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Tolcapone, a potent aggregation inhibitor for the treatment of familial leptomeningeal amyloidosis

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ABBREVIATIONS

TTR, transthyretin; thyroxine, T₄; FAP, familial amyloid polyneuropathy; FAC, familial amyloid cardiomyopathy; ITC, isothermal titration calorimetry; K_d, dissociation constant; nH, Hill coefficient; HBP, halogen binding pocket.

DATABASES

PDB codes for A25T-TTR, V30G-TTR, and Y114C-TTR bound to tolcapone are 6TXV, 6TXW and 6XTK, respectively.

KEYWORDS

transthyretin; amyloidosis; protein aggregation; crystal structures; tolcapone

ABSTRACT

Heredity transthyretin amyloidosis (ATTR) are diseases characterized by the extracellular deposition of transthyretin (TTR) amyloid fibrils. Highly destabilizing TTR mutations cause leptomeningeal amyloidosis, a rare, but fatal, disorder in which TTR aggregates in the brain. The disease remains intractable, since liver transplantation, the reference therapy for systemic ATTR, does not stop mutant TTR production in the brain. In addition, despite current pharmacological strategies have shown to be effective against *in vivo* TTR aggregation by stabilizing the tetramer native structure and precluding its dissociation, they display low brain permeability. Recently, we have repurposed tolcapone as a molecule to treat systemic ATTR. Crystal structures and biophysical analysis converge to demonstrate that tolcapone binds with high affinity and specificity to three unstable leptomeningeal TTR variants, stabilizing them and, consequently inhibiting their aggregation. Because tolcapone is an FDA-approved drug that crosses the blood-brain-barrier, our results suggest that it can translate into a first disease-modifying therapy for leptomeningeal amyloidosis.

INTRODUCTION

The misfolding and subsequent aggregation of proteins into amyloid deposits is associated with the onset of a variety of debilitating human disorders, affecting millions of people worldwide [1]. In transthyretin (TTR) amyloidosis, secreted soluble TTR undergoes a conformational change to generate toxic extracellular deposits that lead to disease. Plasma circulating TTR is synthesised by the liver. The protein can also be found in the cerebrospinal fluid (CSF) and in the eye, produced by the brain's choroid plexus and the retinal pigment epithelial cells [2, 3], respectively. In the blood, TTR is one of the proteins responsible for the transport and delivery of thyroxine (T₄) and retinol to cells, whereas, in the CSF, TTR is the primary T₄ transporter [2, 4].

Amyloidogenesis of wild-type TTR (WT-TTR) is associated with senile systemic amyloidosis, a cardiomyopathy affecting up to 25% of the population over age 80 [5, 6]. Hereditary transthyretin amyloidosis comprises a life-threatening group of rare monogenic diseases inherited in an autosomal dominant fashion, with most affected individuals being heterozygous for one of the approximately 140 disease-associated TTR mutations [7]. Familial amyloid polyneuropathy (FAP) [8, 9] and the familial amyloid cardiomyopathy (FAC) [10] are the most common forms of hereditary amyloidosis. A reduced number of TTR mutations lead to leptomeningeal amyloidosis, a less well characterized and rare phenotype restricted to the central nervous system (CNS) and defined by preferential amyloid deposition in the leptomeningeal vessels, the brain parenchyma and in the eyes. The predominant CNS symptoms include stroke, recurrent subarachnoid hemorrhage, dementia, ataxia, seizures, depression and periods of decreased consciousness, usually with a poor-prognosis [11, 12].

Human TTR is a 55 kDa homotetrameric protein consisting of four β -sheet rich subunits of 127 amino acid residues each, usually labelled from A to D. The AB/CD dimer-dimer interface creates two identical funnel-shaped T₄-binding sites located in opposite sides of the molecule [13, 14]. Scission along this interface is the most probable mechanism of TTR tetramer dissociation,

as it is the weakest of the two interfaces in the tetramer [15] and constitutes the rate-limiting step in TTR aggregation. After tetramer dissociation, the dimers readily convert into monomers, which experiment tertiary structural changes to become aggregation competent units [15, 16]. Disease-associated mutations accelerate TTR amyloidosis by thermodynamic or kinetic destabilization of the protein [17, 18].

By removing the main source of systemic mutant TTR, liver transplantation has become the reference therapy for hereditary TTR amyloidosis [19, 20]. However, these patients continue to produce mutant TTR in the choroid plexus and in the eye. Accordingly, CNS TTR deposition has been reported in more than 500 FAP patients after liver transplantation [21, 22].

The pursuit of non-invasive therapies for TTR amyloidosis has fueled the development of small molecules that prevent tetramer dissociation by binding to the two T₄ pockets, kinetically stabilizing the TTR native state; thus, stalling protein aggregation [23, 24]. Only one of these molecules reached the market for the treatment of TTR amyloidosis, the benzoxazole tafamidis [25].

Tafamidis has been shown to slow down the progression of polyneuropathy [26] and cardiomyopathy [27]. Accordingly, it has been approved in Europe and Japan for the treatment of early-stage FAP [28] and very recently by the Food and Drug Administration (FDA) for the treatment of TTR cardiomyopathy [27]. In contrast, to date, there are no concluding studies on the molecule's ability to halt disease progression in CNS amyloidosis patients. One study suggests that it can attain moderate levels in CSF and the vitreous body [29], whereas others indicate that its blood-brain-barrier (BBB) permeability is too low to reach therapeutic concentrations able to abrogate the aggregation of mutant TTR in the CNS [30, 31].

Using a drug repurposing approach, our group identified tolcapone as a molecule that binds with high affinity and specificity to the two T₄-binding sites of TTR, promoting its stabilization and preventing the formation of amyloid fibrils and cytotoxic oligomeric species [32]. Tolcapone is an FDA-approved molecule for Parkinson's disease that has already passed Phase IIa clinical trials for FAP (ClinicalTrials.gov Identifier: NCT02191826) [33, 34]. In this study, tolcapone induced a clear and robust stabilization of plasmatic TTR in all patients studied, the drug was well tolerated, and treatment raised no safety concerns. Of note, tolcapone penetrates the BBB [35] and preliminary data indicated that it might inhibit the aggregation of highly-destabilized variants [32]. Together, these evidences suggested that tolcapone might find therapeutic application in the rare, but fatal, CNS amyloidosis, which, as described, cannot be treated efficiently by liver transplantation or current pharmacological approaches.

In this work, we have characterized in detail the activity of tolcapone on three TTR variants linked to familial leptomeningeal amyloidosis, A25T-TTR [36], V30G-TTR [37] and Y114C-TTR [38], which are among the most destabilized and fastest dissociating TTR tetramers. Our results indicate that tolcapone binds with high affinity to the T₄-binding sites of these variants, establishing specific contacts with the tetramer interface that stabilize the mutant proteins and, consequently, reduce their aggregation. Overall, these findings converge to demonstrate that tolcapone might become a first broad-spectrum drug to treat TTR-related amyloidosis.

RESULTS AND DISCUSSION

Tolcapone inhibits the aggregation of TTR leptomeningeal amyloidosis-associated variants

As a first step to assess if tolcapone can be considered a candidate for treating familial leptomeningeal amyloidosis, we evaluated its ability to prevent the aggregation of the TTR CNS amyloidosis-associated variants A25T-TTR, V30G-TTR and Y114C-TTR (Fig. 1). TTR solutions were incubated in the absence or presence of tolcapone and aggregation was induced by acidification. The molecule final concentration ranged between 0 and 50 μ M. Orthogonal light scattering at 340 nm was recorded to follow aggregation and normalized to the maximum scattering value in the absence of tolcapone for each individual variant. Interestingly, as shown in Fig. 1, tolcapone inhibits up to 71% the aggregation of A25T-TTR, 76% of V30G-TTR and reaches 100% for Y114C-TTR; the potency for this last mutant being very close to that exerted for WT-TTR. Remarkably, at 5 μ M of tolcapone, which corresponds to less than 2-fold the protein concentration (3.5 μ M), the inhibition was already evident for all the TTR variants. Ten molar equivalents of tolcapone (35 μ M) reduced around 70% of A25T-TTR aggregation, being equivalent to the anti-aggregational activity of T₄ at the same concentration and solution conditions [12].

Tolcapone stabilizes TTR leptomeningeal amyloidosis-associated variants under denaturing conditions

After demonstrating that tolcapone can inhibit the aggregation of TTR variants linked to familial leptomeningeal amyloidosis, we addressed whether this effect depends on native tetramer stabilization, as previously reported for WT-TTR and the FAC-associated V122I-TTR variant [32].

It is well established that in urea TTR monomer unfolding is preceded by tetramer destabilization [39] and once the tetramer is dissociated, tertiary structural changes can be monitored by tryptophan (Trp) intrinsic fluorescence. Accordingly, to evaluate the effect of tolcapone on the stability of our variants, the proteins were incubated with increasing concentrations of urea (0-9.5 M) for 96 h, allowing the solutions to reach the equilibrium [17, 40], both in the absence and presence of tolcapone (20 and 50 μ M). Trp fluorescence was monitored and used to calculate the fraction of unfolded protein at any given urea concentration (Fig. 2). For A25T-TTR, the obtained curves in both conditions are similar, indicating that, surprisingly, tolcapone does not seem to exert a significant stabilizing effect over this variant in the assayed conditions. In stark contrast, for V30G-TTR and Y114C-TTR a clear stabilizing effect is observed in the presence of tolcapone, being specially dramatic in the case of Y114C-TTR, where tolcapone permits > 50% of the protein to remain folded at 9.5 M urea, a stabilization that approaches that exerted on WT-TTR.

The low stabilization observed for A25T-TTR might indicate that in this brain aggregating variant, at the assay protein concentration (1.8 μ M), the tetramer dissociates at such low urea concentrations that the obtained curves report mostly on monomer unfolding. This is consistent with previous findings, indicating that in the 0.72-7.2 μ M protein concentration range the

denaturation curves show a single transition that is concentration independent [18]. This suggests that, at low protein concentrations, A25T-TTR tetramer dissociation and monomer unfolding are not significantly linked. The behavior of A25T-TTR is in stark contrast with that of WT-TTR, for which it is described that tetramer dissociation and monomer unfolding are thermodynamically linked at the protein concentration used in these experiments [18]. Further evidence for these different unfolding mechanisms is provided by the observation that, for A25T-TTR, the urea denaturation curve assessed by resveratrol binding differs significantly from the one monitored by Trp intrinsic fluorescence, whereas for WT-TTR the two curves are nearly coincident (Fig. S1) [12, 18]. Resveratrol has been described to display a substantial increase in its fluorescence quantum yield when bound to at least one of the two T₄-binding sites in the tetramer, being thus a reporter for the native state TTR conformation [17, 18].

V30G-TTR and Y114C-TTR are clearly more stable and the data herein indicates that they would behave like WT-TTR. Indeed, for these two mutants, the denaturation curves in the presence of tolcapone resemble the ones described for molecules that impose kinetic stability on TTR, stabilizing the AB/CD dimer-dimer interface and delaying tetramer dissociation [41, 42].

The analysis emphasizes the complexity of the denaturation mechanism of TTR leptomeningeal-type variants, for which, in contrast to what occurred for WT-TTR and V122I-TTR [32], a direct relationship between tolcapone stabilizing and anti-aggregational effects could not be established. A possible explanation for this lack of correlation would be that the CNS amyloidosis-associated variants are so destabilized that, even in the absence of denaturant, only a fraction of the protein remains in a native tetrameric form in the assay conditions. We evaluated the initial tetramer population for each of the TTR mutants under native conditions using resveratrol binding assays (Fig. 3). The proportion of tetramer under native conditions differed among variants, as calculated from resveratrol binding experiments. A25T-TTR, V30G-TTR and Y114C-TTR displayed 43.7±3.6%, 59.5±5.0% and 96.2±10.2% of tetramer relative to that in WT-TTR, respectively. The calculated tetramer populations fairly correlated with the aggregation inhibitory effect that tolcapone exerts in these mutants.

The influence of the mutations on the quaternary structural stability of the different TTR mutants under native conditions was also assessed by analytical size exclusion chromatography (SEC) at 7 μ M in phosphate buffered saline (PBS) and monitored by absorbance at 280 nm (Fig. 4). This is the lowest TTR concentration at which our chromatography set up has enough detection sensitivity. V30G-TTR and Y114C-TTR appear as a unique species with an elution volume corresponding to that of the WT-TTR tetramer. In contrast, in agreement with a lower tetramer stability, A25T-TTR seems to adopt both higher and lower molecular weight species other than the tetramer, likely corresponding to soluble aggregates and a monomer, respectively. This suggests that A25T-TTR dissociates spontaneously at this concentration, generating unfolded monomers that assemble into the majoritarian high-weight species. If this is the case, the equilibrium between the tetrameric and monomeric forms of the mutant protein should be concentration dependent.

Effectively, when analytical SEC was performed at a higher protein concentration (20 μ M), the elution profile was identical for all the variants, presenting a single peak with an elution volume compatible with the TTR tetramer. This suggests that the amount of initial tolcapone targetable tetramer depends on both the stability of the protein and its concentration in the assay, since both factors impact the equilibrium between the tetrameric and monomeric forms of TTR. It is possible that for highly unstable variants, tolcapone not only would stabilize the available tetramer, but might also displace partially the equilibrium towards the population of the tetrameric form, as a result of Le Chatelier's principle.

Tolcapone binds with high affinity to TTR leptomeningeal amyloidosis-associated variants

The above described data suggests that tolcapone can inhibit the aggregation of even highly destabilized mutants, as long as they remain in the tetrameric state and the molecule can bind to them. We used isothermal titration calorimetry (ITC) to confirm that tolcapone effectively interacts with the brain-associated variants and to determine the mechanisms of binding. The thermodynamic parameters (ΔG , ΔH , $-\Delta S$) for the binding reactions as well as the cooperativity of the interaction were analyzed for the three mutants and WT-TTR according to two models: (1) two identical, but independent binding sites; (2) two identical and cooperative binding sites [43, 44]. Our data indicated that tolcapone binds with high affinity to the first TTR T₄-binding site of all proteins, but they exhibit differences in the degree of cooperativity for the binding to the second T₄ pocket (Table 1 and Fig. 5).

Importantly, although we first reported that tolcapone binds to WT-TTR with a weak negative cooperativity [32], a reevaluation of the protein-ligand interaction in the present study revealed that under the present assay conditions, the interaction is best described as non-cooperative, with $K_d = 29$ nM. Binding of tafamidis to WT-TTR under the same conditions was assessed as a control and, as expected, a strong negative cooperativity was found, displaying $K_{d1} = 9.9$ nM and $K_{d2} = 260$ nM values, consistent with those previously reported [25]. In agreement with three recent studies [32, 45, 46], these data confirm that the concentration of tolcapone required to occupy simultaneously the two WT-TTR T₄ cavities is significantly lower than that of tafamidis in the same conditions, and therefore that tolcapone is a stronger stabilizer of this protein.

The binding of tolcapone to V30G-TTR and Y114C-TTR was also non-cooperative, with K_d s for the two binding sites of 110 nM and 45 nM, respectively, indicating that tolcapone binds more to the less destabilized Y114C-TTR mutant. Indeed, although the binding of tolcapone to Y114C-TTR is slightly weaker than to WT-TTR, the ΔG of the interactions are fairly similar, justifying why, for these initially folded and tetrameric proteins, tolcapone exerts a very similar anti-aggregational effect, being able to abrogate completely TTR deposition at 1:1 T₄-binding sites:tolcapone ratio (Fig. 1). In sharp contrast to the other two leptomeningeal variants, the binding of tolcapone to A25T-TTR displays a high negative cooperativity, reflected in a Hill coefficient (nH) of 0.25. Accordingly, whereas the binding to the first site is strong, with $K_{d1} = 63$ nM, the affinity for the second cavity is significantly decreased with a K_{d2} in the low μ M range (3.1 μ M). Irrespectively of the considered TTR mutant and T₄ pocket, the binding is always enthalpically driven. This is important because it has been suggested that the enthalpic component of binding

plays a predominant role in stabilizing TTR [46] and enthalpic forces correlate with the selectivity of transthyretin binding ligands in plasma [47], explaining why tolcapone binds selectively and stabilizes TTR in the blood of healthy volunteers and patients bearing the polyneuropathy-associated V30M mutation [32, 34]. From our thermodynamic data, the same should be true for the TTR CNS amyloidosis-associated variants in the brain, since the enthalpic component of the binding for these unstable mutants is very similar to the one exhibited for WT-TTR (Fig. 5).

Tolcapone binds TTR leptomeningeal amyloidosis-associated variants at the dimer-dimer interface

The fact that enthalpic contributions dominate the binding of tolcapone to the three mutant proteins, suggests that, as previously shown for WT-TTR [32], there exist specific interactions between the protein and the ligand. To confirm that these destabilized brain variants could accommodate tolcapone in their tetrameric structure, we obtained crystals of A25T-TTR, V30G-TTR and Y114C-TTR in the presence of tolcapone that diffracted beyond 1.60, 1.15 and 1.69 Å resolution, respectively (Fig. 6 and Table S1).

These high-resolution crystal structures clearly indicated the placement of tolcapone in the butterfly-shaped electron density map located in the “weak” dimer-dimer interface of TTR tetramer. As expected, the three new structures were virtually identical to that of WT-TTR in the presence of tolcapone (PDB: 4D7B), with overall root-mean-square deviation values for the C α of 0.20 Å for A25T-TTR (PDB: 6TXV), 0.45 Å for V30G-TTR (PDB: 6TXW) and 0.43 Å for Y114C-TTR (PDB: 6XTK). Accordingly, a closer view into the mutation site suggests that the mutated residues do not significantly disturb the structure of the TTR/tolcapone complex (Fig. S2). This is consistent with the reported data of a vast majority of mutant TTR crystal structures [48]. To the best of our knowledge, the structure of V30G-TTR/tolcapone complex (PDB:6TXW), represents the first one for this protein variant. Val30 maps to the monomer B-strand, our structure indicating that the removal of the side chains of the four buried Val30 residues leads only to minor perturbations of the native tetrameric structure, which are restricted to the vicinity of the mutated residues, e.g. the loss of the intramolecular contacts made with Val71 side-chain in the E-strands. However, the B-factor of Val71 in V30G-TTR remains similar to the one measured in the WT-TTR/tolcapone complex structure, indicating that the mutation does not even affect local flexibility. The structure of the Y114C-TTR/tolcapone complex is the first one to show that a kinetic stabilizer can diffuse into the T4-binding site of this particular mutant.

In all structures, tolcapone binds to the TTR tetramer in the forward mode, with the 4-methyl-phenyl ring occupying the inner binding cavity and the 3,4-dihydroxy-5-nitrophenyl ring of tolcapone oriented to the outer binding cavity. The presence of tolcapone inside of the T4 binding pocket stabilizes the orientation of key residues that form the halogen binding pockets (HBP) HBP1-1' (Lys15 and Leu17), HBP2-2' (Ala108 and Leu110) and HBP3-3' (Ser117 and Thr119), as inferred from their very well-defined electron maps (Fig. S3). This results in virtually identical polar, hydrophobic contacts and electrostatic interactions inside the T4 pocket to those stabilizing the tetramers in WT-TTR/tolcapone complex structure, including a specific hydrogen bond interaction between the carbonyl group of tolcapone and the hydroxyl side chain of Thr119. The

distances between the carbonyl group of tolcapone and the hydroxyl side chain of Thr119 in our structures are similar to the ones observed in the WT-TTR/tolcapone structure and point to the formation of strong hydrogen bonds (Table S2). This bond is buried in the low dielectric context of the T₄ pocket, in such a way that this interaction is predicted to be strong (Fig. 6 and Table S2). Thus, the establishment of hydrogen bonds between the inhibitors and TTR residues in the inner binding cavity seems to be important for the binding affinity [46].

As in the case of WT-TTR, in the leptomeningeal amyloidosis mutants the ϵ -amino groups of Lys15/Lys15' are sandwiched between the hydroxyl groups of the phenyl ring of tolcapone and the carboxylate group of Glu54 (Fig. 6 and Table S2). These specific electrostatic interactions by Lys15 close the cavity around tolcapone hampering the entrance of solvent into the pocket and stabilizing the protein-compound complex.

The new structures highlight how the tolcapone-protein interactions at the T₄-binding site in the “weak” dimer–dimer interface are not significantly affected by the mutations, providing an explanation for the enthalpy driving binding of the TTR CNS amyloidosis-associated variants and suggesting that the observed anti-aggregational activity of the compound responds to a canonical kinetic stabilization of the tetramer once the molecule occupies the hormone-binding sites of the pathogenic proteins.

CONCLUSIONS

In the present study, we have described in detail the activity of tolcapone, a potent kinetic stabilizer originally intended to treat Parkinson’s disease, on TTR variants associated to leptomeningeal amyloidosis, a rare, but fatal form of amyloidosis. We could confirm that A25T-, V30G- and Y114C-TTR variants present lower structural stability compared with WT-TTR, especially A25T-TTR. Mutations impact the tetramer-monomer equilibrium, lowering the ability of these variants to remain in the functional native tetrameric structure, and hence enhancing the protein susceptibility to aggregation. Our study demonstrates that tolcapone binds strongly and canonically at the T₄-binding sites of all the assayed TTR leptomeningeal amyloidosis-associated variants, restoring the structural stability of the tetramer and, consequently, inhibiting the pathogenic protein aggregation. This activity, together with the compound’s ability to pass the BBB, suggests that tolcapone might become a first broad spectrum pharmacologic treatment for all forms of TTR-related amyloidosis, including the devastating CNS forms of the disease.

All in all, our study provides solid biophysical and structural rational for a recent proof of principle clinical trial aimed to test the ability of tolcapone to stabilize TTR in symptomatic and asymptomatic leptomeningeal TTR patients (ClinicalTrials.gov Identifier: NCT03591757).

MATERIALS AND METHODS

TTR expression and purification

The vectors coding for TTR variants were prepared by PCR-site directed mutagenesis using the QuickChange Lightning kit (Agilent technologies, Santa Clara, California, USA). pET28a

vector (Novagen, Addgene, Cambridge, USA) encoding for the WT protein was used as a template.

For protein expression, *Escherichia coli* BL21 (DE3) cells, previously transformed with the corresponding plasmid, were grown in LB medium supplemented with 50 μ g.ml⁻¹ kanamycin at 37 °C to an optical density of 0.8 at 600 nm, and the culture induced with 1 mM isopropyl β -D-thiogalactopyranoside for 16 h. Cells were harvested by centrifugation and sonicated, the soluble fraction was recovered by centrifugation. The TTR-containing fraction was obtained by two consecutive steps of ammonium sulphate precipitation (50 and 90% respectively). The precipitate was solubilized in 25 mM Tris-HCl, pH 8, and dialysed against the same buffer. The sample was loaded onto a HiTrap Q HP column (GE Healthcare, Illinois, USA) and eluted with a linear gradient 0 – 0.5 M NaCl in 25 mM Tris-HCl, pH 8. The TTR-enriched fractions were precipitated in 90% ammonium sulphate and the obtained precipitate redissolved in a small volume of buffer (25 mM Tris, 100 mM NaCl, pH 8). TTR was finally purified by gel filtration chromatography on a HiLoad 26/600 Superdex 75 prep grade column (GE Healthcare) and eluted with 25 mM Tris, 100 mM NaCl. Purest fractions were combined and stored at -20 °C. Protein concentration was determined spectrophotometrically at 280 nm using an extinction coefficient of 77 600 M⁻¹ cm⁻¹. All the TTR variants, including the WT, were expressed and purified following the same procedure.

TTR aggregation inhibition *in vitro*

TTR solutions (7 μ M in 10 mM sodium phosphate, 100 mM KCl, 1 mM EDTA, 1 mM DTT, pH 7.0) were incubated with increasing concentrations of tolcapone for 30 min at 37 °C. Since the stock solutions of tolcapone were prepared in dimethylsulphoxide (DMSO), the percentage of DMSO in all the samples was adjusted to 5% (v/v). After the incubation period, the pH was dropped to 5.0 via dilution 1:1 with acidification buffer (100 mM sodium acetate, 100 mM KCl, 1 mM EDTA, 1 mM DTT, pH 5.0) and the samples were incubated for more 22 h at 37 °C. For WT-TTR and Y114C-TTR, a pH of 4.2 and an incubation period of 72 h were used instead. The formation of aggregates was assessed by light scattering, with the excitation and emission wavelengths set at 340 nm, using a Varian Cary Eclipse Fluorescence Spectrophotometer (Agilent technologies). TTR aggregation was considered maximum in the absence of tolcapone (100% aggregation).

Urea denaturation curves in the absence or presence of tolcapone

TTR solutions at 1.8 μ M in PBS were incubated for 30 min with tolcapone and urea in 50 mM sodium phosphate, 100 mM KCl, pH 7.0, was added to obtain a range of final concentration from 0 to 9.5 M. A control sample was prepared with DMSO, instead of tolcapone. The concentration of urea in the solutions was verified by refractive index, using a Manual Hand-held Refractometer HR901 (A. Krüss Optronic, Germany). After 96 h incubation at RT, the fraction unfolded was determined by intrinsic fluorescence spectroscopy, using a FP-8200 Spectrofluorometer (Jasco, Easton, USA). The samples were excited at 295 nm, allowing the selective excitation of the Trp residues on the protein, and fluorescence emission spectra were

recorded from 310 to 400 nm. Upon denaturation, the Trp residues become more solvent-exposed and the maximum fluorescence changes from 335 nm to 355 nm, approximately. The 355/335 nm fluorescence emission intensity was normalized from minimum (folded) to maximum (unfolded), with the maximum being the sample without tolcapone, and plotted as a function of urea concentration.

Urea mediated transthyretin dissociation measured by resveratrol binding

A25T- and WT-TTR solutions (1.8 μ M in PBS) were incubated with different concentrations of urea buffered with 50 mM sodium phosphate, 100 mM KCl, 1 mM EDTA and 1 mM DTT (pH 7.0) for 96 h at RT. After this time, 1.8 μ l of resveratrol from a 1 mM stock solution in DMSO was added to a 100 μ l sample just before the measurement, in order not to shift the equilibrium towards the tetramer [17]. The incubation time with resveratrol was the same for all samples. The samples were excited at 320 nm and the fluorescence recorded from 350 to 550 nm, using a FP-8200 Spectrofluorometer (Jasco). The fluorescence emission intensity of resveratrol at 394 nm was used to calculate the amount of non-tetrameric TTR and plotted as a function of urea concentration.

Resveratrol binding to TTR

In order to assess resveratrol binding to TTR, 1.8 μ l of resveratrol from a 1 mM stock solution in DMSO were added to 100 μ l TTR samples (1.8 μ M in PBS) and the samples were excited at 320 nm, using a FP-8200 Spectro-fluorometer (Jasco). The resveratrol fluorescence intensity at 394 nm was used as an indicative of the amount of tetrameric TTR present in the samples. The difference between TTR leptomeningeal amyloidosis-associated variants and WT-TTR was analysed by one-way ANOVA Dunnett test using GraphPad Prism 6.0 software (GraphPad Software Inc.). $p < 0.05$ was considered statistically significant.

Analytical size exclusion chromatography

TTR samples (500 μ l at 7 μ M or 100 μ l at 20 μ M, in PBS) were filtered and injected into a Superdex 75 10/300 GL column (GE Healthcare), previously equilibrated with PBS. Elution was done with the same buffer, at a flow rate of 0.4 ml/min, and monitored by absorbance at 280 nm, which was normalized from 0 to 1.

Isothermal titration calorimetry

The thermodynamic parameters that describe the interaction between TTR and tolcapone were determined by ITC, using a MicroCal Auto-iTC200 Calorimeter (MicroCal, Malvern-Panalytical, UK). A25T-, Y114C- and WT-TTR at 5 μ M located in the calorimetric cell were titrated against tolcapone at 100 μ M in the injection syringe in PBS buffer pH 7.0, 100 mM KCl, 1 mM EDTA, 2.5% DMSO, at 25 °C. A stirring speed of 750 rpm. and 2 μ l injections were programmed, with consecutive injections separated by 150 s to allow the calorimetric signal (thermal power) to return to baseline. For V30G-TTR the concentration of protein was increased to 20 μ M, since the concentration of TTR able to bind tolcapone was low. For each protein, two

replicates were performed, and the experimental data was analyzed with a general model for a protein with two ligand-binding sites [43, 44] implemented in Origin 7.0 (OriginLab), which allows to establish whether the two binding sites are identical and independent. Appropriate control blank experiments were performed to test for unwanted heat artefacts or unspecific phenomena.

Crystal structures of TTR/tolcapone complexes

Co-crystals of A25T-TTR/tolcapone, V30G-TTR/tolcapone and of Y114C-TTR/tolcapone were obtained at 18 °C by hanging drop vapor diffusion methods after mixing a tenfold molar excess of ligand with protein to ensure saturation. The reservoir solution contained between 25 and 30% PEG 400, 200 mM CaCl₂, 100 mM HEPES, pH 7.0-7.5. Single crystals appeared after 3 days from equal volumes of protein solution (10 mg.ml⁻¹ in 50 mM Tris-HCl pH 8.0, 100 mM KCl, 1mM EDTA) and reservoir solution. Crystals were cryo-protected in reservoir buffer containing 10% glycerol and flash-frozen in liquid nitrogen before diffraction analysis. Diffraction data were recorded from cryo-cooled crystals (100 K) at the ALBA synchrotron in Barcelona (BL13-XALOC beamline) [49]. Data were integrated and merged using XDS [50] and scaled, reduced and further analyzed using CCP4 [51]. The structure of TTR/tolcapone complex was determined from the X-ray data by molecular replacement using a previous TTR structure (PDB code 1F41) as a model using Phaser [52]. The initial electron density maps showed the butterfly electron density map corresponding to tolcapone in the TTR dimer-dimer interface that was manually built using COOT [53]. Model refinement was performed with Phenix (1.17.1_3660) [52]. Ramachandran analysis shows 98.00% of residues (230) are in preferred regions and only 2.00% of residues (4) are in allowed regions for the A25T-TTR/tolcapone structure. For the V30G-TTR/tolcapone structure, 98.0% of residues (235) are in preferred regions and only 2.0% of residues (4) are in allowed regions. For the Y114C-TTR/tolcapone structure, 96.0% of residues (227) are in preferred regions and only 4.0% of residues (10) are in allowed regions. Refinement and data statistics are provided in Table S1. Structural representations were prepared with PyMOL (DeLano Scientific, LLC).

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHOR CONTRIBUTIONS

FP, SV and IP designed the study. FP, SE, JS, NV and AVC conducted the experiments. FP, NV, DR, SV and IP analyzed the results. FP, NV, AVC and IP prepared the figures. FP, SV and IP wrote the main manuscript text. All authors reviewed the manuscript.

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FIGURE LEGENDS

Fig. 1. Tolcapone effect over the aggregation of TTR leptomeningeal amyloidosis-associated variants. (A) A25T-TTR (in red), (B) V30G-TTR (in blue), (C) Y114C-TTR (in green) and WT-TTR (in black). Light scattering at 340 nm was measured and plotted as percentage of TTR aggregation. The values represent mean \pm s.e.m (n=3).

Fig. 2. Tolcapone effect on urea-induced denaturation of TTR leptomeningeal amyloidosis-associated variants. (A) A25T-TTR, (B) V30G-TTR, (C) Y114C-TTR and (D) WT-TTR. Black squares represent the control without tolcapone, while gold and magenta squares correspond to the samples incubated with 20 and 50 μ M of tolcapone, respectively. The values represent mean \pm s.e.m (n=3).

Fig. 3. Resveratrol binding of TTR leptomeningeal amyloidosis-associated variants. Resveratrol fluorescence at 394 nm when incubated with A25T-TTR (in red), V30G-TTR (in blue), Y114C-

TTR (in green) and WT-TTR (in black). The values represent mean \pm s.e.m (n=3). Statistical differences were found between A25T- and V30G-TTR binding to resveratrol, in comparison to WT-TTR; *p < 0.05 and **p < 0.01.

Fig. 4. Effect of protein concentration on the quaternary structure adopted by TTR leptomeningeal amyloidosis-associated variants. (A-D) Elution profiles of A25T-TTR (in red), V30G-TTR (in blue), Y114C-TTR (in green) and WT-TTR (in black) at 7 (solid line) or 20 μ M (dashed line).

Fig. 5. Calorimetric titration of TTR leptomeningeal amyloidosis-associated variants with tolcapone. (A) The top panel (thermogram) shows how the thermal power changes with time. The bottom panel (binding isotherm) is obtained from the transformation of the thermogram and represents the normalized heat of reaction per injection as a function of the ratio between the molar concentration of tolcapone and protein. The solid line in the binding isotherm corresponds to the best fitting to each curve. (B) Graphical representation of ΔG (dark bars), ΔH (light bars) and $-T\Delta S$ (striped bars) for the binding of tolcapone to the first and second site of A25T-TTR (in red), V30G-TTR (in blue), Y114C-TTR (in green) and WT-TTR (in black). In the case of V30G-, Y114C- and WT-TTR, the binding parameters for both sites are the same.

Fig. 6. Crystal structures of A25T-TTR, V30G-TTR and Y114C-TTR in complex with tolcapone. (A) Global view of TTR variants bound to tolcapone (cartoon representation). The electron density maps of the two tolcapone-binding sites of tolcapone are depicted. The position of the mutated residues in the structure is specified with asterisks. Dashed lines represent the two-fold symmetry axis of the dimer–dimer interface. (B) Detailed and close-up views of one of the TTR tolcapone-binding sites of the variants (PDB: 6TXV, 6TXW, 6XTK); tolcapone and some of the TTR tetramer residues in contact with it are represented by sticks. (C) Close-up views of the electron density map of the mutated residues.

TABLES

Table 1. Thermodynamic parameters obtained by ITC experiments for the binding of tolcapone to TTR leptomeningeal amyloidosis-associated variants*.

	K_d_1 (μ M)	ΔG_1 (kcal/mol)	ΔH_1 (kcal/mol)	$-T\Delta S_1$ (kcal/mol)	K_d_2 (μ M)	ΔG_2 (kcal/mol)	ΔH_2 (kcal/mol)	$-T\Delta S_2$ (kcal/mol)	nH
A25T	0.063	-9.8	-17.1	7.3	3.1	-7.5	-12.3	4.8	0.25
V30G	0.110	-9.5	-12.3	2.8	0.110	-9.5	-12.3	2.8	1
Y114C	0.045	-10.0	-14.4	4.4	0.045	-10.0	-14.4	4.4	1
WT	0.029	-10.3	-8.8	-1.5	0.029	-10.3	-8.8	-1.5	1

* 1 and 2 correspond to the parameters associated with the first and second binding site of TTR, respectively.

SUPPORTING INFORMATION

Table S1. Data collection and refinement statistics of transthyretin variants in complex with tolcapone.

Table S2. Interatomic distances between tolcapone and residues of TTR variants that form HBP1 and -1' and HBP2 and -2'.

Fig. S1. Stereo images of TTR, of the electron density maps of tolcapone and of residues involved in binding.