

Biodegradation of a high-strength wastewater containing a mixture of ammonium, aromatic compounds and salts with simultaneous nitrification in an aerobic granular reactor

Carlos Ramos, María Eugenia Suárez-Ojeda, Julián Carrera*

GENOCOV Research Group. Department of Chemical Engineering, School of Engineering, Universitat Autònoma de Barcelona, Edifici Q, 08193 Bellaterra, Barcelona, Spain

*Corresponding author: Tel. +34 935812141

E-mail address: julian.carrera@uab.cat (J. Carrera)

Abstract

Long-term operation (390 days) of a continuous airlift reactor with aerobic granular biomass was successfully applied to treat a highly complex wastewater composed of: ammonium (1000 mg N L⁻¹), *o*-cresol (100 mg L⁻¹), phenol (100 mg L⁻¹), quinoline (50 mg L⁻¹) and salts (16 g salts L⁻¹). High nitrogen loading rate (1.1 g N L⁻¹ d⁻¹) and organic loading rate of 0.7 (g COD L⁻¹ d⁻¹) were achieved for the simultaneous nitrification and complete biodegradation of the aromatic compounds. The successful operation of the granular airlift reactor can be related to (i) the growth of specialized microorganisms in the aerobic granules and (ii) the continuous feeding regime. Aerobic granules were maintained stable in spite of the high salinity conditions. Dissolved oxygen (DO) concentration and DO/ammonium concentrations ratio were the key parameters to select a suitable effluent for anammox or heterotrophic denitrification via nitrite. Besides, nitrous oxide emissions were related to the DO concentration in the reactor.

Keywords: partial nitrification; phenolic compounds; salinity; nitrous oxide

Introduction

Nowadays, the satisfactory and cost-effective treatment of industrial wastewaters from chemical, petrochemical, coke plant and refineries is a huge challenge since they are complex matrices, containing several organic and inorganic compounds, such as toxic/recalcitrant organic compounds (like aromatic compounds), ammonium and several inorganic salts [1,2]. Industrial wastewaters are often treated by physico-chemical processes; however, these technologies have serious drawbacks [3]: (i) high costs due to the high temperature and pressure conditions applied and chemical requirements, (ii) do not allow complete degradation of the aromatic compounds and (iii) they may produce other hazardous by-products (secondary pollutants).

Biological processes can overcome some of the disadvantages of physico-chemical treatments. However, biological processes can be inhibited by aromatic compounds [4]. Biological treatments based on floccular biomass (such as activated sludge systems) are the main biological technology used at full-scale. Nevertheless, its application for treating complex industrial wastewaters is limited because activated sludge systems are known to be highly inhibited by aromatic compounds [2-5]. Therefore, biological systems with floccular biomass: (i) are poor adjustable to fluctuations in the organic loading, (ii) need large reactor volumes because low loading rates can be applied and (iii) its settling capacity is limited, which means large secondary clarifiers. Biological treatments based on aerobic granules represent an alternative to treat these complex industrial wastewaters [6].

Ammonium can be removed by the application of Biological Nitrogen Removal (BNR) processes via nitrite [7,8], where ammonium is oxidized until nitrite (nitrification) and subsequently, nitrite can be reduced by heterotrophic denitrification via nitrite (denitrification) or by anammox (anaerobic ammonium oxidation) processes [7,8]. Nitrification is usually the

bottleneck for BNR in many wastewater treatment processes because nitrifying bacteria are very sensitive to several factors, such as inhibition by aromatic compounds. Biodegradation of aromatic compounds and nitrification can be simultaneously performed in a single aerobic granular reactor due to the substrates gradients occurring through the granules and the high biomass concentration that can be achieved in this kind of reactor [6,9]. Therefore, a two-stage process composed by a first aerobic granular reactor followed by a second anoxic granular reactor has been proposed as a feasible technology to treat complex industrial wastewaters [9,10]. Until now, this option has only been tested with wastewaters containing ammonium and a single aromatic compound: *o*-cresol [9] or *p*-nitrophenol [10]. However, it is well-known that industrial wastewaters can be composed by a mixture of several aromatic compounds; thus nitrification and biodegradation of a mixture of aromatic compounds should be tested to confirm the technological feasibility of this option. Furthermore, the effect of several parameters should be established to guarantee this feasibility: (i) operation at high salinity conditions, (ii) determination of nitrous oxide emissions from nitrification at different dissolved oxygen (DO) concentrations in the reactor and (iii) assessment of the stability of the morphological characteristics of the aerobic granules at long-term. Therefore, the objective of this study is to demonstrate the feasibility of the biodegradation of a high-strength wastewater containing a mixture of ammonium, aromatic compounds and salts with simultaneous nitrification in an aerobic granular reactor. In this sense, nitrous oxide emissions, stability and characteristics of the aerobic granular biomass at long-term operation and the microbial diversity in the granules will be assessed.

2B Materials and methods

Reactor

An airlift reactor made of glass (2.6 L of working volume) was used (Figure 1). The reactor configuration was as follows: the internal diameter of down-comer was 62.5 mm; the riser had a height of 750 mm and an internal diameter of 42.5 mm and it was at 8 mm from the bottom of the down-comer. At the top of the reactor, a gas-solid-liquid separator allowed the separation of the aerobic granules from the treated wastewater. Compressed air was supplied through an air diffuser placed at the bottom of the reactor at an upflow velocity of 0.2-0.3 cm s⁻¹. Air flow rate in the reactor was regulated manually between 150 to 250 mL min⁻¹ by a rotameter (Aalborg, USA) and it was enough to ensure an appropriate flow in the airlift reactor. The reactor was equipped with DO (Crison DO 6050), temperature (Crison Pt1000) and pH probes (Crison pH 5333) that were connected to a data monitoring system (Crison Multimeter 44). DO was not automatically controlled and varied between 0.5 and 4.0 mg O₂ L⁻¹ (excluding the first 25 days) according to the applied airflow (Figure 2A). A Programmable Logic Controller (PLC) coupled to a Supervisory Control And Data Acquisition (SCADA) system regulated temperature, pH and feeding. pH was maintained at 8.0 ± 0.2 by a regular addition of NaHCO₃ whereas temperature in the reactor was maintained at 30 ± 0.5 °C using a temperature controller coupled with a belt-type heating device (Horst, Germany). Feeding to the reactor was made with a membrane pump (ProMinent Gamma/L).

Inoculum

Aerobic granular sludge from an airlift reactor performing partial nitrification and *o*-cresol biodegradation was used as inoculum. The biomass characteristics were as follows: 1.0-1.5 mm of mean granule size, 40-60 m h⁻¹ (67-100 cm min⁻¹) of settling velocity, sludge

volumetric index at 5 min (SVI₅) of 7-14 mL g⁻¹ TSS and SVI₃₀/SVI₅ ratio of 1.0. The biomass was under starvation period in mineral medium during 30 days at room temperature prior to the start-up of the reactor. More information can be found in Jemaat et al. [9].

Wastewater composition and operational conditions

The airlift reactor was fed continuously with a synthetic wastewater. Throughout the operational period, ammonium concentration was of 1000 ± 39 mg N-NH₄⁺ L⁻¹ (3818 mg NH₄Cl L⁻¹) in the influent and different aromatic compounds were also added. During the first 60 days of operation, hydraulic residence time (HRT) was varied between 6.3 to 0.9 days (by increasing the inflow rate) to increase the applied organic (OLR) and nitrogen (NLR) loading rates. From day-60 onwards, HRT was maintained at 0.9 days. The complexity of the treated wastewater was increased stepwise by introducing the aromatic compounds as follows:

- i) Phase I (from day-22 to day-114). Ammonium (1000 ± 39 mg N L⁻¹) and *o*-cresol (100 ± 6 mg L⁻¹) were simultaneously treated.
 - ii) Phase II (from day-114 to day-195). Phenol was added to the influent; therefore ammonium (1000 ± 39 mg N L⁻¹), *o*-cresol (100 ± 6 mg L⁻¹) and phenol were treated together. Phenol concentration in the influent was increased stepwise from 25 ± 1 to 100 ± 3 mg L⁻¹.
 - iii) Phase III (from day-195 to day-332). Quinoline was added to the influent; therefore ammonium (1000 ± 39 mg N L⁻¹), *o*-cresol (100 ± 6 mg L⁻¹), phenol (100 ± 3 mg L⁻¹) and quinoline were treated together. Quinoline concentration in the influent was increased stepwise from 15 ± 2 to 50 ± 4 mg L⁻¹.
- The aim of the first three phases was to achieve an effluent containing only nitrite, removing completely *o*-cresol, phenol and quinoline for a subsequent denitritation stage.

iv) Phase IV (from day-333 to day-390). The influent was the same than in phase III but the aim was to achieve an effluent with a nitrite/ammonium ratio close to 1 for a subsequent anammox stage, removing completely *o*-cresol, phenol and quinoline.

The chemical oxygen demand to nitrogen ratio (COD/N) in the influent increased from 0.25 to 0.64 throughout the experimental period. The composition of the micronutrients in the synthetic wastewater was (expressed as mg L⁻¹): 48 CH₃COONa; 12.5 C₆H₁₂O₆; 11.9 C₁₂H₂₂O₁₁; 41.0 KH₂PO₄; 176.0 NaCl; 198.0 MgCl₂·7H₂O; 4.0 FeSO₄·7H₂O; 3.0 MnSO₄·H₂O; 4.0 ZnSO₄·7H₂O; 2.0 CuSO₄·5H₂O; 0.02 H₃BO₃; 12.0 CO(NH₂)₂ and 2.0 yeast extract. Acetate, glucose and sucrose were added in low amount as readily biodegradable organic matter. Their contribution in the influent composition was low and only represented: 24, 14 and 11 % of the total COD in the phases I, II and III-IV, respectively. Sodium bicarbonate (11.2 g NaHCO₃ L⁻¹) was added to the influent to ensure enough buffer capacity. As a consequence of the high NH₄Cl and NaHCO₃ concentrations, the influent had high salinity conditions (16 g salts L⁻¹, corresponding to a conductivity of 15 mS cm⁻¹).

Kinetic batch experiments

Several batch experiments were carried out around day-300 to have a deep understanding of the biological processes occurring in the airlift reactor. The experiments were performed in batch mode in a reactor of 300 mL of reaction volume (500 mL of total volume). Air was flowed through a pressure manoreductor and a gas flowmeter to ensure a constant airflow and was supplied from the bottom of the reactor through a diffuser in order to mix the reaction media and maintain the granular structure of the biomass. DO concentration ranged between 2 to 4 mg O₂ L⁻¹. The pH was maintained at 8.0, using phosphate buffer. Temperature was set at

30 ± 1 °C. Aerobic granular biomass from the airlift reactor was used and washed three times with the phosphate buffer before being used in the experiment. A pulse of substrate (ammonium and/or the aromatic compound) was added, and nitrite, nitrate, aromatic compounds and any metabolic intermediate concentrations were followed throughout the reaction time. Four different conditions were studied: (i) only ammonium (60 mg N L⁻¹), (ii) *o*-cresol (13 mg L⁻¹) and phenol (11 mg L⁻¹), (iii) ammonium (60 mg N L⁻¹), *o*-cresol (10 mg L⁻¹) and phenol (10 mg L⁻¹) and (iv) ammonium (60 mg N L⁻¹) and quinoline (15 mg L⁻¹).

Samples and analytical methods

Ammonium, nitrite, nitrate, conductivity, aromatic compounds (phenol, *o*-cresol and quinoline) and metabolic intermediates of the aerobic biodegradation of the aromatic compounds (1,2,4-benzenetriol, hydroquinone, catechol, resorcinol and *p*-nitrocatechol) were determined for the influent and the effluent. Prior to the analysis, the samples were filtered through 0.20 µm membrane of hydrophilic Durapore®.

Ammonium was measured by a continuous flow ammonia analyser (Hach Lange AMTAX sc), based on potentiometric determination. Nitrate and nitrite concentrations were analysed with ionic chromatography using an ICS-2000 Integrated Reagent-Free IC system (DIONEX Corporation), which performs ion analyses using suppressed conductivity detection. Phenol, *o*-cresol, quinoline, 1,2,4-benzenetriol, hydroquinone, catechol, resorcinol and *p*-nitrocatechol were measured by High Performance Liquid Chromatography (HPLC) (the methodology is explained in detail in Supplementary Material). For each aromatic compound, the chemical oxygen demand (COD) was determined as the theoretical oxygen demand, which was calculated as the amount of oxygen required to oxidize a compound to its final oxidation products. Conductivity was measured with a CRISON conductivity device (micro CM 2100)

with independent temperature probe and conductivity cell, and the values are given at 25 °C, according to the method 2510 B of the Standard Methods [11]. Total and volatile suspended solids (TSS and VSS, respectively) were determined in the reactor and in the effluent, according to method 2540 D and 2540 E of the Standard Methods [11], for TSS and VSS, respectively.

The granular biomass was characterised in terms of SVI, size, granule density and settling velocity. SVI was determined using the procedure described in the method 2710 D of the Standard Methods [11]. The size of the granules was measured regularly by using image analysis with an optical microscope (Zeiss Axioskop equipped with a video camera (iAi Protec)), according to Jemaat et al. [9] (more details can be found in Supplementary Material). The distribution of the granules size was measured by a laser particle size analysis system (Mastersizer 2000, Malvern instruments). Density of the granular biomass was determined using the Dextran Blue method [12]. Settling velocity was determined by placing individual granules in a column containing distilled water and measuring the time spent to drop a height of 30 cm.

Granules morphology was measured by using image analysis with an optical microscope (Zeiss Axioskop equipped with a video camera (iAi Protec)) and by Scanning Electron Microscope (SEM). The methodology for SEM can be obtained from Julian et al. [13] (more details can be found in Supplementary Material).

Off-gas grab samples were collected from the headspace of the airlift reactor to determine the nitrous oxide (N₂O) emissions. Nitrous oxide concentration in the gas phase was measured off-line in a gas chromatograph. N₂O emissions were determined at several DO concentrations in the bulk liquid (0.5, 1.0, 1.5 and 4.0 mg O₂ L⁻¹) during the period between day-310 and day-330. The detailed procedure and calculations of nitrous oxide emissions can be found in Supplementary Material.

Microbial diversity analysis

Identification of the microbial population at the day-300 was performed using next-generation sequencing. Total genomic DNA of the aerobic granular biomass was extracted and purified using a PowerBiofilmTM DNA Isolation Kit (MoBio Laboratories, USA); in accordance with the manufacturer's instructions. Paired-end sequencing of the extracted DNA was performed on Roche 454 GS-FLX+ platform by Research and Testing Laboratory (Lubbock, Texas, USA). Bacterial 16S rRNA variable regions V2-V4 were targeted using the primer pair 341F-907R. More details can be found in Supplementary Material. Biodiversity analysis and phylogenetic classification was performed with the methodology explained in detail in Supplementary Material. Relative abundances of reads were determined by taxonomic level. Indices of biological diversity were calculated at 97 % of similitude with the following results: number of reads: 9494, Chao1: 334, Shannon: 2.85 and Eubacteria: 0.51. As can be seen in Figure S1 in Supplementary Material, good coverage of diversity was reached. Fluorescence in-situ hybridization (FISH) coupled with Confocal Laser Scanning Microscopy (CLSM) was used to identify betaproteobacterial ammonia oxidizing (β -AOB) and nitrite oxidizing bacteria (NOB). FISH protocol was adapted from Suárez-Ojeda et al. [14] (more details can be found in Supplementary Material).

Results and discussion

Reactor performance: removal capacity and nitrous oxide emissions

224 The performance of the aerobic granular airlift reactor treating a complex wastewater
 225 composed by ammonium and several aromatic compounds (*o*-cresol, phenol and quinoline) at
 226 high salinity (16 g salts L⁻¹) is presented in Figure 2. In general, throughout the 390 days of
 227 operation, the granular airlift reactor was able to maintain stable nitrification and complete
 228 biodegradation of the aromatic compounds. During the first 21 days of operation, the reactor
 229 was operated in batch mode, adding pulses of ammonium (100 mg N L⁻¹) and *o*-cresol (100
 230 mg L⁻¹) at regular intervals of 24-48 hours. In this way, the activity of the biomass could be
 231 properly reactivated from starvation to perform simultaneous nitrification and biodegradation of
 232 *o*-cresol. On day-21, the operation in continuous feeding was started (Phase I). DO and
 233 [DO]/[N-NH₄⁺] ratio (Figure 2A) were manipulated to achieve successful nitrite accumulation
 234 (Figure 2C) and biodegradation of *o*-cresol (Figure 2D). NLR and OLR were increased
 235 stepwise until reach 1.1 g N L⁻¹ d⁻¹ and 0.3 g COD L⁻¹ d⁻¹, respectively (Figure 2B). From
 236 day-60, the effluent was mainly composed by nitrite (942 ± 32 mg N L⁻¹) and low
 237 concentrations of ammonium and nitrate, 26 ± 19 mg N L⁻¹ and 10 ± 4 mg N L⁻¹, respectively
 238 (Figure 2C). The *o*-cresol removal was always complete without accumulation of
 239 intermediates (Figure 2D).
 240 On day-114, phenol was added to the influent (Phase II). Phenol concentration was increased
 241 stepwise from 25 ± 1 to 100 ± 3 mg L⁻¹ and *o*-cresol was maintained at 100 mg L⁻¹, reaching
 242 an OLR of 0.5 g COD L⁻¹ d⁻¹. The NLR was maintained at 1.1 g N L⁻¹ d⁻¹. During this phase,
 243 neither *o*-cresol or phenol nor metabolic intermediates (1,2,4-benzenetriol, hydroquinone,
 244 catechol, resorcinol and *p*-nitrocatechol) were accumulated in the reactor (Figure 2D). The
 245 effluent was mainly composed by nitrite (838 ± 43 mg N L⁻¹) and the concentrations of
 246 ammonium and nitrate remained low at 27 ± 18 mg N L⁻¹ and 13 ± 6 mg N L⁻¹, respectively
 247 (Figure 2C). The operation of the granular airlift reactor was maintained during other 60 days
 248 at a NLR and OLR of 1.1 g N L⁻¹ d⁻¹ and 0.5 g COD L⁻¹ d⁻¹, respectively.

249 From day-195, quinoline was added to the influent (Phase III). Quinoline concentration was
 250 increased stepwise from 15 ± 2 to 50 ± 4 mg L⁻¹ and phenol and *o*-cresol were keep at 100 mg
 251 L⁻¹ each one, reaching an OLR of 0.7 g COD L⁻¹ d⁻¹. Complete biodegradation of *o*-cresol,
 252 phenol and quinoline took place and no metabolic intermediates were detected (Figure 2D).
 253 The NLR was kept unchanged at 1.1 g N L⁻¹ d⁻¹. As in the previous phases, the aerobic
 254 granular reactor was able to oxidize ammonium to nitrite (844 ± 50 mg N L⁻¹) instead of
 255 nitrate (4 ± 1 mg N L⁻¹); the ammonium concentration in the effluent was 25 ± 27 mg N L⁻¹.
 256 At the end of this phase, a nitrogen mass balance (see Figure S2 in Supplementary Material)
 257 confirmed that the main biological processes related to ammonium removal were: nitrification
 258 (83.8 % of the incoming ammonium), denitrification (8.0 %) and nitrous oxide emissions (4.1
 259 %). Based on this information, there was virtually not denitrification in the aerobic granular
 260 reactor of this study. However, the simultaneous nitrification-denitrification (SND) process
 261 has been reported in other aerobic granular reactors operated at high loading rates and with
 262 readily biodegradable organic matter sources (as acetate) [15,16]. The low extent of
 263 denitrification in the aerobic granular reactor of this study could be related to: (i) the organic
 264 matter sources (*o*-cresol, phenol and quinoline) are recalcitrant compounds and maybe the
 265 aerobic biodegradation of these compounds is favoured compared to their anoxic
 266 biodegradation, (ii) the average granule size reported in granular reactors with SND (3.5-5.0
 267 mm [15,16]) was significantly higher than that achieved in this study (1.0 mm), allowing the
 268 growth of denitrifying bacteria in the inners layer of the granules and (iii) SND is not
 269 favoured in a continuous feeding regimen as the applied in the airlift reactor of this study.
 270 However, alternating oxic-anoxic conditions and the use of sequencing batch reactors can
 271 favour the SND process [16].
 272 In the last phase, from day 333-onward (Phase IV), the aim was to achieve an effluent suitable
 273 for a subsequent anammox reactor, i.e., an effluent composed by a nitrite/ammonium

concentrations ratio close to 1.0 and without aromatic compounds and metabolic intermediates. The strategy to achieve this new effluent was to maintain the NLR and OLR at 1.1 g N L⁻¹ d⁻¹ and 0.7 g COD L⁻¹ d⁻¹, respectively but decreasing the DO concentration in the reactor (by decreasing the airflow) from 1.0 to 0.5 mg O₂ L⁻¹ to limit AOB activity and force the accumulation of ammonium (Figure 2C). After this change, the nitrite concentration decreased to 499 ± 45 mg N L⁻¹ while the ammonium concentration increased to 419 ± 60 mg N L⁻¹. Therefore, the nitrite/ammonium concentrations ratio in the effluent was steadily maintained between 1.0-1.2 during this phase (phase IV, Figure 2C). Considering that the biomass concentration in the reactor was maintained at 3.8 ± 0.6 g VSS L⁻¹ throughout the operational period, the specific NLR in the reactor was 0.29 g N g⁻¹ VSS d⁻¹. Along the reactor operation, the salinity in the effluent remained stable at 14 ± 1 mS cm⁻¹. The difference in the oxygen affinity between AOB and NOB was determined to be the crucial parameter to obtain nitrification in biofilm or granular reactors [17]. If strong oxygen limiting conditions are applied in a granular reactor, AOB outcompete NOB and nitrite oxidation is prevented. To achieve this, the key operational variable to be maintained is the ratio between DO and ammonium concentrations in the bulk liquid [17]. The lower the [DO]/[N-NH₄⁺] ratio, the stronger the oxygen limiting condition in the biofilm [18]. The control of this operational parameter allowed achieving and maintaining nitrification even at high DO concentration, as previously demonstrated with long-term specific experiments [17]. In this study, the [DO]/[N-NH₄⁺] ratio was not automatically controlled but it was always maintained at very low values (0.05 and 0.75 mg O₂ mg⁻¹ N, Figure 2A). These strong oxygen limiting conditions imposed in the reactor for NOB allowed maintaining partial nitrification throughout the study. Four batch experiments were carried out to determine the effect of the simultaneous presence of ammonium and aromatics on the aerobic granular biomass. The results of these batch tests

are shown in Figure 3. In the first test (Figure 3A), only ammonium was added and only nitrite was produced at a specific nitrite production rate of $0.21 \pm 0.04 \text{ g N g}^{-1} \text{ VSS d}^{-1}$. In the second test (Figure 3B), *o*-cresol and phenol were simultaneously added and consumed at the specific degradation rates of $0.30 \pm 0.01 \text{ g COD g}^{-1} \text{ VSS d}^{-1}$ and $0.35 \pm 0.01 \text{ g COD g}^{-1} \text{ VSS d}^{-1}$, respectively. In the third test, ammonium, *o*-cresol and phenol were added together (Figure 3C). Nitrite was not produced until *o*-cresol and phenol were almost completely degraded. Specific nitrite production and *o*-cresol and phenol biodegradations rates were $0.25 \pm 0.06 \text{ g N g}^{-1} \text{ VSS d}^{-1}$, $0.19 \pm 0.05 \text{ g COD g}^{-1} \text{ VSS d}^{-1}$ and $0.24 \pm 0.04 \text{ g COD g}^{-1} \text{ VSS d}^{-1}$, respectively. Finally, in the fourth test, ammonium and quinoline were added together (Figure 3D). In this case, nitrite was simultaneous produced with the biodegradation of quinoline. Specific nitrite production and quinoline biodegradation rates were $0.44 \pm 0.05 \text{ g N g}^{-1} \text{ VSS d}^{-1}$ and $0.14 \pm 0.03 \text{ g COD g}^{-1} \text{ VSS d}^{-1}$, respectively.

These results show the benefits of using a continuous feeding regime in an aerobic granular reactor treating simultaneously ammonium and aromatic compounds [9]. Conventionally, granular biomass is used in sequencing batch reactors (SBRs) where high substrate concentrations can be achieved at the beginning of the SBR cycle. In that case, nitrification could not start until the aromatic compounds were completely consumed, causing an increase in the duration of the SBR cycle and, consequently, a reduction of the efficiency [19,20]. On the contrary, continuous operation can be a suitable option since the concentration of the aromatic compounds inside the reactor (bulk liquid) is expected to be (always) low due to the high removal efficiency, reducing their toxic/inhibitory effect over nitrification or the oxygen limitation caused by the degradation of the organic compounds, which is in agreement with the data of the kinetic tests and the airlift reactor performance.

Nitrous oxide production were measured at several DO concentrations in the bulk liquid (0.5, 1.0, 1.5 and $4.0 \text{ mg O}_2 \text{ L}^{-1}$) during the period between day-310 and day-330 when the airlift

reactor was treating ammonium, *o*-cresol, phenol and quinoline producing a suitable effluent for denitrification (Figure 4). At 0.5 and 1.0 mg O₂ L⁻¹, the lower the DO, the higher the N₂O emissions were. However, between 1.0 to 4.0 mg O₂ L⁻¹, the N₂O emissions remained stable at 4 % of N-converted. N₂O can be produced as an intermediate product of nitrification, mainly due to low DO concentration in the aerobic reactor [21-23]. The observed trend in the N₂O emissions was similar than the obtained by Pijuan et al. [21] in another granular airlift reactor performing nitrification of a real reject wastewater. They also found a significant increase of the N₂O emissions at low DO concentrations (up to 7 % of N-converted at 1 mg O₂ L⁻¹) and a constant percentage of N₂O emissions (2 % of N-converted) for DO concentrations higher than 4 mg O₂ L⁻¹. In this study, the percentage was constant (4 % of N-converted) from 1 mg O₂ L⁻¹ and the maximum value was 11 % of N-converted at 0.5 mg O₂ L⁻¹. The difference in the percentages of N₂O emissions between both studies could be related to differences on: average granular diameter, pH in the reactor, applied airflow, reactor size, etc. The obtained results indicate that the granular airlift reactor of this study should be operated at DO higher than 1.0 mg O₂ L⁻¹ to minimize the N₂O emissions. In this sense, it should be noted that the application of partial nitrification for subsequent anammox step could be limited by the high N₂O emissions (11 % of N-converted) at low DO (0.5 mg O₂ L⁻¹).

Reactor performance: stability of the aerobic granular biomass

The morphological characteristics of the aerobic granular biomass were evaluated throughout the experimental period. Table 1 shows the average size, density and settling velocity of the granular biomass in each of the four phases of the reactor operation. The average granules size decreased from 2.7 ± 0.8 to 1.0 ± 0.1 mm but the percentage of biomass with a diameter higher than 0.2 mm was always higher than 95%, which means that

349 the biomass of the airlift reactor can be considered as mature and stable aerobic granules.
 350 Moreover, density of the granules increased from 57 to 97 g L⁻¹ but there was not a significant
 351 change of the settling velocity.
 352 Throughout the operational period, there was a decrease of the average diameter and an
 353 increase of the granule density. These facts can be related to the supplied aeration because the
 354 airflow was increased together with the increase of the OLR to maintain the DO concentration
 355 high enough to avoid DO limitation for organic matter biodegradation.
 356 The biomass inside the reactor was maintained at 3.8 ± 0.6 g VSS L⁻¹ by withdrawing
 357 granular sludge periodically and achieving a sludge retention time (SRT) of 107 ± 46 days.
 358 The granules had a low VSS/TSS ratio (0.31 ± 0.04), which could be related to the high
 359 salinity (16 g salts L⁻¹) and the high SRT in the reactor. Throughout the reactor operation,
 360 SVI₅ and SVI₃₀ remained at the same low value (12 ± 2 mL g⁻¹ TSS). This value is below 50
 361 mL g⁻¹ TSS, which is considered as a typical value for aerobic granules [9,24,25]. Therefore,
 362 the value of the SVI₃₀/SVI₅ ratio was always one. Moreover, throughout the experimental
 363 period, VSS and TSS concentrations in the effluent were 79 ± 31 mg VSS L⁻¹ and 61 ± 26 mg
 364 TSS L⁻¹, respectively. These values indicate that a good separation of the biomass was
 365 achieved in the reactor since the biomass concentration in the reactor was 3.8 g VSS L⁻¹.
 366 Pictures of the granules were made to study their morphological characteristics. In phase I,
 367 some granules presented a smooth and regular shape and other granules showed an outer
 368 surface covered with filaments, possibly due to the presence of filamentous microorganisms
 369 (Figures 5A and 5B). In subsequent phases, when the OLR was increased due to the
 370 introduction of phenol and quinoline in the influent, almost all the granules were covered with
 371 filamentous microorganisms (see Figures 5C-5H and also Figure S3 in Supplementary
 372 Material). Commonly, the growth of bacteria with filamentous morphology is related to
 373 problems with the settling properties of the sludge [26]. However, in spite of the growth of

374 filamentous bacteria over the surface, the aerobic granules maintained a very good
 375 settleability throughout the study.

376 Another important fact is the stability of the granular biomass at high salinity conditions. The
 377 main salts in the influent were chloride and bicarbonate, giving a salinity of 16 g salts L⁻¹. As
 378 explained before, the aerobic granules presented good settleability at long-term and
 379 consequently, they can be considered also resistant to a high content of salts.

380

381 Bacterial populations in the aerobic granular biomass

382

383 The bacterial communities involved in the simultaneous nitrification and biodegradation of *o*-
 384 cresol, phenol and quinoline at high salinity were analysed by pyrosequencing on day-300
 385 (Figure 6). The most abundant groups at class level were: Betaproteobacteria (80%),
 386 Gammaproteobacteria (10%), Alphaproteobacteria (5%) and Flavobacteria (3%); which
 387 represented the 98% of the analysed sequences. *Comamonas* (65%), *Nitrosomonas* (11%),
 388 *Pseudomonas* (5%), *Stenotrophomonas* (4%), *Bradyrhizobium* (3%) and *Burkholderia* (3%)
 389 were the most abundant genera (Figure 6); representing 91% of the analysed sequences.

390 *Comamonas*, *Nitrosomonas* and *Burkholderia* genera belong to Betaproteobacteria whereas
 391 *Pseudomonas* and *Stenotrophomonas* genera belong to Gammaproteobacteria and
 392 *Bradyrhizobium* genus belong to Alphaproteobacteria.

393 Moreover, on day-300 FISH analyses for AOB and NOB were performed. Positive
 394 hybridization was obtained for β -AOB and the FISH quantification revealed that β -AOB was
 395 71 ± 21 % of the total bacterial population (see Figure S4 in Supplementary Material).
 396 However, NOB was not detected by FISH.

397 Regarding the biodegradation of aromatic compounds, *Comamonas*, *Pseudomonas*,
 398 *Stenotrophomonas*, *Burkholderia* and *Bradyrhizobium* were the main heterotrophic genera

detected and they have been described to be able to biodegrade several aromatic compounds, such as phenol, quinoline and cresols under aerobic conditions [27-30]. Therefore, the results of the pyrosequencing are consistent with the reactor performance and the main identified bacteria can be considered as the responsible microorganisms for biodegradation of phenol, *o*-cresol and quinoline.

It is also remarkable that the main abundant genus in the aerobic granules (*Comamonas*) is able to grow as filamentous bacteria under specific conditions [31,32]. This fact could explain the significant growth of filamentous bacteria over the surface of the aerobic granules (Figure 5). Probably, these filaments were mainly composed by *Comamonas*.

Regarding the oxidation of ammonium, only an AOB genus was detected by pyrosequencing in the aerobic granules (*Nitrosomonas*) while no NOB genus was detected by pyrosequencing or by FISH. This result confirms that the applied $[DO]/[N-NH_4^+]$ ratio (below 0.75 mg O₂ mg⁻¹ N) is a proper condition to achieve the wash-out of NOB and to accumulate nitrite (nitrification) instead of nitrate (conventional nitrification), even with the simultaneous biodegradation of aromatic compounds.

However, the percentage *Nitrosomonas* measured by pyrosequencing (11%) seems to be low to explain the high nitrification capacity of the aerobic granules. Two facts could explain this result: (i) on one hand, the pyrosequencing relates the presence of bacteria and the FISH reveals not only the presence but also the activity of bacteria [33]. In previous studies, it has been described for AOB communities that the most abundant species may not necessarily be the most active ones [33]. The same fact could have happened in this study where *Nitrosomonas* was much less abundant than *Comamonas* (Figure 6). Nevertheless, *Nitrosomonas* could be the most active bacteria, as the FISH analysis revealed; (ii) on the other hand, nitrification is usually attributed to autotrophic bacteria (AOB and NOB). However, some heterotrophic bacteria can also perform heterotrophic nitrification.

Heterotrophic nitrification is still poorly understood and it is usually described as the oxidation of ammonium until nitrate by heterotrophic bacteria [34]. *Comamonas* and *Pseudomonas* have been described as heterotrophic nitrifiers genera [34,35]. However, nitrate was hardly produced in this study while high nitrite accumulation took place (Figure 2C). This means that, if *Comamonas* and *Pseudomonas* played a significant role in the oxidation of ammonium in this study, the heterotrophic nitrification was stopped in nitrite instead of nitrate. *Alcaligenes faecalis* has been identified as a heterotrophic bacterium with the capacity to biodegrade phenol and to oxidize ammonium until nitrite [36], i.e. as a species able to perform heterotrophic nitrification. In the same way, *Comamonas* and *Pseudomonas* genera could also perform heterotrophic nitrification in the conditions applied in this study. According to the above exposed results, the successful operation of the granular airlift reactor for simultaneous nitrification and biodegradation of aromatic compounds can be related to the: (i) growth of specialized microorganisms in the aerobic granules and (ii) the continuous feeding regime.

Conclusions

Simultaneous nitrification and biodegradation of a mixture of aromatic compounds (*o*-cresol, phenol and quinoline) at high salinity were successfully achieved in a continuous airlift granular reactor.

A suitable effluent for denitrification or for anammox was attained by manipulating the DO concentration and the DO/N-NH₄⁺ concentrations ratio in the bulk liquid.

DO concentration had a significant effect on the N₂O emissions and DO higher than 1.0 mg O₂ L⁻¹ should be maintained to avoid high N₂O emissions.

Comamonas and *Pseudomonas* were the main heterotrophic genera detected and they were probably the responsible of biodegradation of aromatic compounds. *Nitrosomonas* genus was the only AOB detected in the aerobic granules while NOB were not detected.

Acknowledgements

This work was supported by the Ministerio de Economía y Competitividad (Spanish Government) through the ONLYBIO project (CTQ2011-24745/PPQ). The authors are members of the GENOCOV group (2009 SGR 815, www.genocov.com). Carlos Ramos wants to thank the Universitat Autònoma de Barcelona for his pre-doctoral fellowship.

References

- [1] Milia S, Cappai G, Perra M, Carucci A. Biological treatment of nitrogen-rich refinery wastewater by partial nitrification (SHARON) process. *Environ Technol* 2012;33:1477-1483.
- [2] Kim S.-S., Kim H.-J. Impact and threshold concentration of toxic materials in the stripped gas liquor on nitrification. *Korean J. Chem. Eng.* 2003; 20: 1103-1110.
- [3] Kim KH, Ihm SK. Heterogeneous catalytic wet air oxidation of refractory organic pollutants in industrial wastewaters: A review. *J Hazard Mater* 2011;186:16-34.
- [4] Al-Khalid T, El-Naas MH. Aerobic Biodegradation of Phenols: A Comprehensive Review. *Crit Rev Env Sci Tec* 2011;42:1631-1690.
- [5] Kim YM, Park D, Lee DS, Park JM. Instability of biological nitrogen removal in a cokes wastewater treatment facility during summer. *J Hazard Mater* 2007;141:27-32.

- 471 [6] Gao D, Liu L, Liang H, Wu WM. Aerobic granular sludge: characterization,
472 mechanism of granulation and application to wastewater treatment. Crit Rev
473 Biotechnol 2011;31:137-152.
- 474 [7] Schmidt I, Sliekers O, Schmid M, Bock E, Fuerst J, Kuenen JG, Jetten MSM, Strous
475 M. New concepts of microbial treatment processes for the nitrogen removal in
476 wastewater. FEMS Microbiol Rev 2003;27:481-492.
- 477 [8] Paredes D, Kusch P, Mbvette TSA, Stange F, Müller RA, Köser H. New aspects of
478 microbial nitrogen transformations in the context of wastewater treatment - A review.
479 Eng Life Sci 2007;7:13-25.
- 480 [9] Jemaat Z, Suárez-Ojeda ME, Pérez J, Carrera J. Partial nitrification and *o*-cresol removal
481 with aerobic granular biomass in a continuous airlift reactor. Water Res 2014;48:354-
482 362.
- 483 [10] Jemaat Z, Suárez-Ojeda ME, Pérez J, Carrera J. Simultaneous nitrification and *p*-
484 nitrophenol removal using aerobic granular biomass in a continuous airlift reactor.
485 Bioresour Technol 2013;150:307-313.
- 486 [11] APHA. Standard Methods for the Examination of Water and Wastewater. 20th edn.
487 Washington: American Water Works Association & Water Environmental Federation;
488 1999.
- 489 [12] Beun JJ, van Loosdrecht MCM, Heijnen JJ. Aerobic granulation in a sequencing batch
490 airlift reactor. Water Res 2002;36:702-712.
- 491 [13] Julián E, Roldán M, Sánchez-Chardi A, Astola O, Agustí G, Luquin M. Microscopic
492 cords, a virulence-related characteristic of *Mycobacterium tuberculosis*, are also
493 present in nonpathogenic mycobacteria. J Bacteriol 2010;192:1751-1760.
- 494 [14] Suárez-Ojeda ME, Montón H, Roldán M, Martín-Hernández M, Pérez J, Carrera J.
495 Characterization of a *p*-nitrophenol-degrading mixed culture with an improved

methodology of fluorescence in situ hybridization and confocal laser scanning
microscopy. *J Chem Technol Biot* 2011;86:1405-1412.

[15] Adav SS, Lee D-J, Lai JY. Microbial community of acetate utilizing denitrifiers in
aerobic granules. *Appl. Microbiol. Biotechnol* 2010;85:753–762.

[16] Adav SS, Lee D-J, Lai J-Y. Biological nitrification–denitrification with alternating
oxic and anoxic operations using aerobic granules. *Appl. Microbiol. Biotechnol*
2009;84:1181–1189.

[17] Bartrolí A, Pérez J, Carrera J. Applying ratio control in a continuous granular reactor
to achieve full nitrification under stable operating conditions. *Environ Sci Technol*
2010;44:8930-8935.

[18] Jemaat Z, Bartrolí A, Isanta E, Carrera J, Suárez-Ojeda ME, Pérez J. Closed-loop
control of ammonium concentration in nitrification: Convenient for reactor operation but
also for modeling. *Bioresour Technol* 2013;128:655-663.

[19] Liu Y, Tay JH, Ivanov V, Moy BYP, Yu L, Tay STL. Influence of phenol on
nitrification by microbial granules. *Process Biochem* 2005;40:3285-3289.

[20] Morita M, Kudo N, Uemoto H, Watanabe A, Shinozaki H. Protective Effect of
Immobilized Ammonia Oxidizers and Phenol-degrading Bacteria on Nitrification in
Ammonia– and Phenol-containing Wastewater. *Eng Life Sci* 2007;7:587-592.

[21] Desloover J, Vlaeminck SE, Clauwaert P, Verstraete W, Boon N. Strategies to
mitigate N₂O emissions from biological nitrogen removal systems. *Curr Opin Biotech*
2012; 23:474-48.

[22] Pijuan M, Torà J, Rodríguez-Caballero A, César E, Carrera J, Pérez J. Effect of
process parameters and operational mode on nitrous oxide emissions from a nitrification
reactor treating reject wastewater. *Water Res* 2014;49:23-33.

- 520 [23] Schneider Y, Beier M, Rosenwinkel KH. Influence of operating conditions on nitrous
521 oxide formation during nitrification and nitrification. *Environ Sci Pollut Res*
522 2014;21:12099-12108.
- 523 [24] Li Y, Liu Y, Xu H. Is sludge retention time a decisive factor for aerobic granulation in
524 SBR?. *Bioresour Technol* 2008;99:7672-7677.
- 525 [25] Liu Y, Tay JH. State of the art of biogranulation technology for wastewater treatment.
526 *Biotechnol Adv* 2004;22:533-563.
- 527 [26] Martins AMP, Pagilla K, Heijnen JJ, van Loosdrecht MCM. Filamentous bulking
528 sludge-a critical review. *Water Res* 2004;38:793–817.
- 529 [27] Cui M, Chen F, Fu J, Sheng G, Sun G. Microbial metabolism of quinoline by
530 *Comamonas* sp. *World J Microb Biot* 2004;20:539-543.
- 531 [28] Ho KL, Chen YY, Lee DJ. Functional consortia for cresol-degrading activated
532 sludges: Toxicity-to-extinction approach. *Bioresour Technol* 2010;101:9000-9005.
- 533 [29] Verma V, Raju SC, Kapley A, Kalia VC, Kanade GS, Dagainawala HF, Purohit HJ.
534 Degradative potential of *Stenotrophomonas* strain HPC383 having genes homologous
535 to dmp operon. *Bioresour Technol* 2011;102:3227-3233.
- 536 [30] Wang Q, Li Y, Li J, Wang Y, Wang C, Wang P. Experimental and kinetic study on the
537 cometabolic biodegradation of phenol and 4-chlorophenol by psychrotrophic
538 *Pseudomonas putida* LY1. *Environ Sci Pollut Res* 2015;22:565-573.
- 539 [31] Gumaelius L, Magnusson G, Pettersson B, Dalhammar G. *Comamonas denitrificans*
540 sp. nov., an efficient denitrifying bacterium isolated from activated sludge. *Int J Syst*
541 *Evol Micr* 2001;51:999-1006.
- 542 [32] Hahn MW, Moore ERB, Höfle MG. Bacterial Filament Formation, a Defense
543 Mechanism against Flagellate Grazing, Is Growth Rate Controlled in Bacteria of
544 Different Phyla. *Appl Environ Microbiol* 1999;65:25-35.

- 545 [33] Fitzgerald CM, Camejo P, Oshlag JZ, Noguera DR. Ammonia-oxidizing microbial
546 communities in reactors with efficient nitrification at low-dissolved oxygen. *Water*
547 *Res* 2015;70:38-51.
- 548 [34] Chen Q, Ni J. Heterotrophic nitrification–aerobic denitrification by novel isolated
549 bacteria. *J Ind Microbiol Biotechnol* 2011;38:1305-1310.
- 550 [35] Liu Y, Li Y, Lv Y. Isolation and characterization of a heterotrophic nitrifier from coke
551 plant wastewater. *Water Sci Technol* 2012;65(11):2084-2090.
- 552 [36] Wittebolle L, Boon N, Vanparys B, Heylen K, De Vos P, Verstraete W. Failure of the
553 ammonia oxidation process in two pharmaceutical wastewater treatment plants is
554 linked to shifts in the bacterial communities. *J Appl Microbiol* 2005;99:997-1006.
555

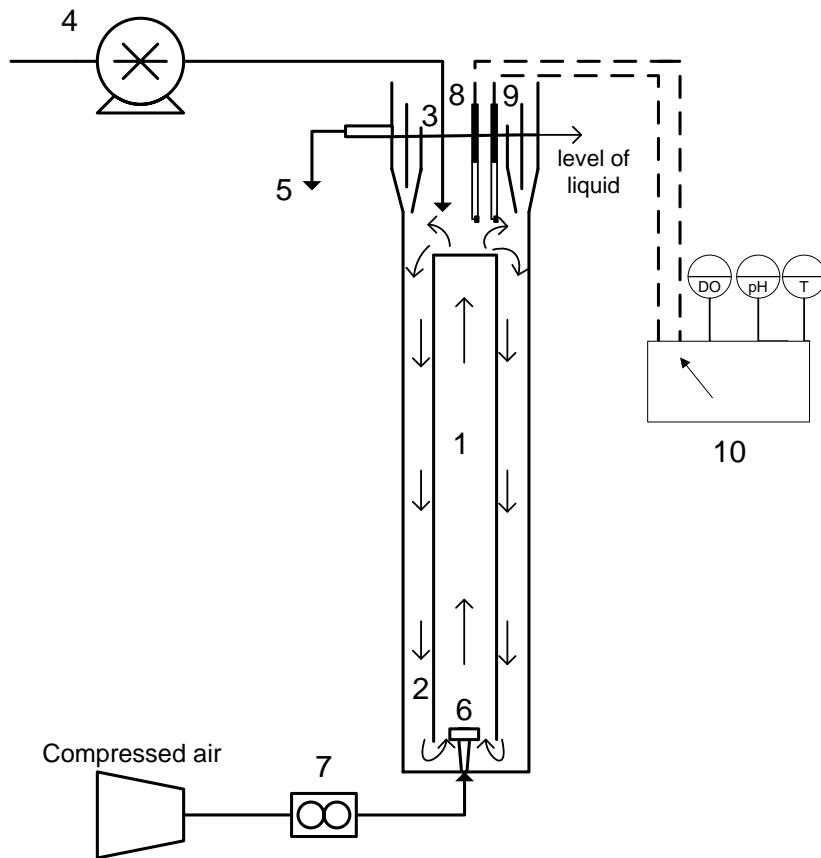


Figure 1. Experimental set-up of the continuous granular airlift reactor. (1) riser; (2) down comer; (3) separator; (4) feed pump; (5) effluent port; (6) air sparger; (7) rotameter; (8) pH probe; (9) DO probe; (10) monitoring panel.

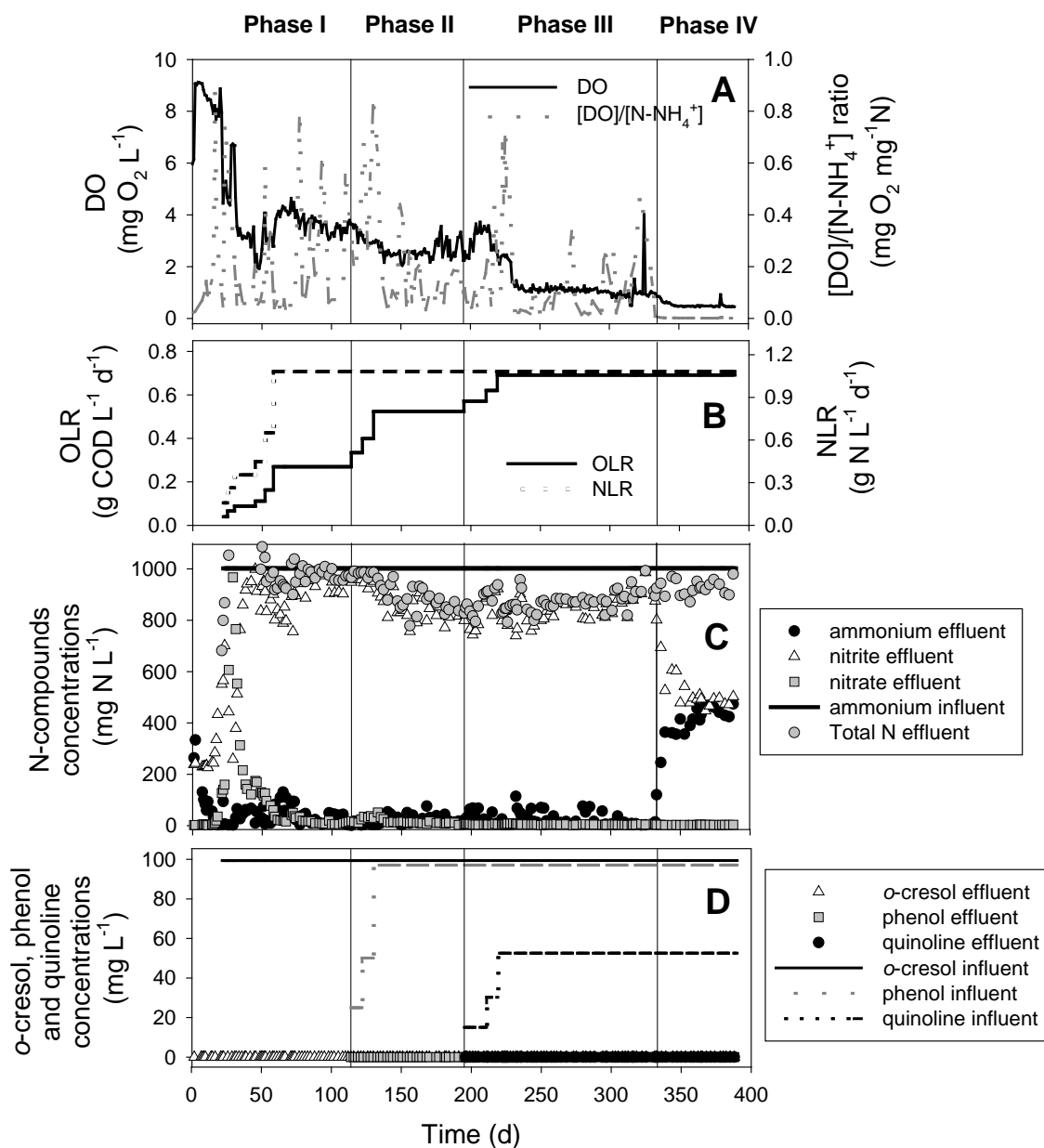


Figure 2. Operational conditions applied and performance of the granular airlift reactor treating a wastewater composed by high concentrations of ammonium and aromatic compounds at high salinity (16 g salts L⁻¹). DO: dissolved oxygen concentration, NLR: nitrogen loading rate and OLR: organic loading rate. Phase I: simultaneous nitrification and biodegradation of *o*-cresol, phase II: simultaneous nitrification and biodegradation of *o*-cresol and phenol, phase III: simultaneous nitrification and biodegradation of *o*-cresol, phenol and quinoline and phase IV: simultaneous partial nitrification and biodegradation of *o*-cresol, phenol and quinoline.

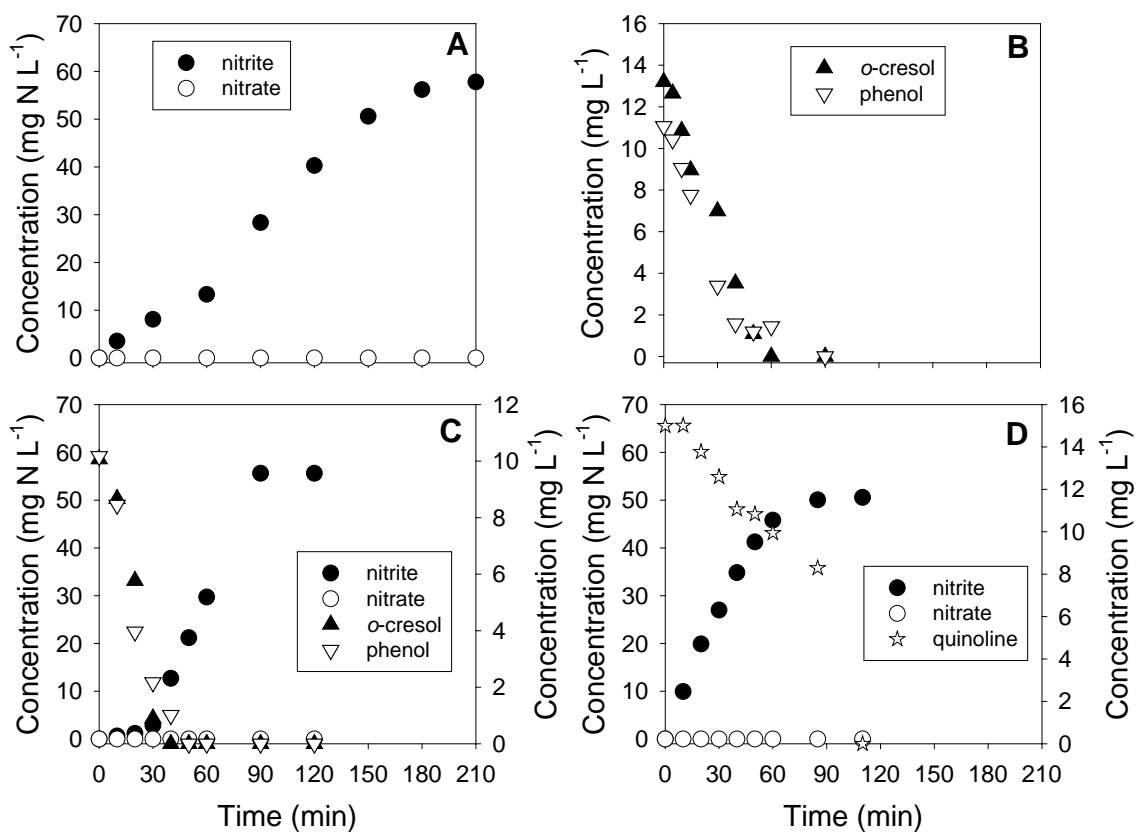


Figure 3. Kinetic study of the granular biomass performing nitrification and biodegradation of aromatic compound. A: ammonium (60 mg N L⁻¹); B: ammonium (60 mg N L⁻¹) plus *o*-cresol (13 mg L⁻¹) and phenol (11 mg L⁻¹); C: ammonium (60 mg N L⁻¹) plus *o*-cresol (10 mg L⁻¹) and phenol (10 mg L⁻¹); D: ammonium (60 mg N L⁻¹) plus quinoline (15 mg L⁻¹).

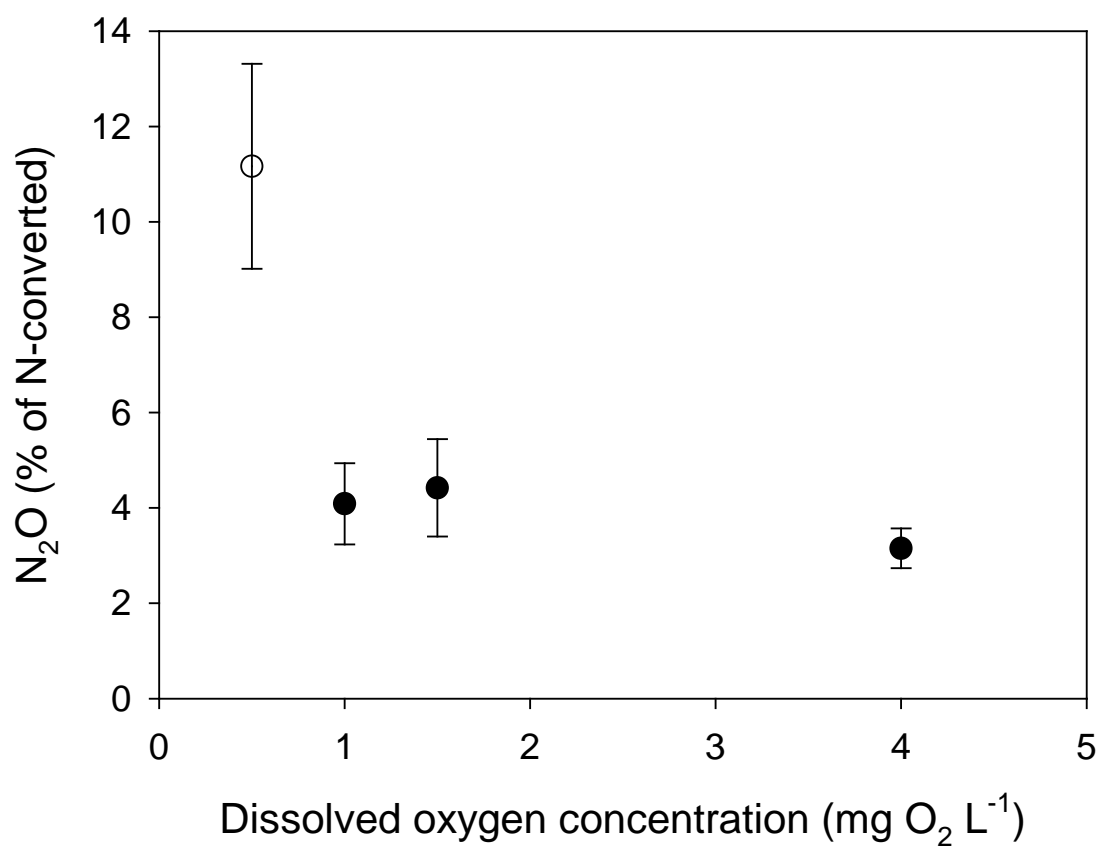


Figure 4. Nitrous oxide production under different dissolved oxygen concentrations in the granular airlift reactor treating ammonium, *o*-cresol, phenol and quinoline at high salinity (16 g salts L⁻¹).

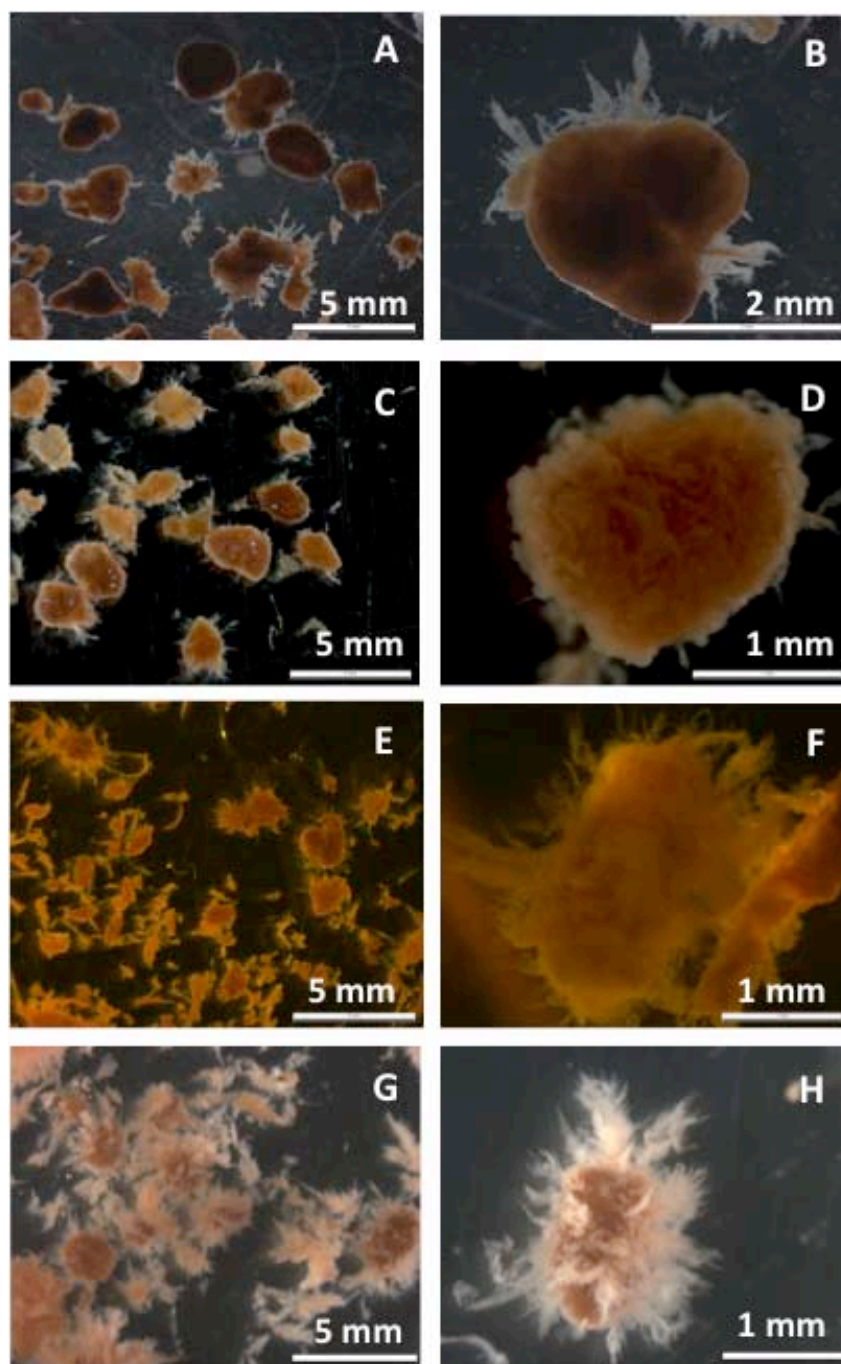


Figure 5. Pictures of the granules in the airlift reactor treating a wastewater composed by high ammonium and aromatic compounds concentrations. A and B treating ammonium and *o*-cresol with 97 % nitrite in the effluent; C and D treating ammonium, *o*-cresol and phenol with 97 % nitrite in the effluent; E and F treating ammonium, *o*-cresol, phenol and quinoline with 97 % nitrite in the effluent and G and H treating ammonium, *o*-cresol, phenol and quinoline with 54 % nitrite in the effluent.

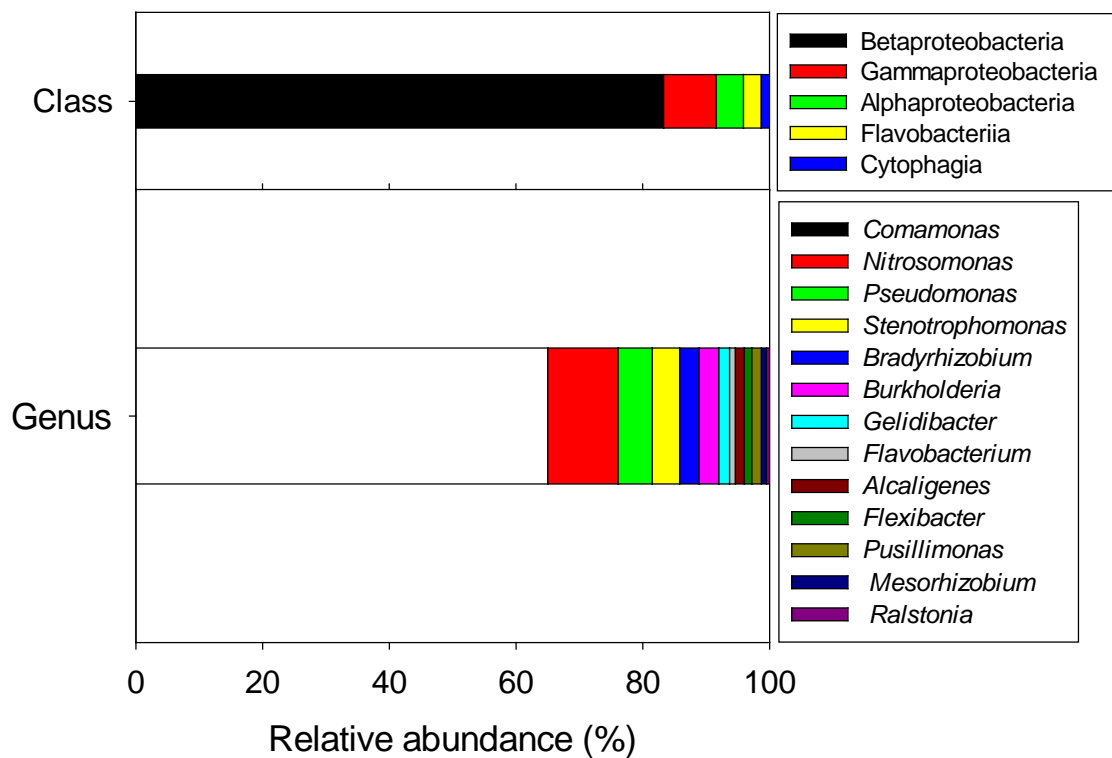


Figure 6. Bacterial distribution at genus and class level in the aerobic granules in the airlift reactor at day-300. The percentages are referred to the relative abundance of bacteria defined as the number of sequences affiliated with that taxon divided by the total number of sequences of the sample.

Table 1. Morphological characterization of the granular biomass along the operation of the airlift reactor performing nitrification and biodegradation of aromatic compounds. The nitrogen loading rate was always $1.1 \text{ g N L}^{-1} \text{ d}^{-1}$.

Phase	Organic compounds in the influent	OLR ($\text{g COD L}^{-1} \text{ d}^{-1}$)	Nitrite effluent (%)	Settling velocity (m h^{-1})	Average granule size (mm)	Granule density ($\text{g VSS L}_{\text{particle}}^{-1}$)
I	<i>o</i> -cresol	0.3	97	57 ± 19	2.7 ± 0.8	57 ± 0
II	<i>o</i> -cresol + phenol	0.5	97	50 ± 23	2.4 ± 0.6	73 ± 4
III	<i>o</i> -cresol + phenol	0.7	97	35 ± 9	1.5 ± 0.4	95 ± 3
IV	+ quinoline	0.7	55	52 ± 17	1.0 ± 0.1	97 ± 3