



Breaking New Ground: Exploring the Promising Role of Solid-State Fermentation in Harnessing Natural Biostimulants for Sustainable Agriculture

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Abstract: Agriculture has been experiencing a difficult situation because of limiting factors in its production processes. Natural biostimulants (NBs) have emerged as a novel alternative. This study reviews NBs produced through solid-state fermentation (SSF) from organic waste, focusing on processes and production methods. The aim is to highlight their potential for improving agricultural productivity and promoting sustainable agriculture. Through a literature review, the effects of NBs on crops were summarized, along with the challenges associated with their production and application. The importance of standardizing production processes, optimizing fermentation conditions, and assessing their effects on different crops is emphasized. Furthermore, future research areas are introduced, such as enhancing production efficiency and evaluating the effectiveness of SSF-produced NBs in different agricultural systems. In conclusion, SSF-produced NBs offer a promising alternative for sustainable agriculture, but further research and development are needed to maximize their efficacy and to enable large-scale implementation.

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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). **Keywords:** natural biostimulant; solid-state fermentation; organic waste; sustainable agriculture; crop improvement

1. Introduction

One of the main challenges in agriculture is achieving global zero hunger [1]. Therefore, sustainable agriculture is a viable method to ensure food security. In this regard, the Food and Agriculture Organization of the United Nations (FAO) envisions providing nutritious and accessible food for all while preserving natural resources to meet current and future needs. Sustainable agriculture also aims to benefit producers in terms of economic development [1]. In conventional agriculture, reducing the intensive use of agrochemicals is a significant challenge that negatively impacts soil health, water scarcity, and biodiversity [2]. In this context, natural biostimulants (NBs) have emerged as alternatives to sustainable agriculture. NBs are derived from products such as microorganisms, plant extracts, and seaweed extracts and can be classified into three main groups based on their source and content: humic substances (HS), hormone-containing products (HCP), and amino-acid-containing products (AACP). HCP, such as seaweed extracts, contain various active substances for plant growth, including auxins, cytokinins, and their derivatives [3]. These products contain biologically active compounds that stimulate plant physiological processes and promote growth, development, and resistance to biotic and abiotic stresses [4–7]. NBs offer significant advantages because they are derived from natural sources, such as waste materials, plant extracts, and microorganisms [8,9], making them more environmentally sustainable than chemical products based on synthetic compounds. Furthermore, NBs are generally safer for the environment and human health than chemical products, which can be harmful [10]. NBs also have the potential to promote beneficial

interactions with soil microorganisms, unlike chemical products that lack this capacity [11]. Additionally, NBs can serve as an easier alternative to chemicals in order to comply with regulations and restrictions in many countries [6,12,13]. Given these issues, NBs present themselves as a promising alternative in agriculture.

Various production methods exist, including solid-state fermentation (SSF), a technology conducted in the absence or near absence of free water, allowing the use of solid materials as substrates for enhanced biotransformation. SSF has been reported as a promising eco-technology for the production of bio-based products, and studies have demonstrated the successful pilot-scale production of NBs using plant biomass as a support and carbon source for different microorganisms. These production processes are performed under controlled conditions, including temperature, humidity, and airflow, to optimize NB synthesis [14,15]. Furthermore, the utilization of organic waste as a substrate in the SSF process has gained attention, primarily involving various solid biodegradable materials derived from agricultural and forestry byproducts and waste [16]. NBs obtained through SSF have shown biostimulant effects on crop development, including physical parameters such as germination, growth, stem length, leaf count, root dry weight, leaf area, biomass production, macronutrients, and micronutrients [17]. They have also demonstrated positive effects on root development in forest species [18]. Therefore, NBs produced through SSF represent an emerging alternative to the limitations of conventional biostimulants, including their negative impact on agricultural sustainability, the need to reduce the impact of waste on the environment, and the desire to limit the use of synthetic compounds in agriculture [19].

This review addresses the production of NBs through SSF using organic waste as a promising approach for sustainable agriculture. Furthermore, these NBs have the potential to enhance plant growth and development while reducing reliance on conventional chemical products. To achieve this, the existing literature was reviewed to assess the effectiveness and limitations of NB production through SSF.

2. Materials and Methods

Methodology

This review article involved the selection of scientific articles from the following scientific databases: SpringerLink (https://link.springer.com/, accessed on 25 May 2023), Science Direct (https://www.sciencedirect.com/, accessed on 25 May 2023), Wiley (https://onlinelibrary.wiley.com, accessed on 25 May 2023), ProQuest (https://www.proquest. com/, accessed on 25 May 2023), Patent Inspiration (https://www.patentinspiration.com/, accessed on 25 May 2023), and Web of Science (https://www.webofscience.com/, accessed on 25 May 2023). Boolean operators (AND and OR) were used to obtain more accurate results. The following keywords were used: "solid state fermentation and biostimulant", "solid state fermentation and auxins", "solid state fermentation and biostimulant name". Literature from the past 30 years was included in the article review.

Articles were selected based on the following inclusion criteria: relevance of the publication to the topic and selected years. The following criteria were considered: type of NB, substrate, microorganisms, optimal conditions, and effects on crops. We aimed to address these research questions by collecting and analyzing relevant studies, considering the latest trends in NB production through SSF using organic waste.

3. Relevant Sections

3.1. Definition and Types of Biostimulants

NBs are derived from natural sources such as microorganisms, plant residues, and seaweed, among others [20]. These products contain biologically active compounds that stimulate plant physiological processes, promoting plant growth, development, and resistance to biotic and abiotic stresses [10]. However, biostimulants include a wide range of compounds, as highlighted by the European Biostimulants Industry Council (EBIC) and the Biological Products Industry Alliance (BPIA) [14]. The EBIC defines plant biostimulants as

substances or microorganisms that stimulate natural processes to enhance nutrient uptake, efficiency, stress tolerance, and crop quality. They do not a have direct pesticidal action and are not regulated by pesticide laws. BPIA defines biostimulants as diverse materials that improve crop vigour, quality, yield, and tolerance to abiotic stresses by facilitating nutrient uptake, enhancing soil microorganism development, and stimulating root growth to increase water-use efficiency [12,13]. This growth is in line with an increase in scientific support for the use of biostimulants as agricultural inputs for various plant species [21].

Currently, there are various types of NBs, including those produced by SSF, which can serve as a starting point for future research (Table 1).

Natural Product	Type of NB	Molecules Present	Action Mode	Biostimulant Effect	SSF-Relevant Origin	Refs.
	Auxins	3-indoleacetic Acid (IAA)	Promotes cell elongation	Stimulates cell elongation and rooting	Produced by SSF	[22,23]
		Indole Propionic Acid (AIP)	Promotes vegetative growth and cell division	Stimulates growth, flowering, and rooting in plants	Not produced by SSF	[24,25]
		Zeatin	Stimulates cell division and vegetative growth	Promotes growth and development of plants	Not produced by SSF	[26-28]
Hormone- Containing Products (HCP)	Cytokinins	Kinetin	Stimulates cell division and vegetative growth	Improves the quality of the crops, increasing the size and weight of the fruits	Produced by SSF and vermicompost	[29–31]
	Abscisic Acid (ABA)	ABA	Regulates stress responses and plant development	Improves stress tolerance and fruit ripening	Produced by SSF	[32,33]
		Gibberellin A3 (GA3)	Stimulates growth and vigor in plants	Inducts germination and flowering	Produced by SSF	[34–36]
	Gibberellins	Gibberellin A4 (GA4)	Promotes plant growth and development	Stimulates germination, development of lateral shoots, and flowering	Produced by SSF	[37,38]
		Alginic Acids	Improves nutrient absorption and stimulates enzyme activity	Increases growth and resistance to abiotic stress	Produced by SSF	[39–41]
Seaweed Extract (AM)	AM	Fucoidan	Improves the defense mechanisms of plants	Increases resistance to abiotic stress	Produced by SSF	[42-45]
		Oligosaccharides	Stimulates physiological responses in plants	Improves immune response and growth	Produced by SSF	[46-49]
	Humic and	Humic Acids	Improves soil structure and nutrient availability	Stimulates root growth and nutrient absorption	Produced by SSF	[50–53]
Humic Substances	Fulvic Acids (AHF)	Humic Acids	Stimulates plant growth and development	Improves nutrient uptake and stress resistance.	Produced by SSF	[54,55]
Amino-Acid-	Amino Acids	L-proline	Regulates plant stress and development	Enhances stress tolerance and resistance	Produced by SSF	[56–58]
Containing Products (AACP)	Peptides	Low Molecular Weight Peptides	Stimulates plant growth and development	Improves plant nutrition and growth	Produced by SSF	[59-61]
	Siderophores	Siderophores	Binds to Fe and is solubilized	Improves absorption and mobilization of Fe	Produced by SSF	[62-64]
Other NBs	Chitosan Fungal	Chitosan Fungal	Promotes plant growth, cell division, increases enzyme activity, and improves nutrient transport	Presents biostimulant activity in seed germination	Produced by SSF	[65,66]

Table 1. Types of NBs, mode of action, and effects produced by SSF.

3.2. Advantages of Natural Biostimulants over Conventional Ones

In this regard, NBs obtained through SSF have emerged as an alternative to conventional biostimulants, primarily because of their positive impact on agricultural sustainability, reduced environmental waste, and limited use of synthetic compounds in agriculture [46].

NBs obtained by SSF from organic waste are gaining interest because of their numerous advantages over conventionally synthesized biostimulants [47]. This article reviews and compares the advantages of NBs in terms of effectiveness, safety, sustainability, and environmental benefits. Among these advantages, the following can be highlighted.

3.2.1. Sustainability and Environmental Impact

The importance of NBs as a sustainable option in agriculture lies in their renewable origin and lower environmental impact than chemical biostimulants [21].

Generally, the use of NBs has a positive environmental impact [19,48,49]. They can help to reduce or rationalize the amount of synthetic fertilizers and pesticides needed to grow plants [67–69]. For example, some NBs can have a positive effect on microbial communities in the soil and can be beneficial for agricultural practices [11]. In terms of environmental impact, NBs extracted from microorganisms are non-toxic and do not pollute the environment [70,71]. In addition, because they are obtained from natural sources, their production is more sustainable than that of chemical biostimulants.

3.2.2. Security

In contrast to the risks associated with the chemicals used in chemical biostimulants, NBs tend to be safer for both the environment and human health [72].

3.2.3. Broad Spectrum of Activity

NBs have a wide spectrum of activities, which implies multiple benefits for plants in terms of growth, nutrient absorption, stress resistance, flowering, and fruiting quality [20,73].

3.2.4. Positive Interactions

NBs promote beneficial interactions with soil microorganisms, improving soil health and favoring more balanced and productive agricultural systems [49,74].

3.2.5. Regulatory Compliance

NBs offer an easier option for complying with government regulations and restrictions on the use of chemicals in agriculture, which has become more relevant in many countries [6].

3.3. Production Processes of NBs by SSF

Thus, SSF is a promising method for NBs production. SSF produces a variety of bioactive products that promote plant growth, development, and responses to abiotic and biotic stress conditions [75,76]. In this chapter, the processes used to obtain natural biostimulants through SSF were explored, highlighting their importance and efficacy in sustainable agriculture.

3.3.1. Substrate Selection in NB Production by SSF

The appropriate choice of substrates is a crucial step in the production of NBs by SSF [77]. Substrates provide a source of nutrients, energy, and bioactive compounds for microorganisms during fermentation. [78]. The most commonly used substrates in SSF include agricultural residues, agro-industrial waste, food industry by-products, and lignocellulosic materials [16]. These substrates are rich in nutrients and can be degraded by microorganisms, allowing the production of beneficial metabolites [79].

3.3.2. Substrate Pretreatment

Pretreatment of substrates is necessary to improve their composition and nutrient availability. Pretreatment may involve steps such as crushing, grinding, sieving, pH adjustment, sterilization, and addition of nutritional agents [75,80,81]. These steps aim to optimize the conditions for microbial growth and production of desired metabolites [82]. Pretreatment can also facilitate the degradation of substrates and increase fermentation efficiency [83].

3.3.3. Microorganisms for NB Production by SSF and Inoculation

Microorganisms play a fundamental role in the production of NBs by SSF, as they are responsible for substrate degradation and synthesis of bioactive metabolites [84]. In this section, we will focus on the different microorganisms used in this process and their relevance to NB production.

Examples of microorganisms used in SSF for NB production include bacteria, fungi, and yeasts. Each type of microorganism possesses specific characteristics that can influence biostimulant production.

The inoculation of microorganisms is a crucial step in the production of NBs by SSF [18]. Beneficial microorganism strains such as bacteria, fungi, and yeast are selected for their ability to degrade substrates and produce bioactive metabolites. These microorganisms were pre-cultivated under optimal conditions and then inoculated into substrates to initiate SSF [78,84]. The choice of suitable microorganisms and their interactions during SSF influence the composition and final quality of the biostimulant [18].

3.3.4. Control of SSF Conditions

Control of SSF conditions is essential for obtaining high-quality biostimulants through SSF. Parameters such as the temperature, humidity, pH, C/N ratio, moisture content, and process duration must be monitored and adjusted accordingly. These conditions affect the growth and metabolism of microorganisms [15,18]. The precise control of SSF conditions ensures the optimization and quality of the biostimulant.

The production of natural biostimulants through SSF involves the selection of suitable substrates, pretreatment of substrates, inoculation of microorganisms, and control of SSF conditions. These processes are crucial for obtaining high-quality NBs that can promote plant growth.

3.3.5. SSF Bioreactors in NB Production

The use of SSF bioreactors has proven to be a promising technique for improving NB production. These systems allow for better control of fermentation conditions and higher efficiency in obtaining high-quality biostimulants [15].

SSF bioreactors can be designed to maintain optimal cultivation conditions, including temperature, humidity, aeration, and water content [85]. The appropriate selection of the bioreactor depends on various factors, such as the type of microorganism, substrate used, and desired production scale [86]. Common types of SSF bioreactors include fixed-bed, fluidized-bed, and packed-bed bioreactors [79]. The implementation of SSF bioreactors in NB production represents a significant improvement in the efficiency and quality of biostimulants, contributing to a more sustainable and productive agriculture [87,88]. Table 2 presents examples of substrates commonly used in the production of NBs by SSF, together with their characteristics and advantages, microorganism selection, production mode, and bioreactor type.

Substrate	Characteristics and Advantages of Substrate	Microorganism Selection	Production Mode	Bioreactor Type	Refs.
Crop Residues	Abundant local availability, nutrient source, and microorganism support	Bacteria, Fungi	Batch, Continuous, Fed-Batch	Fixed-Bed, Packed-Bed	[89–91]
Agroindustrial Waste	Waste valorization and reduced environmental impact	Filamentous Fungi	Batch, Continuous	Fluidized-Bed, Packed-Bed	[47,92]
Food Residues	Rich in nutrients and organic matter, avoids food waste	Bacteria, Filamentous Fungi	Batch, Fed-Batch	Fixed-Bed, Packed-Bed	[37,93]
Plant Residues	High content of bioactive compounds and phytohormones	Bacteria, Filamentous Fungi	Batch, Continuous	Fluidized-Bed, Packed-Bed	[94,95]
Algal Biomass	Rich in bioactive compounds and auxins	Microalgae	Batch, Fed-Batch	Bubble-Column	[96,97]
Wood Residues	Sustainable source with lignocellulosic content	Filamentous Fungi	Fed-Batch, Continuous	Fluidized-Bed, Packed-Bed	[98,99]
Residual Sludge	Reduces waste volume and provides rich source of nutrients	Bacteria, Filamentous Fungi	Batch, Continuous	Plug-Flow, Packed-Bed	[100,101]
Fishery Waste	Utilization of waste from the fishing industry	Filamentous Fungi	Batch, Continuous	Packed-Bed	[102,103]
Brewery Waste	Valorization of waste from brewing processes	Filamentous Fungi	Continuous	Packed-Bed	[104,105]
Citrus Waste	Abundant source of bioactive compounds and antioxidants	Filamentous Fungi	Batch, Fed-Batch	Fixed-Bed, Packed-Bed	[33,106]
Coffee Residues	Rich in bioactive compounds and promotes soil health	Filamentous Fungi	Batch, Continuous	Packed-Bed	[107]
Rice Husk	Rich in organic matter and bioactive substances	Filamentous Fungi	Fed-Batch, Continuous	Packed-Bed	[108]

Table 2. Comparison of substrates, microorganism selection, production mode, and ioreactor type inNB production by SSF.

4. Methods of NB Production

In this section, the production methods used to obtain NBs through SSF are addressed. The type of biostimulant, microorganisms used in this process, and the optimal conditions of SSF for its production will be described.

4.1. Microorganisms Used in NB Production

In the production of NBs through SSF, various beneficial microorganisms play key roles in substrate degradation and the synthesis of metabolites. Examples of microorganisms used include bacteria, fungi, and yeast. Each type of microorganism possesses specific characteristics that can influence biostimulant production. See Table 2.

4.2. Characteristics of SSF for NB Production

SSF is used to produce natural biostimulants. In this process, microorganisms are cultivated on solid substrates, such as agricultural residues or by-products of the food industry. During fermentation, microorganisms secrete enzymes and bioactive metabolites that transform the compounds present in the substrate into forms that are readily assimilated by plants [18].

The biological activity determines the production of NBs and warrants particular attention in future research. Table 3 presents examples of substrate microorganisms used to obtain different natural biostimulants (NBs) through SSF.

				Pretreatment			SF Conditions		
NB	Substrate	Microorganism	Trituration	рН	Sterilization	Moisture %	Temperature °C	Effect of NBs on Crop	Refs.
IAA	Pruning Waste + Grass	Trichoderma harzianum	1 cm	6.8	2 times	74	25		[15]
IAA	Yuca Bagasse Soy Bran Wheat Bran Sorghum Dried Distiller's Grains Corn Dried Distiller's Grains	Aspergillus flavipes Aspergillus ustus Bacillus subtilis Bacillus megaterium Bacillus amyloliquefaciens Trichoderma atroviride Trichoderma koningii Trichoderma harzianum	0.5, 1.0 y > 1.0 mm			50	Room Temperature	Clon IPB2 Eucalyptus grandis and Eucalyptus urophylla Increasing Rooting	[14,18]
Kinetin	Cow Dung + Leaf Litter	Selenomonas ruminantium	2–5 mm	6.9		70–75	25 ± 3		[29]
ABA	Millet Rice	Botrytis cinerea	Millet and Rice		1 time		26.5–25.5		[32]
GA3	Rice Bran	Gibberella fujikuroi			50 °C	65.95%	28 ± 2		[109]
GA3	Corn Cob Residues	Aspergillus niger		5.1		24%			[110]
GA3	Citric Pulp	Fusarium moniliforme LPB03 + Gibberella fujikuroi		5.5–5.8		75	29		[91]
Alginic Acids	Apple Peels	Azotobacter vinelandii, NRRL-14641	0.1 mm	7	60 °C	70	37.5		[39]
Alginic Acids	Sargassum Macroalgae	Cunninghamella echinulate Aspergillus niger Penicillium oxalicum		7–8.5	1 time 121 °C	65–75	28–30		[40]
Fucoida	Seaweed Fucus Vesiculosus	Aspergillus niger Mucor sp				80	30		[42]
Oligosaccharides	Soybean Meal	-					Room Temperature	Effect on Germination	[111]
Chitin Oligosac- charides	Powder of Molting of Mealworms	Talaromyces allahabadensis Hi-4 Talaromyces funiculosus		6			40		[112]

Table 3. Methods of NB production by SSF.

Table 3. Cont.

			Pretreatment			Optimal SSF Conditions			
NB	Substrate	Microorganism	Trituration	pН	Sterilization	Moisture %	Temperature °C	Effect of NBs on Crop	Refs.
Humic Acid	Oil Palm Empty Fruit Bunch	Trichoderma reesei		6		64–72	30		[50,113]
Fulvic Acid	Sugarcane Bagasse	Trichoderma Sp.				70	20		[114]
L-proline	Wheat Straw Ice Straw Wheat Bran Corn Cob Corn Stover	Fomitopsis sp.	Small Pieces	5.5			25–30		[56]
Low Molecular Weight Peptides	Chickpeas	Bacillus subtilis							[60]
Siderophores	Soybean Protein Meal	Lactobacillus plantarum					37		[115]
Chitosan Fungal	Sweet Potato	Gongronella butleri USDB 0201					28		[66]

As detailed in previous chapters, NBs have a significant impact on crop growth, development, and yield. The following are examples of observed effects on different aspects of crop production, supported by scientific studies.

4.3.1. Improvement of Plant Growth and Development

The application of NBs promotes root growth, increases plant biomass, improves plant architecture, and enhances seed germination and seedling emergence. These effects are attributed to the presence of specific molecules in NBs, such as low molecular weight peptides, gibberellic acid (GA3), and indole-3-acetic acid (IAA) [116–119].

Table 4 summarizes the effects of NBs on crop growth and development.

Table 4. Effect of NBs on improving plant growth and development.

Сгор	NB Туре	Effect	Scale	Refs.
Arabidopsis thaliana	Low Molecular Weight Peptides	Increase in plant biomass	Laboratory	[120]
Sesame	GA3	Improvement of plant architecture	Laboratory	[121]
Rice	GA3	Improvement of plant architecture	Laboratory	[122]
Tomato Pepper Seed Arabidopsis Orchid	IAA	Promotion of seed germination and seedling emergence	Greenhouse Laboratory	[17,123,124]

4.3.2. Increased Resistance to Adverse Conditions

In addition to improving plant growth and development, NBs also enhance the resilience of crops against adverse conditions. It has been observed that certain molecules present in NBs, such as ABA and seaweed polysaccharides, contribute to increased tolerance to abiotic stress, enhanced disease and pest resistance, and protection against oxidative stress [125,126].

Table 5 summarizes some NBs and their effects on resistance to adverse conditions.

Crop	NB Туре	Effect	Scale	Refs.
Orange Tobacco Corn	ABA	Abiotic stress tolerance	Laboratory	[127–129]
Strawberry Bean Vine Cucumber	Seaweed Polysaccharides	Resistance to diseases and pests	Field	[130–133]

4.3.3. Effect of NBs on Improving Crop Quality

In this section, we will explore scientific studies that have investigated the influence of different NBs on improving the quality of various crops. Aspects such as nutritional content, physical appearance, shelf life, and resistance to stress will be addressed (Table 6). These findings provide a solid foundation for understanding the potential of NBs for enhancing crop quality and open new perspectives for their application in sustainable agriculture.

Crop	NB Туре	Effect	Scale	Refs.
Gerbera Tectona Grandis Peas Yarrow	Humic Acid	Increased nutrient concentration	Greenhouse	[134–137]
Tomato Apple	Amino Acids	Improved organoleptic quality	Greenhouse	[138–140]
Soy Petunia Flowers Lettuce	Cytokinins	Delayed tissue senescence	Greenhouse	[141–143]

Table 6. Effect of NBs on enhancing resistance to adverse conditions.

4.3.4. Optimization of Nutrient Use Efficiency

In this section, we focus on optimizing nutrient use efficiency in crops through the use of NBs. Nutrient use efficiency is a key factor in agricultural production as it directly influences the absorption, assimilation, and utilization of nutrients by plants. NBs have been demonstrated to be an effective tool for improving this efficiency and maximizing crop yield. Table 7 presents evidence of how NBs enhance nutrient use efficiency.

Table 7. Effect of NBs on optimal nutrient use.

Crop	NB Type	Effect	Scale	Refs.
Tomato		Improvement of		
Strawberries	Alginic Acids	nutrient availability	Greenhouse	[144–146]
Peanut		in the soil		
French Marigold	Oligosaccharides	Reduced nutrient losses	Greenhouse	[147,148]

4.3.5. Effect NBs on Agricultural Productivity

NBs are a promising tool for enhancing crop efficiency and productivity as well as addressing current challenges in agriculture. In this section, examples of studies demonstrating the positive effects of natural biostimulants on agricultural productivity are presented, highlighting the results obtained in different crops and the NBs involved (Table 8).

Table 8. Effect of NBs on agricultural productivity.

Crop	NB Туре	Effect of Productivity on Crops	Scale	Refs.
Corn	Seaweed Extract	Increases grain yield, crop residue, and improves nutritional quality	Field	[149–151]
Grapes	Seaweed Extract	Increases grape production, improves stress resistance, and increases polyphenol content	Greenhouse	[152–154]
Tomato	Seaweed Extract	Increases fruit yield and quality	Greenhouse	[155–157]
Lettuce	Seaweed Extract	Higher yield increase and increases shoot growth	Greenhouse	[158–160]
Strawberries	Seaweed Extract	Improves fruit quality and flavor, higher yield	Greenhouse	[132,161]
Onion	Seaweed Extract	Increases bulb diameter and weight	Field	[162,163]
Potato	Seaweed Extract	Increases tuber yield and quality	Field	[164,165]
Corn	IAA	Stimulates vegetative growth and increases grain production	Greenhouse	[166–168]
Lettuce	IAA	Increases biomass	Greenhouse	[169]
Potato	IAA	Promotes tuber growth and improves yield	Greenhouse	[170–172]
Onion	IAA	Increases bulb size and enhances production	Greenhouse Laboratory	[173–175]
Quinoa	IAA	Boosts grain yield and improves quality	Field	[176,177]

Crop	NB Туре	Effect of Productivity on Crops	Scale	Refs.
Wheat	IAA	Stimulates plant growth and increases yield	Field	[178,179]
Tomato	IAA	Improves rooting, increases fruit production, and enhances antioxidant content	Greenhouse	[180,181]
Soybean	IAA	Improves root development and increases production	Greenhouse	[182,183]
Rice	IAA	Promotes rooting and improves yield	Field	[184,185]
Broad Beans	IAA	Stimulates vegetative growth and increases production	Greenhouse	[183,186]
Grapes	IAA	Enhances root formation and increases yield	Greenhouse	[187–189]
Corn	Cytokinins	Stimulates cell division and increases yield	Greenhouse	[190,191]
Rice	Cytokinins	Promotes grain growth and improves yield	Greenhouse	[192,193]
Wheat	Cytokinins	Increases the number of grains per spike and improves production	Field	[194–196]
Soybean	Cytokinins	Improves vegetative growth and increases production	Greenhouse	[197,198]
Tomato	Cytokinins	Stimulates flower formation and increases yield	Greenhouse	[28,199]
Potato	Cytokinins	Promotes tuber development and improves yield	Field	[200,201]
Grapes	Cytokinins	Enhances cluster size and quality	Greenhouse	[202,203]
Strawberry	Cytokinins	Increases stolon formation and improves production	Greenhouse	[204,205]
Strawberry	Cytokinins	Stimulates bud break and improves yield	Greenhouse	[206]
Citrus	Cytokinins	Increases fruit size and improves production	Greenhouse	[207,208]
Onion	Humic Acids	Enhances bulb yield, improves quality and disease resistance	Greenhouse	[209,210]
Corn	Humic Acids	Improves nutrient absorption and increases yield	Greenhouse	[28,211]
Wheat	Humic Acids	Increases grain size and weight	Greenhouse	[212,213]
Rice	Humic Acids	Boosts the number of spikes and improves production	Greenhouse	[214,215]
Tomato	Humic Acids	Enhances fruit quality and increases yield	Greenhouse	[216,217]
Beans	Humic Acids	Improves vegetative growth and increases production	Field	[218]
Onion	Humic Acids	Increases bulb size and quality	Greenhouse	[219,220]
Carrot	Humic Acids	Promotes root development and improves production	Greenhouse	[221]
Lettuce	Humic Acids	Stimulates leaf growth and increases yield	Greenhouse	[222]

Table 8. Cont.

4.4. Limitations and Challenges of NBs by SSF

Despite the benefits of NBs in sustainable agriculture, some limitations and challenges need to be considered. These aspects can affect their practical application and widespread adoption in agricultural production. Some of the main limitations and challenges of this study are as follows.

4.4.1. Standardization Issues in NB Production by SSF

In this section, we address some standardization issues that may arise in the process of NB production by SSF. Although SSF offers advantages in terms of cost, efficiency, and small-scale production, there are challenges that need to be addressed to achieve standardized and consistent production of high-quality biostimulants [223]. The following are some common limitations.

Substrate variability: the choice of substrate used in SSF can vary depending on the type of microorganism and production objective. However, the chemical composition and physical properties of substrates can vary, which could affect the quality of NBs.

Control of SSF conditions: SSF conditions, such as temperature, humidity, pH, and substrate/microorganism ratio, are crucial for the growth and activity of microorganisms. Without proper control of these conditions, there may be variations in the production of bioactive metabolites and enzymes [79], which can affect the quality and efficacy of NBs.

Scalability of production: the large-scale production of NBs by SSF can be challenging because of the need to maintain optimal fermentation conditions and ensure the quality of the final product. Scalability of production requires optimization of fermentation parameters, selection of suitable equipment, and design of efficient processes that meet quality standards and market demands [47].

Addressing these standardization issues in the production of NBs by SSF will require a combination of scientific research, development of new methodologies, collaboration between academia, industry, and regulatory bodies, and the adoption of good manufacturing practices. These efforts will contribute to ensuring the quality, consistency, and efficacy of NBs produced by SSF, thereby facilitating their reliable and sustainable application in agriculture.

4.4.2. Challenges in the Application of NBs from SSF in Sustainable Agriculture

In this chapter, we explore some difficulties that may arise in the application of NBs produced by SSF in sustainable agriculture. Although NBs offer numerous benefits for improving crop performance and quality, as shown in Table 7, there are still specific challenges related to their application in sustainable agricultural systems. The following are some possible difficulties.

Regulation and Standards: the lack of updated regulations in many countries regarding the use of NBs can hinder their application in sustainable agriculture, as evidenced by a critical analysis [224]. The lack of clear definitions and standards can create uncertainty regarding dosing and the frequency of application, which could hinder their widespread adoption.

Interaction with other inputs: the interaction of NBs with other inputs can be complex and may require adjustments in application practices to avoid possible negative interactions or decrease in product efficacy [225]. In sustainable agriculture, it is common to use multiple inputs such as organic fertilizers, biological pesticides, and beneficial microorganisms.

Adaptability to different crops and agronomic conditions: NBs can have different effects depending on crop type and agronomic conditions [20]. Some NBs may work more effectively on certain crops or at certain phenological stages, requiring a detailed understanding of their mode of action and proper adaptation to the specific conditions of each crop.

Farmer capacity building: the adoption of NBs in sustainable agriculture may require increased awareness and knowledge among farmers [226]. It is important to educate farmers about the benefits and proper use of NBs, as well as providing training and technical assistance to maximize their effectiveness on crops.

Overcoming these difficulties in the application of NBs produced by SSF in sustainable agriculture requires a comprehensive approach involving researchers, farmers, businesses, and the government. It is important to encourage the research and development of best practices, establish clear regulations, and promote training and awareness among key players in the agricultural supply chain.

4.4.3. Factors Limiting the Effectiveness of Natural Biostimulants Produced by SSF in Different Crops

The effectiveness of NBs produced by SSF can be influenced by various factors in different crops. Some of these factors include the genetic variability of crop varieties, environmental conditions, such as temperature and humidity, and nutrient availability in the soil. Additionally, NBs produced by SSF interact with other agricultural inputs, such as fertilizers and pesticides. NBs are not a universal solution and should be combined with good agricultural practices such as crop rotation and proper soil management, which can affect their effectiveness [169,227]. Further research is needed to better understand the response of different crops to NBs produced by SSF and to optimize SSF conditions, valorizing waste to maximize their benefits in sustainable agriculture.

5. Conclusions and Future Research Perspectives

5.1. Conclusions

In this section, we present our conclusions and future research perspectives regarding the production of NBs from SSF. In this review, we have analyzed the use of NBs in agriculture, their production by SSF, and their effects on crops. The main conclusions derived from this study are as follows:

NBs are a promising tool to improve crop development and performance. Their use can contribute to more sustainable agriculture by reducing reliance on synthetic chemicals.

SSF is an efficient technique for producing NBs from organic substrates. This method offers several advantages, such as the valorization of agricultural and agro-industrial waste.

NBs act through various bioactive molecules, such as auxins, cytokinins, alginic acids, humic acids, and other compounds. These molecules can modulate physiological and metabolic processes in plants, improving nutrient uptake, rooting, biotic and abiotic stress tolerance, and crop quality.

However, challenges and limitations still need to be addressed to maximize the effectiveness of NBs. These include standardization of production, optimization of dosages and application, adaptations to different crops and environmental conditions, and understanding interactions with other agricultural inputs.

5.2. Future Research Prospects

The following are future research perspectives. A multidisciplinary approach is required to advance the field of NBs from SSF. Some promising areas of research include the following.

Further studies are needed on the mechanisms of action of NBs at the molecular and cellular levels. This will help to better understand how they interact with plants and modulate specific physiological processes.

Research on the optimization of NB production processes produced by SSF. This involves improving the substrates, selecting efficient microorganisms, and optimizing SSF conditions to obtain high-quality and consistent products.

Investigation of the effectiveness of NBs in different agricultural systems and environmental conditions. This includes field and greenhouse studies that analyze the impact of biostimulants on various crops, regions, and agricultural practices.

Research on the interaction of NBs with other agricultural inputs, such as bio-fertilizers and bio-pesticides is needed to optimize their combined use and minimize potential negative effects.

In conclusion, NBs produced by SSF have significant potential for improving agricultural productivity and promoting sustainable farming practices. However, further research, development, and innovation are needed to overcome these challenges and maximize their efficacy for different crops and environmental conditions. An integrated approach that combines scientific research, collaboration among different stakeholders, and the implementation of science-based agricultural practices is essential to fully harness the benefits of NBs in sustainable agriculture.

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Abbreviations

Natural biostimulants	(NBs)
Solid-state fermentation	(SSF)
The European Biostimulants Industry Council	(EBIC)
Humic substances	(HS)
Hormone-containing products	(HCP)
Amino-acid-containing products	(AACP)
Indole-3-acetic acid	(IAA)
Abscisic acid	(ABA)

References

- 1. 2.4.1 Agricultural Sustainability | Sustainable Development Goals | Food and Agriculture Organization of the United Nations. Available online: https://www.fao.org/sustainable-development-goals/indicators/241/en/ (accessed on 23 May 2023).
- 2. Sumberg, J.; Giller, K.E. What Is 'Conventional' Agriculture? *Glob. Food Secur.* 2022, 32, 100617. [CrossRef]
- Kauffman, G.L.; Kneivel, D.P.; Watschke, T.L. Effects of a Biostimulant on the Heat Tolerance Associated with Photosynthetic Capacity, Membrane Thermostability, and Polyphenol Production of Perennial Ryegrass. Crop Sci. 2007, 47, 261–267. [CrossRef]
- 4. Wong, W.S.; Tan, S.N.; Ge, L.; Chen, X.; Letham, D.S.; Yong, J.W.H. The Importance of Phytohormones and Microbes in Biostimulants: Mass Spectrometric Evidence and Their Positive Effects on Plant Growth. *Acta Hortic.* **2016**, *1148*, 49–60. [CrossRef]
- Boivin, S.; Fonouni-Farde, C.; Frugier, F. How Auxin and Cytokinin Phytohormones Modulate Root Microbe Interactions. *Front. Plant Sci.* 2016, 7, 1240. [CrossRef]
- 6. du Jardin, P. Plant Biostimulants: Definition, Concept, Main Categories and Regulation. Sci. Hortic. 2015, 196, 3–14. [CrossRef]
- 7. Sauer, M.; Robert, S.; Kleine-Vehn, J. Auxin: Simply Complicated. J. Exp. Bot. 2013, 64, 2565–2577. [CrossRef]
- 8. Rady, M.M.; Desoky, E.-S.M.; Elrys, A.S.; Boghdady, M.S. Can Licorice Root Extract Be Used as an Effective Natural Biostimulant for Salt-Stressed Common Bean Plants? *S. Afr. J. Bot.* **2019**, *121*, 294–305. [CrossRef]
- 9. Kurepin, L.V.; Zaman, M.; Pharis, R.P. Phytohormonal Basis for the Plant Growth Promoting Action of Naturally Occurring Biostimulators. J. Sci. Food Agric. 2014, 94, 1715–1722. [CrossRef] [PubMed]
- 10. Posmyk, M.M.; Szafrańska, K. Biostimulators: A New Trend towards Solving an Old Problem. Front. Plant Sci. 2016, 7, 748.
- 11. Hellequin, E.; Monard, C.; Chorin, M.; Le bris, N.; Daburon, V.; Klarzynski, O.; Binet, F. Responses of Active Soil Microorganisms Facing to a Soil Biostimulant Input Compared to Plant Legacy Effects. *Sci. Rep.* **2020**, *10*, 13727. [CrossRef]
- 12. EBIC—The European Biostimulants Industry Council. Available online: https://biostimulants.eu/ (accessed on 8 March 2022).
- 13. Biological Products Industry Alliance | Advancing Knowledge About Biopesticides & Biostimulants. Available online: https://www.bpia.org/ (accessed on 23 April 2023).
- 14. do Prado, D.Z.; Okino-Delgado, C.H.; Zanutto-Elgui, M.R.; da Silva, R.B.G.; Pereira, M.S.; Jahn, L.; Ludwig-Müller, J.; da Silva, M.R.; Velini, E.D.; Fleuri, L.F. Screening of Aspergillus, Bacillus and Trichoderma Strains and Influence of Substrates on Auxin and Phytases Production through Solid-State Fermentation. *Biocatal. Agric. Biotechnol.* **2019**, *19*, 101165. [CrossRef]
- 15. Ghoreishi, G.; Barrena, R.; Font, X. Using Green Waste as Substrate to Produce Biostimulant and Biopesticide Products through Solid-State Fermentation. *Waste Manag.* **2023**, 159, 84–92. [CrossRef]
- 16. Chen, H. Biotechnology Principles of Solid State Fermentation. Mod. Solid State Ferment. 2013, 23–74. [CrossRef]
- 17. Sanchez-Montesinos, B.; Dianez, F.; Moreno-Gavira, A.; Gea, F.J.; Santos, M. Role of Trichoderma Aggressivum f. Europaeumas Plant-Growth Promoter in Horticulture. *Agronomy* **2020**, *10*, 1004. [CrossRef]
- Do Prado, D.Z.; Oliveira, S.L.; Okino-Delgado, C.H.; Auer, S.; Ludwig-Müller, J.; da Silva, M.R.; da Costa Fernandes, C.J.; Carbonari, C.A.; Zambuzzi, W.F.; Fleuri, L.F. Aspergillus Flavipes as a Novel Biostimulant for Rooting-Enhancement of Eucalyptus. J. Clean. Prod. 2019, 234, 681–689. [CrossRef]
- 19. Puglia, D.; Pezzolla, D.; Gigliotti, G.; Torre, L.; Bartucca, M.L.; Del Buono, D. The Opportunity of Valorizing Agricultural Waste, Through Its Conversion into Biostimulants, Biofertilizers, and Biopolymers. *Sustainability* **2021**, *13*, 2710. [CrossRef]
- 20. Zulfiqar, F.; Casadesús, A.; Brockman, H.; Munné-Bosch, S. An Overview of Plant-Based Natural Biostimulants for Sustainable Horticulture with a Particular Focus on Moringa Leaf Extracts. *Plant Sci.* **2020**, 295, 110194. [CrossRef]
- 21. Calvo, P.; Nelson, L.; Kloepper, J.W. Agricultural Uses of Plant Biostimulants. Plant Soil 2014, 383, 3–41. [CrossRef]
- Yu, X.; Li, Y.; Cui, Y.; Liu, R.; Li, Y.; Chen, Q.; Gu, Y.; Zhao, K.; Xiang, Q.; Xu, K.; et al. An Indoleacetic Acid-Producing Ochrobactrum Sp. MGJ11 Counteracts Cadmium Effect on Soybean by Promoting Plant Growth. J. Appl. Microbiol. 2017, 122, 987–996. [CrossRef]
- Karnwal, A.; Dohroo, A. Effect of Maize Root Exudates on Indole-3-Acetic Acid Production by Rice Endophytic Bacteria under Influence of L-Tryptophan. F1000Research 2018, 7, 112. [CrossRef] [PubMed]
- Do, T.C.V.; Tran, D.T.; Le, T.G.; Nguyen, Q.T. Characterization of Endogenous Auxins and Gibberellins Produced by *Chlorella* sorokiniana TH01 under Phototrophic and Mixtrophic Cultivation Modes toward Applications in Microalgal Biorefinery and Crop Research. J. Chem. 2020, 2020, e4910621. [CrossRef]

- Ma, D.; Liu, B.; Ge, L.; Weng, Y.; Cao, X.; Liu, F.; Mao, P.; Ma, X. Identification and Characterization of Regulatory Pathways Involved in Early Flowering in the New Leaves of Alfalfa (*Medicago sativa* L.) by Transcriptome Analysis. *BMC Plant Biol.* 2021, 21, 8. [CrossRef] [PubMed]
- 26. Rusmin, D.; Basmal, J.; Kusumawati, R.; Darwati, I. Improving the Growth of Clove Seedlings by the Application of Seaweed Waste as Organic Fertilizers. *IOP Conf. Ser. Earth Environ. Sci.* **2020**, *418*, 012029. [CrossRef]
- 27. Jain, P.; Farooq, B.; Lamba, S.; Koul, B. Foliar Spray of Moringa Oleifera Lam. Leaf Extracts (MLE) Enhances the Stevioside, Zeatin and Mineral Contents in Stevia Rebaudiana Betoni. S. Afr. J. Bot. 2020, 132, 249–257. [CrossRef]
- 28. Basra, S.M.A.; Lovatt, C.J. Exogenous Applications of Moringa Leaf Extract and Cytokinins Improve Plant Growth, Yield, and Fruit Quality of Cherry Tomato. *HortTechnology* **2016**, *26*, 327–337. [CrossRef]
- 29. Ravindran, B.; Wong, J.W.C.; Selvam, A.; Sekaran, G. Influence of Microbial Diversity and Plant Growth Hormones in Compost and Vermicompost from Fermented Tannery Waste. *Bioresour. Technol.* 2016, 217, 200–204. [CrossRef]
- Ravindran, B.; Contreras-Ramos, S.M.; Sekaran, G. Changes in Earthworm Gut Associated Enzymes and Microbial Diversity on the Treatment of Fermented Tannery Waste Using Epigeic Earthworm Eudrilus Eugeniae. *Ecol. Eng.* 2015, 74, 394–401. [CrossRef]
- Ali, H.M.; Khan, H.Z.; Afzal, I. Exogenous application of growth promoting substances improves growth, yield and quality of spring maize (*Zea mays* L.) hybrids under late sown conditions. *Bull. Biol. Allied Sci. Res.* 2017, 2017, 9. [CrossRef]
- Qi, H. Method for Producing Abscisic Acid by Solid State Fermentation of Fungi 2013. Available online: https://patents.google. com/patent/CN103409474A/en (accessed on 6 June 2023).
- Vandenberghe, L.P.S.; Pandey, A.; Carvalho, J.C.; Letti, L.A.J.; Woiciechowski, A.L.; Karp, S.G.; Thomaz-Soccol, V.; Martínez-Burgos, W.J.; Penha, R.O.; Herrmann, L.W.; et al. Solid-State Fermentation Technology and Innovation for the Production of Agricultural and Animal Feed Bioproducts. *Syst. Microbiol. Biomanuf.* 2021, 1, 142–165. [CrossRef]
- 34. Rodrigues, C.; Vandenberghe, L.P.D.S.; De Oliveira, J.; Soccol, C.R. New Perspectives of Gibberellic Acid Production: A Review. *Crit. Rev. Biotechnol.* **2012**, *32*, 263–273. [CrossRef]
- Yang, S.; Xie, J.; Hu, N.; Liu, Y.; Zhang, J.; Ye, X.; Liu, Z. Bioconversion of Gibberellin Fermentation Residue into Feed Supplement and Organic Fertilizer Employing Housefly (*Musca domestica* L.) Assisted by Corynebacterium variabile. *PLoS ONE* 2015, 10, e0110809. [CrossRef] [PubMed]
- Camara, M.C.; Vandenberghe, L.P.S.; Rodrigues, C.; de Oliveira, J.; Faulds, C.; Bertrand, E.; Soccol, C.R. Current Advances in Gibberellic Acid (GA3) Production, Patented Technologies and Potential Applications. *Planta* 2018, 248, 1049–1062. [CrossRef]
- 37. Brückner, B.; Blechschmidt, D. The Gibberellin Fermentation. Crit. Rev. Biotechnol. 1991, 11, 163–192. [CrossRef]
- Maria, C.; Machado, M.; Soccol, C.R. Gibberellic Acid Production. In *Current Developments in Solid-State Fermentation*; Pandey, A., Soccol, C.R., Larroche, C., Eds.; Springer: New York, NY, USA, 2008; pp. 277–301. ISBN 978-0-387-75213-6.
- Saeed, S.; Mehmood, T.; Irfan, M. Statistical Optimization of Cultural Parameters for the Optimized Production of Alginic Acid Using Apple (Malus Domestica) Peels through Solid-State Fermentation. *Biomass Conv. Bioref.* 2023, 13, 1269–1277. [CrossRef]
- 40. Dos Santos Silva, M.C.; De Farias Silva, C.E.; dos Santos, L.M.; Medeiros, J.A.; Vieira, R.C.; de Souza Abud, A.K.; Almeida, R.M.R.G.; Tonholo, J. Alginate Lyase Produced by Filamentous Fungus Through Solid State Fermentation Using Sargassum from the Brazilian Coast. *Waste Biomass Valor.* 2022, *13*, 2947–2962. [CrossRef]
- 41. Wozniak, E.; Blaszczak, A.; Wiatrak, P.; Canady, M. Biostimulant Mode of Action; Wiley Online Library: Hoboken, NJ, USA, 2020.
- Rodríguez-Jasso, R.M.; Mussatto, S.I.; Sepúlveda, L.; Agrasar, A.T.; Pastrana, L.; Aguilar, C.N.; Teixeira, J.A. Fungal Fucoidanase Production by Solid-State Fermentation in a Rotating Drum Bioreactor Using Algal Biomass as Substrate. *Food Bioprod. Process.* 2013, 91, 587–594. [CrossRef]
- 43. Bergé, J.P. Marine Biotechnology: An Overview of Leading Field. In Proceedings of the IXth ESMB Meeting, Nantes, France, 12–14 May 2002.
- Patel, J.S.; Selvaraj, V.; More, P.; Bahmani, R.; Borza, T.; Prithiviraj, B. A Plant Biostimulant from Ascophyllum Nodosum Potentiates Plant Growth Promotion and Stress Protection Activity of Pseudomonas Protegens CHA0. *Plants* 2023, *12*, 1208. [CrossRef]
- Zou, P.; Yang, X.; Yuan, Y.; Jing, C.; Cao, J.; Wang, Y.; Zhang, L.; Zhang, C.; Li, Y. Purification and Characterization of a Fucoidan from the Brown Algae Macrocystis Pyrifera and the Activity of Enhancing Salt-Stress Tolerance of Wheat Seedlings. *Int. J. Biol. Macromol.* 2021, 180, 547–558. [CrossRef]
- 46. Baltazar, M.; Correia, S.; Guinan, K.J.; Sujeeth, N.; Bragança, R.; Gonçalves, B. Recent Advances in the Molecular Effects of Biostimulants in Plants: An Overview. *Biomolecules* **2021**, *11*, 1096. [CrossRef]
- Mattedi, A.; Sabbi, E.; Farda, B.; Djebaili, R.; Mitra, D.; Ercole, C.; Cacchio, P.; Del Gallo, M.; Pellegrini, M. Solid-State Fermentation: Applications and Future Perspectives for Biostimulant and Biopesticides Production. *Microorganisms* 2023, *11*, 1408. [CrossRef]
 Colla, G.; Rouphael, Y. Biostimulants in Horticulture. *Sci. Hortic.* 2015, *196*, 1–2. [CrossRef]
- 49. Tarafdar, J.C. Chapter 15—Biostimulants for Sustainable Crop Production. In *New and Future Developments in Microbial Biotechnology and Bioengineering*; Singh, H.B., Vaishnav, A., Eds.; Elsevier: Amsterdam, The Netherlands, 2022; pp. 299–313, ISBN 978-0-323-85579-2.
- Solid-State Fermentation for Humic Acids Production by a Trichoderma Reesei Strain Using an Oil Palm Empty Fruit Bunch as the Substrate | SpringerLink. Available online: https://link.springer.com/article/10.1007/s12010-013-0668-2 (accessed on 27 May 2023).

- 51. Zhang, Y.; Dou, S.; Hamza, B.; Ye, S.; Zhang, D. Mechanisms of Three Fungal Types on Humic-Like Substances Formation during Solid-State Fermentation of Corn Straw. *Int. J. Agric. Biol.* **2020**, *23*, 970–976.
- 52. Yang, Y.; Wang, L.; Zhang, Y.; Li, L.; Shi, X.; Liu, X.; Ren, X.; Dou, S. Transformation of Corn Stalk Residue to Humus-like Substances during Solid-State Fermentation. *Sustainability* **2019**, *11*, 6771. [CrossRef]
- Bettoni, M.M.; Mogor, Á.F.; Pauletti, V.; Goicoechea, N. Growth and Metabolism of Onion Seedlings as Affected by the Application of Humic Substances, Mycorrhizal Inoculation and Elevated CO₂. Sci. Hortic. 2014, 180, 227–235. [CrossRef]
- 54. Hölker, U.; Höfer, M. Solid Substrate Fermentation of Lignite by the Coal-Solubilizing Mould, Trichoderma Atroviride, in a New Type of Bioreactor. *Biotechnol. Lett.* **2002**, *24*, 1643–1645. [CrossRef]
- 55. Canellas, L.P.; Olivares, F.L.; Aguiar, N.O.; Jones, D.L.; Nebbioso, A.; Mazzei, P.; Piccolo, A. Humic and Fulvic Acids as Biostimulants in Horticulture. *Sci. Hortic.* 2015, *196*, 15–27. [CrossRef]
- Deswal, D.; Khasa, Y.P.; Kuhad, R.C. Optimization of Cellulase Production by a Brown Rot Fungus Fomitopsis Sp. RCK2010 under Solid State Fermentation. *Bioresour. Technol.* 2011, 102, 6065–6072. [CrossRef] [PubMed]
- 57. Sodhi, H.K.; Sharma, K.; Gupta, J.K.; Soni, S.K. Production of a Thermostable α-Amylase from Bacillus Sp. PS-7 by Solid State Fermentation and Its Synergistic Use in the Hydrolysis of Malt Starch for Alcohol Production. *Process Biochem.* 2005, 40, 525–534. [CrossRef]
- Lucini, L.; Rouphael, Y.; Cardarelli, M.; Canaguier, R.; Kumar, P.; Colla, G. The Effect of a Plant-Derived Biostimulant on Metabolic Profiling and Crop Performance of Lettuce Grown under Saline Conditions. *Sci. Hortic.* 2015, *182*, 124–133. [CrossRef]
- 59. Asri, N.M.; Muhialdin, B.J.; Zarei, M.; Saari, N. Low Molecular Weight Peptides Generated from Palm Kernel Cake via Solid State Lacto-Fermentation Extend the Shelf Life of Bread. *LWT* **2020**, *134*, 110206. [CrossRef]
- 60. Li, W.; Wang, T. Effect of Solid-State Fermentation with Bacillus Subtilis Lwo on the Proteolysis and the Antioxidative Properties of Chickpeas. *Int. J. Food Microbiol.* **2021**, *338*, 108988. [CrossRef] [PubMed]
- 61. Cai, C.; Hua, Y.; Liu, H.; Dai, X. A New Approach to Recycling Cephalosporin Fermentation Residue into Plant Biostimulants. *J. Hazard. Mater.* **2021**, *413*, 125393. [CrossRef] [PubMed]
- 62. Prabhu, G.N.; Bindu, P. Optimization of Process Parameters for Siderophore Production Under Solid State Fermentation Using Polystyrene Beads as Inert Support. *JSIR* 2016, 75, 621–625.
- 63. Le, H.; ZongHao, Y.; Can, C.; ChunYan, L.; Juan, L.; ZhongKe, S. Enhancing Iron Uptake and Alleviating Iron Toxicity in Wheat by Plant Growth-Promoting Bacteria: Theories and Practices. *Int. J. Agric. Biol.* **2020**, *23*, 190–196.
- 64. Marschner, H.; Römheld, V.; Kissel, M. Different Strategies in Higher Plants in Mobilization and Uptake of Iron. *J. Plant Nutr.* **1986**, *9*, 695–713. [CrossRef]
- Stanley-Raja, V.; Senthil-Nathan, S.; Chanthini, K.M.P.; Sivanesh, H.; Ramasubramanian, R.; Karthi, S.; Shyam-Sundar, N.; Vasantha-Srinivasan, P.; Kalaivani, K. Biological Activity of Chitosan Inducing Resistance Efficiency of Rice (*Oryza sativa* L.) after Treatment with Fungal Based Chitosan. *Sci. Rep.* 2021, *11*, 20488. [CrossRef] [PubMed]
- 66. Nwe, N.; Chandrkrachang, S.; Stevens, W.F.; Maw, T.; Tan, T.K.; Khor, E.; Wong, S.M. Production of Fungal Chitosan by Solid State and Submerged Fermentation. *Carbohydr. Polym.* **2002**, *49*, 235–237. [CrossRef]
- 67. Rouphael, Y.; Colla, G. Editorial: Biostimulants in Agriculture. Front. Plant Sci. 2020, 11, 40. [CrossRef]
- 68. Lau, S.-E.; Teo, W.F.A.; Teoh, E.Y.; Tan, B.C. Microbiome Engineering and Plant Biostimulants for Sustainable Crop Improvement and Mitigation of Biotic and Abiotic Stresses. *Discov. Food* **2022**, *2*, *9*. [CrossRef]
- 69. Ben Mrid, R.; Benmrid, B.; Hafsa, J.; Boukcim, H.; Sobeh, M.; Yasri, A. Secondary Metabolites as Biostimulant and Bioprotectant Agents: A Review. *Sci. Total Environ.* **2021**, 777, 146204. [CrossRef]
- Caccavo, V.; Forlano, P.; Mang, S.M.; Fanti, P.; Nuzzaci, M.; Battaglia, D.; Trotta, V. Effects of Trichoderma Harzianum Strain T22 on the Arthropod Community Associated with Tomato Plants and on the Crop Performance in an Experimental Field. *Insects* 2022, 13, 418. [CrossRef]
- 71. Ferrigo, D.; Raiola, A.; Rasera, R.; Causin, R. Trichoderma Harzianum Seed Treatment Controls Fusarium Verticillioides Colonization and Fumonisin Contamination in Maize under Field Conditions. *Crop Prot.* **2014**, *65*, 51–56. [CrossRef]
- 72. Xu, L.; Geelen, D. Developing Biostimulants from Agro-Food and Industrial By-Products. Front. Plant Sci. 2018, 9, 1567. [CrossRef]
- 73. Francesca, S.; Arena, C.; Mele, B.H.; Schettini, C.; Ambrosino, P.; Barone, A.; Rigano, M.M. The Use of a Plant-Based Biostimulant Improves Plant Performances and Fruit Quality in Tomato Plants Grown at Elevated Temperatures. *Agronomy* **2020**, *10*, 363. [CrossRef]
- 74. Nanda, S.; Kumar, G.; Hussain, S. Utilization of Seaweed-Based Biostimulants in Improving Plant and Soil Health: Current Updates and Future Prospective. *Int. J. Environ. Sci. Technol.* **2022**, *19*, 12839–12852. [CrossRef]
- 75. da Silva, L.C.A.; Honorato, T.L.; Cavalcante, R.S.; Franco, T.T.; Rodrigues, S. Effect of PH and Temperature on Enzyme Activity of Chitosanase Produced Under Solid Stated Fermentation by *Trichoderma* spp. *Indian J. Microbiol.* 2012, 52, 60–65. [CrossRef] [PubMed]
- 76. Akram, N.A.; Saleem, M.H.; Shafiq, S.; Naz, H.; Farid-ul-Haq, M.; Ali, B.; Shafiq, F.; Iqbal, M.; Jaremko, M.; Qureshi, K.A. Phytoextracts as Crop Biostimulants and Natural Protective Agents—A Critical Review. Sustainability 2022, 14, 14498. [CrossRef]
- Vassilev, N.; Vassileva, M.; Lopez, A.; Martos, V.; Reyes, A.; Maksimovic, I.; Eichler-Löbermann, B.; Malusà, E. Unexploited Potential of Some Biotechnological Techniques for Biofertilizer Production and Formulation. *Appl. Microbiol. Biotechnol.* 2015, 99, 4983–4996. [CrossRef]
- 78. Chen, H. Modern Solid State Fermentation; Springer: Berlin/Heidelberg, Germany, 2013. [CrossRef]

- Singhania, R.R.; Patel, A.K.; Soccol, C.R.; Pandey, A. Recent Advances in Solid-State Fermentation. *Biochem. Eng. J.* 2009, 44, 13–18. [CrossRef]
- Szabo, O.E.; Csiszar, E.; Koczka, B.; Kiss, K. Ultrasonically Assisted Single Stage and Multiple Extraction of Enzymes Produced by Aspergillus Oryzae on a Lignocellulosic Substrate with Solid-State Fermentation. *Biomass Bioenergy* 2015, 75, 161–169. [CrossRef]
- 81. Yadav, J.S.; Tripathi, J.P. Optimization of Cultivation and Nutrition Conditions and Substrate Pretreatment for Solid-Substrate Fermentation of Wheat Straw ByCoriolus Versicolor. *Folia Microbiol.* **1991**, *36*, 294–301. [CrossRef] [PubMed]
- 82. Kunamneni, A.; Permaul, K.; Singh, S. Amylase Production in Solid State Fermentation by the Thermophilic Fungus Thermomyces Lanuginosus. J. Biosci. Bioeng. 2005, 100, 168–171. [CrossRef]
- 83. Elibol, M.; Moreira, A.R. Optimizing Some Factors Affecting Alkaline Protease Production by a Marine Bacterium Teredinobacter Turnirae under Solid Substrate Fermentation. *Process Biochem.* **2005**, *40*, 1951–1956. [CrossRef]
- Alias, C.; Bulgari, D.; Gobbi, E. It Works! Organic-Waste-Assisted Trichoderma Spp. Solid-State Fermentation on Agricultural Digestate. *Microorganisms* 2022, 10, 164. [CrossRef] [PubMed]
- Raghavarao, K.S.M.S.; Ranganathan, T.V.; Karanth, N.G. Some Engineering Aspects of Solid-State Fermentation. *Biochem. Eng. J.* 2003, 13, 127–135. [CrossRef]
- General Considerations about Solid-State Fermentation Processes | SpringerLink. Available online: https://link.springer.com/ chapter/10.1007/978-0-387-75213-6_2 (accessed on 23 July 2023).
- Kamilova, F.; Okon, Y.; de Weert, S.; Hora, K. Commercialization of Microbes: Manufacturing, Inoculation, Best Practice for Objective Field Testing, and Registration. In *Principles of Plant-Microbe Interactions: Microbes for Sustainable Agriculture*; Lugtenberg, B., Ed.; Springer International Publishing: Cham, Switzerland, 2015; pp. 319–327, ISBN 978-3-319-08575-3.
- Kiruba, N.J.M.; Saeid, A. An Insight into Microbial Inoculants for Bioconversion of Waste Biomass into Sustainable "Bio-Organic" Fertilizers: A Bibliometric Analysis and Systematic Literature Review. Int. J. Mol. Sci. 2022, 23, 13049. [CrossRef]
- Chilakamarry, C.R.; Sakinah, A.M.M.; Zularisam, A.W.; Sirohi, R.; Khilji, I.A.; Ahmad, N.; Pandey, A. Advances in Solid-State Fermentation for Bioconversion of Agricultural Wastes to Value-Added Products: Opportunities and Challenges. *Bioresour. Technol.* 2022, 343, 126065. [CrossRef]
- Weber, F.J.; Tramper, J.; Rinzema, A. A Simplified Material and Energy Balance Approach for Process Development and Scale-up of Coniothyrium Minitans Conidia Production by Solid-State Cultivation in a Packed-Bed Reactor. *Biotechnol. Bioeng.* 1999, 65, 447–458. [CrossRef]
- 91. de Oliveira, J.; Rodrigues, C.; Vandenberghe, L.P.S.; Câmara, M.C.; Libardi, N.; Soccol, C.R. Gibberellic Acid Production by Different Fermentation Systems Using Citric Pulp as Substrate/Support. *BioMed Res. Int.* 2017, 2017, e5191046. [CrossRef]
- Bulgari, D.; Alias, C.; Peron, G.; Ribaudo, G.; Gianoncelli, A.; Savino, S.; Boureghda, H.; Bouznad, Z.; Monti, E.; Gobbi, E. Solid-State Fermentation of *Trichoderma* Spp.: A New Way to Valorize the Agricultural Digestate and Produce Value-Added Bioproducts. J. Agric. Food Chem. 2023, 71, 3994–4004. [CrossRef]
- Oh, Y.-K.; Hwang, K.-R.; Kim, C.; Kim, J.R.; Lee, J.-S. Recent Developments and Key Barriers to Advanced Biofuels: A Short Review. *Bioresour. Technol.* 2018, 257, 320–333. [CrossRef]
- 94. Nitayapat, N.; Prakarnsombut, N.; Lee, S.J.; Boonsupthip, W. Bioconversion of Tangerine Residues by Solid-State Fermentation with Lentinus Polychrous and Drying the Final Products. *LWT Food Sci. Technol.* **2015**, *63*, 773–779. [CrossRef]
- Vassileva, M.; Malusá, E.; Eichler-Löbermann, B.; Vassilev, N. Aspegillus Terreus: From Soil to Industry and Back. *Microorganisms* 2020, 8, 1655. [CrossRef]
- Dos Santos, V.Z.; Vieira, K.R.; Nass, P.P.; Zepka, L.Q.; Jacob-Lopes, E. Application of Microalgae Consortia/Cocultures in Wastewater Treatment. In *Recent Advances in Microbial Degradation*; Ahamed, M.I., Prasad, R., Eds.; Environmental and Microbial Biotechnology; Springer: Singapore, 2021; pp. 131–154, ISBN 9789811605185.
- 97. Tong, C.Y.; Honda, K.; Derek, C.J.C. A Review on Microalgal-Bacterial Co-Culture: The Multifaceted Role of Beneficial Bacteria towards Enhancement of Microalgal Metabolite Production. *Environ. Res.* **2023**, 228, 115872. [CrossRef]
- Berninger, T.; González López, Ó.; Bejarano, A.; Preininger, C.; Sessitsch, A. Maintenance and Assessment of Cell Viability in Formulation of Non-Sporulating Bacterial Inoculants. *Microb. Biotechnol.* 2018, 11, 277–301. [CrossRef]
- Malusá, E.; Sas-Paszt, L.; Ciesielska, J. Technologies for Beneficial Microorganisms Inocula Used as Biofertilizers. Sci. World J. 2012, 2012, e491206. [CrossRef] [PubMed]
- 100. Fuess, L.T.; Lens, P.N.L.; Garcia, M.L.; Zaiat, M. Exploring Potentials for Bioresource and Bioenergy Recovery from Vinasse, the "New" Protagonist in Brazilian Sugarcane Biorefineries. *Biomass* **2022**, *2*, 374–411. [CrossRef]
- Vassileva, M.; Malusà, E.; Sas-Paszt, L.; Trzcinski, P.; Galvez, A.; Flor-Peregrin, E.; Shilev, S.; Canfora, L.; Mocali, S.; Vassilev, N. Fermentation Strategies to Improve Soil Bio-Inoculant Production and Quality. *Microorganisms* 2021, 9, 1254. [CrossRef] [PubMed]
- Zheng, Z.; Shetty, K. Cranberry Processing Waste for Solid State Fungal Inoculant Production. *Process Biochem.* 1998, 33, 323–329. [CrossRef]
- Pandey, A.; Soccol, C.R.; Mitchell, D. New Developments in Solid State Fermentation: I-Bioprocesses and Products. *Process Biochem.* 2000, 35, 1153–1169. [CrossRef]
- Malik, T.; Rawat, S. Biotechnological Interventions for Production of Flavour and Fragrance Compounds. In Sustainable Bioeconomy: Pathways to Sustainable Development Goals; Venkatramanan, V., Shah, S., Prasad, R., Eds.; Springer: Singapore, 2021; pp. 131–170, ISBN 9789811573217.

- 105. Hassan, G.; Shabbir, M.A.; Ahmad, F.; Pasha, I.; Aslam, N.; Ahmad, T.; Rehman, A.; Manzoor, M.F.; Inam-Ur-Raheem, M.; Aadil, R.M. Cereal Processing Waste, an Environmental Impact and Value Addition Perspectives: A Comprehensive Treatise. *Food Chem.* 2021, 363, 130352. [CrossRef] [PubMed]
- 106. Finkler, A.T.J.; Biz, A.; Pitol, L.O.; Medina, B.S.; Luithardt, H.; Luz, L.F.d.L.; Krieger, N.; Mitchell, D.A. Intermittent Agitation Contributes to Uniformity across the Bed during Pectinase Production by Aspergillus Niger Grown in Solid-State Fermentation in a Pilot-Scale Packed-Bed Bioreactor. *Biochem. Eng. J.* 2017, 121, 1–12. [CrossRef]
- 107. Berovic, M. Cultivation of Medicinal Mushroom Biomass by Solid-State Bioprocessing in Bioreactors. In Solid State Fermentation: Research and Industrial Applications; Steudler, S., Werner, A., Cheng, J.J., Eds.; Advances in Biochemical Engineering/Biotechnology; Springer International Publishing: Cham, Switzerland, 2019; pp. 3–25, ISBN 978-3-030-23675-5.
- 108. Khairy, M.F.A.; Mohamed, A.A.I.; Khlil, M.M.N. Development of bioreactor to enrich the protein of agricultural residues. *Misr J. Agric. Eng.* **2015**, *32*, 1625–1640. [CrossRef]
- 109. Werle, L.B. Obtention Gibberellic Acid by Solid State Ferment. Employing Brew. Residue Crude Rice Brand Substrates. Master's Thesis, Federal University of Santa Maria, Santa Maria, Brazil, 2017.
- Monrroy, M.; García, J.R. Gibberellic Acid Production from Corn Cob Residues via Fermentation with *Aspergillus Niger*. J. Chem. 2022, 2022, e1112941. [CrossRef]
- 111. Jain, B.M.; Badve, M.P. A Novel Process for Synthesis of Soybean Protein Hydrolysates and Study of Its Effectiveness as a Biostimulant and Emulsifier. *Chem. Eng. Process. Process Intensif.* **2022**, *174*, 108880. [CrossRef]
- Wei, X.; Sui, Z.; Guo, M.; Chen, S.; Zhang, Z.; Geng, J.; Xiao, J.; Huang, D. The Potential of Degrading Natural Chitinous Wastes to Oligosaccharides by Chitinolytic Enzymes from Two Talaromyces Sp. Isolated from Rotten Insects (Hermetia Illucens) under Solid State Fermentation. *Braz. J. Microbiol.* 2023, 54, 223–238. [CrossRef]
- Volpi, M.P.C.; Corzo, I.J.M.; Bastos, R.G.; Santana, M.H.A. Production of Humic Acids by Solid-State Fermentation of Trichoderma Reesei in Raw Oil Palm Empty Fruit Bunch Fibers. 3 Biotech 2019, 9, 393. [CrossRef]
- 114. Ghanavati, H.; Ramezanipour, N.; Jouzani, G.S.; Kowsari, M.; Valijanian, E.; Nikrad, M.; Mostajeran, F.; Tahmasbi, M. Submerged Fermentation as a Suitable Solution to Produce Humic and Fulvic Acids from Sugarcane Bagasse. *Sci. Iran.* 2022, 29, 3554–3569. [CrossRef]
- 115. Amadou, I.; Le, G.-W.; Shi, Y.-H.; Gbadamosi, O.S.; Kamara, M.T.; Jin, S. Optimized Lactobacillus Plantarum Lp6 Solid-State Fermentation and Proteolytic Hydrolysis Improve Some Nutritional Attributes of Soybean Protein Meal. *J. Food Biochem.* 2011, 35, 1686–1694. [CrossRef]
- 116. López-Bucio, J.; Pelagio-Flores, R.; Herrera-Estrella, A. Trichoderma as Biostimulant: Exploiting the Multilevel Properties of a Plant Beneficial Fungus. *Sci. Hortic.* 2015, 196, 109–123. [CrossRef]
- 117. Kaya, C.; Ashraf, M.; Dikilitas, M.; Tuna, A.L. Alleviation of Salt Stress-Induced Adverse Effects on Maize Plants by Exogenous Application of Indoleacetic Acid (IAA) and Inorganic Nutrients—A Field Trial. *Aust. J. Crop Sci.* 2013, *7*, 249–254.
- Richards, D.E.; King, K.E.; Ait-ali, T.; Harberd, N.P. How gibberellin regulates plant growth and development: A Molecular Genetic Analysis of Gibberellin Signaling. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 2001, 52, 67–88. [CrossRef] [PubMed]
- 119. Brian, P.W. Effects of Gibberellins on Plant Growth and Development. *Biol. Rev.* **1959**, *34*, 37–77. [CrossRef]
- Soper, F.M.; Paungfoo-Lonhienne, C.; Brackin, R.; Rentsch, D.; Schmidt, S.; Robinson, N. Arabidopsis and Lobelia Anceps Access Small Peptides as a Nitrogen Source for Growth. *Funct. Plant Biol.* 2011, *38*, 788–796. [CrossRef] [PubMed]
- 121. Sheng, C.; Song, S.; Zhou, W.; Dossou, S.S.K.; Zhou, R.; Zhang, Y.; Li, D.; You, J.; Wang, L. Integrating Transcriptome and Phytohormones Analysis Provided Insights into Plant Height Development in Sesame. *Plant Physiol. Biochem.* 2023, 198, 107695. [CrossRef] [PubMed]
- 122. Luo, L.; Xie, Y.; Yu, S.; Yang, J.; Chen, S.; Yuan, X.; Guo, T.; Wang, H.; Liu, Y.; Chen, C.; et al. The DnaJ Domain-Containing Heat-Shock Protein NAL11 Determines Plant Architecture by Mediating Gibberellin Homeostasis in Rice (*Oryza sativa*). *New Phytol.* 2023, 237, 2163–2179. [CrossRef] [PubMed]
- 123. Tsavkelova, E.A.; Cherdyntseva, T.A.; Klimova, S.Y.; Shestakov, A.I.; Botina, S.G.; Netrusov, A.I. Orchid-Associated Bacteria Produce Indole-3-Acetic Acid, Promote Seed Germination, and Increase Their Microbial Yield in Response to Exogenous Auxin. *Arch. Microbiol.* **2007**, *188*, 655–664. [CrossRef]
- Egamberdieva, D. Alleviation of Salt Stress by Plant Growth Regulators and IAA Producing Bacteria in Wheat. *Acta Physiol. Plant.* 2009, *31*, 861–864. [CrossRef]
- Florido Bacallao, M.; Bao Fundora, L. Tolerancia a Estrés Por Déficit Hídrico En Tomate (*Solanum lycopersicum* L.). *Cultiv. Trop.* 2014, 35, 70–88.
- 126. Drobek, M.; Frąc, M.; Cybulska, J. Plant Biostimulants: Importance of the Quality and Yield of Horticultural Crops and the Improvement of Plant Tolerance to Abiotic Stress—A Review. *Agronomy* **2019**, *9*, 335. [CrossRef]
- 127. Luo, P.; Shen, Y.; Jin, S.; Huang, S.; Cheng, X.; Wang, Z.; Li, P.; Zhao, J.; Bao, M.; Ning, G. Overexpression of Rosa Rugosa Anthocyanidin Reductase Enhances Tobacco Tolerance to Abiotic Stress through Increased ROS Scavenging and Modulation of ABA Signaling. *Plant Sci.* 2016, 245, 35–49. [CrossRef]
- 128. Yazdani, M.; Croen, M.G.; Fish, T.L.; Thannhauser, T.W.; Ahner, B.A. Overexpression of Native ORANGE (OR) and OR Mutant Protein in Chlamydomonas Reinhardtii Enhances Carotenoid and ABA Accumulation and Increases Resistance to Abiotic Stress. *Metab. Eng.* 2021, 68, 94–105. [CrossRef] [PubMed]

- Zhang, L.; Gao, M.; Hu, J.; Zhang, X.; Wang, K.; Ashraf, M. Modulation Role of Abscisic Acid (ABA) on Growth, Water Relations and Glycinebetaine Metabolism in Two Maize (*Zea mays* L.) Cultivars under Drought Stress. *Int. J. Mol. Sci.* 2012, *13*, 3189–3202. [CrossRef] [PubMed]
- Mukherjee, A.; Patel, J.S. Seaweed Extract: Biostimulator of Plant Defense and Plant Productivity. Int. J. Environ. Sci. Technol. 2020, 17, 553–558. [CrossRef]
- 131. Berthon, J.-Y.; Michel, T.; Wauquier, A.; Joly, P.; Gerbore, J.; Filaire, E. Seaweed and microalgae as major actors of blue biotechnology to achieve plant stimulation and pest and pathogen biocontrol—A review of the latest advances and future prospects. *J. Agric. Sci.* **2021**, *159*, 523–534. [CrossRef]
- 132. Kapur, B.; Sarıdaş, M.A.; Çeliktopuz, E.; Kafkas, E.; Kargı, S.P. Health and Taste Related Compounds in Strawberries under Various Irrigation Regimes and Bio-Stimulant Application. *Food Chem.* **2018**, *263*, 67–73. [CrossRef]
- 133. Jaulneau, V.; Lafitte, C.; Corio-Costet, M.-F.; Stadnik, M.J.; Salamagne, S.; Briand, X.; Esquerré-Tugayé, M.-T.; Dumas, B. An Ulva Armoricana Extract Protects Plants against Three Powdery Mildew Pathogens. Eur. J. Plant. Pathol. 2011, 131, 393–401. [CrossRef]
- 134. Khaled, H.; Fawy, H.A. Effect of Different Levels of Humic Acids on the Nutrient Content, Plant Growth, and Soil Properties under Conditions of Salinity. *Soil Water Res.* 2011, *6*, 21–29. [CrossRef]
- 135. Fagbenro, J.A.; Agboola, A.A. Effect of Different Levels of Humic Acid on the Growth and Nutrient Uptake of Teak Seedlings. *J. Plant Nutr.* **1993**, *16*, 1465–1483. [CrossRef]
- 136. Khan, R.U.; Khan, M.Z.; Khan, A.; Saba, S.; Hussain, F.; Jan, I.U. Effect of Humic Acid on Growth and Crop Nutrient Status of Wheat on Two Different Soils. J. Plant Nutr. 2018, 41, 453–460. [CrossRef]
- 137. Bayat, H.; Shafie, F.; Aminifard, M.H.; Daghighi, S. Comparative Effects of Humic and Fulvic Acids as Biostimulants on Growth, Antioxidant Activity and Nutrient Content of Yarrow (*Achillea millefolium* L.). *Sci. Hortic.* **2021**, 279, 109912. [CrossRef]
- 138. Maach, M.; Boudouasar, K.; Akodad, M.; Skalli, A.; Moumen, A.; Baghour, M. Application of Biostimulants Improves Yield and Fruit Quality in Tomato. *Int. J. Veg. Sci.* 2021, 27, 288–293. [CrossRef]
- 139. Chanthini, K.M.-P.; Senthil-Nathan, S.; Pavithra, G.-S.; Asahel, A.-S.; Malarvizhi, P.; Murugan, P.; Deva-Andrews, A.; Sivanesh, H.; Stanley-Raja, V.; Ramasubramanian, R.; et al. The Macroalgal Biostimulant Improves the Functional Quality of Tomato Fruits Produced from Plants Grown under Salt Stress. *Agriculture* 2023, 13, 6. [CrossRef]
- 140. Graziani, G.; Ritieni, A.; Cirillo, A.; Cice, D.; Di Vaio, C. Effects of Biostimulants on Annurca Fruit Quality and Potential Nutraceutical Compounds at Harvest and during Storage. *Plants* **2020**, *9*, 775. [CrossRef]
- 141. Zwack, P.J.; Rashotte, A.M. Cytokinin Inhibition of Leaf Senescence. Plant Signal. Behav. 2013, 8, e24737. [CrossRef] [PubMed]
- 142. Chang, H.; Jones, M.L.; Banowetz, G.M.; Clark, D.G. Overproduction of Cytokinins in Petunia Flowers Transformed with PSAG12-IPT Delays Corolla Senescence and Decreases Sensitivity to Ethylene. *Plant Physiol.* **2003**, *132*, 2174–2183. [CrossRef]
- 143. Lara, M.E.B.; Garcia, M.-C.G.; Fatima, T.; Ehneß, R.; Lee, T.K.; Proels, R.; Tanner, W.; Roitsch, T. Extracellular Invertase Is an Essential Component of Cytokinin-Mediated Delay of Senescence. *Plant Cell* **2004**, *16*, 1276–1287. [CrossRef]
- 144. Spinelli, F.; Fiori, G.; Noferini, M.; Sprocatti, M.; Costa, G. A Novel Type of Seaweed Extract as a Natural Alternative to the Use of Iron Chelates in Strawberry Production. *Sci. Hortic.* **2010**, *125*, 263–269. [CrossRef]
- 145. Kumari, R.; Kaur, I.; Bhatnagar, A.K. Enhancing Soil Health and Productivity of *Lycopersicon esculentum* Mill. Using Sargassum Johnstonii Setchell & Gardner as a Soil Conditioner and Fertilizer. *J. Appl. Phycol.* **2013**, 25, 1225–1235. [CrossRef]
- 146. Meng, C.; Gu, X.; Liang, H.; Wu, M.; Wu, Q.; Yang, L.; Li, Y.; Shen, P. Optimized Preparation and High-Efficient Application of Seaweed Fertilizer on Peanut. *J. Agric. Food Res.* **2022**, *7*, 100275. [CrossRef]
- 147. Matthews, S.; Ali, A.; Siddiqui, Y.; Supramaniam, C.V. Plant Bio-Stimulant: Prospective, Safe and Natural Resources. *J. Soil Sci. Plant Nutr.* **2022**, 22, 2570–2586. [CrossRef]
- 148. Zeljković, S.; Parađiković, N.; Maksimović, I.; Teklić, T.; Kojić, M.T. Growth and Nutrient Status of French Marigold (*Tagetes Patula* L.) under Biostimulant Application. N. Z. J. Crop Hortic. Sci. 2022, 1–11. [CrossRef]
- 149. Basavaraja, P.K.; Yogendra, N.D.; Zodape, S.T.; Prakash, R.; Ghosh, A. Effect of Seaweed Sap as Foliar Spray on Growth and Yield of Hybrid Maize. *J. Plant Nutr.* **2018**, *41*, 1851–1861. [CrossRef]
- 150. Pal, A.; Dwivedi, S.K.; Maurya, P.K.; Kanwar, P. Effect of Seaweed Saps on Growth, Yield, Nutrient Uptake and Economic Improvement of Maize (Sweet Corn). J. Appl. Nat. Sci. 2015, 7, 970–975. [CrossRef]
- 151. Ertani, A.; Francioso, O.; Tinti, A.; Schiavon, M.; Pizzeghello, D.; Nardi, S. Evaluation of Seaweed Extracts from Laminaria and Ascophyllum Nodosum Spp. as Biostimulants in Zea Mays L. Using a Combination of Chemical, Biochemical and Morphological Approaches. Front. Plant Sci. 2018, 9, 428. [CrossRef]
- 152. Hu, M.; McClements, D.J.; Decker, E.A. Antioxidant Activity of a Proanthocyanidin-Rich Extract from Grape Seed in Whey Protein Isolate Stabilized Algae Oil-in-Water Emulsions. J. Agric. Food Chem. 2004, 52, 5272–5276. [CrossRef] [PubMed]
- 153. Efecto Del Extracto de Células de Alga Verde Como Aerosol Foliar Sobre El Crecimiento Vegetativo, El Rendimiento y La Calidad de Las Bayas de Vides Superiores. Available online: https://www.researchgate.net/publication/237566381_Effect_of_Green_Alga_Cells_Extract_as_Foliar_Spray_on_Vegetative_Growth_Yield_and_Berries_Quality_of_Superior_Grapevines (accessed on 3 June 2023).
- 154. Arioli, T.; Mattner, S.W.; Hepworth, G.; McClintock, D.; McClinock, R. Effect of Seaweed Extract Application on Wine Grape Yield in Australia. *J. Appl. Phycol.* **2021**, *33*, 1883–1891. [CrossRef]
- 155. Jayaraman, J.J.J.; Ali, N. Use of Seaweed Extracts for Disease Management of Vegetable Crops. In *Sustainable Crop Disease Management Using Natural Products*; CABI: Wallingford, UK, 2015; pp. 160–183. [CrossRef]

- 156. Murtic, S.; Oljaca, R.; Murtic, M.S.; Vranac, A.; Akagic, A.; Civic, H. Cherry Tomato Productivity as Influenced by Liquid Organic Fertilizer under Different Growth Conditions. J. Cent. Eur. Agric. 2018, 19, 503–516. [CrossRef]
- 157. Demir, N.; Dural, B.; Yildirim, K. Effect of Seaweed Suspensions on Seed Germination of Tomato, Pepper and Aubergine. *J. Biol. Sci.* **2006**, *6*, 1130–1133.
- 158. Yusuf, R.; Kristiansen, P.; Warwick, N. Effect of Two Seaweed Products and Equivalent Mineral Treatments on Lettuce (*Lactuca sativa L.*) Growth. J. Agron. 2019, 18, 100–106. [CrossRef]
- Di Mola, I.; Cozzolino, E.; Ottaiano, L.; Giordano, M.; Rouphael, Y.; Colla, G.; Mori, M. Effect of Vegetal- and Seaweed Extract-Based Biostimulants on Agronomical and Leaf Quality Traits of Plastic Tunnel-Grown Baby Lettuce under Four Regimes of Nitrogen Fertilization. Agronomy 2019, 9, 571. [CrossRef]
- 160. Nardelli, A.E.; Chiozzini, V.G.; Braga, E.S.; Chow, F. Integrated Multi-Trophic Farming System between the Green Seaweed Ulva Lactuca, Mussel, and Fish: A Production and Bioremediation Solution. *J. Appl. Phycol.* **2019**, *31*, 847–856. [CrossRef]
- 161. Righini, H.; Roberti, R.; Baraldi, E. Use of Algae in Strawberry Management. J. Appl. Phycol. 2018, 30, 3551–3564. [CrossRef]
- 162. Abbas, M.; Anwar, J.; Zafar-ul-Hye, M.; Iqbal Khan, R.; Saleem, M.; Rahi, A.A.; Danish, S.; Datta, R. Effect of Seaweed Extract on Productivity and Quality Attributes of Four Onion Cultivars. *Horticulturae* **2020**, *6*, 28. [CrossRef]
- Almaroai, Y.A.; Eissa, M.A. Role of Marine Algae Extracts in Water Stress Resistance of Onion Under Semiarid Conditions. J. Soil Sci. Plant Nutr. 2020, 20, 1092–1101. [CrossRef]
- 164. Prajapati, A.; Jain, S.; Chongtham, S.; Maheshwari, M.; Patel, C.; Patel, R.; Patel, C.; Singh, N.; Prajapati, A. Evaluation of Seaweed Extract on Growth and Yield of Potato. *Environ. Ecol.* **2016**, *34*, 605–608.
- Dziugieł, T.; Wadas, W. Possibility of Increasing Early Crop Potato Yield with Foliar Application of Seaweed Extracts and Humic Acids. J. Cent. Eur. Agric. 2020, 21, 300–310. [CrossRef]
- 166. Manpuhro, N.; Dawson, J. Influence of Indole Acetic Acid (IAA) and Boron on Growth and Yield of Maize (*Zea mays*. L). *Int. J. Plant Soil Sci.* **2023**, *35*, 33–41. [CrossRef]
- 167. Hagaggi, N.S.A.; Mohamed, A.A.A. Enhancement of *Zea mays* (L.) Growth Performance Using Indole Acetic Acid Producing Endophyte Mixta Theicola Isolated from Solenostemma Argel (Hayne). *S. Afr. J. Bot.* **2020**, *134*, 64–71. [CrossRef]
- 168. Marag, P.S.; Suman, A. Growth Stage and Tissue Specific Colonization of Endophytic Bacteria Having Plant Growth Promoting Traits in Hybrid and Composite Maize (*Zea mays* L.). *Microbiol. Res.* **2018**, 214, 101–113. [CrossRef] [PubMed]
- 169. Visconti, D.; Fiorentino, N.; Cozzolino, E.; Woo, S.L.; Fagnano, M.; Rouphael, Y. Can Trichoderma-Based Biostimulants Optimize N Use Efficiency and Stimulate Growth of Leafy Vegetables in Greenhouse Intensive Cropping Systems? *Agronomy* 2020, 10, 121. [CrossRef]
- 170. Kondhare, K.R.; Patil, A.B.; Giri, A.P. Auxin: An Emerging Regulator of Tuber and Storage Root Development. *Plant Sci.* **2021**, 306, 110854. [CrossRef]
- Romanov, G.A.; Aksenova, N.P.; Konstantinova, T.N.; Golyanovskaya, S.A.; Kossmann, J.; Willmitzer, L. Effect of Indole-3-Acetic Acid and Kinetin on Tuberisation Parameters of Different Cultivars and Transgenic Lines of Potato in Vitro. *Plant Growth Regul.* 2000, 32, 245–251. [CrossRef]
- 172. Ekin, Z. Integrated Use of Humic Acid and Plant Growth Promoting Rhizobacteria to Ensure Higher Potato Productivity in Sustainable Agriculture. *Sustainability* **2019**, *11*, 3417. [CrossRef]
- 173. Hye, M.; Haque, M.; Karim, M. Influence of Growth Regulators and Their Time of Application on Yield of Onion. *Pak. J. Biol. Sci.* **2002**, *5*, 1021–1023. [CrossRef]
- 174. Bista, D.; Sapkota, D.; Paudel, H.; Adhikari, G. Effect of Foliar Application of Growth Regulators on Growth and Yield of Onion (*Allium cepa*). Int. J. Hortic. Sci. Technol. 2022, 9, 247–254. [CrossRef]
- 175. Gupta, S.; Stirk, W.A.; Plačková, L.; Kulkarni, M.G.; Doležal, K.; Van Staden, J. Interactive Effects of Plant Growth-Promoting Rhizobacteria and a Seaweed Extract on the Growth and Physiology of *Allium cepa* L. (Onion). *J. Plant Physiol.* 2021, 262, 153437. [CrossRef]
- 176. Mahdi, I.; Fahsi, N.; Hafidi, M.; Allaoui, A.; Biskri, L. Plant Growth Enhancement Using Rhizospheric Halotolerant Phosphate Solubilizing Bacterium Bacillus Licheniformis QA1 and Enterobacter Asburiae QF11 Isolated from Chenopodium Quinoa Willd. *Microorganisms* 2020, *8*, 948. [CrossRef]
- 177. Azarakhsh, M.R.; Bagherieh-Najjar, M.B.; Sadeghipour, H.R.; Raeisi, S. Improved Grain Yield by Phytohormones-Driven Suppression of Pod Abscission and Revitalization of Source-Sink Relationships in Soybean. *Int. J. Plant Prod.* 2022, 16, 467–481. [CrossRef]
- 178. Hanaa, H.; Safaa, A. Foliar Application Foliar Application of IAA at Different Growth Stages and Their Influenced on Growth and Productivity of Bread Wheat (*Triticum aestivum* L.). J. Phys. Conf. Ser. 2019, 1294, 092029. [CrossRef]
- 179. Çakmakçı, R.; Erat, M.; Erdoğan, Ü.; Dönmez, M.F. The Influence of Plant Growth–Promoting Rhizobacteria on Growth and Enzyme Activities in Wheat and Spinach Plants. J. Plant Nutr. Soil Sci. 2007, 170, 288–295. [CrossRef]
- Colla, G.; Rouphael, Y.; Di Mattia, E.; El-Nakhel, C.; Cardarelli, M. Co-Inoculation of Glomus Intraradices and Trichoderma Atroviride Acts as a Biostimulant to Promote Growth, Yield and Nutrient Uptake of Vegetable Crops. J. Sci. Food Agric. 2015, 95, 1706–1715. [CrossRef]
- 181. Contreras-Cornejo, H.A.; Macías-Rodríguez, L.; Cortés-Penagos, C.; López-Bucio, J. Trichoderma Virens, a Plant Beneficial Fungus, Enhances Biomass Production and Promotes Lateral Root Growth through an Auxin-Dependent Mechanism in Arabidopsis. *Plant Physiol.* 2009, 149, 1579–1592. [CrossRef] [PubMed]

- Tandon, S.; Dubey, A. Effects of Biozyme (*Ascophyllum Nodosum*) Biostimulant on Growth and Development of Soybean [*Glycine max* (L.) Merill]. *Commun. Soil Sci. Plant Anal.* 2015, 46, 845–858. [CrossRef]
- 183. Marathe, R.; Phatake, Y.; Shaikh, A.; Shinde, B.; Gajbhiye, M. Effect of IAA Produced by Pseudomonas Aeruginosa 6a (Bc4) on Seed Germination and Plant Growth of Glycin Max. *J. Exp. Biol. Agric. Sci.* **2017**, *5*, 351–358. [CrossRef]
- Susilowati, D.N.; Riyanti, E.I.; Setyowati, M.; Mulya, K. Indole-3-Acetic Acid Producing Bacteria and Its Application on the Growth of Rice. *AIP Conf. Proc.* 2018, 2002, 020016. [CrossRef]
- 185. Li, Z.; Zhang, X.; Zhao, Y.; Li, Y.; Zhang, G.; Peng, Z.; Zhang, J. Enhancing Auxin Accumulation in Maize Root Tips Improves Root Growth and Dwarfs Plant Height. *Plant Biotechnol. J.* **2018**, *16*, 86–99. [CrossRef]
- 186. Husen, A.; Iqbal, M.; Aref, I.M. Plant Growth and Foliar Characteristics of Faba Bean (*Vicia faba* L.) as Affected by Indole-Acetic Acid under Water-Sufficient and Water-Deficient Conditions. J. Environ. Biol. 2017, 38, 179–186. [CrossRef]
- Hamidon, A.; Shah, R.M.; Razali, R.M.; Lob, S. Effect of different types and concentration of rooting hormones on momordica cochinensis (gac fruit) root vine cuttings. *Malays. Appl. Biol.* 2020, 49, 127–132. [CrossRef]
- 188. Sabir, A. Improvement of Grafting Efficiency in Hard Grafting Grape Berlandieri Hybrid Rootstocks by Plant Growth-Promoting Rhizobacteria (PGPR). *Sci. Hortic.* 2013, *164*, 24–29. [CrossRef]
- Shahzad, K.; Siddiqi, E.H.; Ahmad, S.; Zeb, U.; Muhammad, I.; Khan, H.; Zhao, G.-F.; Li, Z.-H. Exogenous Application of Indole-3-Acetic Acid to Ameliorate Salt Induced Harmful Effects on Four Eggplants (*Solanum melongena* L.) Varieties. *Sci. Hortic.* 2022, 292, 110662. [CrossRef]
- Lur, H.-S.; Setter, T.L. Endorsperm Development of Maize Defective Kernel (Dek) Mutants. Auxin and Cytokinin Levels. Ann. Bot. 1993, 72, 1–6. [CrossRef]
- Rady, M.M.; Talaat, N.B.; Abdelhamid, M.T.; Shawky, B.T.; Desoky, E.-S.M. Maize (*Zea mays* L.) Grains Extract Mitigates the Deleterious Effects of Salt Stress on Common Bean (*Phaseolus vulgaris* L.) Growth and Physiology. *J. Hortic. Sci. Biotechnol.* 2019, 94, 777–789. [CrossRef]
- 192. Xiao, Y.; Liu, D.; Zhang, G.; Gao, S.; Liu, L.; Xu, F.; Che, R.; Wang, Y.; Tong, H.; Chu, C. Big Grain3, Encoding a Purine Permease, Regulates Grain Size via Modulating Cytokinin Transport in Rice. *J. Integr. Plant Biol.* **2019**, *61*, 581–597. [CrossRef] [PubMed]
- 193. Yin, W.; Xiao, Y.; Niu, M.; Meng, W.; Li, L.; Zhang, X.; Liu, D.; Zhang, G.; Qian, Y.; Sun, Z.; et al. ARGONAUTE2 Enhances Grain Length and Salt Tolerance by Activating BIG GRAIN3 to Modulate Cytokinin Distribution in Rice. *Plant Cell* 2020, 32, 2292–2306. [CrossRef] [PubMed]
- 194. Yang, D.; Li, Y.; Shi, Y.; Cui, Z.; Luo, Y.; Zheng, M.; Chen, J.; Li, Y.; Yin, Y.; Wang, Z. Exogenous Cytokinins Increase Grain Yield of Winter Wheat Cultivars by Improving Stay-Green Characteristics under Heat Stress. *PLoS ONE* **2016**, *11*, e0155437. [CrossRef]
- 195. Zaheer, M.S.; Raza, M.A.S.; Saleem, M.F.; Erinle, K.O.; Iqbal, R.; Ahmad, S. Effect of Rhizobacteria and Cytokinins Application on Wheat Growth and Yield under Normal vs Drought Conditions. *Commun. Soil Sci. Plant Anal.* **2019**, *50*, 2521–2533. [CrossRef]
- Liu, Y.; Liao, Y.; Liu, W. High Nitrogen Application Rate and Planting Density Reduce Wheat Grain Yield by Reducing Filling Rate of Inferior Grain in Middle Spikelets. Crop J. 2021, 9, 412–426. [CrossRef]
- 197. Nagel, L.; Brewster, R.; Riedell, W.E.; Reese, R.N. Cytokinin Regulation of Flower and Pod Set in Soybeans (*Glycine max* (L.) Merr.). *Ann. Bot.* 2001, *88*, 27–31. [CrossRef]
- 198. Kron, A.P.; Souza, G.M.; Ribeiro, R.V. Water Deficiency at Different Developmental Stages of *Glycine Max* Can Improve Drought Tolerance. *Bragantia* **2008**, 67, 43–49. [CrossRef]
- 199. Mady, M.A. Effect of foliar application with salicylic acid and vitamin e on growth and productivity of tomato (*Lycopersicon esculentum*, Mill.) Plant. J. Plant Prod. **2009**, 34, 6715–6726. [CrossRef]
- Caldiz, D.O. Seed Potato (*Solanum tuberosum* L.) Yield and Tuber Number Increase after Foliar Applications of Cytokinins and Gibberellic Acid under Field and Glasshouse Conditions. *Plant Growth Regul* 1996, 20, 185–188. [CrossRef]
- 201. Pavlista, A.D. Growth Regulators Increased Yield of Atlantic Potato. Am. J. Potato Res 2011, 88, 479–484. [CrossRef]
- 202. Carvajal-Millán, E.; Carvallo, T.; Orozco, J.A.; Martínez, M.A.; Tapia, I.; Guerrero, V.M.; Rascón-Chu, A.; Llamas, J.; Gardea, A.A. Polyphenol Oxidase Activity, Color Changes, and Dehydration in Table Grape Rachis during Development and Storage As Affected by N-(2-Chloro-4-Pyridyl)-N-Phenylurea. *J. Agric. Food Chem.* 2001, 49, 946–951. [CrossRef] [PubMed]
- Peppi, M.C.; Fidelibus, M.W. Effects of Forchlorfenuron and Abscisic Acid on the Quality of 'Flame Seedless' Grapes. *HortScience* 2008, 43, 173–176. [CrossRef]
- Qiu, Y.; Guan, S.C.; Wen, C.; Li, P.; Gao, Z.; Chen, X. Auxin and Cytokinin Coordinate the Dormancy and Outgrowth of Axillary Bud in Strawberry Runner. BMC Plant Biol. 2019, 19, 528. [CrossRef]
- Dale, A.; Elfving, D.C.; Chandler, C.K. Benzyladenine and Gibberellic Acid Increase Runner Production in Dayneutral Strawberries. *HortScience* 1996, 31, 1190–1194. [CrossRef]
- Costa, G.; Corelli-Grappadelli, L.; Bucchi, F. Studies on apple fruit abscission and growth as affected by cytokinins. *Acta Hortic*. 2001, 243–252. [CrossRef]
- Kumari, S.; Bakshi, P.; Sharma, A.; Wali, V.; Jasrotia, A.; Kour, S. Use of Plant Growth Regulators for Improving Fruit Production in Sub Tropical Crops. Int. J. Curr. Microbiol. Appl. Sci. 2018, 7, 659–668. [CrossRef]
- 208. Ferrer, C.; Martiz, J.; Saa, S.; Cautín, R. Increase in Final Fruit Size of Tangor (*Citrus reticulata* × *C. sinensis*) Cv W. Murcott by Application of Benzyladenine to Flowers. *Sci. Hortic.* **2017**, 223, 38–43. [CrossRef]
- Yousif, K.H. Application Method of Potassium Humate on Growth and Yield of Green Onion (*Allium cepa* L.). Sci. J. Univ. Zakho 2014, 2, 323–328. [CrossRef]

- 210. Forotaghe, Z.A.; Souri, M.K.; Jahromi, M.G.; Torkashvand, A.M. Influence of Humic Acid Application on Onion Growth Characteristics under Water Deficit Conditions. *J. Plant Nutr.* **2022**, *45*, 1030–1040. [CrossRef]
- Kaya, C.; Akram, N.A.; Ashraf, M.; Sonmez, O. Exogenous Application of Humic Acid Mitigates Salinity Stress in Maize (*Zea mays* L.) Plants by Improving Some Key Physico-Biochemical Attributes. *Cereal Res. Commun.* 2018, 46, 67–78. [CrossRef]
- 212. Shafi, M.I.; Adnan, M.; Fahad, S.; Wahid, F.; Khan, A.; Yue, Z.; Danish, S.; Zafar-ul-Hye, M.; Brtnicky, M.; Datta, R. Application of Single Superphosphate with Humic Acid Improves the Growth, Yield and Phosphorus Uptake of Wheat (*Triticum aestivum* L.) in Calcareous Soil. *Agronomy* 2020, 10, 1224. [CrossRef]
- 213. Lamlom, S.F.; Irshad, A.; Mosa, W.F.A. The Biological and Biochemical Composition of Wheat (*Triticum aestivum*) as Affected by the Bio and Organic Fertilizers. *BMC Plant Biol.* **2023**, *23*, 111. [CrossRef] [PubMed]
- Mindari, W.; Sasongko, P.E.; Kusuma, Z.; Syekhfani; Aini, N. Efficiency of Various Sources and Doses of Humic Acid on Physical and Chemical Properties of Saline Soil and Growth and Yield of Rice. *AIP Conf. Proc.* 2018, 2019, 030001. [CrossRef]
- Khedr, R.A.; Sorour, S.G.R.; Aboukhadrah, S.H.; El Shafey, N.M.; Elsalam, H.E.A.; El-Sharnouby, M.E.; El-Tahan, A.M. Alleviation of Salinity Stress Effects on Agro-Physiological Traits of Wheat by Auxin, Glycine Betaine, and Soil Additives. *Saudi J. Biol. Sci.* 2022, 29, 534–540. [CrossRef]
- Suh, H.Y.; Yoo, K.S.; Suh, S.G. Effect of Foliar Application of Fulvic Acid on Plant Growth and Fruit Quality of Tomato (*Lycopersicon* esculentum L.). Hortic. Environ. Biotechnol. 2014, 55, 455–461. [CrossRef]
- 217. Yildirim, E. Foliar and Soil Fertilization of Humic Acid Affect Productivity and Quality of Tomato. *Acta Agric. Scand. Sect. B-Soil Plant Sci.* 2007, 57, 182–186. [CrossRef]
- 218. Hemida, K.A.; Eloufey, A.Z.A.; El-Yazal, M.A.S.; Rady, M.M. Integrated Effect of Potassium Humate and α-Tocopherol Applications on Soil Characteristics and Performance of *Phaseolus Vulgaris* Plants Grown on a Saline Soil. *Arch. Agron. Soil Sci.* 2017, 63, 1556–1571. [CrossRef]
- Kandil, A.A.; Sharief, A.E.; Fathalla, F.H. Onion yield as affected by foliar application with amino and humic acids under nitrogen fertilizer levels. Crop Prod. 2013, 2, 62–72.
- Sruthi, B.E.S. Influence of organic manures on yield, quality and economics of aggregatum onion (*Allium cepa*. L. var. aggregatum). J. Pharmacogn. Phytochem. 2019, 8, 1768–1770.
- 221. Omar, M.; Ramadan, A. Response of Carrot (*Daucus carota* L.) to Foliar Application of Potassium Fertilizers and Some Soil Amendments under Clay Soil Conditions. *J. Soil Sci. Agric. Eng.* **2018**, *9*, 197–202. [CrossRef]
- 222. Raheem, S.M.; Al-Jaf, H.I.; Tofiq, G.K. Influence of Foliar and Soil Application of Humic Acid on Growth and Yield of Lettuce. *Euphrates J. Agric. Sci.* 2018, *10*, 199–204.
- 223. Srivastava, N.; Srivastava, M.; Ramteke, P.W.; Mishra, P.K. Chapter 23—Solid-State Fermentation Strategy for Microbial Metabolites Production: An Overview. In *New and Future Developments in Microbial Biotechnology and Bioengineering*; Gupta, V.K., Pandey, A., Eds.; Elsevier: Amsterdam, The Netherlands, 2019; pp. 345–354, ISBN 978-0-444-63504-4.
- Kapoore, R.V.; Wood, E.E.; Llewellyn, C.A. Algae Biostimulants: A Critical Look at Microalgal Biostimulants for Sustainable Agricultural Practices. *Biotechnol. Adv.* 2021, 49, 107754. [CrossRef] [PubMed]
- Ali, O.; Ramsubhag, A.; Jayaraman, J. Biostimulant Properties of Seaweed Extracts in Plants: Implications towards Sustainable Crop Production. *Plants* 2021, 10, 531. [CrossRef] [PubMed]
- 226. Malik, A.; Mor, V.S.; Tokas, J.; Punia, H.; Malik, S.; Malik, K.; Sangwan, S.; Tomar, S.; Singh, P.; Singh, N.; et al. Biostimulant-Treated Seedlings under Sustainable Agriculture: A Global Perspective Facing Climate Change. *Agronomy* **2021**, *11*, 14. [CrossRef]
- 227. Hasanuzzaman, M.; Parvin, K.; Bardhan, K.; Nahar, K.; Anee, T.I.; Masud, A.A.C.; Fotopoulos, V. Biostimulants for the Regulation of Reactive Oxygen Species Metabolism in Plants under Abiotic Stress. *Cells* **2021**, *10*, 2537. [CrossRef]

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