HCN z-grad probe. The temperature for all measurements was set to 298 K.

In all experiments, the inter-pulse Δ ($=1/(2 * ^1J(\text{CH}))$) delays were set to 3.5 ms (optimized to $^1J(\text{CH})=145$ Hz). Gradient ratios for G1:G2:G3 were set to 80:20.1:13, measured as percentage of the absolute gradient strength of 53.5 G/cm. Sine bell shaped gradients of 1 ms of duration and followed by a recovery delay of 100 μs were used. ^1H saturation during the entire pre-scan delay was accomplished applying a 2.5 kHz WALTZ-16 modulated pulse train. Broadband ^{13}C decoupling during acquisition was achieved applying a 8 kHz GARP modulated pulse train. All experiments were acquired and processed using the echo/anti-echo protocol where the gradient G1 was inverted for every second FID. An scaling factor $k = 8$ were used for the correlation experiments for all compounds. The J-resolved spectra were acquired omitting the $t_{1/2} - 180(^1\text{H}) - t_{1/2}$ element in the pulse sequence of Fig. 1 and reducing the spectral width in the indirect ω_1 dimension to 500 Hz.

For spectra of Figs. 3, 2 scans were accumulated for each one of the 256 t_1 increments and the number of data points in t_2 was set to 2048. The recycle delay was set to 1 s for ω_1 -HSQC type experiments (Fig. 3A and C) and 3 s for ω_1 -iINEPT type experiments (Fig. 3B and D). Spectra 3A and 3B were acquired with an spectral window of 5000 Hz (in ω_2) and 20,000 Hz (in ω_1) giving a FID resolution of 2.4 and 9.8 Hz, respectively. Prior to Fourier-transformation of each data, zero filling to 4096 in ω_2 , 1024 in ω_1 and a $\pi/2$ -shifted sine-squared window function in both dimensions were applied. After applying zero filling the digital resolution was 1.2 and 2.4 Hz, respectively. In spectra of Figs. 3C and 3D, the spectral window in ω_1 dimension was reduced to 500 Hz giving a FID resolution of 2.4 Hz (in ω_2) and 1.9 Hz (in ω_1). After applying zero filling the digital resolution was 1.2 and 0.5 Hz, respectively.

In the ω_1 -iINEPT-J experiments recorded on **2** in isotropic and anisotropic media (Fig. 5), a recycle delay of 3 s was used, 4 scans were accumulated for each one of the 256 t_1 increments and the number of data points in t_2 was set to 2048. Both of them were acquired with an spectral window of 3600 Hz (in ω_2) and 500 Hz (in ω_1) giving a FID resolution of 1.8 and 1.9 Hz, respectively. Prior to Fourier-transformation of each data, zero filling to 4096 in ω_2 , 1024 in ω_1 and a $\pi/2$ -shifted sine-squared window function in both dimensions were applied. After applying zero filling the digital resolution was 0.9 and 0.5 Hz, respectively. In the ω_1 -iINEPT experiment recorded on **3** (Fig. 6), a recycle delay of 3 s was used, 4 scans were recorded for each one of the 256 t_1 increments and the number of data points in t_2 was set to 2048 in all the experiments. Data were acquired with an spectral window of 2000 Hz (in ω_2) and 12,500 Hz (in ω_1) giving a FID resolution of 1.0 and 6.1 Hz, respectively. Prior to Fourier-transformation of each data, zero filling to 4096 in ω_2 , 1024 in ω_1 and a $\pi/2$ -shifted sine-squared window function in both dimensions were applied. After applying zero filling the digital resolution was 0.5 and 1.5 Hz, respectively.

Acknowledgments

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Appendix A. Supplementary material

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.jmr.2014.02.003>.

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