

1 **Organic Cultivation of Jasmine and Tea Increases C**

2 **Sequestration by Changing Plant and Soil Stoichiometry**

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23 **ABSTRACT**

24

25 **RATIONALE**

26 Organic cultivation methods would be a good alternative to conventional cultivation, avoiding
27 the use of industrial fertilizer and reducing the risk of eutrophication, but its impacts on soil
28 elemental composition and stoichiometry warrants to be clearly stated.

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30 **OBJECTIVES**

31 This study was conducted to determine the effects of long-term organic cultivation on soil
32 elemental composition, stoichiometry and carbon storing capacity and CO₂ emissions in the
33 plant-soil systems of jasmine and tea plantations in Fujian and other regions in China.

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35 **METHODS**

36 We examined the impact of organic cultivation on the concentrations, contents and
37 stoichiometric relationships among carbon (C), nitrogen (N), phosphorus (P), and potassium
38 (K).

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40 **RESULTS**

41 Organic cultivation was associated with lower plant N and P concentrations, and P
42 mineralomasses and with higher total plant C:N, C:P, C:K, and N:P ratios and higher soil N
43 and P concentrations and contents at some depths. Organic cultivation was thus associated
44 with a shift of P from plants to soil and with a higher nutrient-use efficiency in biomass
45 production, mainly of P. Soil CO₂ emissions were higher under organic cultivation, but the
46 soil was able to accumulate more C with no changes in C storage in plant biomass, suggesting
47 that organic cultivation could increase the overall C sequestration, thereby mitigating climate

48 change and enhancing soil nutrient content.

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50 CONCLUSIONS

51 Our results thus showed that the organic cultivation of jasmine and tea in Fujian can improve
52 soil fertility and C accumulation, reduce the use of industrial fertilizers and phytosanitary
53 products, and improve product quality without loss of economical profits **KEYWORDS**

54 Nitrogen; phosphorus; N:P; organic cultivation; stoichiometry; Globally Important
55 Agricultural Heritage Systems

56

57 Abbreviations

58 C, carbon; N, nitrogen, P, phosphorus; K, potassium;

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69 INTRODUCTION

70 China is the world's largest producer of tea, with 1.849×10^6 ha⁻¹ of cultivation producing
71 1.359×10^6 t of tea annually (You et al., 2013). Tea is an important cash crop in the subtropical
72 hilly region of China and is mainly distributed in the red soil area, which is one of the most
73 important continental ecosystems in China (You et al., 2013; Wang et al., 2014). Jasmine tea
74 is unique, and China is the only country that has mastered the critical scenting technologies.
75 Protecting this production system is thus important for the protection and inheritance of
76 Chinese culture and traditional technologies. More than half of the jasmine tea in China is
77 produced in Fuzhou Province (Xu et al., 2001; Yang et al., 2008; Xu, 2012). The system for
78 culturing jasmine and other tea plants near the city of Fuzhou was added in 2014 to the United
79 Nation's Globally Important Agricultural Heritage Systems due to its long historic, ecological
80 and cultural function in this region (Lin et al., 2014; Wang et al., 2014; Ren et al., 2015). The
81 climate is very favorable for this activity, and the method for scenting the tea was developed
82 here more than 1000 years ago (Qian, 2011; Xu, 2012).

83 Substituting common agricultural methods based on the intensive use of industrial
84 fertilizers and the chemical control of crop pests by less environmentally aggressive methods
85 is a challenge for the future viability of the extensive cultivation of crops such as jasmine and
86 common teas. This is especially relevant in China, where the pollution associated with the
87 rapid development has had severe environmental impacts. Organic agriculture does not use
88 genetically engineered organisms, synthetic pesticides, industrial fertilizers, growth regulators,
89 feed additives or other substances in order to maintain sustainable and stable agricultural
90 production systems (AQSIQ and SAC, 2011). Organic cultivation in China is currently the
91 most important method for simultaneously improving production quality and soil fertility
92 (Deng et al., 2010). The impacts and consequences of the application of mid-term organic
93 cultivation on C and nutrient allocation and stoichiometry in the plant-soil system, however,

94 are poorly known. This information would provide the tools for introducing new management
95 strategies (such as the controlled use of chemical fertilizers) to achieve long-term optimal
96 nutrient conditions for the system as a whole, including an equilibrium among soil quality,
97 crop yield and quality and the pollution/eutrophication risk from the leaching of excess
98 exchangeable soil nutrients.

99 The present study was conducted in subtropical jasmine and tea fields in Fujian
100 Province, China. We chose fields that had long been cultivated using common and organic
101 methods to ensure that any soil differences were due at least partially to the long-term
102 differences between the two cultivation types. Plantations for the production of organic and
103 common jasmine and tea have different basic strategies of crop management (Table S1). We
104 then (1) studied the soil pH, texture of soils in the two cultivation types (2) studied the
105 nutrient concentrations, contents and stoichiometric ratios of the plants and soils in the two
106 cultivation types, (3) examined the relationship between cultivation type and the soil-plant
107 capacity to store C and of CO₂ emissions, and (4) the overall shifts in plant-soil stoichiometry
108 and soil texture among the two crop species under different cultivation type.

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119 **MATERIALS AND METHODS**

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121 **Study area**

122 This study was conducted in the jasmine and tea culture system near the city of Fuzhou,
123 which is one of the globally important agricultural heritage systems (Fig. S1). The system
124 includes Jin'an and Cangshan districts and Minhou, Lianjiang and Yongtai counties, Changle
125 County in Fuzhou City, Fujian Province, P.R. China, at 118°08'-120°31' E and 25°15'-
126 26°29'N. The climate is subtropical with mean annual temperatures and precipitation of
127 19.7 °C and 1349 mm, respectively. The frost-free period is >300 days. The soil type is red
128 soil. The system is in a hilly area of agroforestry eco-systems in southeastern China.
129 Mountainous and hilly areas cover 72.7% of the region with complex topography. Green tea
130 and jasmine cultivation provide 30% of total household income, and migrant labor and trade
131 provide the remainder. As stated above, the jasmine in Fuzhou is mostly planted in riverside
132 wetlands and shoals (Fig. S2). From high to low elevation, one can see, in the following order,
133 tea plants, trees, buildings, jasmine plants and waterways. Cultivated jasmine and tea trees
134 can enhance water and soil conservation in many ways (Wang et al., 2014). Jasmine trees are
135 mostly planted on the plains and shoals along rivers. They thus prevent rainwater from
136 directly scouring the riverside, thereby mitigating soil and water erosion. Tea trees are planted
137 in terraced fields. The trees enhance the infiltration of water into the soil and decrease the
138 amount and speed of surface-water runoff and thus the scouring of the soils on the slopes,
139 thereby contributing greatly to soil and water conservation (Wang et al., 2014). Jasmine and
140 tea cultivation also improve air quality and increase carbon (C) fixation, oxygen release and
141 nutrient storage (Ren et al., 2014; Wang et al., 2014). Jasmine and tea trees, together with
142 their diversified microclimates, have contributed to the topographic complexity of these
143 regions of China.

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145 **Experimental design**

146 We established 12 plots on a jasmine and a tea plantation to determine the associations
147 between organic cultivation and the concentrations and ratios of plant and soil C, nitrogen
148 (N), phosphorus (P) and potassium (K). Three plots (1 m² each) were randomly selected at
149 each of the organic jasmine, common jasmine, organic tea and common tea sampling
150 locations at the two sites (two types of plantation × two types of cultivation = four stands). We
151 collected aboveground biomass from stands of the organic and common cultivation plots at
152 the jasmine and tea plantations. We randomly sampled the aboveground biomass from three
153 randomly selected sub-quadrats (1 × 1 m) in each stand. Soil and plant samples were collected
154 in March 2013, which was within the growth period.

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156 **Collection and analysis of soil and plant samples**

157 One soil profile (width, 1 m; length, 1 m; depth, 0.5 m) was excavated in each plot. Samples
158 were collected with a small sampler (length, 0.3 m; diameter 0.1 m) from each of five soil
159 layers (0-10, 10-20, 20-30, 30-40 and 40-50 cm) at the center and on both sides of the soil
160 pits. These three samples from each layer were bulked to form one sample per layer. A total of
161 60 soil samples (two types of plantation × two types of cultivation × three plots × five layers)
162 were thus collected.

163 In the laboratory, the samples were air-dried, roots and visible plant remains were
164 removed and the samples were finely ground in a ball mill. The soil C and N concentrations
165 were determined using a Vario MAX CN Elemental Analyzer (Elementar Scientific
166 Instruments, Hanau, Germany). Total soil P concentration was determined by perchloric-acid
167 digestion followed by ammonium-molybdate colorimetry and measurement using a UV-2450
168 spectrophotometer (Shimadzu Scientific Instruments, Kyoto, Japan) and total K concentration

169 was determined by FP 640 flame photometry (Shanghai Electronic Technology Instruments,
170 Shanghai, China).

171 Soil bulk density was measured from three 5×3 cm cores per layer, salinity was
172 measured with a DDS-307 conductivity meter (Boqu Scientific Instruments, Shanghai,
173 China), pH was measured with an 868 pH meter (Orion Scientific Instruments, Minnesota,
174 USA), particle size (clay, silt and sand) was measured by a Mastersizer 2000 laser particle-
175 size analyzer (Malvern Scientific Instruments, Suffolk, UK), water content was measured by
176 the drying method (Lu, 1999) and C (CO₂) release was determined by the incubation method
177 (Wang et al., 2010). Briefly, 30 g of fresh soil were placed into 120-mL incubation bottles.
178 The bottles were sealed with rubber stoppers and incubated at 20 °C for three days. Five
179 milliliters of gas were extracted from the headspaces four times a day. CO₂ concentration was
180 determined by GC-2014 gas chromatography (Shimadzu Scientific Instruments, Kyoto,
181 Japan).

182 Aboveground plant samples were collected from a consistent height to reduce the
183 potential effects of site-specific confounding variables. The biomass was sorted into leaves
184 and branches. Belowground biomass was collected from the sample sub-quadrats. All plant
185 material was gently washed with water and then oven-dried to a constant weight (80 °C for
186 24-36 h) and weighed. A total of 36 plant samples (two types of plantation \times two types of
187 cultivation \times three plots \times three organs) were thus collected.

188 The plant C and N concentrations were determined using a Vario EL III Elemental
189 Analyzer (Elementar Scientific Instruments, Hanau, Germany). The P concentrations of the
190 plants were measured using the molybdate-blue reaction (Lu, 1999) with a UV-2450
191 spectrophotometer (Shimadzu Scientific Instruments, Kyoto, Japan). K concentration was
192 determined by FP 640 flame photometry (Shanghai Electronic Technology Instruments,
193 Shanghai, China).

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195 **C, N, P and K content and release**

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197 C, N, P, and K content for the 0-50 cm profiles were estimated using the equation (Mishra et
198 al., 2010):

$$199 \quad C_s = \sum_{j=1}^n c_m \times \rho_b \times D$$

200 where C_s is C, N, P, or K content (kg m^{-2}); j is the soil-depth interval (1, 2, ... n); C_m is the C,
201 N, P, or K concentration (g kg^{-1}); ρ_b is the bulk density (kg m^{-3}); D is the thickness of each
202 layer (m) and n is the number of layers.

203 C release was estimated using the equation (Wassmann et al., 1998):

$$204 \quad P = \frac{dc}{dt} \cdot \frac{V_H}{W_s} \cdot \frac{MW}{MV} \cdot \frac{T_{st}}{T_{st} + T}$$

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206 where P is the rate of C release ($\mu\text{g}^{-1} \text{g}^{-1} \text{d}^{-1}$), dc/dt is the recorded change in the mixing ratio
207 of C (CO_2) in the headspace over time ($\text{mmol mol}^{-1} \text{d}^{-1}$), V_H is the volume of the headspace
208 (L), W_s is the dry weight of the soil (g), MW is the molecular weight of CO_2 (g), MV is the
209 molecular volume (L), T is the air temperature (K), and T_{st} is the standard temperature (K).

210 Most C release from the wetland soil in the study area was in the form of CO_2 (Wang et al.,
211 2010). We also expected that the main form of C release would not be CH_4 , because some of
212 the land uses were not wetlands and thus had no anaerobic periods, so we only determined
213 CO_2 release.

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215 **Statistical analyses**

216 We used general linear models (GLM) with plantation type (tea and jasmine), cultivation type
217 (common and organic) and soil depth as independent categorical variables and with the soil

218 and plant variables as dependent continuous variables. We also used paired-samples t-tests to
219 compare the variables between common versus organic cultivation within each plantation
220 type. We used Statistica 8.0 (StatSoft, Inc., Tulsa, USA) for the analyses. The relationships
221 among the soil variables were examined by Pearson correlation analysis.

222 We also performed multivariate statistical analyses using a general discriminant analysis
223 (GDA) to determine the overall differences in soil traits in the tea and jasmine plantations
224 with common and organic cultivation. We also took into account the component of the
225 variance due to the different soil depths as an independent categorical variable. Discriminant
226 analyses consist of a supervised statistical algorithm that derives an optimal separation
227 between groups established a priori by maximizing between-group variance while minimizing
228 within-group variance (Raamsdonk et al., 2001). GDA is thus an adequate tool for identifying
229 the variables most responsible for the differences among groups while controlling the
230 component of the variance due to other categorical variables. The GDAs were performed
231 using Statistica 6.0 (StatSoft, Inc., Tulsa, USA).

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243 **RESULTS**

244 **Univariate analyses**

245 **Soil pH and texture**

246 Organic cultivation was associated with higher soil pH for both the jasmine (0-20 cm) and tea
247 (0-50 cm) plantations (Fig. S3). Soil pH in the jasmine plantations was higher under organic
248 than common cultivation. Soil texture was less sandy and contained higher proportions of clay
249 under organic cultivation (>30 cm, Fig. S3). This difference was larger in the tea than the
250 jasmine plantations, consistent with the longer time of organic tea (30 years) than jasmine
251 (five years) cultivation.

252

253 **Soils and plant nutrient and C concentration, content and stoichiometry**

254 The organic cultivation of jasmine had higher soil C and P concentrations at 0-50 cm, higher
255 soil N concentrations at 10-20 and 40-50 cm and lower K concentrations at 20-50 cm than
256 common cultivation. The organic cultivation of tea had higher soil C concentrations at 10-20
257 and 40-50 cm, higher N concentrations at 10-30 and 40-50 cm and higher P and K
258 concentrations at 0-50 cm than common cultivation (Fig. 1). The soils under organic
259 cultivation at all depths had higher P ($F=39.7$, $P<0.00001$) and K ($F=11.1$, $P=0.0015$)
260 concentrations relative to the soils under common cultivation. The soils in the jasmine
261 plantations had higher P ($F=176$, $P<0.00001$) and K ($F=26.7$, $P<0.0001$) concentrations than
262 those in the tea plantations regardless of the cultivation type (Fig. 1).

263 These differences in soil elemental concentrations associated with the cultivation type
264 were greater than the differences in soil stoichiometry. Soil C:N, C:K, N:K, and P:K ratios
265 were higher and soil C:P and N:P ratios lower at most depths under organic than common
266 cultivation in the jasmine plantations (Fig. 2). Soil C:N and C:K ratios at most depths were
267 lower under organic than common cultivation in the tea plantations (Fig. 2). The GLM

268 indicated that the C:P, C:K, N:P, and N:K ratios were lower in the jasmine than the tea soil
269 profiles (Table S2).

270 Organic cultivation had higher soil P contents (Mg ha^{-1}) at >30 cm in the jasmine
271 plantations and with higher C, N, and P contents at 10-20 cm in the tea plantations (Fig. 3). C
272 and N contents were lower and P and K contents were higher throughout the soil profiles in
273 the jasmine than the tea plantations (Table S2). Soil P concentration was strongly and
274 positively correlated ($R=0.69$, $P<0.0001$) with clay concentration (Table S3), showing that the
275 higher clay concentration with organic cultivation was associated with the higher P
276 concentrations.

277 Organic cultivation was associated with lower N, P, and K concentrations in the leaves,
278 stems and roots and with lower C concentrations in the leaves and higher C concentrations in
279 the roots of jasmine trees relative to common cultivation (Fig. 4). Organic tea cultivation had
280 lower foliar and root C and N concentrations, and stem N concentrations and higher foliar P
281 and K concentrations, stem C, N and P concentrations, and root P concentrations (Fig. 4).
282 Foliar C:N, C:P, C:K, N:K, and P:K ratios were higher and N:P ratios were lower in jasmine
283 trees under organic than common cultivation (Fig. 5). Stem C:N, C:P, C:K, N:P, N:K, and P:K
284 ratios and root C:N, C:P, and C:K ratios were higher and root N:P, N:K and P:K ratios were
285 lower in jasmine trees under organic cultivation (Fig. 5).

286 C:P, N:K, and N:P ratios were lower and C:N and P:K ratios were higher in the leaves,
287 stems and roots of the tea trees under organic than common cultivation (Fig. 5). Foliar and
288 stem C:K ratios were lower in the tea trees under organic than common cultivation (Fig. 5).

289 None of the interactions between plantation type and cultivation type for biomasses
290 and mineralomasses were significant (Table S4). Root biomasses were higher in the jasmine
291 than the tea plantations. K contents were higher in all organs, N contents were higher in stems
292 and roots, and C and P contents were higher in roots in the jasmine than the tea plantations

293 (Table S4). Plants under organic cultivation had lower P contents in all organs and lower K
294 contents in leaves and stems (Table S4). C, N and K mineralomasses were higher in the
295 jasmine than the tea plantations. Total P mineralomasses were lower under organic cultivation
296 (Table S5). The total mineralomass C:N, C:P, N:P, C:K, and P:K ratios were higher in the tea
297 than the jasmine plantations, whereas the total mineralomass C:N, C:P,N:P, C:K, and N:K
298 ratios were higher and P:K ratios were lower under organic than common cultivation (Table
299 S5).

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301 **Soil CO₂ emission**

302 Soil CO₂ emissions were higher with organic cultivation from 0-10 cm in the jasmine
303 plantations and at depths >20 cm in the tea plantations (Fig. 6). CO₂ emissions throughout the
304 soil profile were higher in the tea plantations under common cultivation (Table S2).

305

306 **Multivariate analysis. Overall differences among crop types and cultivation methods**

307 The GDA showed that the soil variables separated all four combinations of plantation ×
308 cultivation type (jasmine with common cultivation, jasmine with organic cultivation, tea with
309 common cultivation and tea with organic cultivation) (Table S6, Fig. 7). Soil total C, N, and P
310 concentrations, N:K and P:K ratios and C and N contents had significant loadings in the
311 model (Table S7).

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318 **DISCUSSION**

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320 **Soil texture**

321 Organic cultivation was correlated with the higher proportion of clay than sand in these soils.
322 This effect was greater in the tea than the jasmine plantations, also consistent with the longer
323 period of organic cultivation in the tea plantations. These results thus strongly suggest that
324 organic cultivation contributed to the enrichment of clay in the soils, thereby changing the soil
325 texture and contributing to the capacity of the soil to store/release nutrients. Moreover, the
326 higher soil P concentrations and lower soil N:P ratios in the soils under organic cultivation in
327 tea crops were correlated with the higher proportion of clay than sand in these soils. Thus, the
328 results strongly suggested that the observed changes in soil elemental composition associated
329 to organic cultivation should be due at least in part to the related increase of soil clay
330 concentration.

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332 **Plant and soil nutrients concentrations, content and stoichiometry**

333 Organic cultivation was not associated with changes in plantation biomasses but was
334 associated with changes in plant and soil nutrient concentrations and stoichiometric ratios.
335 Organic cultivation had higher soil N and P concentrations and contents in both the jasmine
336 and tea plantation at some soil depths. Under organic cultivation tea and jasmine plants had
337 higher C:N, whereas tea had lower C:P ratios and jasmine higher C:P ratios in total biomass
338 thus suggesting higher N- and P-use efficiency in jasmine and higher N- and lower P-use
339 efficiency in tea. Soil elemental ratios differed between the two cultivation types. P
340 concentration was proportionally higher than C and N concentrations under organic
341 cultivation in both plantation types, whereas C:N and P:K ratios differed between the two
342 cultivation types, depending on the plantation type. Jasmine trees had higher allocations of

343 biomass and nutrients to the roots, higher overall C, N, and K contents and lower C:nutrients
344 and N:P ratios than the tea trees.

345 In jasmine whereas P concentrations were higher in soil under organic cultivation, they
346 were lower in biomass, further suggesting an increase of P use efficiency. Differently, P
347 concentrations increased in both soil and biomass in in tea cropland organic cultivation. These
348 results provide thus evidences that the association of organic cultivation with plant-soil
349 stoichiometry in croplands depends of cropland type. However, under organic cultivation we
350 observed a decrease of soil total K concentrations in certain soil depths in jasmine croplands
351 and an increase in tea croplands. The different effects of organic cultivation on soil texture
352 between the two cultivation methods seems to be underlying these different responses
353 between the two studied species. The higher clay concentrations observed in tea soils under
354 organic cultivation are consistent with the increases in soil K contents, because K^+ is strongly
355 retained by clay in soils (Cofie and Pleysier, 2004; Blank, 2010) and at the same time clay is a
356 primary source of new K^+ (Askegaard et al., 2003; Blank, 2010). The different potential
357 leaching/sedimentary balance in the bottom of the valleys in the river flooding areas in the
358 case of jasmine croplands with respect to the soils of the top of the mountain in the case of
359 teas opens different scenarios and interactions with organic cultivation. In any case, both the
360 soil total N and P concentrations increase under organic cultivation in both crops. Increases in
361 P and other nutrients such as Zn and Cu in soil have been associated with the organic
362 cultivation of cotton (*Gossypium hirsutum* L.) (Blaise et al., 2004). Organic cultivation has
363 been widely demonstrated to be able to improve the chemical traits of plantation types and
364 their nutritional quality, including higher vitamin concentrations and contents of total
365 phenolics and soluble sugars (del Amor et al., 2008; Hallman, 2012; Lombardo et al., 2012).
366 Organic cultivars have been associated with decreases in plant biomass N and P
367 concentrations, also in agreement with our results (López et al., 2013). In contrast to our

368 results, however, the organic cultivation of strawberries did not induce changes in
369 macronutrient concentrations between plant tissues (Hargreaves et al., 2008).

370

371 **Plant and soil carbon concentrations, content and stoichiometry**

372 The higher levels of C stored in the soil despite the higher CO₂ soil emission together with no
373 significant difference in C content in total plant biomasses strongly suggested that organic
374 cultivation stored more C at the level of the plant-soil system. The long-term organic
375 cultivation (5 years in the jasmine plantations) and common cultivation of jasmine
376 accumulated 70.8 and 68.2 Mg ha⁻¹ of organic carbon, respectively. The long-term organic
377 cultivation (30 years in the tea plantations) and common cultivation of tea accumulated 71.5
378 and 69.4 Mg ha⁻¹ of organic carbon, respectively.

379 Thus, despite the high soil CO₂ emissions in the organic cultivation type, the higher
380 soil C concentration strongly suggests that organic cultivation increases the sequestration of C
381 and thus in turn may help mitigating climate change. Previous studies have reported similar
382 results (Chirinda et al., 2010; Lehtinen et al., 2014). Higher soil respiration, frequently
383 measured in organic cultivars, has been generally correlated with soil fertility, texture, higher
384 concentrations of C and higher diversity and density of soil microbes and fauna (Pimentel et
385 al., 2011; Lehtinen et al., 2014).

386

387 **Overall effects of organic cultivation in plant-soil stoichiometry, carbon balance and** 388 **yield**

389 The higher soil C, N, P and clay concentrations in organic plantations in tea crops are
390 consistent with the expected positive and synergic link widely observed among organic matter
391 concentration, clay concentration, fertility, stability and resistance against erosion (Wagner et
392 al., 2007; Wuddivira et al., 2009; Soinne et al., 2014). The cementing potentials of clay and

393 organic matter are important for the stability of soil aggregates for preventing erosion and
394 leaching (Wagner et al., 2007; Wuddivira et al., 2009; Abdollahi et al., 2014; Peng et al.,
395 2015). Sources of organic matter improve the stability of clay aggregates and therefore
396 improve nutrient levels and C-retention capacity (Soinne et al., 2014). The application of
397 organic matter increases clay aggregation that in turn has a positive feed-back on organic
398 matter and aggregate stabilization (Djajadi et al., 2012).

399 These improvements in soil nutrient contents are unfortunately associated with lower
400 yields. The average yields of jasmine flowers in the study area are 6.2 and 12.0 t ha⁻¹ y⁻¹ in
401 organic and common cultivation, respectively, and the average yields of tea are 4.2 and 6.0 t
402 ha⁻¹ y⁻¹, respectively. Organic cultivation has been widely associated with lower yields than
403 the corresponding common cultivars (López et al., 2013; Yousef et al., 2015), but not all
404 yields are lower (Seidler-Lozykowska et al., 2015). The price of organic products is,
405 however, >2-3-fold higher than the price of products from common cultivation. If the yield of
406 an organic cultivation type is less than half, but the price is more than double, then total
407 benefits will improve. Moreover, the results support organic cultivation as a very important
408 method for improving mid- to long-term soil fertility and provide further evidence supporting
409 the plans of Chinese government of promoting organic cultivation as a useful tool for
410 improving the safety of crop production and for decreasing the negative environmental effects
411 of the intensive use of inorganic-industrial fertilizers and organic compounds against pests
412 without decrease farmer's economy.

413 The higher accumulation of C and nutrients in the soil together with the higher clay contents suggested that
414 the continuous use of organic cultivation instead of traditional cultivation can increase soil fertility and
415 improve production capacity for the mid-term. Studies in other cropland systems have reported that soil
416 fertility, soil C and nutrient concentrations and even yield have continuously increased after several years
417 of applying organic fertilizers under organic cultivation (Rasool et al., 2007; Zingore et al., 2008). Our
418 results are consistent with the premise that organic cultivation will be important for the future development

419 of sustainable agriculture in China. However, the continuous use of organic cultivation improved soil
420 nutrient contents but decreased yield and some plant nutrient concentrations (particularly in Jasmine plants).
421 The effect of supplementation with industrial fertilizer after several years of strictly organic cultivation
422 should thus be investigated. The most logical hypothesis would be that the improved soil conditions after
423 years of organic cultivation would favor a better use of moderate amounts of nutrients from industrial
424 fertilizers and would thus improve yields.

425 **CONCLUSSIONS**

426 Organic cultivation affects soil texture under tea but not under jasmine crops.

427 Soil total N and P concentrations are higher under organic cultivation in both studied species.

428 Organic cultivation shifted P from plants to soil in tea crops, whereas in jasmine crops this
429 was not observed.

430 Jasmine plants under organic cultivation presented higher C:P ratios, so P-use efficiency
431 increased, and the contrary was observed in tea crops. Differently, both cropland species
432 presented higher C:N ratios under organic cultivation and thus N-use efficiency increased in
433 them both.

434 The increase in C stored in soil in both crop types, together with the non-significant decrease
435 in the C stored in plant biomass, suggest that organic cultivation is able to increase C fixation,
436 despite the increase in soil respiration associated with organic cultivation.

437 The lower accumulation of P in biomass of Jasmine plants under organic cultivation was not
438 associated with a decrease in biomass; instead it was related with a great decrease (50%) of
439 flowers production, suggesting a decoupling between vegetative and reproductive
440 productivity

441 The results gave consistent support to the fact that organic cultivation is a very important
442 method for improving mid- to long-term soil fertility without decreasing farmer's economy.

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444

445 **CONFLICTS OF INTEREST**

446 The authors declare no conflicts of interest.

447

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584 **Figure legends**

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586 **Fig.1.** Concentrations (mean \pm S.E.) of soil C (a, b), N (c, d), P (e, f) and K (g, h) in the
587 jasmine and tea plantations. Different letters indicate significant differences ($P < 0.05$) between
588 organic and common cultivation in a paired-samples t-test. Error bar indicates standard error
589 of the mean of triplicate measurements. (Green color indicates data corresponding to common
590 cultivation method, whereas red color indicates data corresponding to organic cultivation
591 method).

592 **Fig.2.** Soil C:N (a, b), C:P (c, d), C:K (e, f), N:P (g, h), N:K (i, j) and P:K (k, l) ratios (mean \pm
593 S.E.) in the jasmine and tea plantations. Different letters indicate significant differences
594 ($P < 0.05$) between organic and common cultivation in a paired-samples t-test. Error bar
595 indicates standard error of the mean of triplicate measurements. (Green color indicates data
596 corresponding to common cultivation method, whereas red color indicates data corresponding
597 to organic cultivation method).

598 **Fig.3.** Soil C (a, b), N (c, d), P (e, f) and K (g, h) contents (mean \pm S.E.) in the jasmine and tea
599 plantations. Different letters indicate significant differences ($P < 0.05$) between organic and
600 common cultivation in a paired-samples t-test. Error bar indicates standard error of the mean
601 of triplicate measurements. (Green color indicates data corresponding to common cultivation
602 method, whereas red color indicates data corresponding to organic cultivation method).

603 **Fig.4.** Plant C (a, b), N (c, d), P (e, f) and K (g, h) concentrations (mean \pm S.E.) in the jasmine
604 and tea plantations. Different letters indicate significant differences ($P < 0.05$) between organic
605 and common cultivation in a paired-samples t-test. Error bar indicates standard error of the
606 mean of triplicate measurements. (Green color indicates data corresponding to common
607 cultivation method, whereas red color indicates data corresponding to organic cultivation
608 method).

609 **Fig.5.** Plant C:N (a, b), C:P (c, d), C:K (e, f), and N:P (g, h) ratios (mean \pm S.E.) in the

610 jasmine and tea plantations. Different letters indicate significant differences ($P < 0.05$) between
611 organic and common cultivation in a paired-samples t-test. Error bar indicates standard error
612 of the mean of triplicate measurements. (Green color indicates data corresponding to common
613 cultivation method, whereas red color indicates data corresponding to organic cultivation
614 method).

615 **Fig.6.** Emissions (mean \pm S.E.) of C (as CO₂) (a, b) in soils from jasmine and tea cultivation.
616 Different letters indicate significant differences ($P < 0.05$) between organic and common
617 cultivation in a paired-samples t-test. Error bar indicates standard error of the mean of
618 triplicate measurements. (Green color indicates data corresponding to common cultivation
619 method, whereas red color indicates data corresponding to organic cultivation method).

620 **Fig.7.** Biplot of the standardized canonical discriminate function coefficients for the first two
621 roots representing the various grouping dependent factors corresponding to the plant
622 communities. JC, = jasmine common cultivation; JO = jasmine organic cultivation; TC, = tea
623 common cultivation; TO, = tea organic cultivation.

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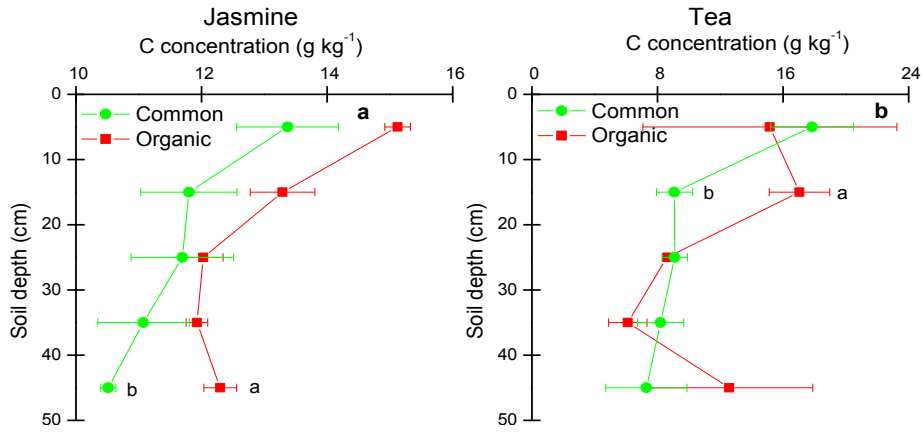
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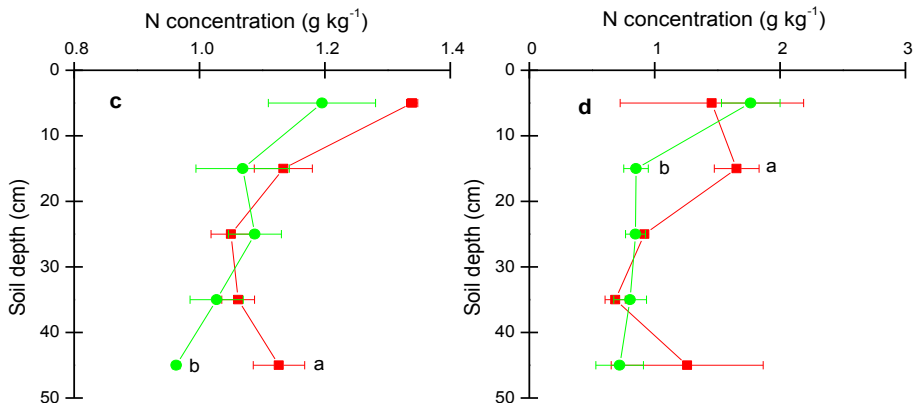
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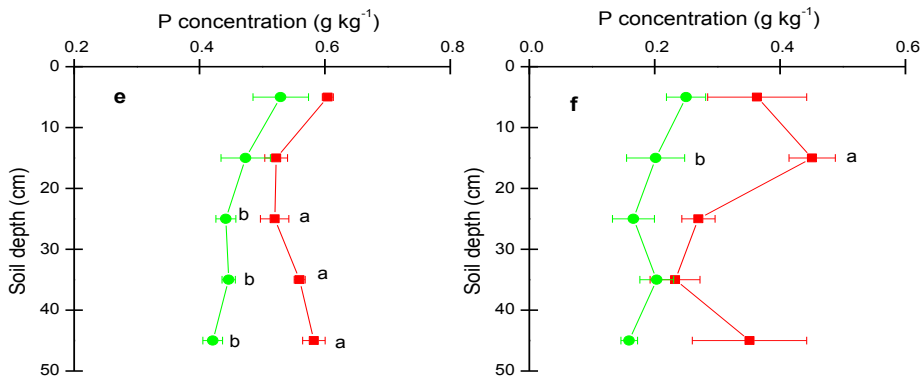
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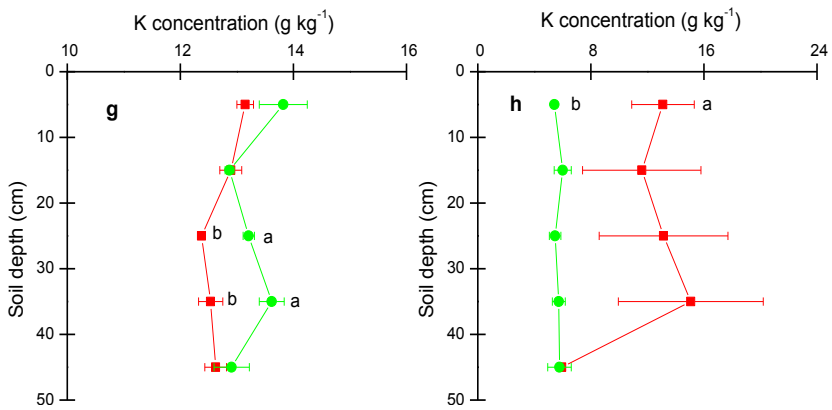
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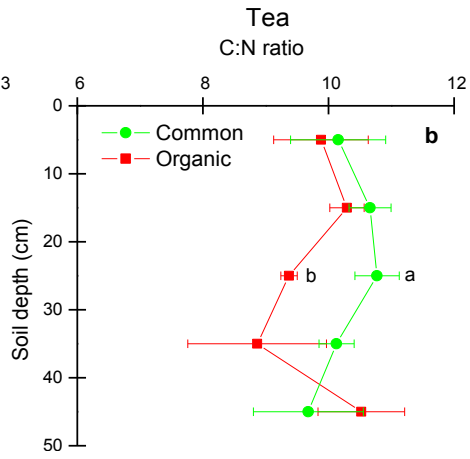
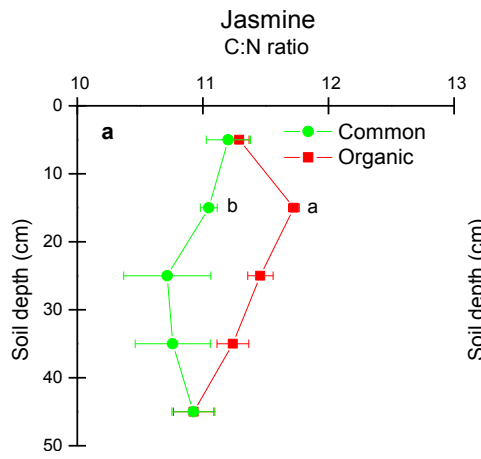
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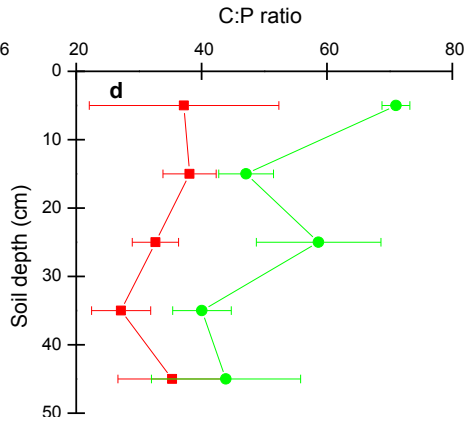
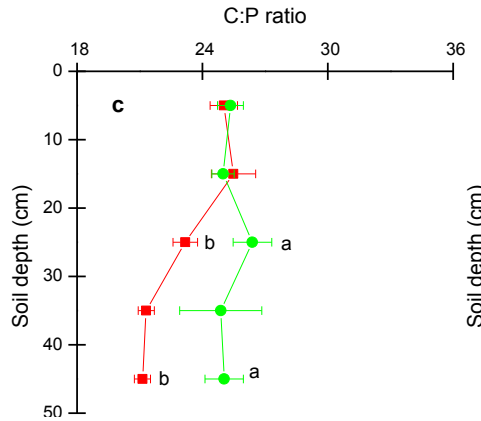
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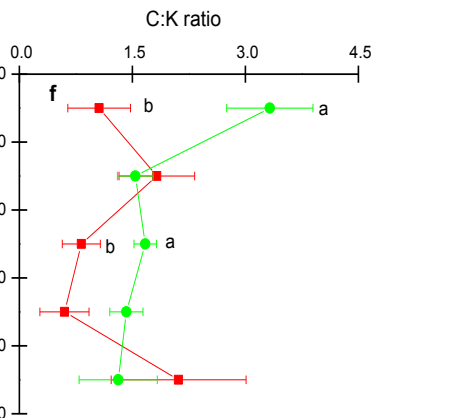
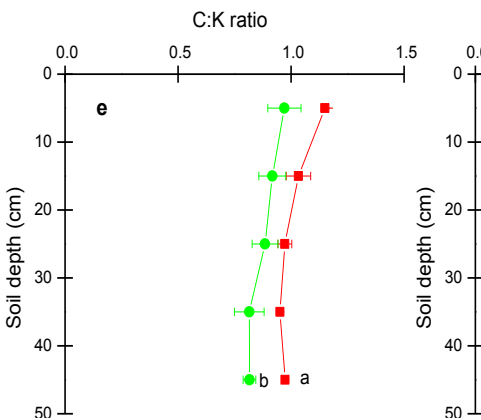
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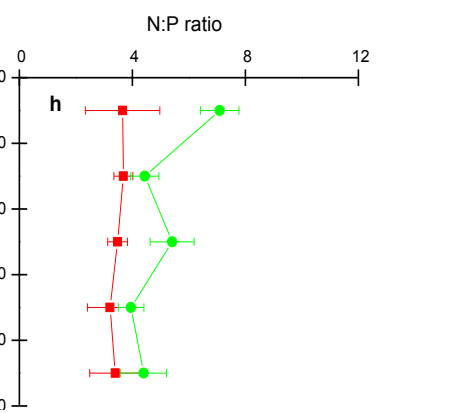
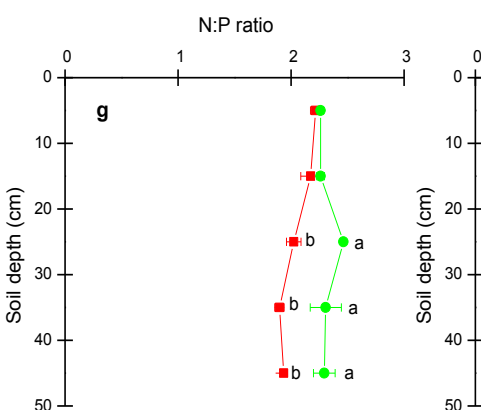
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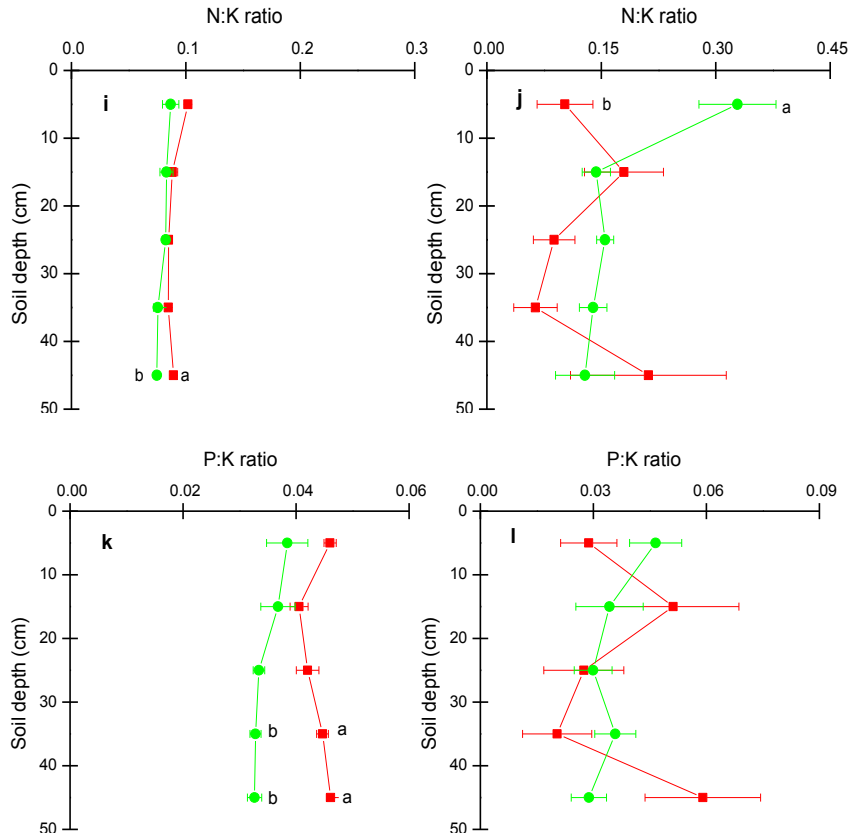


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647 **Fig.2.**

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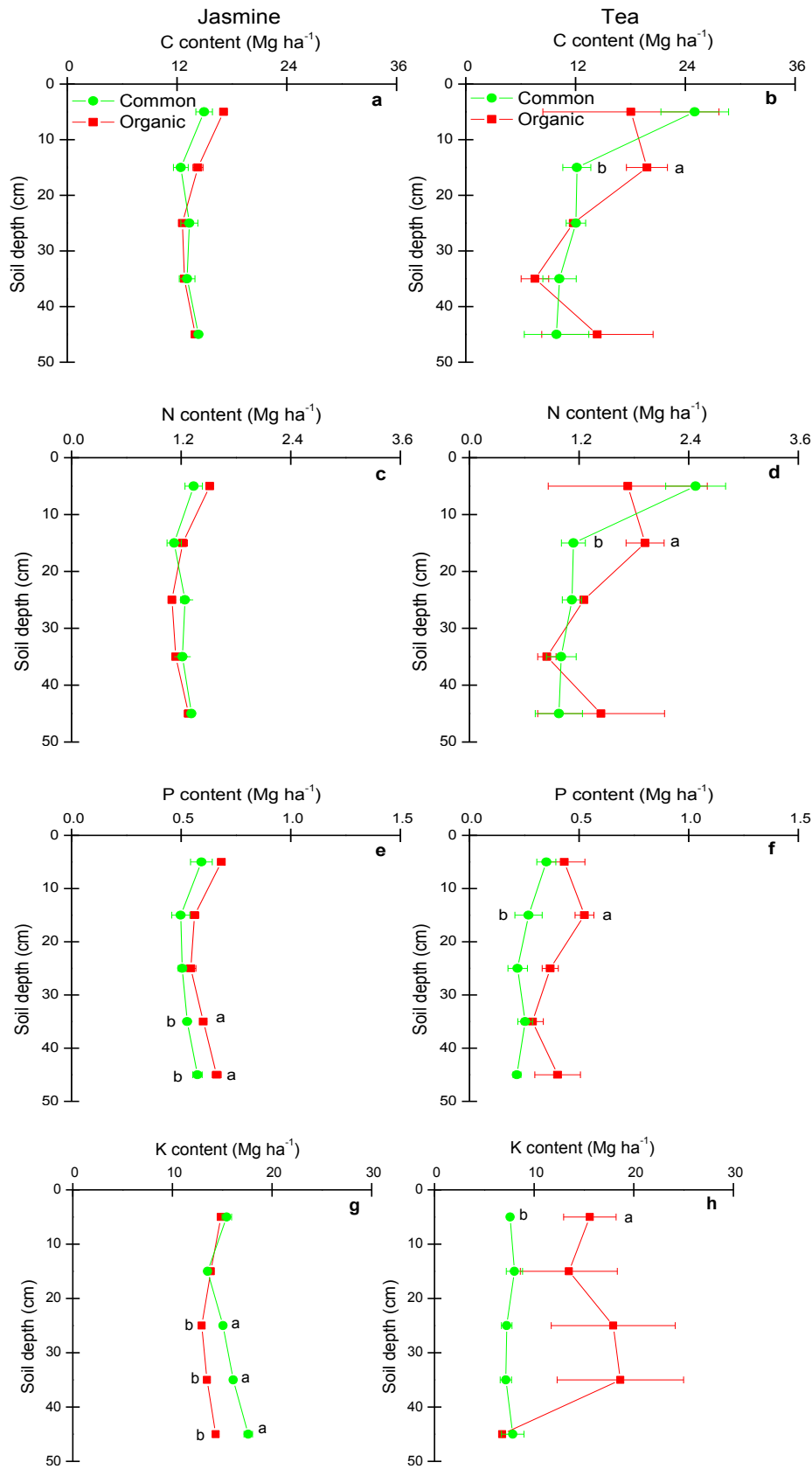
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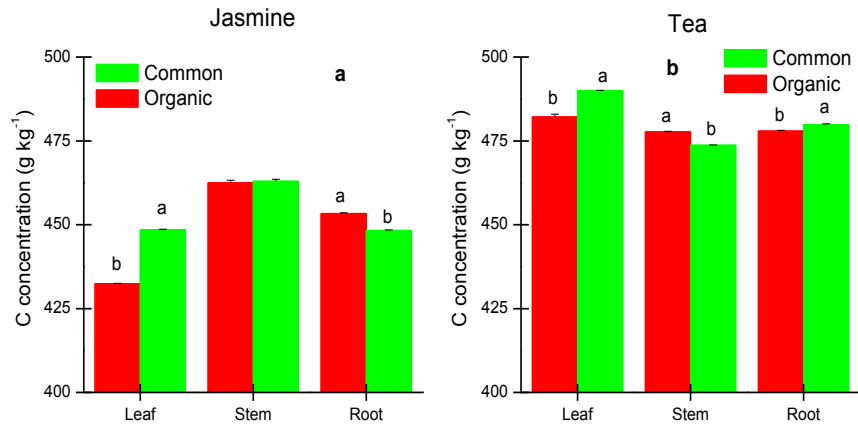
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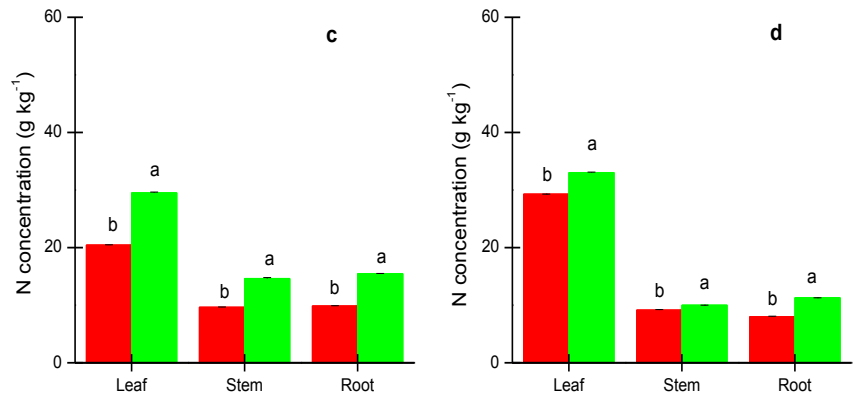
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Fig.3.

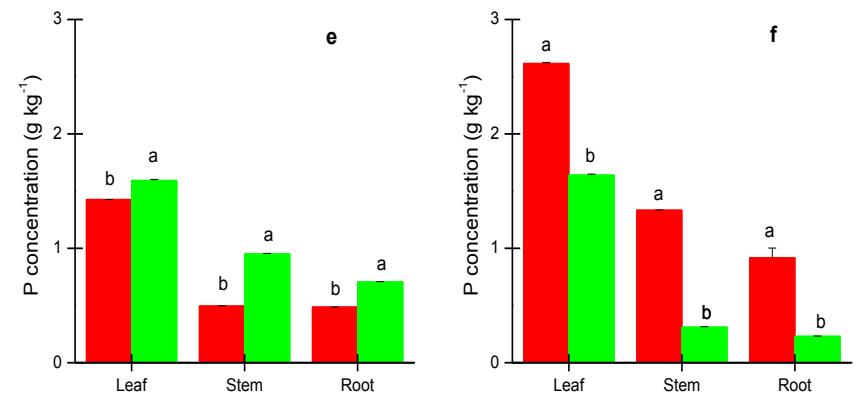
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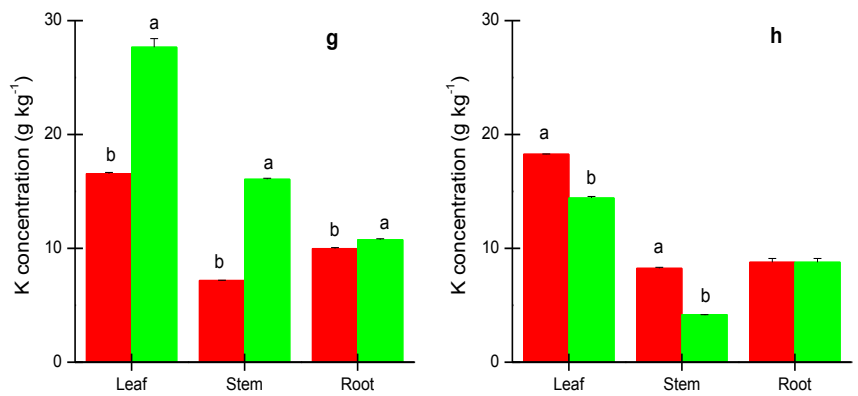
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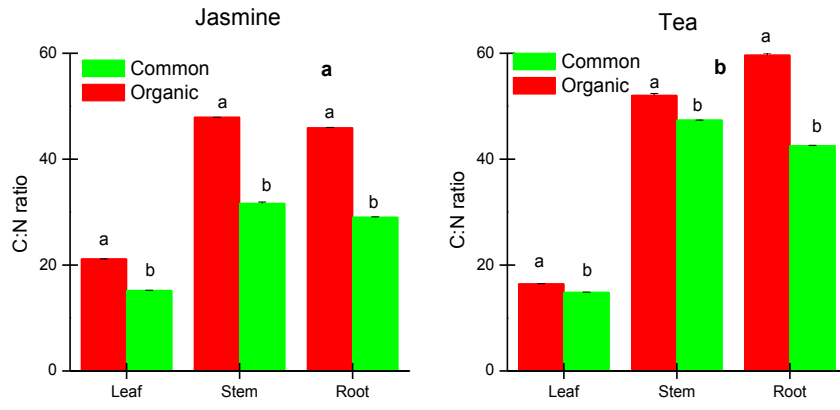


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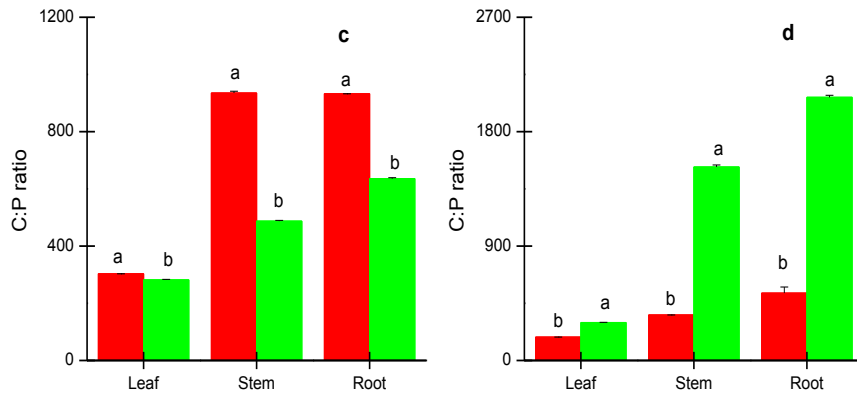


669 **Fig.4.**

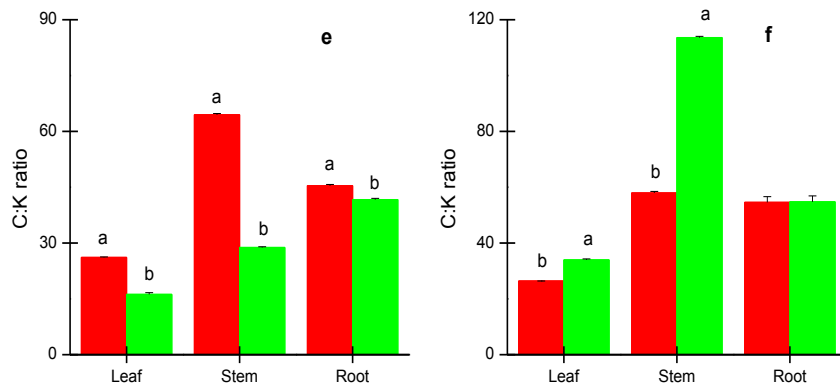
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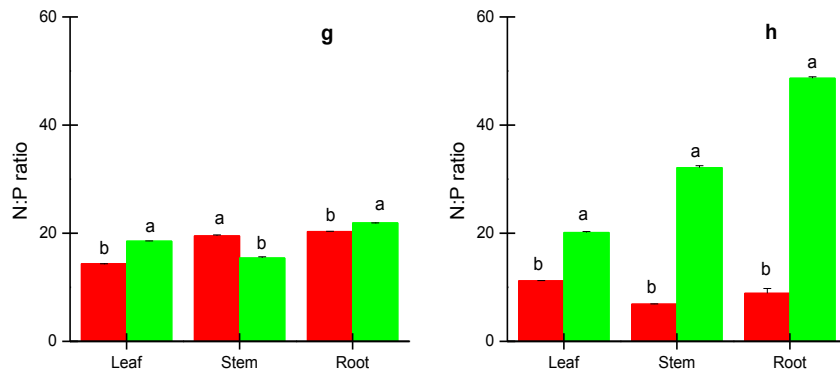
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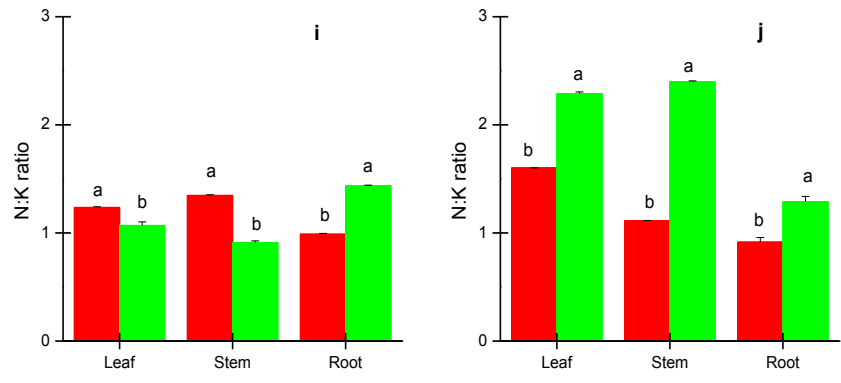
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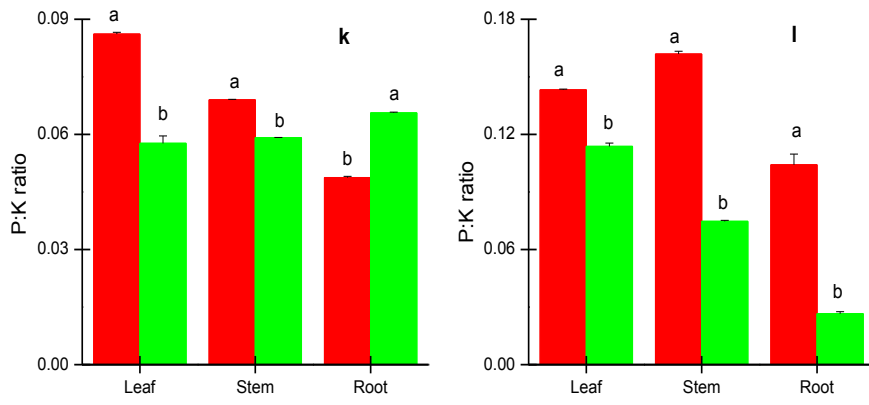
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677 **Fig.5.**

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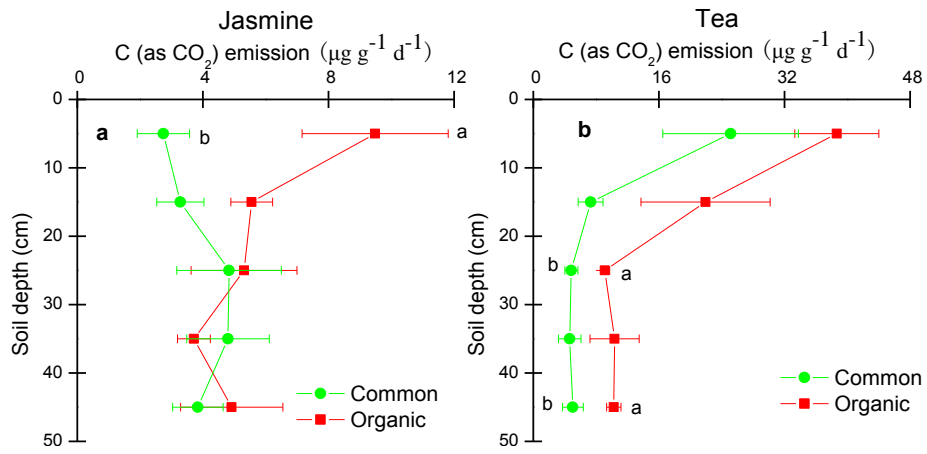
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685 **Fig.6.**

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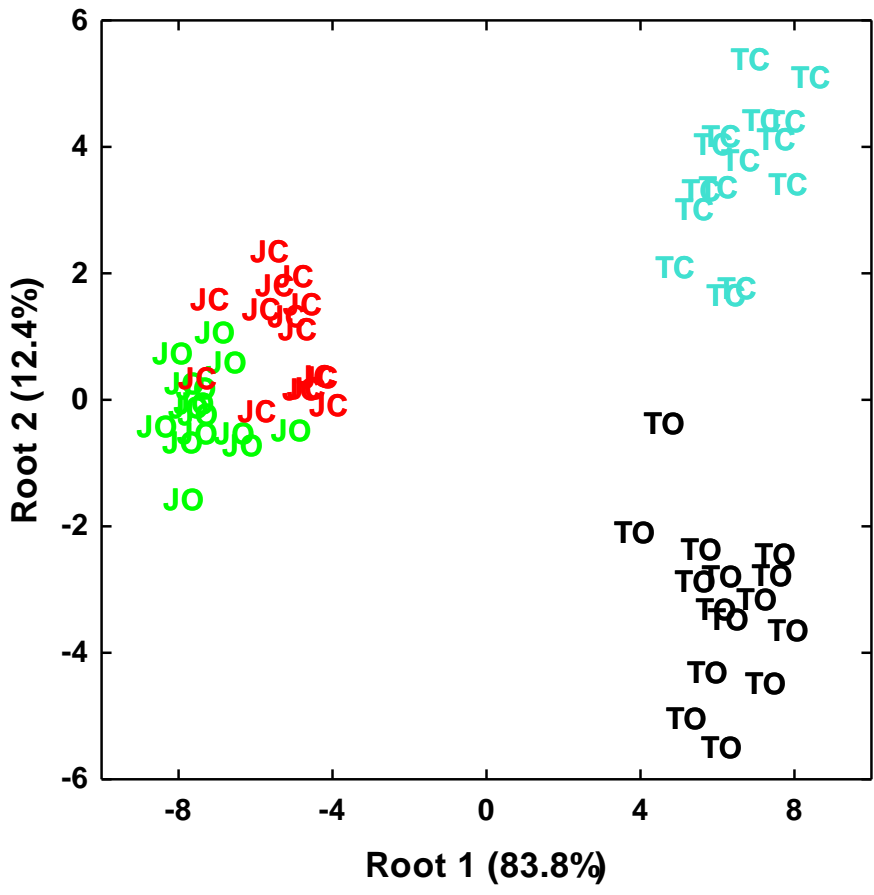
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696 **Fig.7.**