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1 Comparative anatomy of the carotid canal in the Miocene small-bodied catarrhine *Pliobates*  
2 *cataloniae*

3

#### 4 **Abstract**

5 The small-bodied Miocene catarrhine *Pliobates cataloniae* (11.6 Ma, Spain) displays a  
6 mosaic of catarrhine symplesiomorphies and hominoid synapomorphies that hinders  
7 deciphering its phylogenetic relationships. Based on cladistic analyses it has been  
8 interpreted as a stem hominoid or as a pliopithecoid. Intriguingly, the carotid canal  
9 orientation of *Pliobates* was originally described as hylobatid-like. The variation in carotid  
10 canal morphology among anthropoid clades shown in previous studies suggests that this  
11 structure might be phylogenetically informative. However, its potential for phylogenetic  
12 reconstruction among extinct catarrhines remains largely unexplored. Here we quantify the  
13 orientation, proportions, and course of the carotid canal in *Pliobates*, extant anthropoids  
14 and other Miocene catarrhines (*Epipliopithecus*, *Victoriapithecus*, and *Ekembo*) using 3D  
15 morphometric techniques. We also compute phylogenetic signal and reconstruct the  
16 ancestral carotid canal course for main anthropoid clades. Our results reveal that carotid  
17 canal morphology embeds strong phylogenetic signal but mostly discriminates between  
18 platyrrhines and catarrhines, with an extensive overlap among extant catarrhine families.  
19 The analyzed extinct taxa display a quite similar carotid canal morphology more closely  
20 resembling that of extant catarrhines. Nevertheless, our results for *Pliobates* highlight some  
21 differences compared with the pliopithecoid *Epipliopithecus*, which displays a somewhat  
22 more platyrrhine-like morphology. In contrast, *Pliobates* appears as derived toward the  
23 modern catarrhine condition as the stem cercopithecoid *Victoriapithecus* and the stem  
24 hominoid *Ekembo*, which more closely resemble one another. Moreover, *Pliobates* appears

25 somewhat derived toward the reconstructed ancestral hominoid morphotype, being more  
26 similar than other Miocene catarrhines to the condition of great apes and the hylobatid  
27 *Symphalangus*. Overall, our results rule out previously noted similarities in carotid canal  
28 morphology between *Pliobates* and hylobatids, but do not show particular similarities with  
29 pliopithecoids either—as opposed to extant and other extinct catarrhines. Additional  
30 analyses will be required to clarify the phylogenetic relationships of *Pliobates*, particularly  
31 given its dental similarities with dendropithecids.

32

33 **Keywords:** Internal carotid artery; Petrosal; Geometric morphometrics; Fossil primates;  
34 Phylogeny; Evolution.

35

## 36 1. Introduction

### 37 1.1. *Pliobates* and catarrhine evolution

38 Crown catarrhines include two main extant clades distinguished at the superfamily rank:  
39 Old World monkeys (Cercopithecoidea) and apes and humans (Hominoidea). Earliest stem  
40 catarrhines have been reported from the latest Eocene (34.5–29.5 Ma) of Afro-Arabia  
41 (Seiffert, 2006, 2012; Seiffert et al., 2010). In turn, earliest crown catarrhines (both  
42 cercopithecoids and hominoids) date back to the late Oligocene (~25 Ma) of East Africa  
43 (Stevens et al., 2013)—in accordance with molecular estimates indicating that the  
44 cercopithecoid–hominoid divergence occurred during the Oligocene (Chatterjee et al., 2009;  
45 Perelman et al., 2011; Springer et al., 2012; Finstermeier et al., 2013; Pozzi et al., 2014).  
46 Catarrhines did not disperse into Eurasia until later in the early Miocene, following the  
47 closure of the Tethys Seaway and the establishment of intermittent land bridges at ~19 Ma  
48 (Harzhauser et al., 2007; Harrison, 2013). Several catarrhine lineages eventually dispersed

49 from Afro-Arabia into Eurasia (Roos et al., 2019; Gilbert et al., 2020a), including:  
50 pliopithecoids (considered an Eurasian clade of stem catarrhines, first recorded ~18–17 Ma;  
51 Harrison and Gu, 1999; Begun, 2002, 2017; Harrison, 2013); hylobatids (first recorded ~13.8–  
52 12.5 Ma; Gilbert et al., 2020b); large-bodied hominoids (presumably hominids, first recorded  
53 ~16.5–14 Ma; Casanovas-Vilar et al., 2011; Harrison, 2017); and various lineages of  
54 cercopithecoid primates, successively recorded at different times in Eurasia, namely  
55 colobines (~8.5 Ma; Alba et al., 2015a), cercopithecins (~8–6.5 Ma; Gilbert et al., 2014),  
56 macaques (~5.9–5.3 Ma; Alba et al., 2014a), and geladas (1.6–1.2 Ma; Alba et al., 2014b).

57 A panoply of small-bodied, putative stem catarrhine genera (including, but not restricted  
58 to, dendropithecids) are recorded in the early to middle Miocene of Africa (Harrison, 2010,  
59 2013). Some Miocene Eurasian taxa have previously been linked to some of these African  
60 stem catarrhines—implying yet another catarrhine dispersal event from Africa—but such  
61 claims have been mostly rejected by reinterpreting them as pliopithecoids or hylobatids  
62 (Harrison, 2016 and references therein). The most likely exception is some isolated dental  
63 remains from Pakistan (~17–16 Ma; Barry et al., 1987; Bernor et al., 1988), which are most  
64 similar to African dendropithecids (Harrison, 2016) but might ultimately prove to be stem  
65 pliopithecoids (Sankhyan et al., 2017). In turn, as remarked by Roos et al. (2019), the small-  
66 bodied catarrhine *Pliobates* (middle/late Miocene of Spain, 11.6 Ma; Alba et al., 2015b)  
67 might denote an additional out-of-Africa dispersal event of catarrhines unless it is  
68 interpreted as a pliopithecoid. This taxon was originally recovered by Alba et al.'s (2015b)  
69 cladistic analysis as a stem hominoid morphologically more derived than *Proconsul* s.l.  
70 (currently split into *Proconsul* s.s. and *Ekembo*; McNulty et al., 2015) and subsequently  
71 reinterpreted as a pliopithecoid by different cladistic analyses (Nengo et al., 2017; Gilbert et  
72 al., 2020b), as first suggested by Benefit and McCrossin (2015). However, given the mosaic

73 nature of its cranial and postcranial skeleton—combining plesiomorphic, stem catarrhine-  
74 like features (including dental resemblances with dendropithecids) with crown hominoid  
75 synapomorphies (Alba et al., 2015b)—the possibility remains that *Pliobates* is a late  
76 descendant of an African stem catarrhine lineage.

77       Deciphering the phylogenetic position of *Pliobates* would have important implications for  
78 our current understanding of catarrhine evolution and paleobiogeography during the  
79 Miocene. If ultimately shown to be a pliopithecoid or as a member of a different stem  
80 catarrhine lineage, it would imply remarkable cranial and postcranial convergences with  
81 crown hominoids; in turn, if interpreted as a non-pliopithecoid stem catarrhine or as a stem  
82 hominoid, it would indicate that pliopithecoids were not the only small catarrhines to  
83 disperse from Africa into Europe. Unfortunately, determining the evolutionary history and  
84 phylogenetic relationships of small-bodied catarrhines is still hindered by the fragmentary  
85 nature of their fossil remains and the incompleteness of their fossil record—with long ghost  
86 lineages for both pliopithecoids and hylobatids. Deciphering the phylogenetic position of  
87 extinct taxa ultimately requires performing cladistic analyses with all available evidence from  
88 multiple anatomical regions. However, as mentioned above, previous attempts in this regard  
89 have yielded contradictory results for *Pliobates* (compare Alba et al., 2015b with Nengo et  
90 al., 2017 and Gilbert et al., 2020b). Therefore, additional research on the various characters  
91 included in these analyses is required to refine their definition and scoring for both extant  
92 and extinct taxa. With this aim in mind, here we focus on one of the most intriguing features  
93 included in the original diagnosis of *Pliobates*: the orientation of the carotid canal, which was  
94 originally described as most closely resembling hylobatids than either pliopithecoids or other  
95 hominoids (Alba et al., 2015b). Given the potential implications of such similarities—if  
96 interpreted as a hylobatid synapomorphy or hominoid symplesiomorphy—we decided to

97 analyze them quantitatively by means of a novel morphometric approach devised to  
98 measure the size and shape of the carotid canal. To the best of our knowledge, this is the  
99 first study to quantitatively assess the orientation, proportions, and course of this  
100 anatomical structure in extant and extinct anthropoids.

101

## 102 *1.2. The carotid canal in primate phylogenetics*

103 All haplorrhines and non-cheirogaleid lemuriforms have an internal carotid artery  
104 enclosed by a bony canal (termed 'carotid canal') that derives from the anterior lamina of  
105 the petrosal plate (Bugge, 1974, 1980; Cartmill et al., 1981; MacPhee and Cartmill, 1986; Kay  
106 et al., 2008; Boyer et al., 2016). The carotid canal runs from the posterolaterally located  
107 external carotid foramen to the endocranial space (Bugge, 1980; MacPhee and Cartmill,  
108 1986; Kay et al., 2008; Boyer et al., 2016). In haplorrhines, the carotid canal displays a  
109 'perbullar pathway': it is located in the primary medial wall of the auditory bulla and runs  
110 through the middle ear cavity, passing through the transverse septum that separates the  
111 tympanic cavity from the anterior accessory cavity (Cartmill et al., 1981; MacPhee and  
112 Cartmill 1986; Kay et al., 2008). The canal then opens more anteromedially into the  
113 endocranial surface, with an opening located either on the basisphenoid or between the  
114 basisphenoid and petrosal (Boyer et al., 2016). In adult anthropoids, one of the two  
115 branches of the internal carotid artery (the stapedial artery) is missing and only the  
116 promontory artery remains patent (Bugge 1974, 1980; Rosenberger and Szalay, 1980;  
117 MacPhee and Cartmill, 1986; Kay et al., 2008; Boyer et al., 2016). Therefore, the carotid  
118 canal of anthropoid primates corresponds to a unique and well-developed bony enclosure.

119 Low homoplasy is one of the most important criteria for choosing phylogenetically  
120 informative morphological characters (Lieberman, 1999). Among characters least affected by

121 homoplasy (convergence, parallelism, reversal, and homoiology), soft-tissue traits have  
122 previously been proven more efficient in producing accurate primate phylogenies than hard-  
123 tissue traits (Gibbs et al., 2000, 2002; Diogo and Wood, 2011). Given that vessel and nerve-  
124 related characters are soft-tissue traits, this rationale might potentially apply to the carotid  
125 canal as well. A few previous studies have included carotid canal features in character-taxon  
126 matrices devised for cladistic analysis, focusing on the position of the carotid foramen in the  
127 bulla as well as on the overall direction and orientation of the canal itself (Beard and  
128 MacPhee, 1994; Ross, 1994; Shoshani et al., 1996; Ross et al., 1998; Kay et al., 2008; Alba et  
129 al., 2015b). These studies aimed at resolving phylogenetic relationships among higher-rank  
130 primate clades (Beard and MacPhee, 1994; Ross, 1994; Shoshani et al., 1996; Ross et al.,  
131 1998; Kay et al., 2008) and/or deciphering the phylogenetic relationships of particular fossil  
132 taxa (Ross et al., 1998; Kay et al., 2008; Alba et al., 2015b). Nonetheless, some of them found  
133 differences between platyrrhines and catarrhines, as well as among some catarrhine groups.  
134 Based on these studies, the morphology of the carotid canal seems promising for  
135 phylogenetic reconstruction in catarrhine primates. However, the exact phylogenetic  
136 potential of this structure for distinguishing among catarrhine subclades and, hence,  
137 clarifying the relationships of extinct taxa remains unexplored.

138 In the light of all the considerations above, the aim of this paper is threefold: (1)  
139 investigate the orientation, proportions, and course of the carotid canal in a wide sample of  
140 extant anthropoid primates, to test its potential for phylogenetic inference in extinct  
141 catarrhines; (2) re-evaluate previously noted differences and similarities among several  
142 catarrhine clades; and (3) describe the carotid canal morphology of *Pliobates* and compare it  
143 with that of extant and fossil catarrhines, so as to gain additional insight on the controversial  
144 phylogenetic relationships of this genus. To do so, we collected  $\mu$ CT scans of both fossil taxa

145 and an extant comparative sample, and developed a newly devised semiautomatic iterative  
146 protocol to obtain comparable 3D carotid canal surfaces (i.e., 3D models). These are  
147 analyzed by means of traditional morphometrics and three-dimensional geometric  
148 morphometric (3DGM) and imaging techniques, to quantify the orientation, proportions,  
149 and course of the carotid canal within the petrosal in a standardized manner.

150

## 151 **2. Materials and methods**

### 152 *2.1. Materials*

153 Studied sample This study focuses on the cranium of the holotype (IPS58443) of *Pliobates*  
154 *cataloniae*, which consists of a partial skeleton (Alba et al., 2015b). The carotid canal is  
155 entirely preserved in the right temporal fragment (IPS58443.1; Alba et al., 2015b: Fig. 1A). In  
156 contrast, in the left temporal fragment (same specimen) the inferiormost part of the canal is  
157 broken, with most of its lateral and anterior walls missing along a length of >1.5 mm.  
158 IPS58443.1 was originally scanned at a resolution of 95  $\mu\text{m}$  (Alba et al., 2015b). To explore  
159 fine bony structures, the specimen was scanned again by X-ray microtomography at the  
160 Centro Nacional de Investigación sobre la Evolución Humana (CENIEH, Burgos, Spain) using a  
161 Phoenix V|Tome|X s240  $\mu\text{CT}$  scanner with the following parameters: 0.35 mA current, 170  
162 kV voltage, 0.2 mm Cu filter, and a magnification of 9.52. The final reconstructed volume has  
163 an isometric voxel size of 21  $\mu\text{m}$ .

164 Fossil comparative sample Three Miocene catarrhines are included in the comparative  
165 sample: the Miocene pliopithecoid *Epipliopthecus vindobonensis* (see Zapfe, 1961), the stem  
166 cercopithecoid (victoriapithecoid) *Victoriapithecus macinnesi* (see review in Benefit and  
167 McCrossin, 1997), and the stem hominoid (proconsulid) *Ekembo heseloni* (see review in  
168 McNulty et al., 2015). There are four available fragmentary temporal bones of



169 *Epipliopithecus* (Zapfe, 1961): NMB OE 303 (right and left temporals, individual II), NHMW  
170 1970/1397/0002 (right temporal, individual III; holotype), and NHMW 1970/1397/0003 (left  
171 temporal, individual III; holotype). The carotid canal is only sufficiently preserved in the  
172 latter specimen, which includes a portion of the bulla, the external acoustic meatus, and  
173 most of the petrosal (Zapfe, 1961: Fig. 28). In contrast, the anterior end of the canal is  
174 missing from the remaining specimens. The petrosal of the holotype was scanned at the  
175 Vienna  $\mu$ CT lab (V $\mu$ -CTL) using a Viscom X8060  $\mu$ CT scanner with the following parameters:  
176 0.24 mA current, 120 kV voltage, 0.5 mm Cu filter, and a magnification of 7.50. The final  
177 reconstructed volume has an isometric voxel size of 22  $\mu$ m. *Ekembo heseloni* is represented  
178 by the right temporal specimen KNM-RU 2036al (Davis and Napier, 1963: Fig. 1; Alba et al.,  
179 2015b: Fig. 4C), which is part of the holotype (Walker et al., 1993), while *V. macinnesi* is  
180 represented by the left temporal of the cranium KNM-MB 29100 (Benefit and McCrossin,  
181 1997: Fig. 1; Benefit, 1999: Fig. 3; Jablonski and Frost, 2010: Fig. 23.1). The  $\mu$ CT scans of  
182 KNM-RU 2036al and KNM-MB 29100 have isometric voxel sizes of 64  $\mu$ m and 44  $\mu$ m,  
183 respectively. They were both kindly made available for this study by the Department of Earth  
184 Sciences, National Museums of Kenya (which holds the copyright) and the Department of  
185 Human Evolution, Max Plank Institute for Evolutionary Anthropology, Leipzig, Germany.

186 Extant comparative sample Our extant comparative sample consists of  $\mu$ CT scans of 127  
187 crania belonging to 41 anthropoid species (13 platyrrhines and 28 catarrhines) from 36  
188 genera, representing all extant hominoid genera as well as all cercopithecoid subtribes and  
189 platyrrhine (sub)families (Table 1; Supplementary Online Material [SOM] Table S1). A  
190 minimum of three individuals per genus are included. Ontogenetic changes in basicranial  
191 angulation and elongation—somewhat related to each other and potentially affecting  
192 petrosal morphology—continue up to adulthood as measured by M<sup>3</sup> eruption (Lieberman

193 and McCarthy, 1999; Lieberman et al., 2000). Therefore, all individuals included in the  
194 comparative sample are fully adult (exhibiting the M<sup>3</sup> in occlusion, or the M<sup>2</sup> in  
195 callitrichines). Most of the  $\mu$ CT scans included in our extant comparative sample were  
196 downloaded from MorphoSource (<https://www.morphosource.org>; Table 1; SOM Table S1).  
197 Three  $\mu$ CT scans of specimens from the AMNH were kindly provided by Sergio Almécija,  
198 while seven additional  $\mu$ CT scans of specimens housed in the Institut Català de Paleontologia  
199 Miquel Crusafont (Sabadell, Spain) were scanned at the CENIEH. The  $\mu$ CT resolution for the  
200 specimens included in the extant comparative sample ranges from 27 to 131  $\mu$ m (Table 1;  
201 SOM Table S1).

202

## 203 *2.2. Morphometric methods*

204 Segmentation For each specimen, the carotid canal was virtually extracted through  
205 semiautomatic threshold-based segmentation in Avizo v. 7.0 (Visualization Sciences Group,  
206 Mérignac). The right carotid canal was segmented for most specimens; when this was not  
207 possible, the left canal was segmented and mirrored. Both extremities of the resulting 3D  
208 canal surfaces (i.e., 3D models) were cut along homologous planes to allow their comparison  
209 among different individuals. First, to cut the posterior end of the canal, we used a best-fit  
210 plane based on landmarks placed on its external aperture (Fig. 1a, l; SOM Fig. S1a, b). Then,  
211 the anterior limit of the carotid canal was identified by determining a landmark on the  
212 anterior part of the petrosal (the ‘intersection ridge–groove’ [IRG]; Fig. 1b–l; SOM Fig. S1c, d;  
213 see SOM S1) and cut through such a homologous point using a script coded in R v. 4.0.2 (R  
214 Core Team, 2020; see SOM file S1).

215 Alignment and canal orientation The orientation of the carotid canal within the petrosal was  
216 quantified by means of two 2D angles representing the canal superior orientation along the

217 (para)sagittal plane (2DYZA) and its medial orientation along a transverse plane (2DXYA)—  
218 which can be related to canal 'orientation' and 'direction' sensu Alba et al. (2015b: Tables S5  
219 and S6), respectively. Each was computed as the angle between the vector connecting the  
220 canal endpoints (from posterior to anterior) and a standard reference vector. Both the canal  
221 vector and the reference vector were computed after performing a Procrustes alignment  
222 based on petrosal/tympanic landmarks (Fig. 2; Table 2) and applying the translation-scaling-  
223 rotation parameters resulting from this alignment to the two canal endpoints (SOM S1). This  
224 procedure allowed us to retain the information on the position, orientation and shape  
225 variation of the canal inside the petrosal. For both angles, the standard reference vector  
226 corresponds to a 2D vector parallel to the anteroposterior cranial axis (from posterior to  
227 anterior), in lateral view for 2DYZA and in superior view for 2DXYA (Fig. 3). Aligning the  
228 carotid canal on the basis of petrosal/tympanic landmarks instead of the cranium as a whole  
229 enables the analysis of fragmentary fossils and has several advantages (see SOM S1 for  
230 further details), while being consistent with previous studies (Beard and MacPhee, 1994;  
231 Ross, 1994; Shoshani et al., 1996; Ross et al., 1998; Kay et al., 2008; Alba et al., 2015b) that  
232 assessed carotid foramen locations relative to the bulla instead of general cranial axes or  
233 planes.

234 Volume and length The volume ( $V$ , in  $\text{mm}^3$ ) of the 3D canal surfaces was computed using the  
235 module 'Surface Area Volume' in Avizo. To calculate canal length ( $L$ , in mm), we used  
236 equidistant slices between the two canal endpoints (which correspond to type III landmarks;  
237 Bookstein, 1991). These slices were used to generate 8 additional equidistant semilandmarks  
238 that follow the canal streamline using the 'digit.curve' function of 'Geomorph' v. 3.3.1  
239 (Adams et al., 2019), summing a total of 10 canal landmarks, which define the canal  
240 streamline. This canal landmark configuration thus corresponds to a total of 10 canal

241 streampoints. Canal length was taken as the sum of the length of the resulting nine canal  
242 segments (see SOM S1 for further details).

243 Robusticity index Carotid canal proportions were measured by a dimensionless index of  
244 canal robusticity as  $L/V^{1/3}$ . Even though the carotid canal is a hollow structure, we refer to  
245 this index as 'robusticity' because it measures how thick the canal is on average relative to  
246 its length. Low values of the index denote robust canals, whereas high values denote slender  
247 canals. The same terms have been applied to other hollow cranial structures, such as the  
248 semicircular canals of the bony labyrinth (Urciuoli et al., 2020, 2021a, 2021b). The diameter  
249 (and hence the cross-sectional area) of the carotid canal varies along its length, so that using  
250 the cube root of the volume as the denominator is equivalent to relying on the hypothetical  
251 average cross-sectional area. We used the cube root of the volume to make the index  
252 dimensionless (i.e., a shape variable), and verified that the robusticity index was free from  
253 allometric (size-scaling) effects by computing a phylogenetic generalized least-squares  
254 (PGLS) regression of  $\ln L$  against  $\ln V^{1/3}$  on the species means of the whole sample. We  
255 further computed a major axis regression of the same parameters, using the function  
256 'lmodel2' of the R package 'lmodel2' v. 1.7-3 (Legendre, 2018). The objective was to test  
257 whether length and volume of the carotid canal covary in a linear (i.e., isometric) or non-  
258 linear (i.e., allometric) fashion, and to do so under the assumptions that the independent  
259 variable is measured with error (major axis) or without error (PGLS). Note that, under  
260 geometric similarity, linear dimensions and volumes are expected to be correlated with an  
261 allometric slope of 1/3 (Gould, 1966). However, this does not apply here because we used  
262 the cube root of the canal volume, so that allometry would imply an allometric slope  
263 significantly different from 1.

264 Canal course configuration To assess carotid canal course within the petrosal, we applied the  
265 translation-scaling-rotation parameters resulting from the alignment of the  
266 petrosal/tympanic landmarks described above to the 10 canal landmark configuration. As for  
267 canal orientation, we were able to retain the position, orientation and shape variation of the  
268 canal inside the petrosal. Therefore, the postalignment 10 canal landmark configuration  
269 represents the course of the carotid canal within the petrosal (SOM S1) and will be  
270 thereafter referred to as 'carotid canal course configuration'.

271 Size scaling To better understand the influence of size-scaling (allometric) effects on carotid  
272 canal shape, we computed multiple bivariate regressions using PGLS. On the one hand, we  
273 relied on allometric regressions of canal measurements (as measured by  $L$  and  $V^{1/3}$ ) against  
274 overall canal size (as measured by centroid size [CS]; see Section 2.3 below) and body size (as  
275 measured by body mass [BM]); the allometric regression between CS and BM was also  
276 computed. On the other hand, we computed regressions between carotid canal shape—  
277 including not only the above-mentioned index of canal robusticity, but also the multivariate  
278 shape variables derived by means of between-group principal component analysis (bgPCA;  
279 see Section 2.3 below)—against log-transformed carotid canal size (as measured by  $V^{1/3}$  and  
280 CS) and BM. To minimize the effects of body size dimorphism, for each taxon we relied  
281 exclusively on average female BM taken from the literature (Smith and Jungers, 1997;  
282 Delson et al., 2000; Turner et al., 2018). Natural logarithms ( $\ln$ ) were used in all instances.  
283 For allometric regressions between metrical linear variables, the null hypothesis of isometry  
284 was rejected when unity was excluded from the 95% CI of the allometric slope. In contrast,  
285 for bivariate regressions of shape vs. size, isometry was rejected when the regression was  
286 significant (i.e., when the slope was significantly different from zero).

287

288 *2.3. Between-group principal component analyses*

289 Carotid canal course variation among clades was examined using a bgPCA of the carotid  
290 canal course configurations using the 'groupPCA' function of the R package 'Morpho' v. 2.8  
291 (Schlager, 2017). We used the following four main crown anthropoid clades as grouping  
292 factor: platyrrhines, cercopithecids, hylobatids, and hominids, with fossils projected onto the  
293 morphospace a posteriori. The bgPCA analysis was repeated based on a catarrhine-only  
294 subsample to see if it provided a better discrimination among extant catarrhine families  
295 once the variance introduced by platyrrhines was removed.

296 As explained in the preceding section, allometric effects on canal course configuration  
297 were assessed by means of PGLS regressions of between-group principal component (bgPCs)  
298 against log-transformed canal size ( $\ln CS$  and  $\ln V^{1/3}$ ) or log-transformed body size ( $\ln BM$ )—  
299 see Section 2.2 for further details.

300 We verified that the bgPCA grouping structure was not spurious by comparing our bgPCA  
301 results with those of a cross-validated bgPCA (Cardini et al., 2019). This procedure iteratively  
302 resamples the data set by excluding one individual at a time and using the remaining ( $n - 1$ )  
303 individuals to compute the model that is then used to classify the omitted individual. A  
304 cross-validated bgPCA significantly reduces the distortion of mean group differences, so that  
305 its comparison with the non-cross-validated bgPCA allows one to ascertain whether (or to  
306 what extent) the latter is affected by spurious grouping effects. As further recommended by  
307 Cardini and Polly (2020), we also relied on a permutational analysis of variance  
308 (PERMANOVA) to (1) explore the significance ( $p$ ) of group mean differences and (2) compute  
309 the variance ( $R^2$ ) explained by them. The permutations consist in randomly resampling  $n$   
310 times the data set (thus modifying group composition, as the individuals are permuted),  
311 each time recomputing the distance between the centroids of all pairs of newly formed

312 groups. Based on this set of permutations, here consisting of between-group Euclidean  
313 distances, both  $p$  and  $R^2$  were computed within the full space of canal course configurations  
314 as well as within the bgPCA spaces (both standard and cross-validated bgPCs). As in the case  
315 of the cross-validated bgPCA scatterplot, comparing the  $p$  and  $R^2$  values of the bgPCA  
316 (before and after cross-validation) with those of the raw canal course configuration  
317 coordinates permits verifying if the former inflates the differences among groups. These  
318 computations were done using the 'lm.rpp' function of the R package 'RRPP' v. 0.6.1  
319 (Collyer and Adams, 2018).

320 The discrimination among a priori defined groups was evaluated on the basis of correctly  
321 classified individuals after cross-validation. Posterior probabilities of group membership  
322 were computed for fossil specimens based on the squared Mahalanobis distances between  
323 their bgPC scores (bgPC1 and bgPC2) and those of extant group centroids using the  
324 'typprobClass' function of 'Morpho'. Finally, a neighbor-joining (NJ) cluster was computed  
325 using a matrix of Euclidean distances based on the raw coordinates of the canal course  
326 configurations with the 'nj' function of the R package 'phangorn' v. 2.5.5 (Schliep, 2011).

327

#### 328 *2.4. Statistical analyses*

329 Box and whisker plots were employed to assess the variation in carotid canal orientation  
330 (2DYZA and 2DXYA), proportions (index  $L/V^{1/3}$ ), and course (bgPC1 and bgPC2). To test for  
331 significant differences among extant groups, we used Kruskal-Wallis non-parametric tests  
332 and Bonferroni-corrected Mann-Whitney post hoc pairwise comparisons performed in R.  
333 When significant differences were found, *Pliobates* and other extinct genera were compared  
334 with the distributions of extant groups using z-scores—computed as  $z = (\text{individual value} -$

335 group mean) / SD—to assess if the fossil specimens fall within the variation of the latter ( $|z|$   
336  $\leq 1.96$ ).

337

## 338 2.5. Phylogenetically informed methods

339 Phylogenetic tree To compute PGLS regressions and the phylomorphospaces, as well as to  
340 quantify phylogenetic signal, we used a time-calibrated phylogenetic tree among the  
341 investigated taxa. The tree for the extant taxa was primarily derived from the 10kTrees v.3  
342 website (Arnold et al., 2010), although some species that were not included there were  
343 added a posteriori following Meyer et al. (2011) and Springer et al. (2012)—see SOM S1 for  
344 further details. Two different hypotheses for the extinct genera were considered, except for  
345 *Victoriapithecus*, which was invariably considered a stem cercopithecoid. Under hypothesis 1  
346 (SOM Fig. S2a), *Epipliopithecus*, *Ekembo*, and *Pliobates* were considered successive stem  
347 hominoids, following the results of Alba et al.'s (2015b) cladistic analysis. In contrast, under  
348 hypothesis 2 (SOM Fig. S2b), only *Ekembo* was considered a stem hominoid, while both  
349 *Pliobates* and *Epipliopithecus* were considered stem catarrhines (pliopithecoids), following  
350 the results of other cladistic analyses (Nengo et al., 2017; Gilbert et al., 2020b). The tip and  
351 divergence dates for the extinct taxa are detailed in SOM S1.

352 Phylogenetic signal To assess the phylogenetic signal embedded in carotid canal course, we  
353 computed Pagel's  $\lambda$  (Pagel, 1999) and Blomberg's K (Blomberg et al., 2003) statistics using  
354 the R package 'phytools' v. 0.7-70 (Revell, 2012). These metrics compare the variance in the  
355 phylogenetic tree tips relative to that expected under a Brownian motion evolutionary  
356 model. Pagel's  $\lambda$  measures the covariance among related species and ranges from 0 (no  
357 phylogenetic signal) to 1 (high phylogenetic signal). In turn, Blomberg's K reflects to what  
358 extent the phylogenetic tree matches the variance in the data (and where variance is



359 concentrated):  $K \approx 1$  means that trait evolution approximates the expectations under a  
360 Brownian motion model;  $K \ll 1$  means that closely related taxa resemble each other less  
361 than expected under a Brownian motion model (such that variance is mostly found within  
362 clades); finally,  $K \gg 1$  means that closely related units resemble each other more than  
363 expected under a Brownian motion model (such that variance is mostly found among  
364 clades).

365 Phylomorphospace and reconstruction of ancestral morphotypes To quantify and visualize  
366 patterns of carotid canal course variation along the branches of the phylogeny, we produced  
367 a phylomorphospace by projecting the phylogenetic tree onto the two first bgPCs  
368 (Sidlauskas, 2008). In a phylomorphospace, the taxon centroids are connected to one  
369 another in the morphospace following the phylogenetic relationships given by the chosen  
370 phylogeny. In the case of fossil taxa, because there are competitive hypotheses regarding  
371 their phylogenetic relationships, various time-calibrated cladograms were devised a priori—  
372 resulting in a different phylomorphospace for each phylogenetic hypothesis. Scores of the  
373 internal nodes—last common ancestors (LCAs)—were estimated based on a maximum  
374 likelihood method for continuous characters using the 'fastAnc' function of 'phytools'. To  
375 explore the extent to which extant and extinct taxa differ from the various LCAs, we rotated  
376 and translated the bgPC scores estimated for the LCAs back into the Kendall's shape space  
377 (i.e., configuration space) to obtain their landmarks coordinates and thus visualize the  
378 estimated ancestral carotid canal course conformations.

379

### 380 3. Results

381 The 3D carotid canal surface of *Pliobates* (SOM File S2) is compared with those of  
382 *Ekembo*, *Epipliopithecus*, and *Victoriapithecus*, as well as selected taxa from the extant

383 comparative sample in Figure 4. Individual data for the variables describing the orientation,  
384 proportions, and course of the carotid canal in the extant comparative sample are reported  
385 in SOM Table S2.

386

### 387 3.1. Carotid canal orientation

388 For 2DYZA, the angle values range from ca. 30° to 90° (Fig. 5a; Tables 3 and 4). On this  
389 basis, and taking into account the mean extant clade differences (SOM Tables S3 and S4), we  
390 defined three different states (potentially usable in future cladistic analyses): oblique (<55°);  
391 subvertical (55–65°); and vertical (>65°). Hominoids have smaller angles than platyrrhines  
392 and cercopithecids, i.e., a less vertically oriented canal, whereas cercopithecids have the  
393 highest angles, corresponding to vertical canals (Fig. 5a; Table 3; SOM Tables S3 and S4).  
394 *Pliobates* has the lowest value among Miocene catarrhines and does not significantly differ  
395 from the hylobatid range of variation (Fig. 5a; Tables 4 and 5). *Epipliopithecus* and *Ekembo*  
396 display very similar values to one another and most closely fit with the hominoid variation,  
397 although they only significantly differ from cercopithecids (Fig.5a; Tables 4 and 5). In  
398 contrast, *Victoriapithecus* has a higher angle, more similar to the monkey condition (i.e., a  
399 more vertical canal; Fig.5a; Table 4), although it does not significantly differ from any extant  
400 group (Table 5).

401 In the case of 2DXYA, the values range from ca. -4° to 45° (Fig. 5a; Tables 3 and 4; see  
402 SOM S1 regarding the interpretation of negative angles). On this basis, and taking into  
403 account the mean differences among extant clades (SOM Tables S3 and S4), we defined two  
404 discrete states: anteriorly oriented ( $\leq 30^\circ$ ); and medially oriented ( $> 30^\circ$ ). Despite some  
405 overlap, platyrrhines display a significantly higher 2DXYA—i.e., a more medially (less  
406 anteriorly) directed canal—than the three catarrhine families, which do not differ

407 significantly from one another (Fig. 5b; Table 3; SOM Tables S3 and S4). Among the fossil  
408 specimens, *Ekembo* shows the highest 2DXYA whereas *Victoriapithecus* shows the lowest  
409 (Fig.5b; Table 4). Based on the z-scores (Table 5), *Ekembo* displays a higher angle than extant  
410 hominoids, *Victoriapithecus* and *Pliobates* show a lower angle than extant platyrrhines, and  
411 *Epipliopithecus* displays a higher angle than hylobatids.

412

### 413 3.2. Carotid canal proportions and body size scaling

414 Robusticity index The two allometric regressions between L and  $V^{1/3}$  are significant ( $p <$   
415 0.001; Table 6), but the PGLS regression indicates slight negative allometry, whereas the  
416 major axis regression does not allow us to exclude isometry. This suggests that the canal  
417 robusticity index  $L/V^{1/3}$  is only slightly affected by intrinsic size-scaling effects, i.e., that  
418 carotid canal length and volume covary in an almost linear fashion when geometric similarity  
419 is considered. This is further confirmed by the lack of significant correlation between  $L/V^{1/3}$   
420 against CS (Table 6). When separate allometric regressions of L and  $V^{1/3}$  against CS are  
421 computed, it emerges that  $V^{1/3}$  and CS display an isometric relationship, whereas L and CS  
422 display a minimally positive allometric relationship (with a slope very close to 1). Overall, this  
423 suggests that these three variables are suitable proxies for the size of the carotid canal and  
424 that the robusticity index is only very slightly affected, if at all, by intrinsic size-scaling effects  
425 related to the overall size of the canal.

426 Based on the robusticity index, and despite some overlap, platyrrhines display  
427 significantly slenderer canals than catarrhine families, while hominids also display slenderer  
428 canals than both cercopithecids and hylobatids, with the latter showing the stoutest canals  
429 (Fig. 5c; Table 3; SOM Tables S3 and S4). *Ekembo* and *Epipliopithecus* exhibit similar indices,  
430 while *Victoriapithecus* shows a somewhat higher value but similarly overlaps with all

431 catarrhine families (Fig. 5c; Table 4), and *Pliobates* shows an even higher value and falls  
432 outside the hylobatid range (Fig. 5c; Table 4). However, none of the fossil specimens  
433 significantly differs from any extant clade (Table 5).

434 Body-size allometry The fact that the robusticity index is not markedly affected by the size of  
435 the canal itself notwithstanding, either carotid canal size and/or proportions could still be  
436 affected by body size-scaling effects as measured by regressions against BM (Table 6). Both L  
437 and CS display a negative allometric relationship relative to BM—slightly below the  
438 theoretical isometric slope of  $1/3$  predicted by geometric similarity—whereas, in contrast,  
439  $V^{1/3}$  fits well with the prediction of geometric isometry. This suggests that the volume of the  
440 carotid canal is a better proxy for BM than either L or CS if size-scaling effects are not  
441 corrected for, because the former variable is directly proportional to BM in a linear fashion,  
442 whereas both L and CS display negative allometry relative to BM. The similar allometric  
443 pattern of L and CS is not surprising, because the latter was computed from the carotid canal  
444 configurations—which are sets of canal streampoints—and, hence, it only contains  
445 information about carotid canal length (but not volume).

446 Differences in size scaling between L and V are reflected in the existence of a mild  
447 negatively allometric relationship when L is considered the dependent variable, although  
448 such a relationship is not significant when both variables are considered to display error (i.e.,  
449 isometry cannot be excluded in the major axis regression; Table 6). Differences in scaling  
450 between L and V relative BM, in any case, are reflected in the regression between the  
451 robusticity index and BM (Table 6), which is of inverse proportionality (i.e., a negative slope):  
452 since the numerator (L) of the index increases slightly slower than the denominator ( $V^{1/3}$ )  
453 relative to BM, the index tends to decrease as the body size increases. However, it is

454 noteworthy that despite being significant, the correlation between the robusticity index and  
455 body size only explains 14% of the variance (Table 6).

456

### 457 *3.3. Carotid canal course within the petrosal*

458 Principal components analysis The bgPCA discriminates between platyrrhines and  
459 catarrhines but not among catarrhine families (Fig. 6). The cross-validated bgPCA (SOM Fig.  
460 S3) is virtually identical to the standard bgPCA, and the permutational ANOVA results (SOM  
461 Table S5) show that mean group differences are significant in all instances. The percentage  
462 of variance explained by bgPCAs only increases slightly as compared with the raw data—  
463 indicating that grouping structure is not spurious. The percentage of correctly classified  
464 individuals by the bgPCA (Table 7) is moderately high (79%), even after cross-validation  
465 (77%), although much higher in the case of platyrrhines (92%) and hominids (91%) than for  
466 cercopithecoids (63%) and hylobatids (64%).

467 bgPC1 (Fig. 6a) explains most of the variance (94%), is not significantly correlated with CS  
468 ( $p = 0.21$ ; Table 6), and shows a strong phylogenetic signal at  $p < 0.001$  ( $\lambda = 0.95$ ;  $K = 3.28$ ).  
469 Although bgPC1 is not significantly correlated to  $\ln$  CS, it displays significant allometry  
470 relative to both  $V^{1/3}$  and BM, but only explains 19% and 25% of the shape variance,  
471 respectively (Table 6). This axis discriminates platyrrhines (negative scores) from catarrhines  
472 (positive scores) at  $p < 0.001$  (Fig. 5d; Table 3; SOM Tables S3 and S4) because of longer  
473 canals that originate more inferolaterally in the petrosal of the former (Figs. 6b, 7; SOM Figs.  
474 S4–S6). In contrast, catarrhines display shorter canals that originate more superiorly and are  
475 overall located more medially in the petrosal (Figs. 6b and 7; SOM Figs. S4–S6). Extant  
476 catarrhine families extensively overlap along bgPC1 and do not significantly differ in terms of  
477 canal length and foramen/canal superoinferior and mediolateral position (Fig. 5d; Table 3;

478 SOM Tables S3 and S4). *Pliobates* and the remaining fossil taxa display similar (moderately  
479 negative) bgPC1 scores that are intermediate between those of extant catarrhines and  
480 extant platyrrhines (Figs. 5d and 6a; Table 4) and only slightly overlap with cercopithecids  
481 and hominids or, in the case of *Epipliopithecus*, also marginally with platyrrhines. Their canal  
482 configuration (SOM Figs. S4–S6) is thus somewhat shorter and more medially located than in  
483 platyrrhines, although *Epipliopithecus* more closely resembles the platyrrhine condition than  
484 *Pliobates*, *Victoriapithecus*, and *Ekembo* do.

485 bgPC2 (Fig. 6a) explains only 5% of variance, is significantly correlated with CS,  $V^{1/3}$ , and  
486 BM (explaining respectively 58%, 46%, and 35% of the shape variance; Table 6), and shows  
487 significant phylogenetic signal at  $p < 0.001$  ( $\lambda = 0.99$ ;  $K = 0.62$ ). Variation along bgPC2 (Figs.  
488 6b, 7) reflects the canal position on the petrosal (more anterior toward negative values, and  
489 more posterior toward positive scores). Platyrrhines and catarrhines extensively overlap  
490 along this axis, but hominids display significantly more positive values than other groups  
491 despite considerable overlap (Fig. 5e; SOM Figs. S4–S6; SOM Tables S3 and S4). Along bgPC2,  
492 *Epipliopithecus*, *Victoriapithecus* and *Ekembo* are found on slightly negative scores and  
493 overlap with platyrrhines, cercopithecids and hylobatids, whereas *Pliobates* has a slightly  
494 positive score that further overlaps with hominids (Figs. 5e and 6a; Table 4). The canal  
495 configuration of *Pliobates* is thus somewhat more posteriorly located than in the other  
496 extinct taxa (SOM Figs. S4–S6), particularly *Epipliopithecus*.

497 Z-scores (Table 5) indicate that, for bgPC1, *Epipliopithecus* significantly differs from all  
498 extant groups, while other extinct taxa match the hominid distribution only. In contrast, for  
499 bgPC2, *Epipliopithecus*, *Victoriapithecus* and *Ekembo* only significantly differ from hominids,  
500 whereas *Pliobates* matches the variation of the four extant groups. When both bgPCs are  
501 considered simultaneously, the analyzed extinct taxa differ from the four extant anthropoid

502 groups except for *Pliobates*, which matches the hominid variation. This is confirmed by  
503 posterior probabilities of group membership (Table 8), according to which *Epipliopithecus*,  
504 *Victoriapithecus*, and *Ekembo* are classified as cercopithecoids—although *Epipliopithecus*  
505 falls outside their range of variation ( $p = 0.018$ )—whereas *Pliobates* is classified as a hominid  
506 and only significantly differs from platyrrhines. According to these results, the canal course  
507 of both *Ekembo* and *Victoriapithecus* fit better the cercopithecoid variation, whereas  
508 *Pliobates* fits better with hominids, and *Epipliopithecus* does not fit well with any of the  
509 extant groups analyzed. This notwithstanding, the results of the NJ cluster analysis (Fig. 8)  
510 indicate that *Pliobates* and *Epipliopithecus* are most similar to one another. Indeed, the four  
511 analyzed extinct taxa are grouped together in a cluster of their own, distinct from the  
512 subcluster of great apes and that of platyrrhines.

513 The discrimination among the catarrhine families in the catarrhine-only bgPCA is slightly  
514 better than in the analysis based on the whole sample but the three groups still overlap to  
515 an important extent (SOM Fig. S7). The first axis explains most of the variance (85.33%) and  
516 is the only one that discriminates between hominids and the remaining groups—as in the  
517 whole sample analysis, in which only one axis (bgPC2) discriminates between them. The  
518 results for the fossils in the catarrhine-only analysis are not more conclusive than for the  
519 analysis including platyrrhines, with only minor differences. In particular, *Epipliopithecus* falls  
520 closer to the hylobatid centroid and is classified as a hylobatid ( $p = 0.54$ ), although it displays  
521 no significant differences with either cercopithecoids ( $p = 0.30$ ) or hominids ( $p = 0.29$ );  
522 *Victoriapithecus* still matches better the cercopithecoid range of variation ( $p = 0.22$ ), as it is  
523 the case for *Ekembo* ( $p = 0.73$ ), while *Pliobates* is classified first as a hominid ( $p = 0.39$ ) but  
524 also falls within the range of hylobatids ( $p = 0.14$ )—apparently owing to the more hominid-  
525 like condition of *Symphalangus* as compared to other hylobatids.

526 Phylomorphospace The results of the phylomorphospace (Fig. 9; SOM Fig. S8) and the  
527 reconstructed ancestral morphotypes (Fig. 10; SOM Fig. S9) are virtually identical  
528 irrespective of the hypothesis considered. The crown platyrrhine LCA displays a very long  
529 canal that originates inferiorly and is located very laterally in the petrosal (Fig. 10e; SOM Fig.  
530 S9e), whereas the crown catarrhine LCA displays a condition almost indistinguishable from  
531 the mean anthropoid configuration (Fig. 10f; SOM Fig. S9f). The inferred crown  
532 cercopithecoïd (Fig. 10g; SOM Fig. S9g) and crown hominoid (Fig. 10h; SOM Fig. S9h)  
533 ancestral morphotypes are not very divergent from one another or from the catarrhine LCA,  
534 but both are more derived than the latter by displaying a somewhat shorter and more  
535 medially and posteriorly located canal. The crown hylobatid (Fig. 10i; SOM Fig. S9i) and  
536 crown hominid (Fig. 10j; SOM Fig. S9j) LCAs more clearly diverge from the crown hominoid  
537 LCA in different directions. Thus, the hylobatid LCA displays a higher bgPC1 score  
538 overlapping with extant hylobatids and cercopithecoïds, due to a shorter and more medially  
539 located canal. In contrast, the hominid LCA displays a higher bgPC2 than the crown hominoid  
540 LCA (like the extant hylobatid *Symphalangus*) due to a more posteriorly located canal.

541 The four analyzed extinct taxa are broadly similar to the crown catarrhine LCA's condition  
542 (Fig. 9). Indeed, both *Victoriapithecus* (Fig. 10d) and *Ekembo* (Fig. 10c) most closely resemble  
543 the crown catarrhine LCA rather than the crown ceropithecoïd (Fig. 10g) or hominoid (Fig.  
544 10h) LCAs. *Epipliopithecus* (Fig. 8b) displays a slightly more platyrrhine-like condition in  
545 bgPC1 but is nevertheless very distinct from the crown platyrrhine LCA (Fig. 10e) and closer  
546 to the inferred crown catarrhine ancestral condition. *Pliobates* (Fig. 10a) closely resembles  
547 the crown catarrhine LCA in canal length and carotid foramen/canal superoinferior and  
548 mediolateral position (as reflected in bgPC1), but unlike *Victoriapithecus* and *Ekembo* it  
549 appears derived along bgPC2 toward the reconstructed ancestral hominid condition—albeit



550 being more similar along this axis to *Symphalangus* than to extant great apes—due to its  
551 more posteriorly located canal.

552

## 553 **4. Discussion**

### 554 *4.1. Carotid canal orientation*

555 Two different characters were defined by Alba et al. (2015b: Tables S5 and S6) to code  
556 the carotid canal morphology, one describing the canal ‘orientation’ (more or less  
557 horizontal) and another (following Shoshani et al., 1996) describing the canal ‘direction’  
558 (more or less anterior). In Alba et al. (2015b), canal ‘orientation’ was defined as the  
559 inclination between the canal and the basioccipital, whereas canal ‘direction’ was coded  
560 based on the spatial relationship between the imaginary line emerging from the canal and  
561 the foramen magnum. Canal ‘direction’ sensu Alba et al. (2015b) could be anterior or  
562 posterior relative to the foramen magnum, i.e., more laterally or more anteriorly directed,  
563 respectively. To evaluate this character, previous authors (Shoshani et al., 1996) placed a  
564 straight wire inside the carotid canal and noted the point of intersection between the  
565 imaginary line along the wire and the foramen magnum. In Alba et al. (2015b) on the other  
566 hand, this character was evaluated conducting visual inspections on  $\mu$ CT scans, drawing the  
567 course of the canal on basicranium inferior view and noting the point of intersection  
568 between the imaginary line in continuation of the canal course and the foramen magnum.  
569 2DYZA (canal superior orientation in the present study) is comparable with Alba et al.’s  
570 (2015b) canal ‘orientation’ because this angle is measured along the parasagittal plan,  
571 although based on values obtained we prefer to categorize them as  
572 oblique/subvertical/vertical (as opposed to horizontal/oblique/subvertical as in Alba et al.,  
573 2015b). In contrast, 2DXYA (canal medial orientation in the present study) is not directly

574 comparable with Alba et al.'s (2015b) canal 'direction', because while both studies rely  
575 primarily on the line passing through the two canal endpoints (i.e., whole carotid canal), in  
576 the present study the alignment process and the quantitative assessment of this character  
577 (i.e., possibility of having more than two states vs. two qualitative states only in Alba et al.  
578 [2015b]) make that this measure of canal orientation, for a same canal specimen, might  
579 differ between the two studies.

580 *Pliobates* was coded by Alba et al. (2015b) as displaying a horizontal orientation  
581 parallel to the basioccipital, like hylobatids, whereas the stem catarrhine *Saadanius*,  
582 cercopithecoids (including *Victoriapithecus*) and hominids were coded with a subvertical  
583 orientation, and *Epipliopithecus* and *Ekembo* with an intermediate (oblique) condition. Our  
584 results for superior canal orientation (2DYZA) indicate that, in this regard, *Pliobates* does not  
585 differ from hylobatids and display a lower value than *Ekembo* and *Epipliopithecus*. However,  
586 our results further show that both hylobatids and hominids display a rather oblique  
587 orientation (i.e., a horizontal orientation parallel to the basioccipital using the terminology of  
588 Alba et al., 2015b), with *Ekembo* and *Epipliopithecus* resembling hominoids as a whole.  
589 Nonetheless, all these taxa possess a less vertical orientation than platyrrhines (i.e., a  
590 subvertical canal) and, especially, cercopithecoids (including *Victoriapithecus*), in which the  
591 canal is vertical. To sum up, according to Alba et al. (2015b) hylobatids and *Pliobates* had a  
592 horizontal canal, *Ekembo* and *Epipliopithecus* an oblique canal, and cercopithecoids  
593 (including *Victoriapithecus*) and hominids a subvertical canal, whereas according to our  
594 results all hominoids (including *Ekembo*), *Epipliopithecus*, and *Pliobates* have an oblique  
595 canal, platyrrhines a subvertical canal, and cercopithecoids (including *Victoriapithecus*) an  
596 even more vertical canal. Therefore, the superior orientation of the carotid canal is

597 consistent with the cercopithecoid status of *Victoriapithecus*, but it is not informative  
598 regarding the stem catarrhine vs. stem hominoid status of *Pliobates*.

599 In turn, *Pliobates* was coded by Alba et al. (2015b) as resembling *Epipliopithecus* and  
600 hylobatids in the 'direction' of the canal. The condition of all these taxa was coded as  
601 anteriorly directed, the imaginary line emerging from the carotid canal crossing the foramen  
602 magnum more posteriorly than in *Saadanius*, cercopithecids, and hominids, whose canal  
603 would be more laterally directed, as in platyrrhines (see also Shoshani et al., 1996). In  
604 contrast, our results indicate that hylobatids extensively overlap in canal medial orientation  
605 with other extant catarrhines, which only significantly differ from platyrrhines. Furthermore,  
606 *Pliobates* does not specifically resemble hylobatids or *Epipliopithecus*, but rather crown  
607 catarrhines as a whole, including *Victoriapithecus*. On the other hand, *Epipliopithecus* and,  
608 especially, *Ekembo*, exhibit a more platyrrhine-like condition, i.e., a medially directed canal.  
609 To sum up, our results indicate that cercopithecoids (including *Victoriapithecus*), crown  
610 hominoids, and *Pliobates* have an anteriorly oriented canal, whereas platyrrhines, *Ekembo*,  
611 and *Epipliopithecus* have a medially oriented canal.

612

#### 613 4.2. Carotid canal proportions

614 Volumetric proportions—which were not considered by Alba et al. (2015b)—as  
615 measured by the robusticity index of the carotid canal, further confirm the differences in  
616 carotid canal morphology between platyrrhines and catarrhines. Our allometric analyses  
617 indicate that these differences cannot be explained by body size-scaling effects alone. In  
618 particular, the volume of the carotid canal displays an isometric relationships with body size,  
619 whereas length displays a slight negative allometric relationship with the latter. As a result,  
620 larger-bodied taxa tend to display lower values of the robusticity index (i.e., slightly stouter

621 carotid canals). However, correlation with body mass only explains 14% of the variance in  
622 carotid canal robusticity.

623         Based on our results, platyrrhines display on average slenderer canals (i.e., lesser  
624 volume relative to length) than catarrhines. However, as for medial canal orientation,  
625 catarrhine families largely overlap in canal proportions, although great apes display less  
626 robust canals than cercopithecids and hylobatids, and are somewhat intermediate between  
627 them and platyrrhines. The analyzed fossil catarrhines more closely resemble each other in  
628 canal proportions than in canal orientation—being catarrhine-like but further overlapping  
629 with the platyrrhine range of variation. *Pliobates* displays somewhat slenderer canals than  
630 the other extinct taxa and, unlike them, it does not overlap with the hylobatids, even more  
631 clearly than in the case of medial canal orientation thus further reinforcing the rejection of  
632 possible closer similarities with hylobatids the latter family.

633         Our results for the extant taxa are in broad agreement with those previously obtained  
634 by Boyer et al. (2016) on relative internal carotid artery area. Boyer et al. (2016) found  
635 internal carotid area to scale isometrically with cranial area (measured as the square root of  
636 the product between prosthion-inion length and bizygomatic breadth). Among catarrhines,  
637 these authors found a pattern of decreasing robusticity in relative carotid artery area from  
638 hylobatids to hominids, and to cercopithecids, which is the same as we found for the carotid  
639 canal as a whole. The main difference between our results and those of Boyer et al. (2016) is  
640 that, in the latter study, the carotid artery robusticity of platyrrhines was found to be  
641 intermediate between hylobatids and hominids, whereas our results show that platyrrhines  
642 have slenderer canals than catarrhines. This discrepancy is easily explained by the fact that  
643 Boyer et al. (2016) considered internal carotid artery area relative to cranial area, whereas  
644 we relied on the ratio between the length and the volume of the canal. Our results (see

645 below for further discussion) indicate that platyrrhines display relatively longer carotid  
646 canals than catarrhines, thereby resulting in less robust volumetric proportions for the canal  
647 as a whole. Given that Boyer et al. (2016) did not take canal length into account, this  
648 explained the above-mentioned discrepancy for platyrrhines (but not the other groups) as  
649 compared to our results.

650

#### 651 4.3. Carotid canal course

652 Our bgPCA results indicate that the morphology of the canal embeds strong phylogenetic  
653 signal, although most of the variance merely discriminates between platyrrhines and  
654 catarrhines, due to differences in canal length and location reflected in bgPC1. In particular,  
655 extant catarrhines display shorter canals that originate more superiorly than in platyrrhines.  
656 It is noteworthy to stress beforehand that *Pliobates* and hylobatids do not show particular  
657 similarities in bgPC1 scores.

658 Previous studies suggested that the carotid foramen is located ventrally (Ross, 1994; Ross  
659 et al., 1998) and posteriorly (Ross, 1994; Ross et al., 1998; Kay et al., 2008) in both  
660 platyrrhines and catarrhines, whereas Alba et al. (2015b) noted some differences in the  
661 anteroposterior carotid foramen location between hylobatids and other catarrhines (with  
662 the former displaying a more anteriorly located carotid foramen). Previous studies also  
663 noted that platyrrhines display a more medially located foramen than catarrhines (Ross et  
664 al., 1998; Kay et al., 2008). Our bgPC1 results indicate that extant platyrrhines and  
665 catarrhines differ not only regarding the length of the canal, but also the location of the  
666 (external) carotid foramen, such that in catarrhines the entire carotid canal (i.e., not only the  
667 foramen but the whole structure) is more superiorly and medially located within the  
668 petrosal. Our results therefore point to differences between platyrrhines and catarrhines in

669 terms of dorsoventral location of the carotid foramen, contrary to previous studies (Ross,  
670 1994; Ross et al., 1998). This might be attributable to the different methodologies employed  
671 (i.e., because of having performed an alignment on the specimens in the present study).  
672 Similarly, because of the alignment, our conclusions for the location along the mediolateral  
673 axis might be the opposite as those noted by previous authors (Ross et al., 1998; Kay et al.,  
674 2008), who simply recorded foramen position relative to the bulla. Since catarrhines possess  
675 a tubular ectotympanic, catarrhine landmark configurations are shifted more medially. This  
676 is the consequence of alignment optimization of their tympanic landmarks with those of  
677 platyrrhines, whose annular ectotympanic causes their tympanic landmarks to be more  
678 medially positioned. Regardless, we observe an extensive overlap among extant catarrhine  
679 families in the position of the carotid foramen, at least based on bgPC1.

680 Our results for bgPC2 lend some support that hylobatids display on average a more  
681 anteriorly located foramen in the petrosal than hominids. However, rather than indicating a  
682 particular condition for hylobatids, our bgPC2 results suggest that only hominids differ in this  
683 regard from other catarrhines by displaying a more posteriorly located foramen. This  
684 conclusion should be taken with caution, given that bgPC2 shows an extensive overlap  
685 between hominids and the upper range of other crown catarrhines. Furthermore, bgPC2  
686 reflects a smaller amount of variance, embeds lesser phylogenetic signal than bgPC1, and is  
687 more strongly correlated with size—although the percentage of variance explained by  
688 allometry varies depending on whether canal size or body size is considered, our results  
689 clearly indicate that this axis is more strongly influenced by size-scaling than bgPC1.  
690 Therefore, one should be cautious when interpreting similarities among taxa based on  
691 bgPC2 scores.

692 Our 3DGM results for the analyzed fossil taxa do not support particular similarities  
693 between *Pliobates* and hylobatids in terms of overall carotid canal course (contra Alba et al.,  
694 2015b), and indicate instead that *Pliobates* is most similar in this regard to *Epipliopithecus*,  
695 *Victoriapithecus* and *Ekembo*. All these fossil catarrhines appear overall closer to extant  
696 catarrhines in terms of canal length and location in the petrosal, albeit being somewhat  
697 intermediate between platyrrhines and catarrhines. *Epipliopithecus* appears slightly more  
698 platyrrhine-like, whereas the other extinct genera only overlap with the lowest range of  
699 cercopithecids and hominids (but not hylobatids). *Pliobates* only differs slightly from the  
700 remaining fossil catarrhines by more closely resembling hominids and *Symphalangus* along  
701 bgPC2 (reflecting a more posteriorly located foramen/canal). As noted above, bgPC2 is quite  
702 strongly correlated with CS, suggesting that the differences between hominids and the rest  
703 of the sample could merely result from the larger body size of the former. However, this  
704 cannot explain the differences along this axis between *Pliobates* and *Epipliopithecus*, as the  
705 former more closely resembles the large-bodied hominids and the largest-bodied hylobatid  
706 (*Symphalangus*) despite a much lower estimated body size than *Epipliopithecus* (4–5 kg vs.  
707 11–12 kg, respectively; Alba et al., 2015b).

708

#### 709 4.4. Evolutionary implications

710 Alba et al. (2015b) concluded that *Pliobates* displays a mosaic of plesiomorphic (stem  
711 catarrhine-like) cranial and postcranial features and more derived (crown hominoid) cranial  
712 and postcranial synapomorphies, coupled with a dendropithecoid-like dentition and some  
713 cranial similarities with hylobatids. A stem hominoid status more derived than that of  
714 *Ekembo* was supported for *Pliobates* by Alba et al. (2015b) based on the results of their

715 cladistic analysis. Such a phylogenetic placement would support the view that some  
716 purported synapomorphies of crown catarrhines, best exemplified by the completely  
717 ossified tubular ectotympanic, would have been independently acquired independently in  
718 cercopithecoids and hominoids—a possibility taken into account by some previous authors  
719 (Begun, 2002). A stem hominoid status for *Pliobates* would further support that the last  
720 common ancestor of crown hominoids would have been more hylobatid-like than previously  
721 assumed (Alba et al., 2015b). Interestingly, one of the resemblances between *Pliobates* and  
722 hylobatids found by Alba et al. (2015b) was the carotid canal morphology, which is at odds  
723 with the results of our more elaborate analyses of this anatomical structure, which denote a  
724 generalized catarrhine morphology with some closer similarities with extant hominids and  
725 uniquely *Symphalangus* among hylobatids.

726       Contrary to Alba et al. (2015b), subsequent cladistic analyses—based on different taxon-  
727 character matrix—have recovered *Pliobates* as a stem catarrhine more closely related to  
728 pliopithecoids (Nengo et al., 2017; Gilbert et al., 2020b). Based on these results, Gilbert et al.  
729 (2020a: 399) concluded that “*Pliobates* is most likely a derived, late-occurring pliopithecoid  
730 or other stem catarrhine taxon that has converged on some interesting but minor details of  
731 extant hominoid elbows and wrists”. In fact, the similarities in elbow and wrist morphology  
732 between *Pliobates* and hominoids are not minor but quite extensive, more so than those  
733 displayed by *Ekembo* and certainly than those convergently acquired by atelids (Alba et al.,  
734 2015b). However, it is not unconceivable that these similarities—being functionally related  
735 to enhanced forearm rotation and ulnar deviation capabilities—might have independently  
736 evolved between *Pliobates* and crown hominoids. Under such scenario, in which postcranial  
737 similarities between *Pliobates* and crown hominoids would be dismissed as convergences, it  
738 should be taken into account that the dental morphology of *Pliobates* suggests much closer



739 affinities to dendropithecids (particularly *Micropithecus*) than to pliopithecoids (Alba et al.,  
740 2015b). A dendropithecoid status for *Pliobates* would not imply an independent acquisition of  
741 a fully-ossified ectotympanic, but would not automatically resolve the phylogenetic  
742 placement of the former, given the controversies surrounding the phylogenetic relationships  
743 of dendropithecids—considered either stem catarrhines more derived than pliopithecoids  
744 (Harrison, 2010, 2013, 2017; Nengo et al., 2017; Gilbert et al., 2020b) or stem hominoids  
745 more basal than proconsulids (Rae, 1999; Zalmout et al., 2010; Begun, 2015; Alba et al.,  
746 2015b; Rossie and Hill, 2018).

747 Unfortunately, the carotid canal morphology of dendropithecids is currently unknown,  
748 thereby precluding a direct comparison with *Pliobates*. Furthermore, despite substantial  
749 differences between platyrrhines and catarrhines in terms of carotid canal orientation and  
750 proportions as well as carotid canal length and (foramen) location in the petrosal, our results  
751 indicate a considerable overlap among extant catarrhines. As a result, carotid canal  
752 morphology is of limited utility for investigating the phylogenetic position of fossil  
753 catarrhines, although some insights can be drawn for the investigated taxa. Before  
754 discussing the evolutionary implications of our results, however, it should be noted that the  
755 small sample sizes available for extinct taxa make it necessary interpret the differences  
756 among the analyzed fossil specimens by assuming that they are more or less representative  
757 of their respective taxa. Although this caveat is not restricted to the carotid canal  
758 morphology, it should not be forgotten that we cannot know a priori how similar in carotid  
759 canal morphology a particular fossil specimen is to the (unknown) average condition of the  
760 taxon to which it belongs. For example, given the considerable variation that can be  
761 observed in some extant taxa for carotid canal course (e.g., *Aotus*, *Macaca*, *Mandrillus*), the  
762 possibility remains that some of the differences inferred among extinct taxa, or between

763 them and extant catarrhine clades, might vanish (or become more accentuated) if additional  
764 specimens became available. In the meantime, caution is required by bearing in mind that  
765 the reliability of inferences for fossil taxa is limited by the number of specimens available to  
766 them, which often preclude an adequate assessment of intraspecific variability.

767       Based on the currently available specimens, our results indicate that the stem hominoid  
768 *Ekembo* and the stem cercopithecoid *Victoriapithecus* are virtually identical in carotid canal  
769 volumetric proportions and course morphology, although *Ekembo* displays a more  
770 platyrrhine-like medial orientation (like *Epipliopithecus*) and a more hominoid-like superior  
771 orientation (like *Epipliopithecus* and *Pliobates* to some extent). The less medially/more  
772 anteriorly oriented carotid canal in *Pliobates* than in *Ekembo* would be consistent with a  
773 more derived stem hominoid status, but given the modern cercopithecoid-like carotid canal  
774 orientation of *Victoriapithecus*, the condition of extant cercopithecoids and hominoids might  
775 have evolved independently to some extent. In turn, based on the condition of  
776 *Epipliopithecus*, the possession of an obliquely oriented canal in hominoids might represent  
777 a catarrhine symplesiomorphy; in that case, the condition of extant cercopithecoids would  
778 have independently evolved from that of extant platyrrhines. Alternatively, the oblique  
779 orientation of the canal might be synapomorphic for hominoids, in which case the similar  
780 condition of *Epipliopithecus* would be a convergence. Either way, this feature is not  
781 informative regarding the phylogenetic position of *Pliobates*.

782       In terms of carotid canal course, as reflected by our 3DGM analysis, *Pliobates* does not  
783 show particular affinities with hylobatids, and rather resembles the other analyzed extinct  
784 taxa, which are overall more similar to catarrhines but somewhat intermediate between  
785 platyrrhines and both cercopithecoids and hominids. The phylomorphospace shows that both

786 *Victoriapithecus* and *Ekembo* apparently reflect well the inferred ancestral crown catarrhine  
787 morphotype, despite their generally accepted crown catarrhine (respectively, stem  
788 cercopithecoid and stem hominoid) status (e.g., Benefit and McCrossin, 1997; Benefit, 1999;  
789 Alba et al., 2015b; Nengo et al., 2017; Gilbert et al., 2020b). In contrast, *Epipliopithecus*  
790 appears somewhat more plesiomorphic, whereas *Pliobates* appears slightly derived (in the  
791 anteroposterior position of the carotid foramen/canal) toward the great ape condition,  
792 being closer to the inferred ancestral hominid morphotype as well as the hylobatid  
793 *Symphalangus*.

794 In the case of *Epipliopithecus*, our results are in agreement with the widely held opinion  
795 that pliopithecoids are a clade of stem catarrhines, predating the divergence of  
796 cercopithecoids and hominoids (Harrison, 2013; Begun, 2002, 2017; Nengo et al., 2017;  
797 Gilbert et al., 2020a, b; Sankhyan et al., 2017; Harrison et al., 2020; Urciuoli et al., 2021;  
798 contra Alba et al., 2015b). For *Pliobates*, our results are more ambiguous and subject to  
799 interpretation, although it is noteworthy that they do not support a closer relationship with  
800 pliopithecoids—at least, based on the information provided by *Epipliopithecus*. They suggest  
801 instead that, in carotid canal morphology, *Pliobates* is more derived than *Epipliopithecus*  
802 toward the inferred ancestral crown catarrhine morphotype, like *Ekembo* and  
803 *Victoriapithecus*. Furthermore, our results discount very close similarities in carotid canal  
804 morphology between *Pliobates* and hylobatids as a whole, but indicate some resemblances  
805 with the hylobatid *Symphalangus* and great apes. In the latter regard, *Pliobates* is  
806 approximately equidistant from the crown catarrhine and crown hominoid reconstructed  
807 morphotypes. It is uncertain whether this reflects a more derived status of *Pliobates* or an  
808 independent development, although the latter is plausible in light of the *Symphalangus*  
809 condition—if the hylobatid LCA reconstructed in the phylomorphospace is broadly correct,

810 *Symphalangus* must have independently acquired a more great ape-like condition;  
811 alternatively, the condition shared by the hominid LCA and *Symphalangus* might be  
812 plesiomorphic for crown hominoids and the reconstructed hylobatid LCA would reflect a  
813 secondary reversal of the remaining hylobatids. Of course, the morphology of a single  
814 anatomical region cannot provide much insight on the phylogenetic position of a given taxon  
815 among catarrhines, particularly if—as it is the case of the carotid canal—it does not  
816 adequately discriminate among catarrhine clades. However, our results for this structure do  
817 not show any particular resemblances between pliopithecoids and *Pliobates* (beyond those  
818 displayed by all the extinct taxa analyzed), and further fail to contradict the alternative  
819 phylogenetic status of *Pliobates* as a stem hominoid. The phylogenetic status of *Pliobates*  
820 thus seems far from being settled.

821

## 822 **5. Summary and conclusions**

823 Previous research on the primate carotid canal within a phylogenetic context has been  
824 limited to the inclusion of qualitative features related to carotid foramen position in the  
825 bulla and overall direction and orientation of the canal itself in character-taxon matrices  
826 devised for cladistic analysis (Beard and MacPhee, 1994; Ross, 1994; Shoshani et al., 1996;  
827 Ross et al., 1998; Kay et al., 2008; Alba et al., 2015b). From a quantitative viewpoint, only  
828 Boyer et al. (2016) investigated the allometric relationship between carotid canal area and  
829 BM (Boyer et al., 2016), whereas some authors relied on carotid canal area for inferring  
830 encephalic blood flow rates and blood flow metabolism by further taking vertebral canal size  
831 into account (Boyer and Harrington, 2018, 2019; Beaudet et al., 2020). Thus, our study is the  
832 first to rigorously quantify the orientation, direction, overall course and volumetric

833 proportions of the carotid canal in 3D among a large sample of anthropoids. The allometric  
834 regressions performed also represent an advancement in the understanding of body size-  
835 scaling effects on carotid canal morphology. The methodological protocols devised here  
836 enable repeatability among different researchers and thus will enable a refinement of our  
837 conclusions by adding additional extant and fossil specimens in the future. The analyzed  
838 variables are no panacea for clarifying the phylogeny of extinct catarrhines, because various  
839 extant groups overlap to a large extent and also because intraspecific variation will be  
840 difficult to evaluate due to small samples sizes. However, the analyzed variables offer the  
841 prospect to provide taxonomically and phylogenetically relevant information for extinct taxa,  
842 particularly if they are added to character-taxon matrices devised for cladistic analysis  
843 including information from other anatomical areas. Other future directions of work on the  
844 carotid canal should deepen our current understanding about the influence of function, as  
845 well as allometric and phylogenetic constraints, on the evolution of this anatomical  
846 structure. This would be required not only to better interpret carotid canal morphology from  
847 an adaptive viewpoint, but also to derive from it paleobiological inferences for extinct taxa.

848 In this study, we rely on 3D morphometric techniques to quantify the orientation,  
849 proportions, and course of the carotid canal in the small-bodied catarrhine *Pliobates*, and  
850 compare it with extant anthropoids and other Miocene catarrhines (*Epipliopithecus*,  
851 *Victoriapithecus*, and *Ekembo*). Our results reveal that *Pliobates* and the other analyzed  
852 Miocene catarrhines are broadly similar to one another in carotid canal morphology and  
853 more closely resemble extant catarrhines than platyrrhines. Unfortunately, carotid canal  
854 morphology does not adequately discriminate among catarrhine clades, thereby hindering a  
855 more detailed phylogenetic assessment of *Pliobates* on this basis. The difficulty of assessing  
856 intraspecific variation in carotid canal morphology in the extinct taxa, coupled with the lack

857 of marked differences among extant catarrhine families, make it impossible to reach  
858 definitive conclusions about the systematic position of *Pliobates* on this basis alone.  
859 However, our results rule out the previously noted similarities in carotid canal morphology  
860 between *Pliobates* and hylobatids and indicate instead that this extinct catarrhine is broadly  
861 similar to catarrhines as a whole and, in particular, to other Miocene catarrhines (both stem  
862 and crown).

863 It is however noteworthy that the carotid canal of *Pliobates* is not particularly similar to  
864 that of pliopithecoids (as represented by *Epipliothecus*, which appears somewhat more  
865 platyrrhine-like), thereby not supporting (or contradicting) the hypothesis that *Pliobates* is a  
866 pliopithecoid. Like the stem hominoid *Ekembo* and the stem cercopithecoid *Victoriapithecus*,  
867 *Pliobates* appears somewhat more derived than *Epipliothecus* toward the modern  
868 catarrhine condition, and indeed, in some respects, *Pliobates* appears even more derived  
869 toward the ancestral hominoid morphotype than the remaining Miocene catarrhines  
870 analyzed, more closely resembling extant hominids and the hylobatid *Symphalangus*.  
871 Therefore, our results do not enable us to discount the hypothesis that *Pliobates* is a stem  
872 hominoid rather than a stem catarrhine. Given the dental similarities between *Pliobates* and  
873 African dendropithecids, the former might be a late offshot of the latter clade that dispersed  
874 into Eurasia independently from pliopithecoids. Unfortunately, this possibility could not be  
875 directly assessed in this study because the carotid canal morphology of dendropithecids is  
876 unknown. Additional analyses focused on other anatomical (both cranial and postcranial)  
877 areas will be required to further clarify the phylogenetic relationships of *Pliobates*, as well as  
878 to clarify whether its postcranial similarities with crown hominoids might have been  
879 independently acquired.

880

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912

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1129

### 1130 **Figure captions**

1131

1132 **Figure 1.** Schematic illustration of the protocol used in this study for cutting the extremities  
1133 of the carotid canal as exemplified in a *Macaca fascicularis* individual. a) To cut the posterior  
1134 end, a best-fit plane defined for the landmarks placed at the carotid foramen is computed. b)  
1135 To cut the anterior end, the 3D canal surface is first imported as a cloud of points in R. c) The

1136 projection of IRG on the cloud of points (IRG') is computed. d) 360 points are created around  
1137 the axis IRG–IRG'. e, f) 360 possible cutting planes passing through IRG are created (each  
1138 being based on IRG, IRG' and one of the points rotating around the axis), here depicted in  
1139 anterior (e) and superior (f) views. The cross-sectional area obtained by cutting the canal  
1140 using each possible plane is calculated as follows: g–i) the points of the cloud belonging to  
1141 the plane are recorded (g), the most external points (in red) are kept to create a convex hull  
1142 that approximates the shape of the canal contour (h), and a set of triangles is drawn, each of  
1143 them having the same point of the contour as the most acute vertex, and the two  
1144 subsequent points on the contour as the other vertices (i); the total area is the obtained by  
1145 summing those of each individual triangle. j, k) The plane with the smallest cross-sectional  
1146 area (that is, orthogonal to the 3D canal surface and therefore to the canal streamline) (j) is  
1147 used as a reference for cutting the canal at its anterior end (k). l) The centroids of the  
1148 (posterior) carotid foramen landmark set and of the anterior cross-section (i.e., obtained by  
1149 cutting with the orthogonal plane), representing the posterior and anterior endpoints of the  
1150 carotid canal, respectively, are calculated. IRG = Intersection ridge–groove; IRG' = orthogonal  
1151 projection on the 3D carotid canal surface of IRG.

1152

1153 **Figure 2.** Petrosal landmarks used for aligning the specimens as exemplified in a *Macaca*  
1154 *fascicularis* individual in different views: a) right lateral; b) inferior; c) sagittal cross section  
1155 (right-mirrored for visualization purpose); d) superior transverse cross section. Landmarks: 1  
1156 = porion; 2 = inferior external auditory meatus; 3 = posterior external auditory meatus; 4 =  
1157 posterior internal auditory meatus; 5 = anterior internal auditory meatus; 6 = medial  
1158 anterior pyramidal ridge; 7 = intersection ridge-groove; 8 = greater superficial petrosal nerve  
1159 hiatus; 9 = stylomastoid foramen.

1160

1161 **Figure 3.** Graphic representation of the 2D angles defining the carotid canal orientation  
1162 relative to the cranium. a) Virtual external reconstructions of the cranium, in superior (left)  
1163 and lateral (right) views. b) Virtual cross sections of the cranium to visualize the petrosal  
1164 bone, in transverse (left) and sagittal (right) views. c) Representations of the carotid canal  
1165 medial orientation (2DXYA; left) and superior orientation (2DYZA; right) within the cranium,  
1166 rendered in semitransparency; red lines represent the 2D canal vector and its direction,  
1167 orange squares represent the plane along which the angle can vary, and dotted black lines  
1168 represent the standard reference vector. For 2DXYA, the plane of angle variation is a  
1169 transverse plane, whereas for 2DYZA the plane of angle variation is a (para)sagittal plane.  
1170 For both angles, the standard reference vector is a 2D vector parallel to the anteroposterior  
1171 cranial axis (from posterior to anterior). Both angles are calculated between the 2D canal  
1172 vector and the 2D standard reference vector. Because the anthropoid mean configuration is  
1173 extremely similar to the canal course configuration of *Presbytis* (i.e., before alignment), the  
1174 cranium of this taxon is used as a reference for orienting the canal following the cranial axes  
1175 of reference.

1176

1177 **Figure 4.** Virtual reconstructions of carotid canal comparing *Pliobates* with extant  
1178 anthropoids and some Miocene catarrhines: a) *Pliobates* (IPS58443.1); b) *Epipliothecus*  
1179 (NHMW 1970/1397/0003); c) *Ekembo* (KNM-RU 2036al); d) *Victoriapithecus* (KNM-MB  
1180 29100); e) *Alouatta*; f) *Aotus*; g) *Cebus*; h) *Mico*; i) *Plecturocebus*; j) *Cercopithecus*; k)  
1181 *Colobus*; l) *Mandrillus*; m) *Presbytis*; n) *Hylobates*; o) *Hoolock*; p) *Nomascus*; q)  
1182 *Symphalangus*; r) *Gorilla*; s) *Pan*; t) *Pongo*. The canals are shown in medial view with their  
1183 posterior ends horizontally aligned. Scale bars = 5 mm.

1184

1185 **Figure 5.** Box-and-whisker plots of the variables describing the orientation, proportions and  
1186 course of the carotid canal in the fossil specimens and main extant clades analyzed in this  
1187 study: a) 2D YZ angle; b) 2D XY angle; c) index  $L/V^{(1/3)}$ ; d) bgPC1; e) bgPC2. Horizontal lines  
1188 denote medians, boxes depict the interquartile range, whiskers the minimum-maximum  
1189 values excluding outliers, dots denote outliers (beyond 1.5 times above or below the  
1190 interquartile range). Abbreviations: L = length (in mm); V = volume (in  $\text{mm}^3$ ); bgPC =  
1191 between-group principal component; 2DYZA = 2D angle in the YZ plane (canal superior  
1192 orientation; in degrees); 2DXYA = 2D angle in the XY plane (canal medial orientation; in  
1193 degrees).

1194

1195 **Figure 6.** Results of the between-group principal component analysis (bgPCA) based on  
1196 carotid canal course variation among main extant anthropoid clades. a) Bivariate plot of  
1197 bgPC2 vs. bgPC1 (the variance explained by each bgPC is denoted in parentheses). b)  
1198 Minimum (blue) and maximum (red) landmark conformations compared with the  
1199 anthropoid mean configuration (black) for bgPC1 and bgPC2, in superior (top), lateral  
1200 (middle), and anterior (bottom) views (anatomical axes of reference are specified next to  
1201 each view; see Fig. 7 for a representation of the carotid canal within the cranium). Convex  
1202 hulls correspond to: platyrrhines (blue), cercopithecids (red), hylobatids (green), and  
1203 hominids (gray). Abbreviation: bgPC = between-group principal component.

1204

1205 **Figure 7.** Representations of the carotid canal relative to the cranium in superior (left),  
1206 lateral (middle), and anterior (right) views. a) Virtual external reconstruction of the cranium.  
1207 b) Virtual cross sections of the cranium to visualize the petrosal bone (from left to right,

1208 transverse, sagittal, and coronal sections are depicted). c) Virtual reconstructions of the  
1209 carotid canal within the cranium rendered in semitransparency. d, e) Minimum (blue) and  
1210 maximum (red) landmark conformations compared with the anthropoid mean configuration  
1211 (black) for bgPC1 (d) and bgPC2 (e) within the cranium rendered in semitransparency.  
1212 Because the anthropoid mean configuration is extremely similar to the canal course  
1213 configuration of *Presbytis* (i.e., before alignment), the cranium of this taxon is used as a  
1214 reference for orienting the canal following the cranial axes of reference.

1215

1216 **Figure 8.** Neighbor-joining cluster (unrooted) based on the Euclidean distances calculated  
1217 from raw coordinates of the canal course configurations in *Pliobates*, extant anthropoids and  
1218 other Miocene catarrhines. Cophenetic = 0.94.

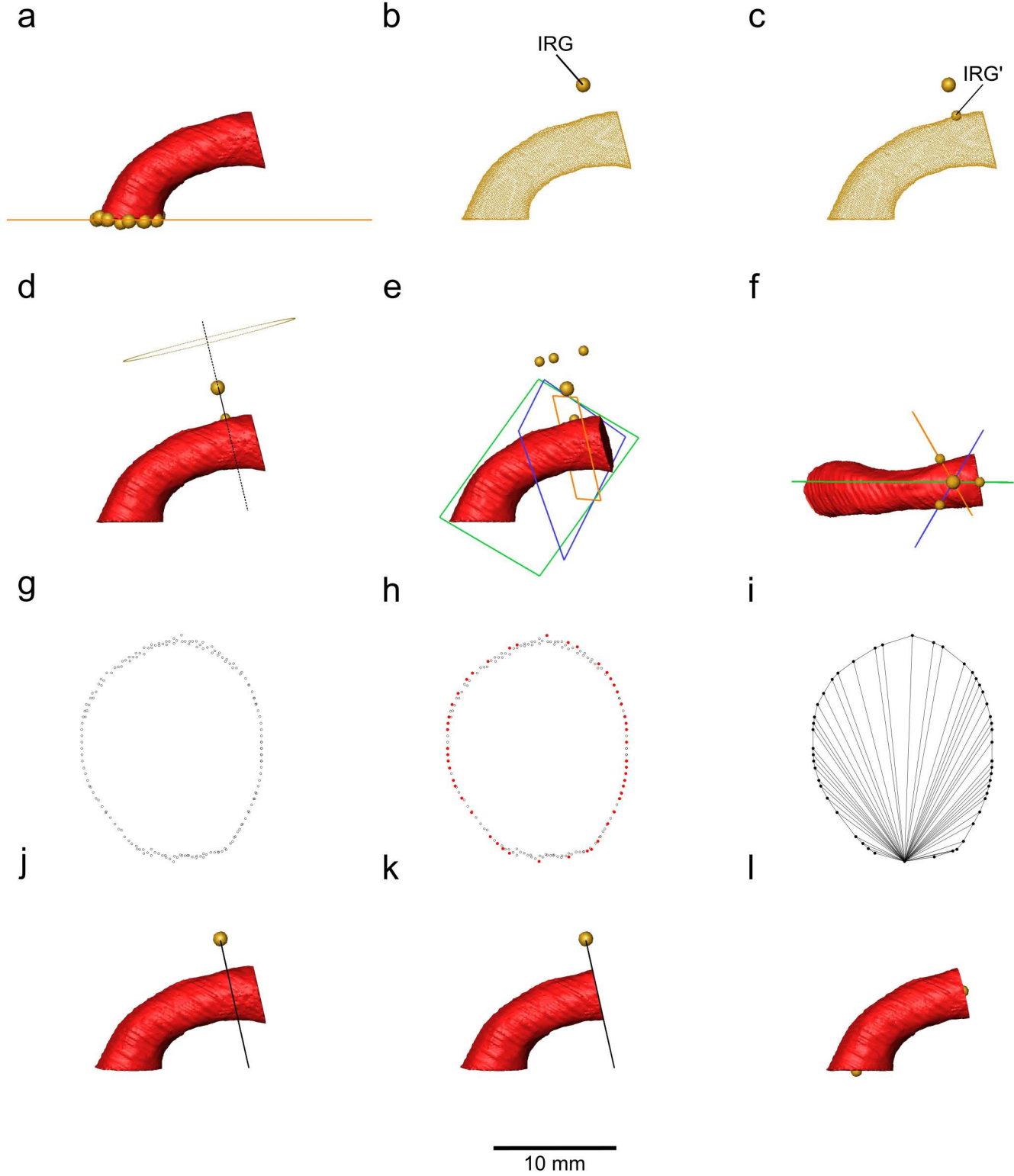
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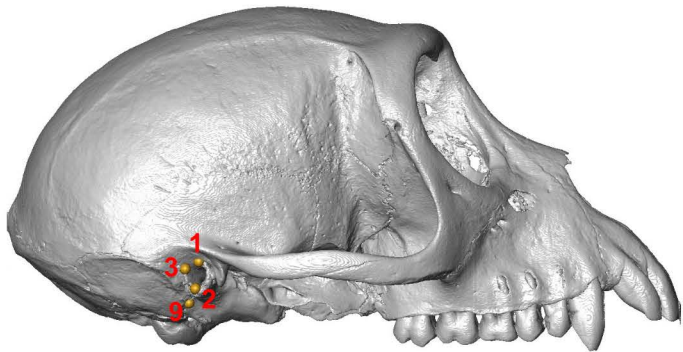
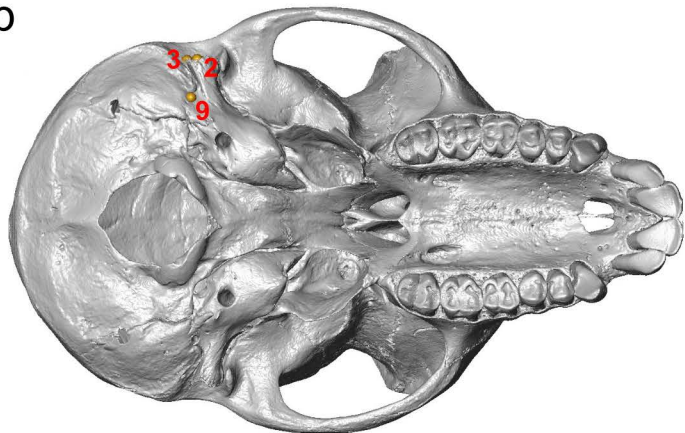
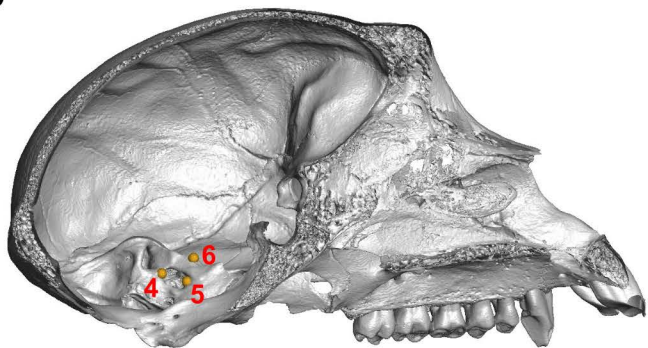
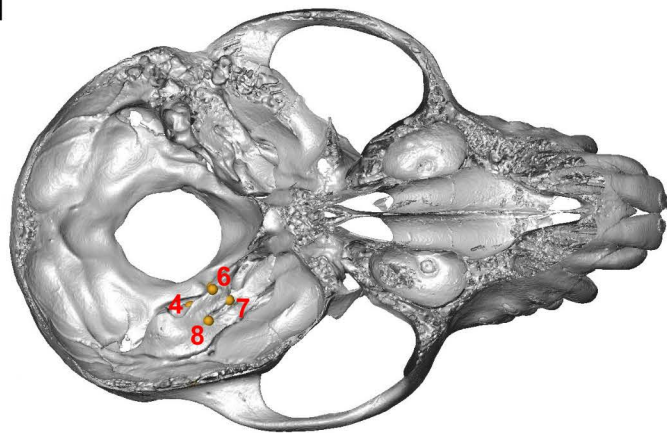
1220 **Figure 9.** Phylomorphospace of carotid canal course variation in extant anthropoids and  
1221 extinct Miocene catarrhines including *Pliobates*. *Epipliopithecus* and *Pliobates* are considered  
1222 sister taxa within a stem catarrhine pliopithecoid clade and *Ekembo* is considered a stem  
1223 hominoid while *Victoriapithecus* is considered a stem cercopithecoid (see SOM Fig. S2b for a  
1224 representation of this phylogenetic hypothesis). Ancestral nodes: 1 = crown platyrrhines; 2 =  
1225 crown catarrhines; 3 = crown cercopithecoids; 4 = crown hominoids; 5 = crown hominids; 6 =  
1226 crown hylobatids. See SOM Figure S8 for a phylomorphospace derived from the alternative  
1227 phylogenetic hypothesis (SOM Fig. S2b).

1228

1229 **Figure 10.** Configurations of the carotid canal course in *Pliobates* compared with other  
1230 Miocene catarrhines and the reconstructed carotid canal course for the last common  
1231 ancestors (LCAs) of various anthropoid clades: a) *Pliobates* (IPS58443.1); b) *Epipliopithecus*

1232 (NHMW 1970/1397/0003); c) *Ekembo* (KNM-RU 2036a); d) *Victoriapithecus* (KNM-MB  
1233 29100); e) crown platyrrhines; f) crown catarrhines; g) crown cercopithecoids; h) crown  
1234 hominoids; i) crown hylobatids; j) crown hominids. The canal courses of the LCAs have been  
1235 inferred based on the phylomorphospace displayed in Figure 9 (see SOM Fig. S9 for the  
1236 inferred LCA configurations derived from the alternative phylogenetic hypothesis). For each  
1237 specimen/LCA, the configurations are displayed in superior (left), lateral (middle), and  
1238 anterior (right) views. The mean configuration computed for the whole anthropoid sample is  
1239 represented by black dots, while configurations for Miocene catarrhines and LCAs are  
1240 represented in beige. The anthropoid mean configuration was used as a reference for  
1241 visualizing all the configurations at the same relative scale.  
1242

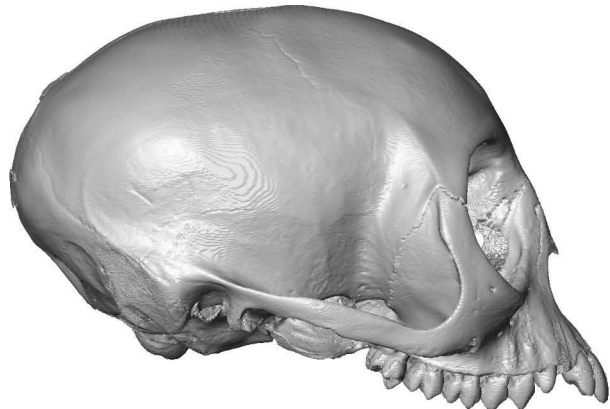
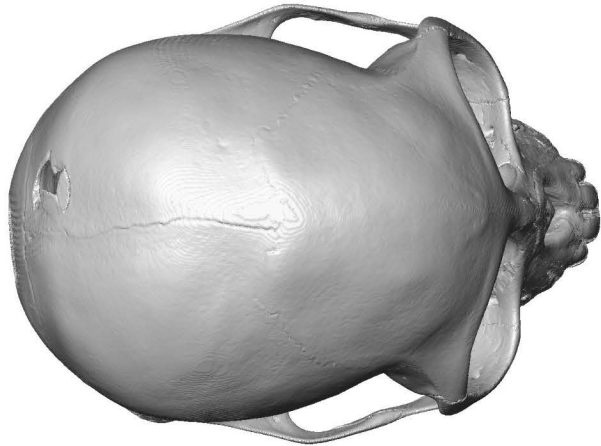


**a****b****c****d**

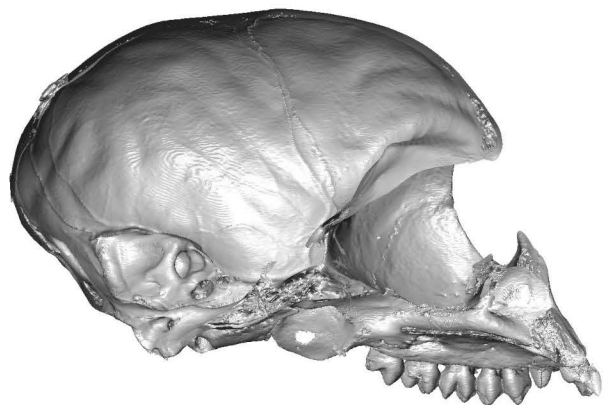
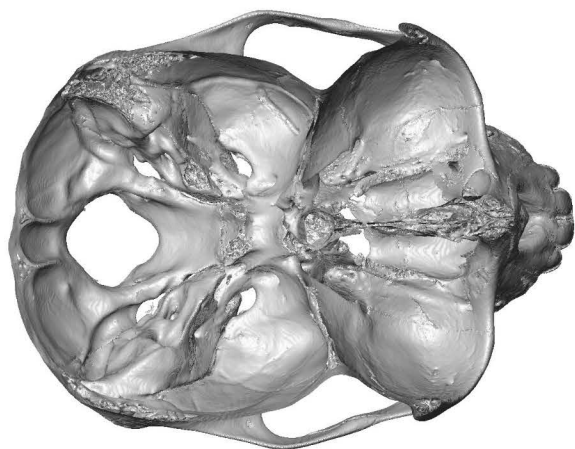
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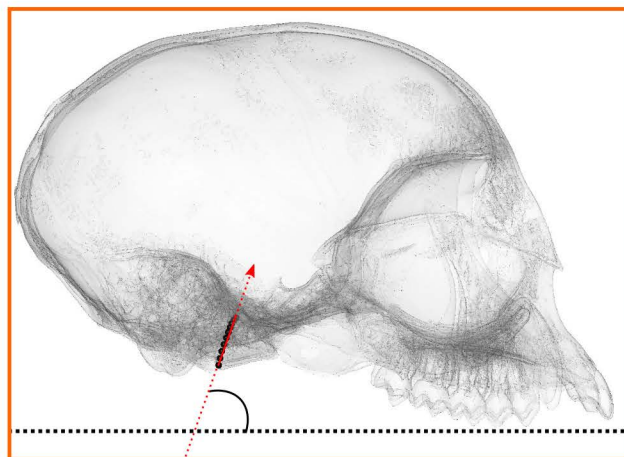
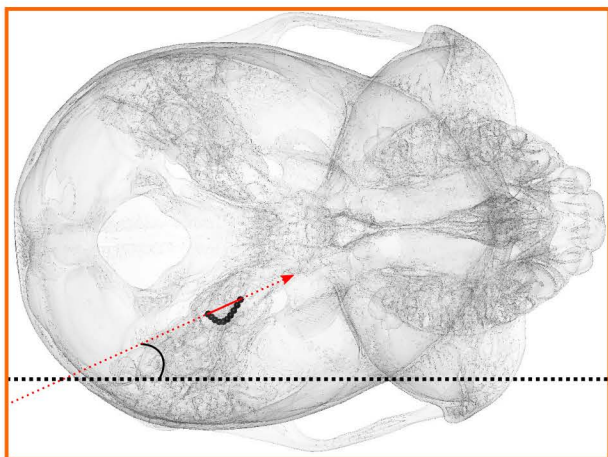
a



b

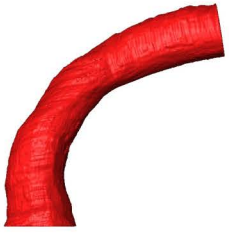


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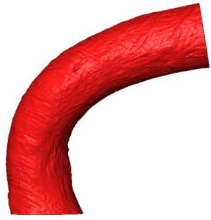


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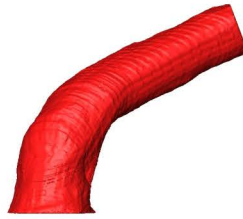
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b) *Epipliopthecus*



c) *Ekembo*



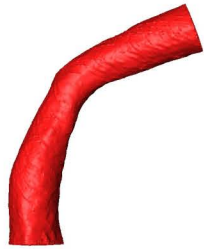
d) *Victoriapithecus*



e) *Alouatta*



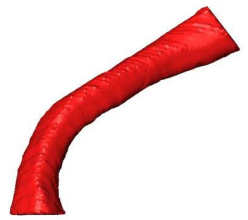
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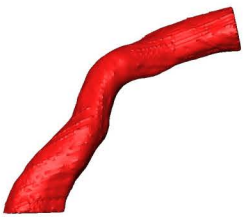
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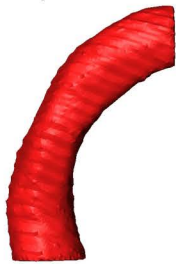
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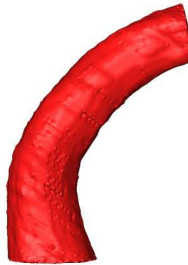
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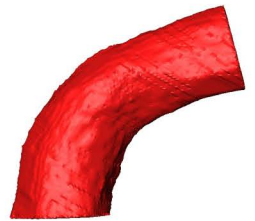
j) *Cercopithecus*



k) *Colobus*



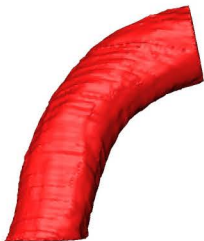
l) *Mandrillus*



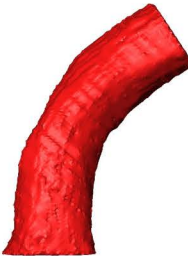
m) *Presbytis*



n) *Hylobates*



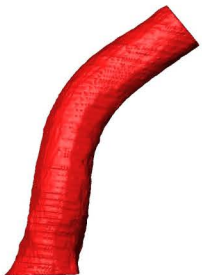
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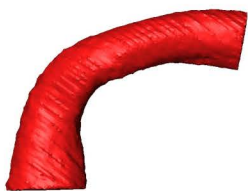
p) *Nomascus*



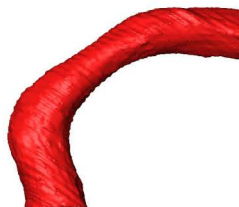
q) *Symphalangus*



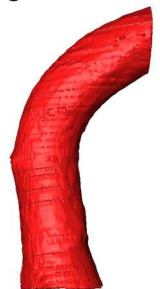
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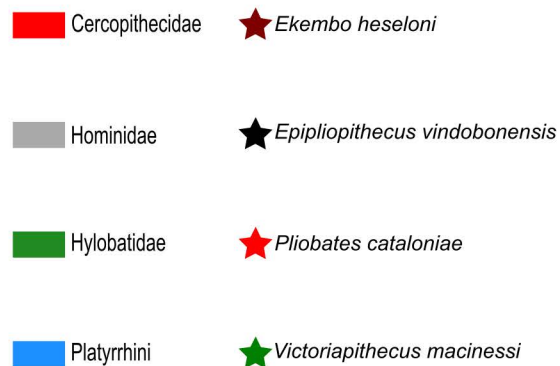
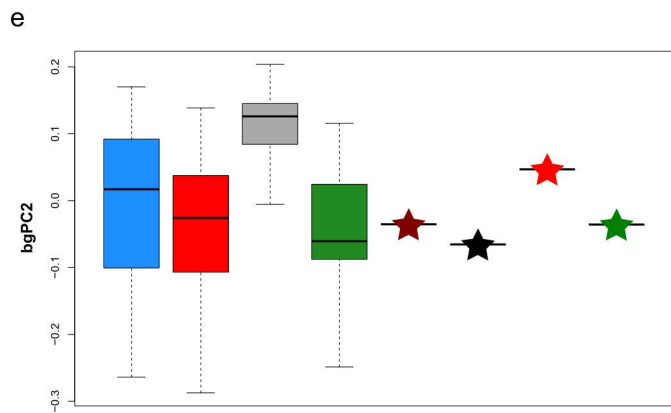
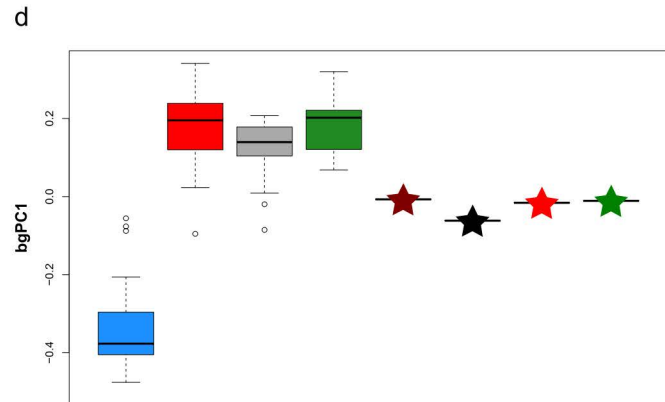
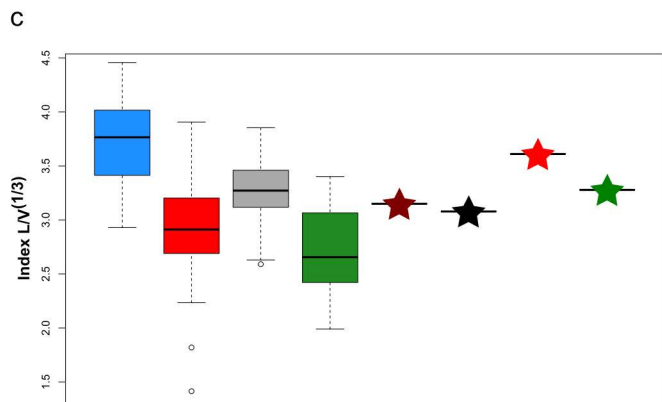
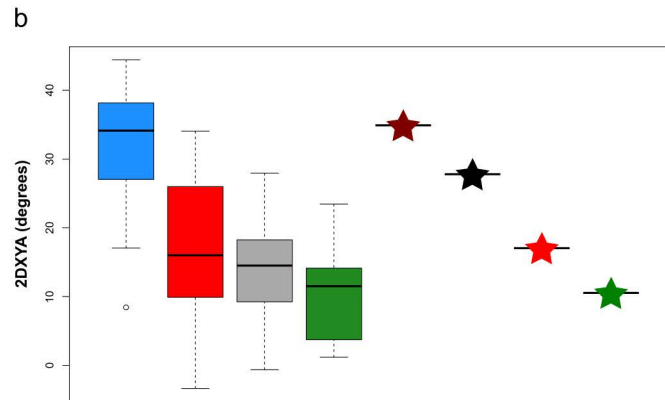
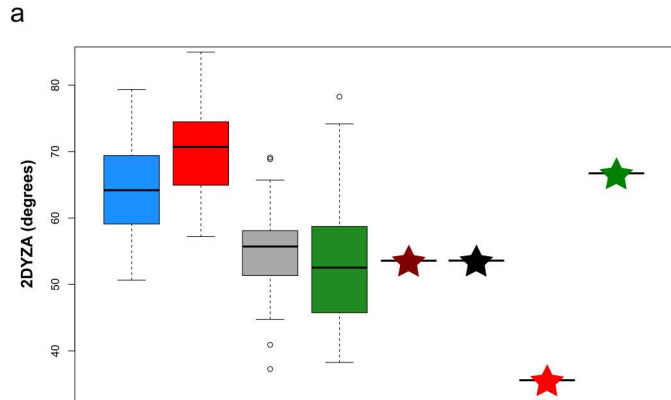


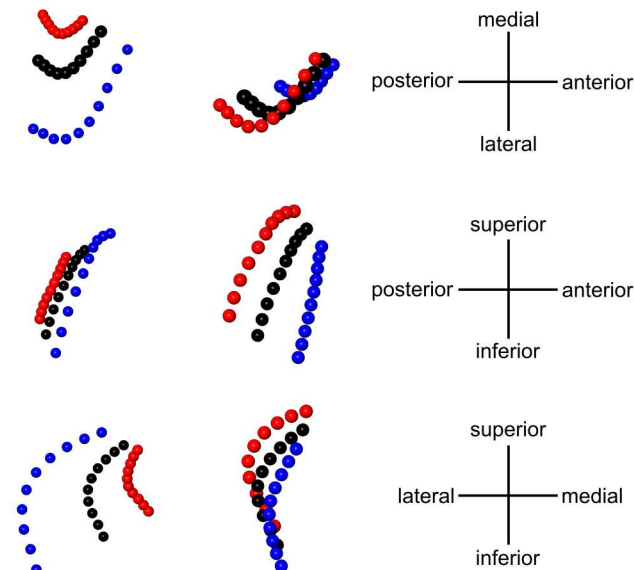
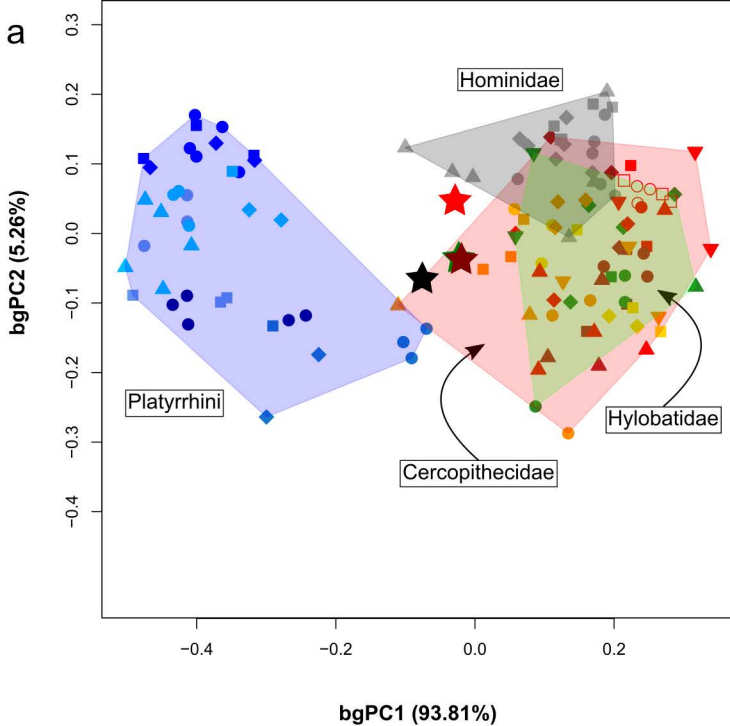
s) *Pan*



t) *Pongo*







- *Alouatta palliata*
- *Ateles geoffroyi*
- ◆ *Lagothrix lagotricha*
- *Aotus griseimembra*
- *Mico argentatus*
- *Saguinus fuscicollis*
- ◆ *Saguinus midas*
- *Cebus imitator*
- *Saimiri sciureus*
- *Chiropotes chiropotes*
- *Pithecia monachus*
- ◆ *Pithecia pithecia*
- ▲ *Plecturocebus cupreus*

- *Cercocebus galeritus*
- *Cercocebus torquatus*
- ◆ *Lophocebus albigena*
- ▲ *Macaca fascicularis*
- ▼ *Mandrillus sphinx*
- *Papio anubis*
- *Theropithecus gelada*
- *Cercopithecus mitis*
- *Chlorocebus pygerythrus*
- ◆ *Erythrocebus patas*
- ▲ *Miopithecus talapoin*

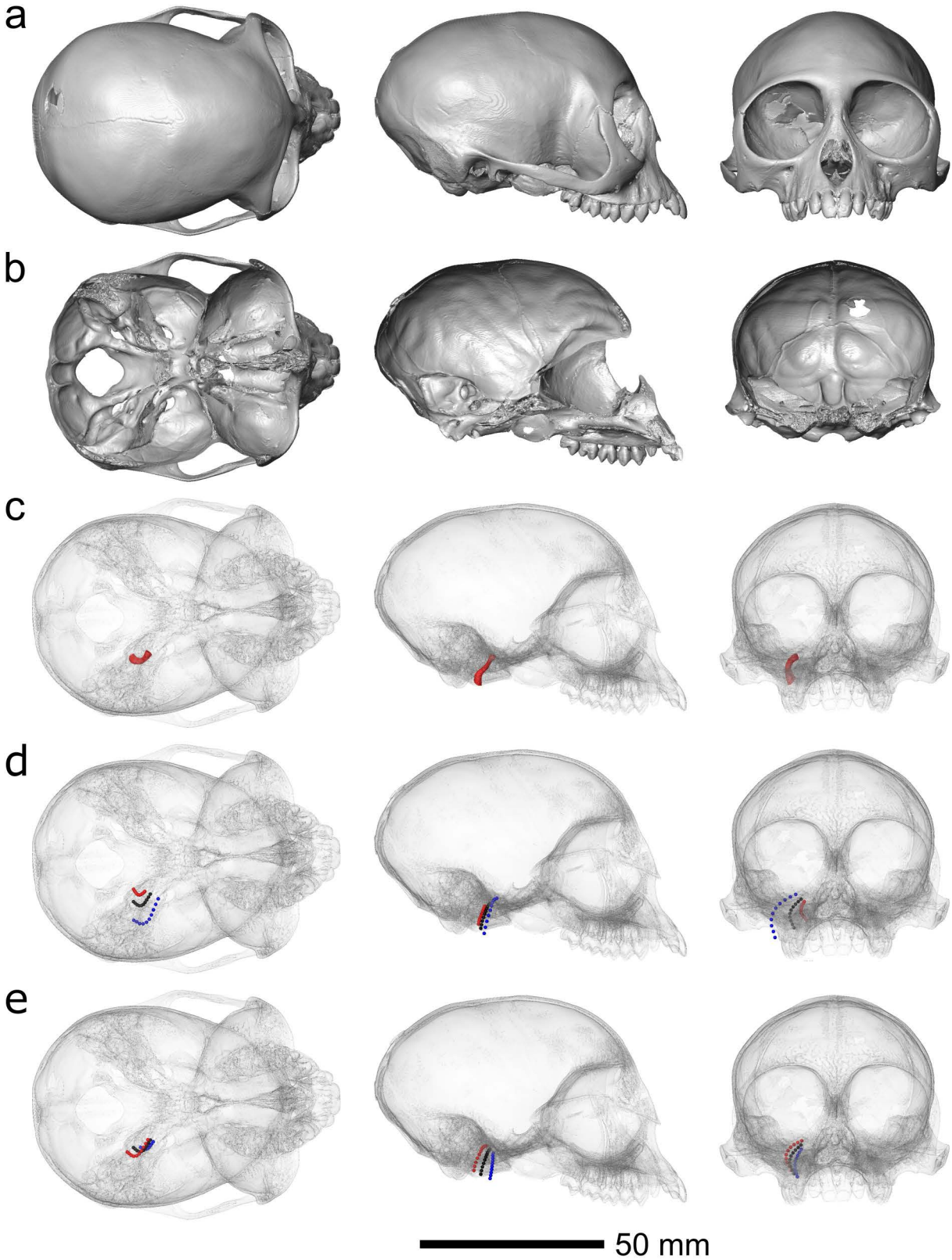
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- *Ptilocolobus badius*
- ◆ *Procolobus verus*
- *Nasalis larvatus*
- *Presbytis rubicunda*
- ◆ *Semnopithecus entellus*
- ▲ *Simias concolor*
- ▼ *Trachypithecus cristatus*

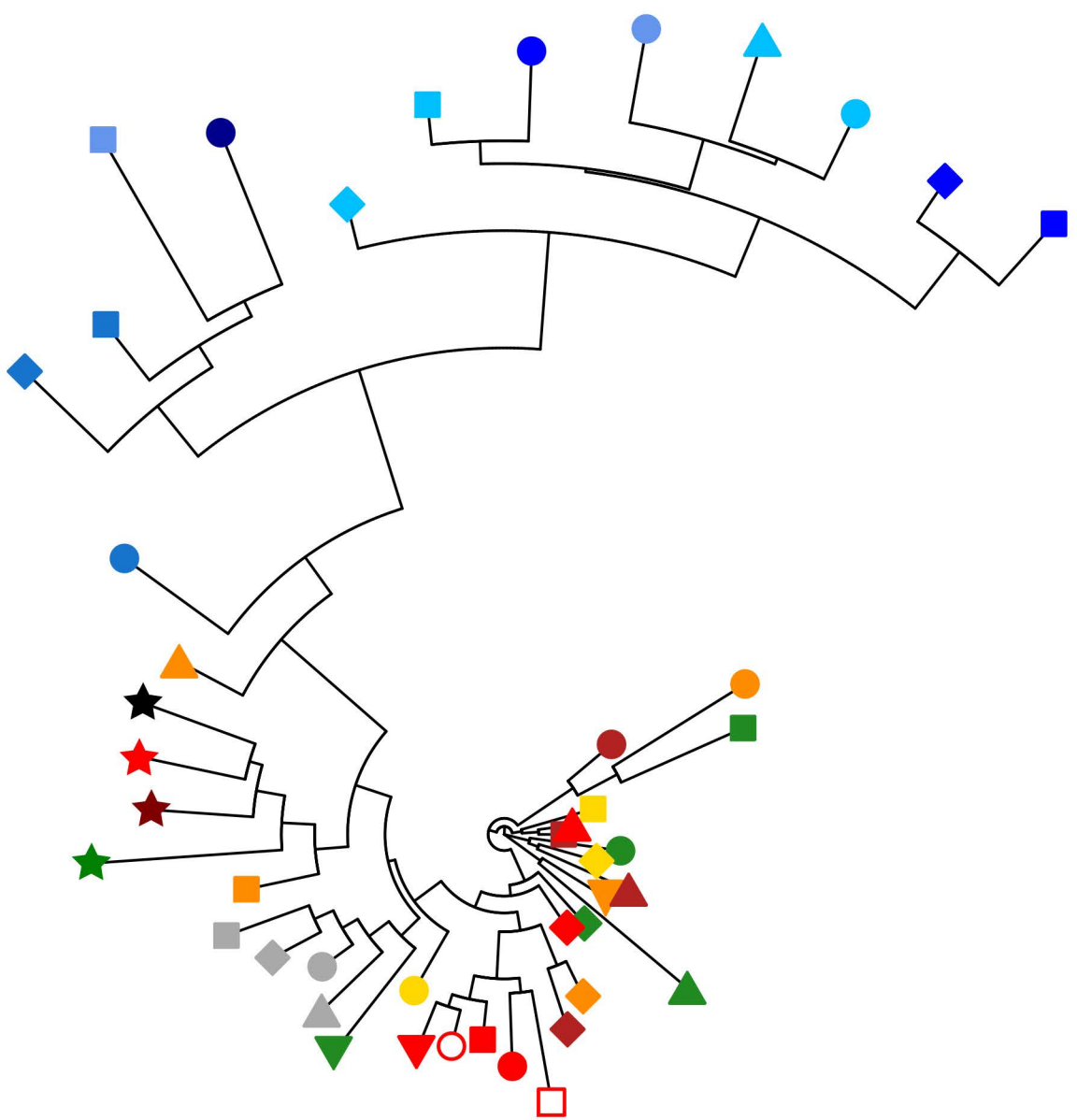
- *Gorilla gorilla*
- *Pan paniscus*
- ◆ *Pan troglodytes*
- ▲ *Pongo sp.*

- ★ *Ekembo heseloni*
- ★ *Epipliopithecus vindobonensis*
- ★ *Pliobates cataloniae*
- ★ *Victoriapithecus macinessi*

- *Hoolock hoolock*
- *Hylobates agilis*
- ◆ *Hylobates lar*
- ▲ *Nomascus leucogenys*
- ▼ *Symphalangus syndactylus*







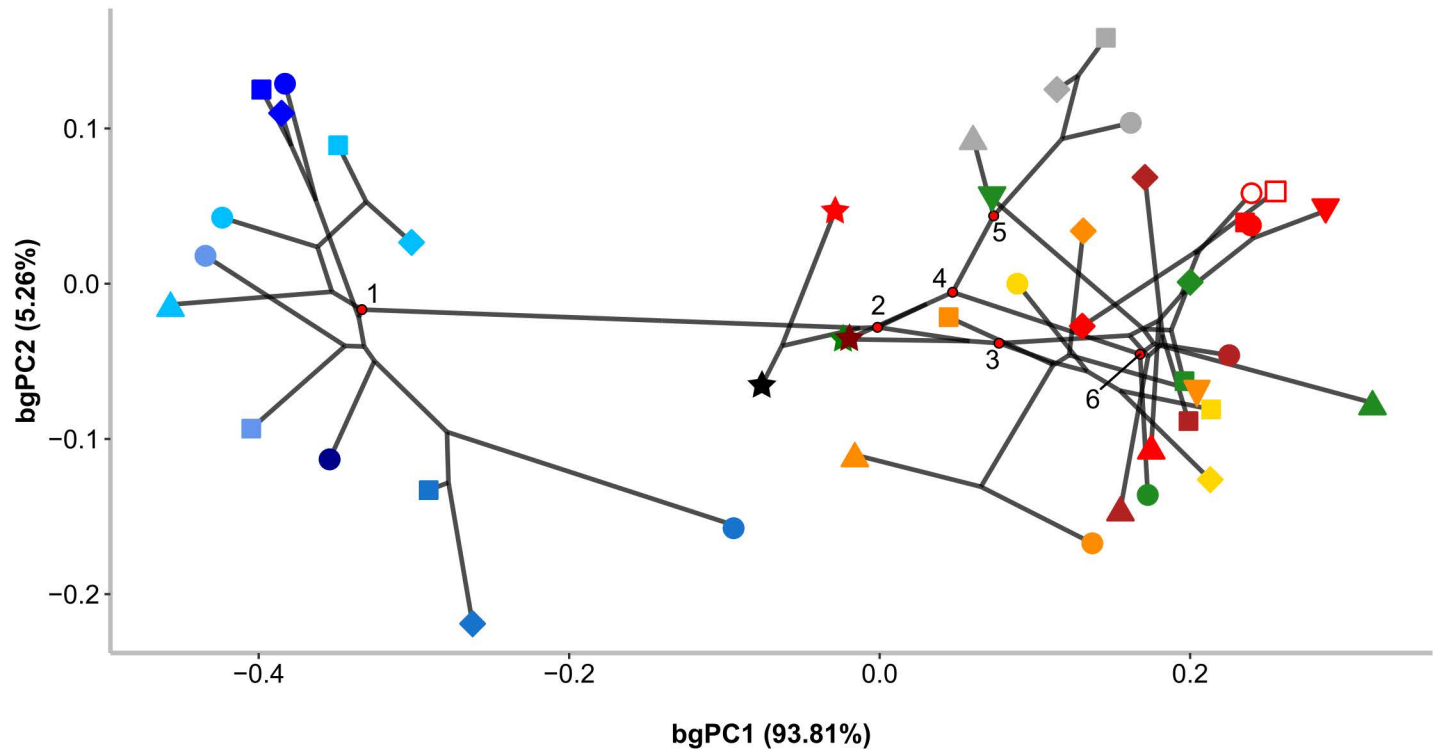
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- *Ateles geoffroyi*
- ◆ *Lagothrix lagotricha*
- *Aotus griseimembra*
- *Mico argentatus*
- *Saguinus fuscicollis*
- ◆ *Saguinus midas*
- *Cebus imitator*
- *Saimiri sciureus*
- *Chiropotes chiropotes*
- *Pithecia monachus*
- ◆ *Pithecia pithecia*
- ▲ *Plecturocebus cupreus*

- *Cercocebus galeritus*
- *Cercocebus torquatus*
- ◆ *Lophocebus albigena*
- ▲ *Macaca fascicularis*
- ▼ *Mandrillus sphinx*
- *Papio anubis*
- *Theropithecus gelada*
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- ★ *Ekembo heseloni*
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- ★ *Victoriapithecus macinessi*
- *Hoolock hoolock*
- *Hylobates agilis*
- ◆ *Hylobates lar*
- ▲ *Nomascus leucogenys*
- ▼ *Symphalangus syndactylus*



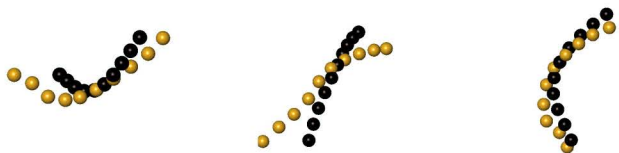
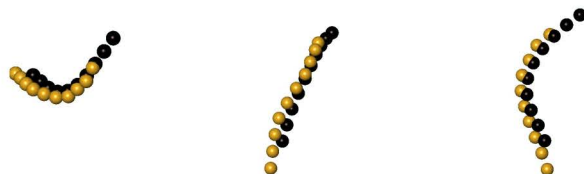
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- *Ateles geoffroyi*
- ◆ *Lagothrix lagotricha*
- *Aotus griseimembra*
- *Mico argentatus*
- *Saguinus fuscicollis*
- ◆ *Saguinus midas*
- *Cebus imitator*
- *Saimiri sciureus*
- *Chiropotes chiropotes*
- *Pithecia monachus*
- ◆ *Pithecia pithecia*
- ▲ *Plecturocebus cupreus*

- *Cercocebus galeritus*
- *Cercocebus torquatus*
- ◆ *Lophocebus albigena*
- ▲ *Macaca fascicularis*
- ▼ *Mandrillus sphinx*
- *Papio anubis*
- *Theropithecus gelada*
- *Cercopithecus mitis*
- *Chlorocebus pygerythrus*
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- ▲ *Miopithecus talapoin*

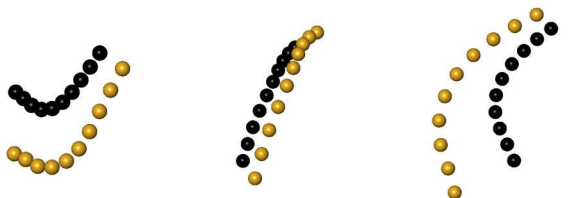
- *Colobus guereza*
- *Ptilocolobus badius*
- ◆ *Procolobus verus*
- *Nasalis larvatus*
- *Presbytis rubicunda*
- ◆ *Semnopithecus entellus*
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- *Hylobates agilis*
- ◆ *Hylobates lar*
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- ▼ *Symphalangus syndactylus*

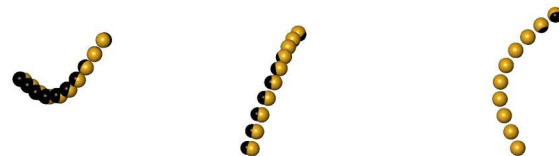
- ★ *Ekembo heseloni*
- ★ *Epipliopthecus vindobonensis*
- ★ *Pliobates cataloniae*
- ★ *Victoriapithecus macinessi*

a) *Pliobates*b) *Epipliopthecus*c) *Ekembo*d) *Victoriapithecus*

e) Crown platyrrhines



f) Crown catarrhines



g) Crown cercopithecoids



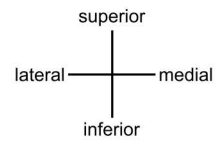
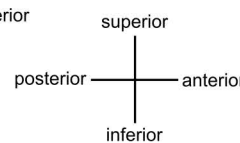
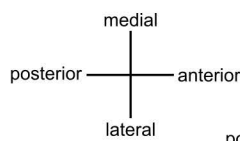
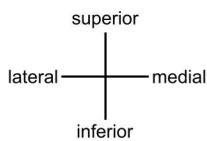
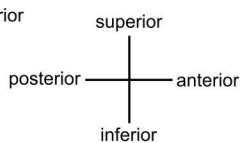
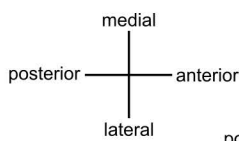
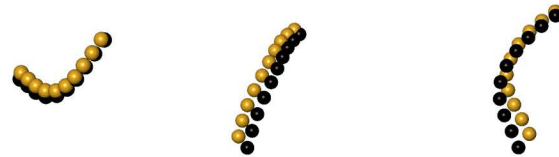
h) Crown hominoids



i) Crown hylobatids



j) Crown hominids





**Table 1**

Extant comparative sample used in this study based on  $\mu$ CT scans. Sample size and sex composition, institutional repository for the specimens, digital repository source, and voxel size range are indicated for each species. See SOM Table S1 for further details on the specimens included.

| Taxon           | Species                        | <i>n</i> | M | F | ? | Repository | Source | Voxel size ( $\mu$ m) |
|-----------------|--------------------------------|----------|---|---|---|------------|--------|-----------------------|
| Platyrrhini     | <i>Alouatta palliata</i>       | 5        | 2 | 3 | 0 | DUEA       | MS     | 52–67                 |
| Platyrrhini     | <i>Aotus griseimembra</i>      | 5        | 2 | 3 | 0 | USNM       | MS     | 34–38                 |
| Platyrrhini     | <i>Ateles geoffroyi</i>        | 3        | 0 | 3 | 0 | AMNH, MCZ  | MS     | 62–63                 |
| Platyrrhini     | <i>Cebus imitator</i>          | 3        | 0 | 3 | 0 | MCZ        | MS     | 80                    |
| Platyrrhini     | <i>Chiropotes chiropotes</i>   | 3        | 1 | 2 | 0 | USNM       | MS     | 47–49                 |
| Platyrrhini     | <i>Lagothrix lagothricha</i>   | 3        | 1 | 2 | 0 | USNM       | MS     | 58–60                 |
| Platyrrhini     | <i>Mico argentatus</i>         | 3        | 2 | 1 | 0 | MCZ        | MS     | 40                    |
| Platyrrhini     | <i>Pithecia monachus</i>       | 1        | 0 | 0 | 1 | MCZ        | MS     | 50                    |
| Platyrrhini     | <i>Pithecia pithecia</i>       | 2        | 1 | 1 | 0 | MCZ        | MS     | 50                    |
| Platyrrhini     | <i>Plecturocebus cupreus</i>   | 5        | 3 | 2 | 0 | AMNH, MCZ  | MS     | 47–51                 |
| Platyrrhini     | <i>Saguinus fuscicollis</i>    | 1        | 0 | 1 | 0 | AMNH       | MS     | 27                    |
| Platyrrhini     | <i>Saguinus midas</i>          | 2        | 2 | 0 | 0 | MCZ        | MS     | 40                    |
| Platyrrhini     | <i>Saimiri sciureus</i>        | 3        | 1 | 1 | 1 | MCZ        | MS     | 47                    |
| Cercopithecidae | <i>Cercocebus torquatus</i>    | 2        | 2 | 0 | 0 | AMNH       | MS     | 90–120                |
| Cercopithecidae | <i>Cercocebus galeritus</i>    | 1        | 1 | 0 | 0 | AMNH, MCZ  | MS     | 70                    |
| Cercopithecidae | <i>Cercopithecus mitis</i>     | 3        | 0 | 3 | 0 | MCZ        | MS     | 80                    |
| Cercopithecidae | <i>Chlorocebus pygerythrus</i> | 3        | 1 | 2 | 0 | SIU        | MS     | 46–54                 |
| Cercopithecidae | <i>Colobus guereza</i>         | 3        | 1 | 2 | 0 | AMNH       | MS     | 66–73                 |
| Cercopithecidae | <i>Erythrocebus patas</i>      | 3        | 2 | 1 | 0 | MCZ        | MS     | 80                    |
| Cercopithecidae | <i>Lophocebus albigena</i>     | 3        | 1 | 2 | 0 | MCZ        | MS     | 90                    |
| Cercopithecidae | <i>Macaca fascicularis</i>     | 5        | 2 | 3 | 0 | MCZ        | MS     | 60–91                 |
| Cercopithecidae | <i>Mandrillus sphinx</i>       | 3        | 2 | 1 | 0 | AMNH, MCZ  | MS     | 75–126                |
| Cercopithecidae | <i>Miopithecus talapoin</i>    | 3        | 3 | 0 | 0 | MCZ        | MS     | 50                    |
| Cercopithecidae | <i>Nasalis larvatus</i>        | 3        | 0 | 3 | 0 | MCZ        | MS     | 71                    |

|                 |                                 |   |   |   |   |                |              |         |
|-----------------|---------------------------------|---|---|---|---|----------------|--------------|---------|
| Cercopithecidae | <i>Papio anubis</i>             | 3 | 3 | 0 | 0 | MCZ            | MS           | 108–118 |
| Cercopithecidae | <i>Ptilocolobus badius</i>      | 3 | 2 | 1 | 0 | MCZ            | MS           | 80–90   |
| Cercopithecidae | <i>Presbytis rubicunda</i>      | 3 | 1 | 2 | 0 | MCZ            | MS           | 80      |
| Cercopithecidae | <i>Procolobus verus</i>         | 2 | 1 | 1 | 0 | AMNH           | MS           | 76–80   |
| Cercopithecidae | <i>Semnopithecus entellus</i>   | 3 | 2 | 1 | 0 | AMNH           | MS           | 52–108  |
| Cercopithecidae | <i>Simias concolor</i>          | 2 | 1 | 1 | 0 | AMNH           | MS           | 56      |
| Cercopithecidae | <i>Theropithecus gelada</i>     | 3 | 2 | 1 | 0 | AMNH           | MS           | 84–120  |
| Cercopithecidae | <i>Trachypithecus cristatus</i> | 3 | 0 | 3 | 0 | MCZ            | MS           | 50      |
| Hominidae       | <i>Gorilla gorilla</i>          | 6 | 1 | 5 | 0 | AMNH, MCZ      | MS           | 106–131 |
| Hominidae       | <i>Pan paniscus</i>             | 5 | 1 | 3 | 1 | AMNH, ICP, MCZ | MS, own data | 66–116  |
| Hominidae       | <i>Pan troglodytes</i>          | 6 | 3 | 3 | 0 | AMNH, ICP, MCZ | MS, own data | 77–111  |
| Hominidae       | <i>Pongo</i> sp.                | 6 | 0 | 6 | 0 | ICP, MCZ       | MS, own data | 77–125  |
| Hylobatidae     | <i>Hoolock hoolock</i>          | 3 | 1 | 2 | 0 | AMNH           | MS           | 52–84   |
| Hylobatidae     | <i>Hylobates agilis</i>         | 1 | 0 | 0 | 1 | ICP            | MS, own data | 52      |
| Hylobatidae     | <i>Hylobates lar</i>            | 4 | 0 | 4 | 0 | MCZ            | MS           | 67      |
| Hylobatidae     | <i>Nomascus leucogenys</i>      | 1 | 1 | 0 | 0 | AMNH           | MS           | 105     |
| Hylobatidae     | <i>Symphalangus syndactylus</i> | 2 | 0 | 2 | 0 | AMNH           | MS           | 62–79   |

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Abbreviations: M = male; F = female; ? = unknown sex; AMNH = American Museum of Natural History, New York, USA; DUEA = Duke University, Evolutionary Anthropology, Durham, USA; ICP = acronym of the collections of the Institut Català de Paleontologia Miquel Crusafont, Sabadell, Spain; MCZ = Museum of Comparative Zoology, Harvard University, Cambridge, USA; MS = MorphoSource (<https://www.morphosource.org>); SIU = Southern Illinois University, Carbondale, USA; USNM = Smithsonian National Museum of Natural History, Washington, D.C., USA.

**Table 2**

Definition of the petrosal/tympanic landmarks used in this paper for aligning the specimens.

| No. | Landmark                           | Definition                                                                                                                |
|-----|------------------------------------|---------------------------------------------------------------------------------------------------------------------------|
| 1   | Porion                             | Most superior point on the margin of the external auditory meatus                                                         |
| 2   | Inferior external auditory meatus  | Most inferior point on the margin of the external auditory meatus                                                         |
| 3   | Posterior external auditory meatus | Most posterior point on the margin of the external auditory meatus                                                        |
| 4   | Posterior internal auditory meatus | Most posterior point on the margin of the internal auditory meatus                                                        |
| 5   | Anterior internal auditory meatus  | Most anterior point on the margin of the internal auditory meatus                                                         |
| 6   | Medial anterior pyramidal ridge    | Most medial point where anterior pyramidal ridge starts running laterally above the fossa for the semilunar ganglion      |
| 7   | Intersection ridge-groove          | Point of intersection between the laterally running anterior pyramidal ridge and the groove of the greater petrosal nerve |
| 8   | Greater petrosal nerve hiatus      | Most superior, anterior point on the margin of the hiatus for the greater petrosal nerve                                  |
| 9   | Stylomastoid foramen               | Center of the stylomastoid foramen                                                                                        |

**Table 3**

Descriptive statistics of the variables describing the orientation, proportions and course of the carotid canal in the main extant anthropoid clades. See SOM Table S2 for the individual values of extant anthropoid specimens.

| Clade           | <i>n</i> | 2DYZA |       |             | 2DXYA |      |             | L/V <sup>(1/3)</sup> |      |           | bgPC1 |      |            | bgPC2 |      |            |
|-----------------|----------|-------|-------|-------------|-------|------|-------------|----------------------|------|-----------|-------|------|------------|-------|------|------------|
|                 |          | Mean  | SD    | Range       | Mean  | SD   | Range       | Mean                 | SD   | Range     | Mean  | SD   | Range      | Mean  | SD   | Range      |
| Platyrrhini     | 39       | 64.06 | 7.33  | 50.66–79.36 | 31.92 | 7.57 | 8.25–44.26  | 3.68                 | 0.39 | 2.90–4.42 | -0.36 | 0.11 | 0.50–0.07  | -0.01 | 0.11 | -0.26–0.17 |
| Cercopithecidae | 54       | 69.71 | 7.42  | 57.2–85     | 16.69 | 10   | -3.54–33.87 | 2.86                 | 0.42 | 1.40–3.87 | 0.17  | 0.08 | -0.11–0.34 | -0.04 | 0.09 | -0.29–0.14 |
| Hominidae       | 23       | 54.90 | 7.83  | 37.27–69.11 | 13.74 | 7.38 | -0.82–27.78 | 3.23                 | 0.31 | 2.57–3.82 | 0.12  | 0.08 | -0.10–0.20 | 0.12  | 0.05 | -0.01–0.20 |
| Hylobatidae     | 11       | 54.51 | 12.39 | 38.27–78.27 | 10.27 | 7.44 | 1.00–23.27  | 2.68                 | 0.47 | 1.97–3.37 | 0.18  | 0.08 | 0.06–0.32  | -0.04 | 0.10 | -0.25–0.12 |

Abbreviations: 2DYZA = 2D angle in the YZ plane (canal superior orientation; in degrees); 2DXYA = 2D angle in the XY plane (canal medial orientation; in degrees); L = length (in mm); V = volume (in mm<sup>3</sup>); bgPC = between-group principal component.

**Table 4**

Variables describing the orientation, proportions and course of the carotid canal in the fossil specimens.

| Species                              | Catalog No.         | 2DYZA | 2DXYA | $L/V^{(1/3)}$ | bgPC1 | bgPC2 |
|--------------------------------------|---------------------|-------|-------|---------------|-------|-------|
| <i>Ekembo heseloni</i>               | KNM-RU 2036al       | 53.58 | 34.73 | 3.12          | -0.02 | -0.03 |
| <i>Epipliopithecus vindobonensis</i> | NHMW 1970/1397/0003 | 53.59 | 27.62 | 3.05          | -0.08 | -0.06 |
| <i>Pliobates cataloniae</i>          | IPS58443.1          | 35.58 | 16.86 | 3.58          | -0.03 | 0.05  |
| <i>Victoriapithecus macinessi</i>    | KNM-MB 29100        | 66.73 | 10.36 | 3.25          | -0.02 | -0.04 |

Abbreviations: 2DYZA = 2D angle in the YZ plane (canal superior orientation; in degrees);

2DXYA = 2D angle in the XY plane (canal medial orientation; in degrees); L = length (in mm);

V = volume (in mm<sup>3</sup>); bgPC = between-group principal component.

**Table 5**

Z-score analysis of the three variables describing the orientation, proportions and course of the carotid canal in the fossil specimens and compared to the variation expressed by the main extant anthropoid clades.<sup>a</sup>

| Variable and species                 | Catalog No.         | Platyrrhini  | Cercopithecidae | Hominidae    | Hylobatidae  |
|--------------------------------------|---------------------|--------------|-----------------|--------------|--------------|
| 2DYZA                                |                     |              |                 |              |              |
| <i>Ekembo heseloni</i>               | KNM-RU 2036aI       | -1.43        | <b>-2.17</b>    | -0.17        | -0.08        |
| <i>Epipliopithecus vindobonensis</i> | NHMW 1970/1397/0003 | -1.43        | <b>-2.17</b>    | -0.17        | -0.07        |
| <i>Pliobates cataloniae</i>          | IPS58443.1          | <b>-3.88</b> | <b>-4.60</b>    | <b>-2.46</b> | -1.52        |
| <i>Victoriapithecus macinnesi</i>    | KNM-MB 29100        | 0.36         | -0.40           | 1.51         | 0.98         |
| 2DXYA                                |                     |              |                 |              |              |
| <i>Ekembo heseloni</i>               | KNM-RU 2036aI       | 0.37         | 1.80            | <b>2.84</b>  | <b>3.29</b>  |
| <i>Epipliopithecus vindobonensis</i> | NHMW 1970/1397/0003 | -0.56        | 1.09            | 1.88         | <b>2.33</b>  |
| <i>Pliobates cataloniae</i>          | IPS58443.1          | <b>-1.99</b> | 0.02            | 0.42         | 0.89         |
| <i>Victoriapithecus macinnesi</i>    | KNM-MB 29100        | <b>-2.85</b> | -0.63           | -0.46        | -0.02        |
| L/V <sup>(1/3)</sup>                 |                     |              |                 |              |              |
| <i>Ekembo heseloni</i>               | KNM-RU 2036aI       | -1.46        | 0.61            | -0.36        | 0.93         |
| <i>Epipliopithecus vindobonensis</i> | NHMW 1970/1397/0003 | -1.64        | 0.44            | -0.58        | 0.78         |
| <i>Pliobates cataloniae</i>          | IPS58443.1          | -0.27        | 1.69            | 1.09         | 1.89         |
| <i>Victoriapithecus macinnesi</i>    | KNM-MB 29100        | -1.13        | 0.91            | 0.04         | 1.20         |
| bgPC1                                |                     |              |                 |              |              |
| <i>Ekembo heseloni</i>               | KNM-RU 2036aI       | <b>3.23</b>  | <b>-2.30</b>    | -1.78        | <b>-2.39</b> |
| <i>Epipliopithecus vindobonensis</i> | NHMW 1970/1397/0003 | <b>2.70</b>  | <b>-2.97</b>    | <b>-2.50</b> | <b>-3.08</b> |
| <i>Pliobates cataloniae</i>          | IPS58443.1          | <b>3.14</b>  | <b>-2.41</b>    | -1.90        | <b>-2.51</b> |
| <i>Victoriapithecus macinnesi</i>    | KNM-MB 29100        | <b>3.19</b>  | <b>-2.35</b>    | -1.83        | <b>-2.45</b> |
| bgPC2                                |                     |              |                 |              |              |
| <i>Ekembo heseloni</i>               | KNM-RU 2036aI       | -0.25        | 0.02            | <b>-3.16</b> | 0.04         |
| <i>Epipliopithecus vindobonensis</i> | NHMW 1970/1397/0003 | -0.51        | -0.31           | <b>-3.78</b> | -0.27        |
| <i>Pliobates cataloniae</i>          | IPS58443.1          | 0.48         | 0.93            | -1.47        | 0.87         |
| <i>Victoriapithecus macinnesi</i>    | KNM-MB 29100        | -0.25        | 0.02            | <b>-3.17</b> | 0.03         |

Abbreviations: 2DYZA = 2D angle in the YZ plane (canal superior orientation; in degrees);  
2DXYA = 2D angle in the XY plane (canal medial orientation; in degrees); L = length (in mm);  
V = volume (in mm<sup>3</sup>); bgPC = between-group principal component.

<sup>a</sup>Significant z-scores at  $p < 0.05$  are bolded.

**Table 6**

Phylogenetic generalized least-squares (PGLS) allometric regressions between carotid canal size variables (L,  $V^{1/3}$  or CS) or between carotid canal size and BM, together with PGLS bivariate regressions between carotid canal shape (as represented by the canal robusticity index or bgPCs) vs. log-transformed carotid canal size or BM.

| Regression             | $p^a$  | $R^2$ | Slope | Slope 95% CI |
|------------------------|--------|-------|-------|--------------|
| ln L vs. ln $V^{1/3}$  | <0.001 | 0.81  | 0.86  | 0.73, 0.99   |
| ln L vs. ln $V^{1/3b}$ | <0.001 | 0.81  | 0.95  | 0.81, 1.10   |
| $L/V^{1/3}$ vs. ln CS  | >0.05  | -0.02 | 0.11  | -0.34, 0.57  |
| ln L vs. ln CS         | <0.001 | 0.99  | 1.04  | 1.02, 1.06   |
| ln $V^{1/3}$ vs. ln CS | <0.001 | 0.81  | 0.99  | 0.84, 1.13   |
| ln L vs. ln BM         | <0.001 | 0.68  | 0.25  | 0.20, 0.31   |
| ln $V^{1/3}$ vs. ln BM | <0.001 | 0.90  | 0.30  | 0.27, 0.34   |
| ln CS vs. ln BM        | <0.001 | 0.69  | 0.25  | 0.19, 0.30   |
| $L/V^{1/3}$ vs. ln BM  | <0.01  | 0.14  | -0.17 | -0.29, -0.05 |
| bgPC1 vs. ln CS        | >0.05  | 0.01  | 0.12  | -0.07, 0.31  |
| bgPC1 vs. ln $V^{1/3}$ | <0.01  | 0.19  | 0.26  | 0.10, 0.41   |
| bgPC1 vs. ln BM        | <0.001 | 0.25  | 0.10  | 0.05, 0.15   |
| bgPC2 vs. ln CS        | <0.001 | 0.58  | 0.21  | 0.16, 0.27   |
| bgPC2 vs. ln $V^{1/3}$ | <0.001 | 0.46  | 0.17  | 0.11, 0.23   |
| bgPC2 vs. ln BM        | <0.001 | 0.35  | 0.05  | 0.04, 0.08   |

Abbreviations: L = carotid canal length (mm);  $V^{1/3}$  = carotid canal volume cube root (mm);  $L/V^{1/3}$  = shape index of canal robusticity; CS = carotid canal centroid size (mm); BM = body mass (g); bgPC = between-group principal component (representing carotid canal course).

<sup>a</sup> Regressions are significant when  $p < 0.05$ .



<sup>b</sup> For  $\ln L$  vs.  $\ln V^{1/3}$ , a major axis regression was also computed to test whether the two variables are isometric when deviation from the best-fit line is minimized simultaneously for both variables (instead of only the dependent variable, in this case  $\ln L$ ). Our results show that, unlike for the PGLS regression (which displays slight negative allometry), isometry cannot be discounted for the major axis regression.

**Table 7**

Number (*n*) and percentages (%) of correctly and incorrectly classified individuals obtained by the between-group principal components analysis after cross-validation. In total, 98 out of 127 (77.2%) individuals were correctly classified (in bold).

|                 | Platyrrhini       | Cercopithecidae   | Hominidae         | Hylobatidae      |
|-----------------|-------------------|-------------------|-------------------|------------------|
| Platyrrhini     | <b>36 (92.3%)</b> | 0 (0%)            | 0 (0%)            | 3 (7.7%)         |
| Cercopithecidae | 1 (1.9%)          | <b>34 (63.0%)</b> | 9 (16.7%)         | 10 (18.5%)       |
| Hominidae       | 0 (0%)            | 2 (8.7%)          | <b>21 (91.3%)</b> | 0 (0.0%)         |
| Hylobatidae     | 0 (0%)            | 3 (27.3%)         | 1 (9.1%)          | <b>7 (63.6%)</b> |

**Table 8**

Posterior probabilities of group membership based on the between-group principal component scores of fossil specimens.<sup>a</sup>

| Species                             | Catalog No.         | Platyrrhini | Cercopithecidae | Hominidae    | Hylobatidae |
|-------------------------------------|---------------------|-------------|-----------------|--------------|-------------|
| <i>Ekembo heseloni</i>              | KNM-RU 2036aI       | <0.001      | <b>0.098</b>    | 0.059        | 0.087       |
| <i>Epipliopthecus vindobonensis</i> | NHMW 1970/1397/0003 | <0.001      | <b>0.018</b>    | 0.009        | 0.016       |
| <i>Pliobates cataloniae</i>         | IPS58443.1          | <0.001      | 0.064           | <b>0.169</b> | 0.056       |
| <i>Victoriapithecus macinessi</i>   | KNM-MB 29100        | <0.001      | <b>0.089</b>    | 0.054        | 0.079       |

<sup>a</sup> Each value corresponds to the probability of having the given score if the specimen was a member of the group considered, not the likelihood of group membership in each of the a priori defined groups given a particular score (the greater the number, the higher the probability). For each specimen the highest probability is bolded.