



Review article

Management of cytomegalovirus in adult solid organ transplant patients: GESITRA-IC-SEIMC, CIBERINFEC, and SET recommendations update



Elisa Ruiz-Arabi^a, Julian Torre-Cisneros^{b,c,*}, Victoria Aguilera^d, Rodrigo Alonso^e, Marina Berenguer^d, Oriol Bestard^f, Marta Bodro^{g,c}, Sara Cantisán^{b,c}, Jordi Carratalà^{h,c}, Juan José Castón^{b,c}, Elisa Cordero^{i,j,c}, Carme Facundo^k, María Carmen Fariñas^{l,c}, Mirian Fernández-Alonso^m, Mario Fernández-Ruiz^{n,c}, Jesús Fortún^{o,c}, María Dolores García-Cosío^p, Sabina Herrera^g, David Iturbe-Fernández^q, Oscar Len^{r,c}, Francisco López-Medrano^{n,c}, María Ovidia López-Oliva^s, Ibai Los-Arcos^r, María Angeles Marcos^{t,c}, Pilar Martín-Dávila^{o,c}, Víctor Monforte^{u,v}, Patricia Muñoz^{w,v}, David Navarro^{x,c}, Aurora Páez-Vega^y, Ana Belén Pérez^{z,c}, Natalia Redondo^{n,c}, Rodríguez Álvarez R.^{aa}, Alberto Rodríguez-Benot^{ab}, Isabel Rodríguez-Goncer^{n,c}, Rafael San-Juan^{n,c}, Javier Sánchez-Céspedes^{i,c}, Maricela Valerio^{w,v}, José Manuel Vaquero^{ac}, Diego Viasus^{ad}, Elisa Vidal^{b,c}, José María Aguado^{c,n,**}

^a Service of Infectious Diseases, Reina Sofía University Hospital, Maimónides Institute for Biomedical Research (IMIBIC), Córdoba, Spain

^b Service of Infectious Diseases, Reina Sofía University Hospital. Maimónides Institute for Biomedical Research (IMIBIC), University of Córdoba, Córdoba, Spain

^c Centro de Investigación Biomédica en Red de Enfermedades Infecciosas (CIBERINFEC), Instituto de Salud Carlos III, Madrid, Spain

^d Hepatology and Liver Transplantation Unit, Hospital Universitario La Fe-IIS La Fe Valencia, CiberEHD and University of Valencia, Spain

^e Lung Transplant Unit, Pneumology Service, Instituto de Investigación Hospital 12 de Octubre (imas12), University Hospital 12 de Octubre, Madrid, Spain

^f Department of Nephrology and Kidney Transplantation, Vall d'Hebron University Hospital-VHIR, Barcelona, Spain

^g Department of Infectious Diseases, Hospital Clínic-IDIBAPS, University of Barcelona, Spain

^h Department of Infectious Diseases, Bellvitge University Hospital-IDIBELL, University of Barcelona, Spain

ⁱ Unit of Infectious Diseases, Microbiology and Parasitology, Instituto de Biomedicina de Sevilla (IBiS), Virgen del Rocío University Hospital, Junta de Andalucía, CSIC, Universidad de Sevilla, Sevilla, Spain

^j Department of Medicine, Faculty of Medicine, Universidad de Sevilla, Spain

^k Department of Nephrology, Fundació Puigvert, Institut de Recerca Sant Pau (IR Sant Pau), RICORS 2024 (Kidney Disease), Barcelona, Spain

^l Department of Infectious Diseases, Hospital Universitario Marqués de Valdecilla-IDIVAL, Universidad de Cantabria, Santander, Spain

^m Microbiology Service, Clínica Universidad de Navarra, IdiSNA, Navarra Institute for Health Research, Pamplona, Spain

ⁿ Unit of Infectious Diseases, University Hospital "12 de Octubre", Instituto de Investigación Hospital "12 de Octubre" (i+12), School of Medicine, Universidad Complutense, Madrid, Spain

^o Service of Infectious Diseases, Ramón y Cajal University Hospital, IRYCIS, Madrid, Spain

^p Department of Cardiology, University Hospital "12 de Octubre", Instituto de Investigación Hospital "12 de Octubre" (i+12), CIBERCV, Madrid, Spain

^q Department of Pneumology, University Hospital Marqués de Valdecilla-IDIVAL, Santander, Spain

^r Department of Infectious Diseases, Vall d'Hebron for Solid Organ Transplantation Research Group, Vall d'Hebron University Hospital, Barcelona, Spain

^s Department of Nephrology, University Hospital La Paz, Madrid, Spain

Abbreviations: ATC, adoptive T-cell; ATG, anti-thymocyte globulin; BAL, bronchoalveolar lavage; CIBERINFEC, Biomedical Research Center Network in Infectious Diseases; CMI, cell-mediated immunity; CMV, cytomegalovirus; CMV Ig, Anti-CMV immunoglobulins; D, donor; EC₅₀, half maximal effective concentration; ELISA, enzyme-linked immunosorbent assays; ELISpot, enzyme-linked immunosorbent spot assays; FOS, foscarnet; GCV, ganciclovir; GESITRA-IC, Group for the Study of Infection in Transplantation and the Immunocompromised Host; GRADE, Grading of Recommendations Assessment, Development and Evaluation system; HSCT, hematopoietic stem cell transplantation; HT, heart transplant; ICS, intracellular cytokine staining; IFN-γ, interferon-gamma; IGRA, interferon-γ release assay; IV, intravenous; KT, kidney transplant; LET, letermovir; LT, lung transplant; mAb, monoclonal antibody; MBV, maribavir; mTOR, mammalian target of rapamycin; NGS, next-generation sequencing; NK, natural killer; PBMC, peripheral blood mononuclear cell; PCR, polymerase chain reaction; QNAT, quantitative nucleic acid amplification test; R, recipient; RCT, randomized controlled trial; SEIMC, Spanish Society of Infectious Diseases and Clinical Microbiology; SET, Spanish Society of Transplantation; SOT, solid organ transplantation; VGCV, valganciclovir; VST, virus-specific T cell.

* Correspondence to: J. Torre-Cisneros, Clinical Unit of Infectious Diseases, Instituto Maimónides de Investigación Biomédica (IMIBIC), Reina Sofía University Hospital, 14004 Córdoba, Spain.

** Correspondence to: J.M. Aguado, Unit of Infectious Diseases, Instituto de Investigación Hospital "12 de Octubre" (i+12) University Hospital "12 de Octubre", Universidad Complutense, 28041 Madrid, Spain.

E-mail addresses: julian.torre@sspa.juntadeandalucia.es (J. Torre-Cisneros), jaguadog1@gmail.com (J.M. Aguado).

<https://doi.org/10.1016/j.trre.2024.100875>

^t Department of Clinical Microbiology, Hospital Clinic, University of Barcelona, ISGlobal Barcelona Institute for Global Health, Barcelona, Spain^u Lung Transplant Program, Department of Pulmonology, Hospital Universitari Vall d'Hebron, Barcelona, Spain^v CIBER Enfermedades Respiratorias (CIBERES), Instituto de Salud Carlos III, Madrid, Spain^w Department of Clinical Microbiology and Infectious Diseases, Hospital General Universitario Gregorio Marañón, Instituto de Investigación Sanitaria Gregorio Marañón, Departamento de Medicina, Universidad Complutense, Madrid, Spain^x Microbiology Service, Clinic University Hospital, INCLIVA Health Research Institute, Valencia, Spain. Department of Microbiology School of Medicine, University of Valencia, Spain^y Maimonides Institute for Biomedical Research of Cordoba (IMIBIC), Reina Sofia University Hospital, University of Cordoba, Spain^z Microbiology Unit, Hospital Universitario Reina Sofia-Maimonides Institute for Biomedical Research (IMIBIC), Cordoba, Spain^{aa} Unit of Infectious Diseases, Hospital Universitario Cruces, Bilbao, Spain^{ab} Department of Nephrology, Reina Sofia University Hospital, Cordoba, Spain^{ac} Unit of Pneumology, Thoracic Surgery, and Lung Transplant, Reina Sofia University Hospital, Cordoba, Spain^{ad} Division of Health Sciences, Faculty of Medicine, Universidad del Norte, Hospital Universidad del Norte, Barranquilla, Colombia

ARTICLE INFO

Keywords:

Cytomegalovirus

SOT

Transplantation

Expert

Recommendations

Infection

Prevention

ABSTRACT

Cytomegalovirus (CMV) infection remains a significant challenge in solid organ transplantation (SOT). The last international consensus guidelines on the management of CMV in SOT were published in 2018, highlighting the need for revision to incorporate recent advances, notably in cell-mediated immunity monitoring, which could alter the current standard of care. A working group including members from the Group for the Study of Infection in Transplantation and the Immunocompromised Host (GESITRA-IC) of the Spanish Society of Infectious Diseases and Clinical Microbiology (SEIMC) and the Spanish Society of Transplantation (SET), developed consensus-based recommendations for managing CMV infection in SOT recipients. Recommendations were classified based on evidence strength and quality using the Grading of Recommendations Assessment, Development and Evaluation (GRADE) system. The final recommendations were endorsed through a consensus meeting and approved by the expert panel.

1. Introduction

Cytomegalovirus (CMV) infection in solid organ transplantation (SOT) recipients continues to pose a significant threat to their health. The last international consensus guidelines on the management of CMV in SOT were published in 2018 [1]. Since then, new anti-CMV drugs and methods to evaluate cell-mediated immunity (CMI) against CMV have been incorporated into clinical practice.

2. Methods

We created a working group composed of members of the Group for the Study of Infection in Transplantation and the Immunocompromised Host (GESITRA-IC) of the Spanish Society of Infectious Diseases and Clinical Microbiology (SEIMC), the Center for the Biomedical Research Network in Infectious Diseases (CIBERINFEC) and the Spanish Society of Transplantation (SET), to reach a consensus on the management of CMV infection in SOT recipients. As an initial step, the coordinators (JTC, JMA) proposed a series of clinical questions that were approved by the GESITRA-IC Board. The selected questions were distributed among different working groups of the expert panel. The MEDLINE, CENTRAL, Embase, and Clinical Trials databases were searched to retrieve relevant studies published until September 30, 2023. Unpublished communications to congresses, clinical cases, and small case series were excluded. The recommendations were classified by level of evidence and graded following the Grading of Recommendations Assessment, Development and Evaluation system (GRADE), which includes an assessment of the strength of the recommendation (i.e., strong, weak) and the quality of evidence (i.e., high, moderate, low, very low) [2–5]. Expert opinion grading was applied to recommendations with very low-quality evidence or those subject to controversy. All recommendations were discussed in The CMV Consensus Meeting held by GESITRA-IC/CIBERINFECT/SET on January 25, 2024, in Cordoba, Spain. Each working group proposed its recommendations and grading. These were discussed globally at the Consensus Meeting and agreed upon by majority vote. Specifically, recommendations based on expert opinion required a 2/3 majority of the expert panel to be approved. The final recommendations received support from patient associations collaborating with SEIMC and SET, in addition to all the members of both societies.

3. Recommendations

3.1. Virological monitoring

Few advances in virological monitoring have been made since our last consensus [6]. Importantly, a specific CMV viral load threshold for initiating preemptive treatment in asymptomatic patients is still pending [1], and different scenarios, including patient individual risk stratification, should be considered.

Quantitative nucleic acid amplification tests (QNATs), mainly those based upon real-time polymerase chain reaction (PCR), are the first choice for monitoring CMV DNAemia. Both whole blood and plasma specimens are equally suitable for monitoring. However, due to variability, it is necessary to use the same specimen for its consecutive monitoring [7]. Commercially available real-time QNAT assays have not been validated for CMV DNA quantitation in specimens other than whole blood and plasma, so the use of these techniques in bronchoalveolar lavage (BAL) or intestinal biopsies may be ancillary to cell-culture-based assays and histopathology. Since no consistent CMV DNA threshold for diagnosis of end-organ disease has been defined, only the lack of detection of CMV DNA in these specimens could have a high negative predictive value [8]. The dynamics of CMV loads over time may be more important in managing CMV infection than any absolute viral load value [9]. Specifically, using CMV DNA doubling time to initiate preemptive therapy may reduce variability across centers that apply different real-time QNATs [10–13]. Finally, virological monitoring during universal prophylaxis should be based on clinical criteria rather than being conducted systematically.

3.1.1. Consensus recommendations

- Insufficient evidence precludes recommending a specific threshold of DNAemia for triggering the inception of preemptive therapy in asymptomatic patients. Alternatively, we recommend individualizing each case based on risk stratification (**strong, low**).
- Real-time PCR assays performed on BAL or gastrointestinal biopsies display a high negative predictive value for diagnosing CMV pneumonitis or CMV gastrointestinal disease (**strong, low**).

Table 1

Summary of recommended primary prevention strategies and their potential modifications based on CMV-CMI monitoring availability.

Organ	Serostatus	Standard of care	Strength of recommendation; Quality of evidence	Potential modifications based on cell-mediated immune monitoring	Strength of recommendation; Quality of evidence
Kidney	D+/R-	Universal prophylaxis with VGCV ^a or LET ^b for 6 months (alternative: preemptive therapy with close monitoring ^c).	Strong; high [14–18,19,20,21]	Test CMV-CMI at the time of discontinuation ^e -If <u>positive</u> : stop prophylaxis and continue with preemptive therapy ^d . -If <u>negative</u> (or indeterminate): continue prophylaxis. Alternatively, switch to preemptive therapy with close monitoring and reassess CMV-CMI monthly.	Weak; moderate-low [22–29]
	R+	3 months of preemptive therapy (alternative: universal prophylaxis with VGCV ^a if close monitoring cannot be guaranteed).	Strong; high [14–16,30,31]	Baseline (pretransplant) or early (between 15–30 days) posttransplant monitoring of CMV-CMI: -If <u>negative</u> (or indeterminate): if D+, consider it as a high-risk (D+/R-) transplant. Test CMV-CMI at the time of discontinuation: -If <u>positive</u> : stop preemptive therapy. -If <u>negative</u> (or indeterminate): continue preemptive therapy with close monitoring and reassess CMV-CMI monthly.	Strong; moderate [32–37]
		Consider universal prophylaxis with LET ^b .	Expert opinion; no evidence available		Expert opinion; not specifically tested in RCTs
	R+ with ATG treatment or increased immunosuppression ^f	Universal prophylaxis with VGCV ^a for at least 3 months. Consider universal prophylaxis with LET ^b .	Strong; moderate [38–42]	Minimum 1 month of universal prophylaxis. Test CMV-CMI from then on: -If <u>positive</u> : stop prophylaxis and continue with preemptive therapy ^d . -If <u>negative</u> (or indeterminate): continue prophylaxis and reassess CMV-CMI monthly.	Strong; high [27,43]
Liver	D+/R- and R+ with increased immunosuppression ^f	Preemptive therapy for 3 months, as long as close monitoring ^c is available (alternative: universal prophylaxis with VGCV ^a). Consider universal prophylaxis with LET ^b .	Strong; high [44–49]	Test CMV-CMI at the time of discontinuation ^e :	Strong; moderate [24,50,27,29]
	R+	Preemptive therapy for 3 months.	Strong; high [46–49]	-If <u>positive</u> : stop prophylaxis and consider continuing with preemptive therapy ^d depending on the individual patient's risk. -If <u>negative</u> (or indeterminate): extend universal prophylaxis or switch to preemptive therapy with close monitoring ^c . Baseline (pretransplant) or early (between 15–30 days) posttransplant monitoring of CMV-CMI: - If <u>negative</u> (or indeterminate): if D+, consider it as a high-risk (D+/R-) transplant. Follow same strategy as in D+/R- transplant recipients at the time of discontinuation ^e .	Weak; low [51,52] Expert opinion; not specifically tested in RCTs
Heart	D+/R- and R+ with increased immunosuppression ^f	Universal prophylaxis for 3 to 6 months with VGCV ^a . Consider universal prophylaxis with LET ^b .	Strong; moderate-low [53,54,55] Expert opinion; no evidence available	Test CMV-CMI at the time of discontinuation ^e : -If <u>positive</u> : stop prophylaxis and consider continuing with preemptive therapy ^d depending on the individual patient's risk. -If <u>negative</u> (or indeterminate): continue prophylaxis and reassess CMV-CMI monthly (alternative: switch to preemptive therapy with close monitoring ^c)	Weak; low [24,56]

(continued on next page)

- Systematic virological monitoring is not recommended during universal prophylaxis (**expert opinion, controversial evidence**).

Table 1 (continued)

Organ	Serostatus	Standard of care	Strength of recommendation; Quality of evidence	Potential modifications based on cell-mediated immune monitoring	Strength of recommendation; Quality of evidence
R+		Universal prophylaxis for 3 months with VGCV ^a (alternative: preemptive therapy with close monitoring ^c).	Strong; moderate [57,58]	Baseline (pretransplant) or early (between 15-30 days) posttransplant monitoring of CMV-CMI: -If negative (or indeterminate): if D+, consider it as a high-risk (D+/R-) transplant.	Expert opinion; no evidence available
		Consider universal prophylaxis with LET ^b .	Expert opinion; no evidence available	Follow same strategy as in D+/R- transplant recipients at the time of discontinuation ^e .	Weak; low [59]
Lung	D+/R-	Universal prophylaxis for 12 months with VGCV ^a (consider continuation with preemptive therapy ^d depending on the individual patient's risk).	Strong; moderate [54,60-62,63,64]	Test CMV-CMI at the time of discontinuation ^e :	Strong; moderate [65,24,66]
		Consider universal prophylaxis with LET ^b .	Expert opinion; no evidence available	-If <u>positive</u> : stop universal prophylaxis and consider continuing with preemptive therapy ^d depending on the individual patient's risk. -If <u>negative</u> (or indeterminate): continue with universal prophylaxis or switch to preemptive therapy with close monitoring ^c and reassess CMV-CMI monthly.	
R+		Universal prophylaxis for at least 6 months with VGCV ^a , followed by preemptive therapy ^d for another 6 months.	Strong; moderate [54,60,62,63,64,67,68]	Baseline (pretransplant) or early (between 15-30 days) posttransplant monitoring of CMV-CMI: -If negative (or indeterminate): if D+, consider it as a high-risk (D+/R-) transplant.	Weak; low [36]
		Consider universal prophylaxis with LET ^b .	Expert opinion; no evidence available	Follow same strategy as in D+/R- transplant recipients at the time of discontinuation ^e .	Strong; moderate [65,69,66,70]
R+ with increased immunosuppression ^f		Universal prophylaxis for at least 6 months with VGCV ^a , followed by preemptive therapy ^d for another 6 months.	Weak; low [71]	Follow same strategy as in D+/R- transplant recipients at the time of discontinuation ^e .	Expert opinion; not specifically tested in RCTs
		Consider universal prophylaxis with LET ^b .	Expert opinion; no evidence available		
Intestinal, multivisceral, composite tissue	High risk no matter D/R serostatus	Universal prophylaxis with VGCV ^a for at least 6 months, followed by an indeterminate period of preemptive therapy ^d . Consider universal prophylaxis with LET ^b .	Weak; low [47,72-74]	Test CMV-CMI at the time of discontinuation ^e :	Expert opinion; no evidence available
Pancreas	High risk regardless of D/R serostatus	Universal prophylaxis with VGCV ^a for 3 to 6 months. Consider universal prophylaxis with LET ^b .	Weak; low [75-77]	Follow same strategy as in high-risk kidney transplant.	Expert opinion; no evidence available
			Expert opinion; no evidence available		
Liver, Heart, Lung, Kidney	R+ on preemptive therapy with asymptomatic viremia	Individualize each case, considering clinical aspects.	Expert opinion; controversial evidence	Test CMV-CMI: -If <u>positive</u> : withhold antiviral treatment.	Weak; low [78,79]

ATG: anti-thymocyte globulin; CMV: cytomegalovirus; CMV-CMI: CMV-specific cell-mediated immunity; D: donor; GCV: ganciclovir; Ig: immunoglobulin; LET: letermovir; R: recipient; RCT: randomized controlled trial; VGCV: valganciclovir.

^a In cases of oral intolerance, consider intravenous GCV.

^b In case of neutropenia or difficulties managing VGCV/GCV.

^c Strict preemptive protocol with weekly viral load monitoring, prompt reporting, and immediate therapy initiation.

^d If preemptive therapy is initiated after completing universal prophylaxis, the frequency of CMV monitoring could be extended to every two weeks.

^e Discontinuation of treatment can occur for various reasons, such as the completion of the recommended universal prophylaxis period, drug toxicity, or clinical criteria.

^f Therapies that induce profound immunosuppression, such as acute rejection treatment or ABO-incompatible protocols.

3.2. Prevention (Table 1)

3.2.1. Standard of care: universal prophylaxis and preemptive therapy

The duration of prophylaxis depends on the type of organ transplanted, the donor (D)/recipient (R) CMV serostatus, and the type of immunosuppression used. Table 1 summarizes recommendations regarding the standard prevention of CMV disease in SOT recipients.

3.2.1.1. Consensus statement. Universal prophylaxis and preemptive therapy are equally effective in preventing CMV disease if a strict preemptive protocol is followed. The decision to choose preemptive therapy is conditioned by the hospital's logistic capacity. The panel of experts advocates for clinicians to follow a strict preemptive protocol with weekly viral load monitoring, prompt reporting, and immediate therapy initiation. If preemptive therapy is initiated after completing universal prophylaxis, the frequency of CMV monitoring could be extended to every two weeks.

The quality of evidence supporting preemptive therapy compared to universal prophylaxis varies across different organ transplant contexts. Of particular concern are the potential indirect effects of viral replication on graft viability, especially in heart and lung transplantation. In the latter, immunoguided prophylaxis has recently provided new insights into the safety of the preemptive strategy (see below).

3.2.1.2. Kidney recipients. In the first randomized controlled trial (RCT) on seropositive (R+) kidney transplant (KT) recipients, it was found that preemptive therapy led to a higher incidence of CMV infection and disease compared to universal prophylaxis. However, they used a low-intense CMV surveillance protocol, and no differences were observed in terms of rejection, graft loss, and death [30,31]. Another meta-analysis in CMV R+ KT recipients showed that universal prophylaxis was better than preemptive therapy in the prevention of CMV disease [14]. However, the universal prophylaxis group required a high number of patients to be treated to achieve this outcome.

A recent RCT performed in CMV mismatch (D+/R-) and R+ recipients found that both regimens were effective in preventing CMV disease without increasing the risk of acute rejection. Nevertheless, preemptive therapy resulted in significantly higher rates of CMV DNAemia and lower neutropenia [15]. This is consistent with the results of a previous systematic review of RCTs, which showed that universal prophylaxis significantly reduced the rates of early CMV infection but led to higher rates of late infection and hematological adverse events [16]. A recently published meta-analysis of preemptive therapy stratified studies according to high or low threshold surveillance protocol in CMV D+/R- KT recipients. They found that a low threshold preemptive therapy (initiation of antiviral at any level of CMV DNAemia) was associated with a significantly lower CMV disease incidence than 6 months of universal prophylaxis or a high threshold preemptive therapy strategy [17]. Moreover, a retrospective study conducted on D+/R- KT recipients found no differences in long-term outcomes such as patient death, graft loss, or death-censored graft loss among both strategies [18].

3.2.1.3. Liver recipients. A single RCT conducted in high-risk (D+/R-) liver transplant recipients reported a lower incidence of CMV disease after one year of follow-up in patients managed with a preemptive approach compared to universal prophylaxis. There were no significant differences in terms of allograft rejection, opportunistic infections, graft loss, neutropenia, and all-cause mortality between both strategies [44]. In a post-hoc analysis of these results, preemptive therapy even showed protection against mortality [45]. Other meta-analyses [46–48] and a systematic review [49] concluded similar efficacy in the prevention of CMV disease of both strategies, regardless of serostatus, with no apparent impact on rejection, opportunistic infections, and mortality.

An increased incidence of CMV infection was observed only in D+/R- patients treated with preemptive therapy. This analysis excluded the only RCT in which this variable was not studied [48]. Finally, a preemptive strategy is also linked to lower overall costs compared to universal prophylaxis in high-risk patients [80,81].

3.2.1.4. Heart recipients. There is a lack of RCTs comparing both strategies in heart transplant (HT) recipients. In these patients, the effect of CMV replication in the development of graft vascular disease is well-documented [53,54], and universal prophylaxis is generally preferred. In fact, a preemptive strategy in R+ patients has been associated with an increased risk of CMV infection and CMV-related hospitalization in retrospective studies and may be associated with worse post-transplant graft outcomes [57,58].

3.2.1.5. Lung recipients. No RCT has compared preemptive versus universal prophylaxis in lung transplant (LT). However, one study has evaluated the outcomes using a hybrid strategy (see immunoguided prevention below) [65], and another is pending publication [82]. The risk of CMV infection and disease is notably elevated in LT recipients, and the indirect effect of CMV replication on graft survival is well-documented [54]. For this reason, universal prophylaxis is preferred [60–62].

3.2.1.6. Small bowel, pancreas, multivisceral, and composite tissue transplant recipients. Although few data have been published, intestinal and multivisceral transplant recipients are considered at high risk of CMV infection, and universal prophylaxis is generally prioritized over preemptive therapy [47,72,73], regardless of serostatus. A similar strategy is suggested for composite tissue transplants [74].

3.2.2. Antivirals for prophylaxis

Once-daily 900 mg of valganciclovir (VGCV), with normal renal function, remains the preferred regimen in any prophylaxis indication [1,6]. However, its utilization may be constrained by significant side effects, particularly neutropenia. In cases of oral intolerance, 5 mg/kg/day of intravenous (IV) ganciclovir (GCV) is an option, although it presents logistical drawbacks. Letermovir (LET) represents an oral alternative due to its similar efficacy and lower toxicity. It has been approved by both the EMA and FDA for primary prophylaxis in D+/R- KT recipients, following a recent RCT where LET was shown to be noninferior to VGCV in preventing CMV disease, with a lower incidence of neutropenia [19]. The dose of LET for prophylaxis is 480 mg per day (240 mg in case of coadministration with cyclosporine), and drugs to prevent herpes simplex and varicella virus infection should be added to this regimen [19]. Even though the use of low-dose VGCV (450 mg/day) was previously reported as an active off-label practice in a third of transplant centers [83], there is still a concern about viral replication progress and the emergence of GCV resistance, especially in high-risk transplant recipients [84–86].

The use of mammalian target of rapamycin (mTOR) inhibitors, alone or in combination with calcineurin inhibitors, as de novo or early conversion immunosuppression maintenance therapy has been shown to significantly reduce the incidence of CMV infection after kidney transplantation [87–90], especially in patients with low/moderate immunological risk and R+, but also in high-risk D+/R patients [88]. Similarly, converting mycophenolic acid to mTOR inhibitors in R+ KT patients after the first episode of treated infection is an effective and safe strategy for preventing recurrent episodes [91]. In heart and lung recipients, RCTs have also demonstrated a reduction in CMV infection rate [92–95]. It has been suggested that mTOR inhibitors have an immunomodulatory effect in R+ recipients, promoting the activation of antiviral memory and effector immune responses [90,96]. However, there is limited data regarding their potential long-term nephrotoxicity.

3.2.2.1. Consensus recommendations

- In patients with leukopenia or difficulties managing GCV/VGCV, LET is the alternative for D+/R- KT recipients (**strong, high**). This could be extended to the rest of the organs (**expert opinion, no evidence available**).
- LET should be the first choice in case of toxicity over low-dose VGCV (**expert opinion, no evidence available**).
- Switching to mTOR inhibitors is recommended in R+ KT recipients with CMV viremia while on preemptive therapy (**strong, high**).
- The use of mTOR inhibitors as de novo or early conversion immunosuppression maintenance therapy is suggested in patients with low/moderate immunological risk (**weak, moderate**).

3.2.3. Use of immunoglobulins in primary CMV disease prevention

Anti-CMV immunoglobulins (CMV Ig) have been used as a prophylactic measure for many years, primarily in combination with antivirals. They are mainly used in D+/R- high-risk patients and in cases of severe hypogammaglobulinemia [97], especially in cardiothoracic transplantation [98,99]. An observational study of heart and lung transplant recipients showed that patients who received combined prophylaxis had less CMV disease, fewer rejections, and better survival than those treated with IV GCV alone. Immunoglobulins have also been associated with a better immune profile in HT recipients [100]. In terms of duration, another observational study showed that there were no differences in the rate of acute rejection between LT recipients treated with 12 months and those treated with 24 months [101]. Data on the use of CMV Ig during hypogammaglobulinemia is limited [102–104]. However, studies on HT recipients have demonstrated the prevention of CMV disease [105–108].

3.2.3.1. Consensus recommendations

- We recommend a personalized administration of CMV Ig, along with antivirals, during the first months post-transplantation for high-risk lung and heart transplant recipients (D+/R-), especially in patients with moderate to severe hypogammaglobulinemia (**weak, moderate-low**).

3.2.4. Immunoguided prevention

3.2.4.1. Immunologic monitoring. Some immune biomarkers, such as absolute lymphocyte count [109–111], natural killer (NK) cell counts [112], CD4+/CD8+ T cell subsets [113,114], total serum immunoglobulin [97] and/or complement levels have been suggested to evaluate the risk of infection during the post-transplant period [115–117]. However, current risk-stratification algorithms are still based on pre-transplant D/R CMV serostatus and post-transplant viral load testing. Hypogammaglobulinemia in the post-transplant period ranges from 12% to 70% in different series [118–122] and is related to a significant increase in the risk of suffering various infections [105,123–125], including CMV [122,126,127].

For initial risk assessment before transplantation, it is recommended the determination of D/R CMV-specific IgG antibodies using serological tests with high sensitivity and specificity. Recent studies have shown controversial data about the quantification of IgG antibodies' usefulness in the pre-transplant setting [128,129]. Determination of CMV IgM is not routinely recommended unless primary CMV infection is suspected.

In the last decade, it has become clearer that CMV-CMI is critical in controlling CMV reactivation [130]. Several methods are currently available. Most of them rely on the detection of interferon (IFN)- γ production or release (interferon- γ release assay [IGRA]) following stimulation of whole blood or peripheral blood mononuclear cells (PBMCs) with CMV-specific proteins (pp65 and IE-1) or CMV-infected dendritic cells [131]. The most common methods include enzyme-linked immunosorbent assays (ELISA), enzyme-linked immunosorbent spot assays (ELISpot), and intracellular cytokine staining (ICS) assays with flow cytometry. In general, all assays measure T-cell mediated effector immune responses of IFN- γ production in response to pp65 and IE-1 [131]. The nature of each assay provides specific advantages and drawbacks related to the technical procedure itself or the biological insight. Moreover, there is no consensus on the appropriate cut-off for defining protection from active CMV infection or disease [22,23,32,33,78,132–134]. The best CMV-CMI assay in SOT is open to debate. Table 2 provides a concise summary of commercially available methods.

Table 2
Overview of commercially available functional methods for measuring CMV-specific T-cell mediated immune response.

Assays	Brand names	Sample required	Turnaround time	Target cells	CMV stimulating antigen	Advantages	Disadvantages
ELISpot (IGRA)	T-SPOT.CMV® T-Track CMV®	PBMCs (10 mL)	24–48 h	CD4+/CD8+/NK/NKT	Immunogenic peptides or proteins covering Pp65 and IE-1 proteins (separately)	Measures both CD4+ and CD8+ response Not limited by HLA CE marking	Requires ELISpot reader Requires PBMC isolation procedure Lack of cut-off standardization Unable to differentiate between CD4+ or CD8+ response
ELISA (IGRA)	QuantiFERON-CMV®	Whole blood (3–5 mL)	24 h	CD8+	Immunogenic peptides mapped within pp65, IE-1, IE-2, pp50 and gB	Standardized Simple to perform in laboratories Full automation CE marking	HLA Class I restricted Sensitive to lymphopenia Only measures CD8+ T cells (not CD4+ T cells)
ICS by flow cytometry	Viracor® CMV T-cell Immunity Panel (TCIP)	PBMCs or whole blood (1–2 mL)	8–10 h	CD4+/CD8+	Usually pp65, IE-1, gB immunogenic peptides (overlapping peptides libraries) or pp65/IE-1/gB proteins	Differentiates between CD4+ and CD8+ response Allows phenotypic characterization Potential to measure a variety of cytokines and cell surface markers	Requires flow cytometer High cost Lack of technical standardization Labour-intensive No CE marking

CE, European conformity; ELISA, enzyme-linked immunosorbent assay; ELISpot, enzyme-linked immunoabsorbent spot assay; gB, glycoprotein B; HLA, human leukocyte antigen; ICS, intracellular cytokine staining; IE, immediate-early; IGRA, Interferon-gamma release assay; NK/NKT, Natural Killer/Natural Killer T Cells; PBMCs, peripheral blood mononuclear cells; pp65, phosphoprotein 65.

Table 3

Observational studies and RCTs that have evaluated the clinical application of CMV-CMI monitoring in different clinical scenarios.

Clinical scenario	Predicted event	Proposed intervention	Observational studies	RCTs	Monitoring method
High-risk patients (D+/R-, T-cell-depleting antibodies, lung transplantation) during antiviral prophylaxis or at the time of discontinuation	Late-onset disease ^a	Prolong antiviral prophylaxis or close monitoring for viremia in patients with no protective CMV-CMI	Yes [22,134,23–25,33,50,66,28,135,29,70,136]	Yes [65,27,43]	QTF-CMV, ELISpot
Early posttransplant (between 15–30 days) monitoring in intermediate-risk (R+) patients	Post-transplant viremia and/or disease	Initiate antiviral prophylaxis in patients with no protective CMV-CMI	Yes [33,34,36,37,51,52]	Yes [32]	QTF-CMV, ELISpot, ICS
Intermediate-risk patients (R+) on preemptive therapy with no concurrent viremia	Subsequent viremia and/or disease	Reduce the frequency and/or discontinue monitoring of viremia if adequate response	Yes [33,34,28,135,137–140,141]	No	QTF-CMV, ELISpot, ICS, MHC-tetramer staining
Intermediate-risk patients (R+) on preemptive therapy with asymptomatic viremia	Spontaneous clearance	Withhold antiviral therapy if adequate response	Yes [78,79]	No	QTF-CMV
Active CMV infection or disease after discontinuation of antiviral treatment	Post-treatment relapse	Initiate secondary antiviral prophylaxis if inadequate response	Yes [142]	Yes [143]	QTF-CMV, ICS

CMV: cytomegalovirus; CMV-CMI: cytomegalovirus-specific cell-mediated immunity; D: donor; ELISpot: enzyme-linked immunosorbent spot assay; ICS: Intracellular cytokine staining; QTF-CMV: QuantiFERON-CMV assay; MHC: major histocompatibility complex; R: recipient; RCT: randomized controlled trial.

^a Refers to the occurrence of CMV disease after discontinuing antiviral prophylaxis with ganciclovir or valganciclovir (usually administered for 100–200 days).

3.2.4.1.1. Consensus recommendations

- During the early post-transplant period, we suggest determining other immune parameters such as absolute lymphocyte count, CD4+/CD8+ T cell subset, and total serum immunoglobulin for a global assessment of the immunological risk of CMV infection (**weak, low**).
- We recommend monitoring IgG levels during the first six months posttransplantation and whenever particularly severe or recurrent infections appear (**strong, moderate**).
- Monitoring CMV-CMI is recommended to optimize the preventive strategy (**strong, moderate**).
- An optimal CMV-CMI test should be cost-effective, reproducible, and applicable to assistance laboratories. No recommendation can be made regarding the best method to be used (**expert opinion, controversial evidence**).

3.2.4.2. Preventive strategies relying on CMV-CMI monitoring. Over the past years, many observational studies and RCTs have investigated the clinical application of CMV-CMI monitoring. This research comprises heterogeneous immune assays, study designs, transplant types, and clinical scenarios (high-risk [D+/R- or T-cell-depletion] or intermediate-risk [R+] patients), which are all summarized in Table 3. Based on these data, we make our recommendations in Table 1 that unify recommended primary preventive strategies by organ and the suggested modifications eventually based on CMV-CMI monitoring availability.

There are four main clinical scenarios in which CMV-CMI monitoring has been applied in CMV prevention. The strategy with the most evidence supports tailoring the duration of antiviral prophylaxis for high-risk patients (D+/R- and R+ treated with T-cell-depleting agents) based on the presence of a protective immune response rather than using a fixed-duration regimen. Observational studies have shown that those with a reactive assay at the end of the planned course of prophylaxis had a lower incidence of late CMV disease than those with a negative (or indeterminate) result [24–26,50,59,69], and even a lower incidence of high-level CMV replication was observed in LT recipients [66]. Three open-labeled RCTs have recently confirmed the safety and effectiveness of this strategy [65,27,43], and another one is pending publication [82]. In a single-center trial, 118 LT recipients were randomly assigned to receive either a fixed 5-month course or a variable length of VGCV

prophylaxis based on QuantiFERON® (QTF)-CMV assay results [65]. The incidence of CMV replication in BAL was significantly lower in the immunoguided arm and there were no differences in secondary outcomes, including acute cellular rejection, chronic lung allograft dysfunction, and death within 18 months of therapy, except for a lower recurrence rate of CMV viremia in the experimental arm [65]. The multicenter non-inferiority TIMOVAL trial [43] randomized 150 R+ KT recipients treated with anti-thymocyte globulin (ATG) to receive a fixed three-month course of VGCV prophylaxis or a variable duration based on the results of the QTF-CMV assay. Patients in the experimental group with a reactive QTF-CMV assay discontinued prophylaxis beyond the first post-transplant month to be subsequently managed by preemptive therapy up to month 3. The 12-month incidence of CMV disease was comparable between the experimental and control arms, as was the occurrence of CMV replication. The duration of prophylaxis was significantly reduced in the immunoguided group, and no differences were observed in terms of adverse events, including acute rejection, except neutropenia rate, which benefited the immunoguided arm [43]. Finally, in an open-label RCT carried out in six Swiss centers [27], they randomized 185 high-risk KT and liver transplant patients (R+ recipients receiving ATG or D+/R-) to receive a fixed-duration VGCV prophylaxis (3 or 6 months, depending on the risk group) or monthly CMV-CMI monitoring with an ELISpot assay (T-Track® CMV), discontinuing prophylaxis in the presence of a positive assay result. The clinically significant CMV infection incidence was similar between both groups, although non-inferiority could not be demonstrated. However, the duration of prophylaxis was shortened in the immunoguided group, and there were no differences in the incidence of acute rejection and allograft/patient survival at 1-year follow-up [27].

A second application of CMV-CMI monitoring allows stratifying the intermediate-risk R+ patients without T-cell-depleting induction therapy according to the presence of protective immunity at baseline or early after transplantation. About one-third of R+ patients have been found to lack a detectable CMV-CMI response before transplantation [34–37,56]. Mounting evidence suggests that this subgroup should be considered functionally comparable to R- patients, which would place them in the high-risk category if they receive an organ from a D+ [33,34,36,37,56,144]. Moreover, an early assessment (between 15–30 days posttransplantation) may provide an increased predictive capacity than the pre-transplant evaluation since some patients with robust preformed responses may lose their functional CMV-CMI following induction

Table 4

Principal clinical trials of CMV vaccines in solid organ transplantation. Adapted from [145].

Vaccine name	Vaccine type	NCT number	Phase	Route	Study cohort	Primary endpoint	Efficacy
Live-attenuated vaccine and disabled-infectious single cycle vaccine							
Towne vaccine	Attenuated Towne strain	NCT00370006 NCT00373412	I	sc	KT KT CMV HR-R (-) / D (+)	CMV infections	- No benefit in CMV (+). In HR, infection rates similar, less severe CMV disease - Infection rates similar, less severe CMV disease
Adjuvanted recombinant protein vaccine							
gB/MF59	MF59-adjuvanted recombinant soluble Towne strain gB	NCT00299260	II	im	Adults awaiting a KT or LT	Safety and immunogenicity	↑ gB-Ab in CMV (+ and -) vs placebo. In HR, ↓ duration of viraemia and number of days of ganciclovir treatment
DNA vaccine							
ASP0113 (VCL-6365, VCL-6368)	Poloxamer formulated, bivalent DNA vaccine expressing HCMV pp65 and gB	NCT01974206	II	im	Adults KT CMV HR R (-)/D (+)	CMV viremia from 100 d - 1y after the first study vaccine injection	Not efficacy in the prevention of CMV viremia in this cohort KT
Viral vectored vaccine							
HB-101	Bivalent vaccine containing 2 replication deficient lymphocytic choriomeningitis viruses expressing CMV pp65 and gB	NCT03629080	II	im	R (-) adults awaiting a KT from D (+)	Safety, immunogenicity and efficacy	• Results pending

CMV: cytomegalovirus; HR: high-risk patients, CMV D: donor CMV; CMV R: recipients CMV; KT: kidney transplant; LT: liver transplant; sc: subcutaneous; im: intramuscular; CMV: cytomegalovirus; gB: glycoprotein B; SOT: solid organ transplantation.

therapy [32,34]. Another application where CMV-CMI could be useful is to initiate secondary antiviral prophylaxis after discontinuation [143,142]. Finally, various observational studies have assessed the role of CMV-CMI monitoring in preemptively managed intermediate-risk (R+) patients to predict protection against CMV infection [78,33,34,28,79,135,137–141], discriminating those who will be able to spontaneously clear the infection.

3.2.4.2.1. Consensus recommendations

- Posttransplant monitoring of CMV-CMI should be used in R+ high-risk patients to guide antiviral prophylaxis (**strong, high**). This strategy may also be applied to D+/R- patients (**weak, low**).
- Early (between 15-30 days) posttransplant monitoring should be used in R+ intermediate-risk patients to stratify the subsequent CMV infection risk (**strong, moderate**).
- Posttransplant CMV-CMI may be monitored in R+ intermediate-risk patients managed by preemptive therapy to predict CMV spontaneous clearance (**weak, moderate**).
- Posttransplant CMV-CMI could be used to predict the risk of recurrence after discontinuation of treatment (**weak, moderate**).

3.2.5. Vaccines

Published phase-II/III RCTs have been performed mainly on healthy and hematopoietic stem cell transplant (HSCT) recipients [145], with few studies testing them on SOT [146]. Vaccines were generally well tolerated, with an adequate immune response. Unfortunately, they did not clearly demonstrate clinical efficacy. Table 4 references the main advanced clinical studies of vaccines in SOT based on the targets or the technology used in vaccine design.

In transplant patients, the current benefits of vaccines apply to a minority of individuals, and the potential effects on graft dysfunction are being explored as secondary endpoints in some ongoing clinical trials. Future aims will involve the selection of multivalent vaccines, primarily focusing on mRNA and viral-vectored vaccines [147], as well as preventive pre-transplant vaccination.

3.2.5.1. Consensus recommendations

- A recommendation regarding the use of CMV vaccines in SOT recipients cannot be made as no vaccine has been approved for use in a clinical setting (**expert opinion, no evidence available**).

3.3. Treatment

3.3.1. Initial treatment and duration

The efficacy of GCV for treating CMV disease has been demonstrated in numerous studies [148–153]. IV GCV must be given as initial treatment in patients with severe or life-threatening CMV disease, those with high viral load, and when gastrointestinal malabsorption is suspected [1,6]. Inadequate dosing can lead to ineffective treatment, resistance, and toxicity [154,155]. Consequently, frequent monitoring of renal function is recommended to guide dosage adjustments. Maribavir (MBV) and LET have not been extensively evaluated as initial treatment in SOT patients and are not approved for this indication [156]. Brincidofovir (BCV, CMX-001), given its safety concerns, and filociclovir (MBX-400 or cyclopropavir), still in the preclinical stage, are also not recommended [157–162]. Therapy with CMV-targeted monoclonal antibodies (mAbs) could be a new therapeutic alternative with potential advantages such as higher target specificity, antiviral activity, and lower toxicity [163]. However, there is still a lack of clinical data, with only one ongoing phase II RCT (NCT04225923) in KT recipients for CMV prevention.

The optimum duration should be individualized and continued until symptoms resolve; viral eradication is assessed by a viral load in one or two consecutive weekly samples and, generally, after a minimum treatment course of two weeks [153,164–166]. Secondary prophylaxis is not routinely recommended for all patients [167,168]. Many risk factors for developing recurrent CMV disease have been described, mostly related to primary disease severity and patient immunosuppression [164–166,169–171]. The therapeutic approach to patients suffering from recurrent CMV disease after a disease-free period should be the same as that used during the first episode [153].

3.3.1.1. Consensus recommendations

- CMV disease should be treated with VGCV (900 mg/12 h) or IV GCV (5 mg/kg/12 h), adjusted according to the glomerular filtration rate (**strong, high**).
- IV GCV should be the initial treatment for patients presenting with life-threatening and severe CMV disease or with gastrointestinal malabsorption (**strong, low**).
- Antiviral treatment should continue until clinical symptoms resolve and viral clearance is achieved (**strong, moderate**).
- Secondary prophylaxis is not routinely recommended (**strong, moderate**).

3.4. Refractory/resistant CMV

3.4.1. Concept of refractory/resistant CMV infection

The definitions of refractory/resistant CMV infection were updated during the revision process. We intend to incorporate these modifications into the final version of the document. Refractory CMV infection is defined as CMV viremia that increases (ie, $>1 \log_{10}$ increase in CMV DNA levels in the same blood compartment from the peak viral load as measured in the same laboratory and/or with the same commercial assay) OR persists ($\leq 1 \log_{10}$ increase or decrease in CMV DNA levels) after at least 2 weeks of appropriate antiviral therapy [172]. Therefore, it is considered a clinical definition based on suboptimal response to treatment criteria. On the other hand, resistant CMV infection is defined as refractory CMV infection, as defined above, in addition to viral genetic alteration that decreases susceptibility to one or more antiviral drugs [172].

3.4.2. Indications for antiviral resistance study

The prevalence of CMV resistance in SOT is low, but its impact on patient outcomes remains high [173,174]. Risk factors include prolonged antiviral drug exposure, younger age, exposure to low levels of GCV or inappropriate antiviral drugs, recipients' negative serostatus, type of transplanted organ, presenting the infection on VGCV prophylaxis, and coadministration of immunosuppressive therapies [175–177].

CMV resistance has been described to all available antivirals, including MBV and LET [178]. After GCV/VGCV exposure, the most common mutations occur in the UL97 kinase gene, followed by the UL54 DNA polymerase gene [175]. The most common UL97 mutations conferring clinical GCV/VGCV resistance are clustered at codons 460, 520, and 590 to 607, typically conferring 5- to 15-fold increases in GCV 50% inhibitory concentrations (EC₅₀), except C592G, which generates 3-fold increases. UL54 mutations typically add to pre-existing UL97 mutations to increase the level of GCV resistance and confer cross-resistance to other anti-CMV drugs [179]. Additionally, after treatment with MBV, it is necessary to evaluate resistance mutations at UL97 and UL27 genes and at UL56, UL89, and UL51 genes if the patient was treated with LET [180,181]. Most diagnostic laboratories test for limited codon ranges, such as 450 to 650 for UL97, 300 to 1000 for UL54, and 229 to 369 for UL56, which cover the most common mutations. However, these approaches could potentially omit some unusual loci where resistance mutations have been reported [182]. Sanger sequencing is the most commonly used method, although its sensitivity is limited to mutations found above 20% frequency in a viral population [183]. Next-generation sequencing (NGS) provides greater sensitivity and a higher resolution for detecting emerging antiviral resistances, although the accurate interpretation of minor variant subpopulations remains a challenge [182,184]. Genotypic testing could be performed on any type of sample [185,186], including plasma and whole blood. Finally, the emergence of new resistance mutants requires their phenotypic confirmation by recombinant phenotyping, which is only performed in reference laboratories [187,188].

3.4.2.1. Consensus recommendations

- Suspect drug resistance in patients with persistent/increasing CMV DNAemia or symptomatic disease despite full-dose therapy at least for 2 weeks, particularly if they have received cumulative antiviral exposure (**strong, high**).
- Sequencing of each genetic locus should be determined based on prior drug exposure (**strong, high**).

3.4.3. Management of refractory/resistant CMV infection

Genotypic analysis should guide treatment whenever possible. In general, immunosuppression should be minimized, including possible mTOR inhibitor conversion. In the case of low viral load and mild clinical symptoms, increasing GCV dose to 10 mg/kg/12 h may be beneficial [189]. This approach seems reasonable in CMV infections with low-level resistance mutations (e.g., C529G of the UL97 gene) and in situations of refractory infection due to malabsorption. However, it poses the risk of relapse due to inefficacy, the appearance of new mutations, and the potential for side effects, particularly neutropenia. Another option is FOS, particularly in cases of high viral load or life-threatening conditions, provided that the recipient does not show renal toxicity. [1]. Nonetheless, the use of FOS has been associated with high mortality and morbidity [190,191]. MBV, with its good oral bioavailability and favorable safety profile, has provided an alternative option for patients with refractory or resistant CMV irrespective of the transplanted organ [192,193]. However, as a possible selection bias in the most recent RCT [193], patients with low viral load and who had little severe damage to other organs were included. Another study on MBV-treated patients reported a 20% resistance rate due to mutations on UL97 (T409M, H411Y, or C480F) [194].

Severe hypogammaglobulinemia has been associated with the development of severe CMV disease episodes that are sometimes refractory to antiviral therapy. Retrospective studies suggested that combined immunoglobulin administration, including patients with refractory CMV, was associated with an improved clinical course [195–199]. However, further studies are required to investigate optimal protocols and their combination with emerging antiviral treatments.

3.4.3.1. Consensus recommendations

- Treatment of resistant CMV should be based on UL97 and UL54 gene analysis (**strong, high**).
- MBV should be used as the first therapeutic option in treating refractory/resistant disease, especially in cases with mild viral load (**strong, high**).
- In patients with GCV resistance, severe CMV disease, and high viral load, we suggest using FOS as initial therapy to decrease viral load and switching to MBV once a moderate-low viral load is achieved (**expert opinion, no evidence available**).

3.5. Adoptive therapy

Adoptive T-cell (ATC) therapy has been successfully employed in HSCT to prevent and treat viral disease [200–202]. In contrast to CMV-specific T cells generated from healthy CMV-seropositive individuals for administration in HSCT [203,204], ATC therapy in SOT relies on the ability to generate CMV-specific T cells from immunosuppressed individuals [205]. However, the development of third-party virus-specific T cell (VST) banks from immunocompetent donors is likely to overcome this issue [206,207].

Most data in SOT are isolated cases describing success in individuals with refractory or resistant CMV infection or limiting forms such as CMV retinitis [208–210]. The most important experience comes from a phase I clinical trial with autologous CMV-specific VST in 13 LT, HT, and KT recipients [211]. It showed an 84% success rate and no serious side

effects, including rejection. Immunosuppressive therapy decreases VST activity. To address this, anti-calcineurin-resistant VSTs have begun to be developed, although they have not yet entered clinical trials [212]. The duration of persistent VST remains relatively unknown and is affected by the characteristics of the infused product (autologous vs. HLA mismatch) and numerous receptor-related factors, such as graft rejection or immunosuppressive medication. Randomized, placebo-controlled trials are needed to assess their actual efficacy and safety.

3.5.1. Consensus recommendations

- Adoptive therapy could be useful as rescue therapy in severe CMV disease refractory to conventional treatment (**weak, very low**). Its implementation as a preventive therapy to replace current drug treatment has not been consistently evaluated in clinical trials (**expert opinion, no evidence available**).

4. Conclusions

Despite the progress made in managing CMV infection, there are still several aspects that need clarification. Incorporating CMV-CMI monitoring into clinical algorithms has been shown to improve the prediction of CMV infection and reduce costs associated with preemptive therapy and the use of antivirals through universal prophylaxis. Additionally, it can help address the challenge of determining a viral load threshold for initiating preemptive treatment in asymptomatic patients. However, further studies are needed to establish a consensus on the appropriate cut-off for defining protection from active CMV infection or disease using different assays. It is also important to conduct additional research on the cost-effectiveness of these monitoring techniques.

While the approval of LET for prophylaxis in D+/R- KT recipients and MBV for refractory/resistant disease represent significant progress in disease control, further advancements, such as developing vaccines, mAbs and adoptive therapy, are necessary to reduce the incidence of this disease. Specifically, more data are needed on the use of letermovir in prophylaxis outside of renal transplantation and the use of maribavir in other clinical scenarios, such as in patients with high viral loads.

Funding

This document has been financed by the Spanish Society of Infectious Diseases and Clinical Microbiology (SEIMC).

Disclosure

Aguado JM has been a consultant to and on the speakers' bureau for Mundipharma, Pfizer, Merck Sharp and Dohme (MSD), and Takeda Pharmaceuticals. Cantás S has received research funding and honoraria for educational activities from Qiagen. Castón JJ has received honoraria for speaking at educational events from Pfizer, MSD, Menarini, Novartis, and Qiagen. Additionally, he has received research grants from Pfizer and Qiagen. Facundo C has received honoraria for serving on the advisory board from MSD. Fernández-Alonso M has received travel support from Roche and Werfen for scientific purposes and additionally has received reagents for research from Qiagen. Fernández-Ruiz M has received honoraria for serving on advisory boards from MSD and BioTech. Herrera S has received honoraria for speaking at educational events from Pfizer, Shionogi and has received travel support from Shire, Pfizer, and Menarini for scientific purposes. Len O has received research grants from Pfizer and MSD and has been a speaker for Pfizer, Astellas, Novartis, and MSD. Los-Arcos I has received honoraria for speaking at educational events from MSD and Pfizer and has received travel support from Gilead, MSD, Pfizer, and Menarini for scientific purposes. Navarro D has been a consultant to and on the speakers' bureau for Roche Pharma, Abbott, Biomerieux, Beckton Dickinson, Takeda, Gilead, MSD, and Pfizer. Torre-Cisneros J has been a consultant to and on the speakers' bureau for MSD, Takeda Pharmaceuticals, Biotest, and

Qiagen. He has also received a research fund from Qiagen. Valerio M has received travel grants from Pfizer, MSD, and Shionogi and received speaker honoraria from Pfizer, MSD, GSK, and Shionogi.

Declaration of competing interest

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

Julian Torre-Cisneros reports administrative support, article publishing charges, travel, and writing assistance were provided by Spanish Society of Infectious Diseases and Clinical Microbiology. Julian Torre-Cisneros reports a relationship with Merck Sharp & Dohme UK Ltd. that includes: consulting or advisory and speaking and lecture fees. Julian Torre-Cisneros reports a relationship with Takeda Pharmaceutical Company Limited that includes: consulting or advisory. Julian Torre-Cisneros reports a relationship with Biotest AG that includes: consulting or advisory and speaking and lecture fees. Julian Torre-Cisneros reports a relationship with QIAGEN GmbH that includes: consulting or advisory, funding grants, and speaking and lecture fees. Jose Maria Aguado reports a relationship with Mundipharma International Limited that includes: consulting or advisory and speaking and lecture fees. Jose Maria Aguado reports a relationship with Pfizer that includes: consulting or advisory and speaking and lecture fees. Jose Maria Aguado reports a relationship with Merck Sharp & Dohme UK Ltd. that includes: consulting or advisory and speaking and lecture fees. Jose Maria Aguado reports a relationship with Takeda Pharmaceutical Company Limited that includes: consulting or advisory and speaking and lecture fees. Sara Cantás reports a relationship with QIAGEN GmbH that includes: funding grants and speaking and lecture fees. Juan Jose Castón reports a relationship with Pfizer Inc. that includes: funding grants and speaking and lecture fees. Juan Jose Castón reports a relationship with Merck Sharp & Dohme UK Ltd. that includes: speaking and lecture fees. Juan Jose Castón reports a relationship with A Menarini International Pharmaceuticals that includes: speaking and lecture fees. Juan Jose Castón reports a relationship with Novartis Pharmaceuticals Corporation that includes: speaking and lecture fees. Juan Jose Castón reports a relationship with QIAGEN GmbH that includes: funding grants and speaking and lecture fees. Carme Facundo reports a relationship with Merck Sharp & Dohme UK Ltd. that includes: consulting or advisory. Mirian Fernandez-Alonso reports a relationship with Roche Diagnostics Corporation that includes: travel reimbursement. Mirian Fernandez-Alonso reports a relationship with Werfen that includes: travel reimbursement. Mirian Fernandez-Alonso reports a relationship with QIAGEN GmbH that includes: non-financial support. Mario Fernandez-Ruiz reports a relationship with Merck Sharp & Dohme UK Ltd. that includes: consulting or advisory. Mario Fernandez-Ruiz reports a relationship with Biotech that includes: consulting or advisory. Sabina Herrera reports a relationship with Pfizer Inc. that includes: speaking and lecture fees and travel reimbursement. Sabina Herrera reports a relationship with Shionogi Inc. that includes: speaking and lecture fees. Sabina Herrera reports a relationship with Shire that includes: travel reimbursement. Sabina Herrera reports a relationship with A Menarini International Pharmaceuticals that includes: travel reimbursement. Oscar Len reports a relationship with Pfizer that includes: funding grants and speaking and lecture fees. Oscar Len reports a relationship with Merck Sharp & Dohme UK Ltd. that includes: funding grants and speaking and lecture fees. Oscar Len reports a relationship with Astellas Pharma Inc. that includes: speaking and lecture fees. Oscar Len reports a relationship with Novartis Pharmaceuticals Corporation that includes: speaking and lecture fees. Ibai Los-Arcos reports a relationship with Pfizer Inc. that includes: speaking and lecture fees and travel reimbursement. Ibai Los-Arcos reports a relationship with Merck Sharp & Dohme UK Ltd. that includes: speaking and lecture fees and travel reimbursement. Ibai Los-Arcos reports a relationship with Gilead Sciences Inc. that includes: travel reimbursement. Ibai Los-Arcos reports a relationship with A Menarini International Pharmaceuticals that includes: travel

reimbursement. David Navarro reports a relationship with Roche Diagnostics Corporation that includes: consulting or advisory and speaking and lecture fees. David Navarro reports a relationship with Abbott that includes: consulting or advisory and speaking and lecture fees. David Navarro reports a relationship with bioMérieux SA that includes: consulting or advisory and speaking and lecture fees. David Navarro reports a relationship with Beckton Dickinson that includes: consulting or advisory and speaking and lecture fees. David Navarro reports a relationship with Gilead Sciences Inc. that includes: consulting or advisory and speaking and lecture fees. David Navarro reports a relationship with Takeda Pharmaceutical Company Limited that includes: consulting or advisory and speaking and lecture fees. David Navarro reports a relationship with Merck Sharp & Dohme UK Ltd. that includes: consulting or advisory and speaking and lecture fees. David Navarro reports a relationship with Pfizer Inc. that includes: consulting or advisory and speaking and lecture fees. Maricela Valerio reports a relationship with Pfizer Inc. that includes: speaking and lecture fees and travel reimbursement. Maricela Valerio reports a relationship with Merck Sharp & Dohme UK Ltd. that includes: speaking and lecture fees and travel reimbursement. Maricela Valerio reports a relationship with Shionogi and Co Ltd. that includes: speaking and lecture fees and travel reimbursement. Maricela Valerio reports a relationship with GSK that includes: speaking and lecture fees. If there are other authors, they declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- [1] Kotton CN, Kumar D, Caliendo AM, Huprikar S, Chou S, Danziger-Isakov L, et al. The third international consensus guidelines on the Management of Cytomegalovirus in solid-organ transplantation. *Transplantation* 2018;102: 900–31. <https://doi.org/10.1097/TP.0000000000002191>.
- [2] Schünemann HJ, Oxman AD, Brozek J, Glasziou P, Jaeschke R, Vist GE, et al. Grading quality of evidence and strength of recommendations for diagnostic tests and strategies. *BMJ* 2008;336:1106–10. <https://doi.org/10.1136/bmj.39500.677199.AE>.
- [3] Guyatt GH, Oxman AD, Kunz R, Jaeschke R, Helfand M, Liberati A, et al. Incorporating considerations of resources use into grading recommendations. *BMJ* 2008;336:1170–3. <https://doi.org/10.1136/bmj.39504.506319.80>.
- [4] Jaeschke R, Guyatt GH, Dellinger P, Schunemann H, Levy MM, Kunz R, et al. Use of GRADE grid to reach decisions on clinical practice guidelines when consensus is elusive. *BMJ* 2008;337:a744. <https://doi.org/10.1136/bmj.a744>.
- [5] Guyatt GH, Oxman AD, Vist GE, Kunz R, Falck-Ytter Y, Alonso-Coello P, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ* 2008;336:924–6. <https://doi.org/10.1136/bmj.39489.470347.AD>.
- [6] Torre-Cisneros J, Aguado JM, Caston JJ, Almenar L, Alonso A, Cantisán S, et al. Management of cytomegalovirus infection in solid organ transplant recipients: SET/GESITRA-SEIMC/REIPI recommendations. *Transplant Rev* 2016;30:119–43. <https://doi.org/10.1016/j.trre.2016.04.001>.
- [7] Vinuesa V, Giménez E, Solano C, Albert E, Torres I, Pérez A, et al. Factors influencing cytomegalovirus DNA load measurements in whole blood and plasma specimens from allogeneic hematopoietic stem cell transplant recipients. *Diagn Microbiol Infect Dis* 2019;94:22–7. <https://doi.org/10.1016/j.dmicrobo.2018.11.012>.
- [8] Huang C, Solis D, Sahoo MK, Pinsky BA. Assessment of an automated cytomegalovirus nucleic acid amplification test using clinical plasma, bronchoalveolar lavage, and tissue specimens. *J Clin Virol* 2023;168:105582. <https://doi.org/10.1016/j.jcv.2023.105582>.
- [9] Peddy V, Bradley BT, Castro AM, Shree R, Colbert BG, Xie H, et al. High-resolution profiling of human cytomegalovirus cell-free DNA in human plasma highlights its exceptionally fragmented nature. *Sci Rep* 2020;10:3734. <https://doi.org/10.1038/s41598-020-60655-6>.
- [10] Vinuesa V, Giménez E, Solano C, Gimeno C, Navarro D. Would kinetic analyses of plasma cytomegalovirus DNA load help to reach consensus criteria for triggering the initiation of preemptive antiviral therapy in transplant recipients? *Clin Infect Dis* 2016;63:1533–5. <https://doi.org/10.1093/cid/ciw608>.
- [11] Atabani SF, Smith C, Atkinson C, Aldridge RW, Rodriguez-Perálvarez M, Rolando N, et al. Cytomegalovirus replication kinetics in solid organ transplant recipients managed by preemptive therapy. *Am J Transplant* 2012;12:2457–64. <https://doi.org/10.1111/j.1600-6143.2012.04087.x>.
- [12] Griffiths PD, Rothwell E, Raza M, Wilmore S, Doyle T, Harber M, et al. Randomized controlled trials to define viral load thresholds for cytomegalovirus pre-emptive therapy. *PLoS One* 2016;11:e0163722. <https://doi.org/10.1371/journal.pone.0163722>.
- [13] Martín-Gandul C, Pérez-Romero P, Mena-Romo D, Molina-Ortega A, González-Roncero FM, Suárez M, et al. Kinetic of the CMV-specific T-cell immune response and CMV infection in CMV-seropositive kidney transplant recipients receiving rabbit anti-thymocyte globulin induction therapy: a pilot study. *Transpl Infect Dis* 2018;20. <https://doi.org/10.1111/tid.12883>.
- [14] Caskurlu H, Karadag FY, Arslan F, Cag Y, Vahaboglu H. Comparison of universal prophylaxis and preemptive approach for cytomegalovirus associated outcome measures in renal transplant patients: a meta-analysis of available data. *Transpl Infect Dis* 2019;21. <https://doi.org/10.1111/tid.13016>.
- [15] Reischig T, Vlas T, Kacer M, Pivoarcikova K, Lysak D, Nemcova J, et al. A randomized trial of Valganciclovir prophylaxis versus preemptive therapy in kidney transplant recipients. *J Am Soc Nephrol* 2023;34:920–34. <https://doi.org/10.1681/ASN.0000000000000090>.
- [16] Raval AD, Kistler K, Tang Y, Murata Y, Snydman DR. Antiviral treatment approaches for cytomegalovirus prevention in kidney transplant recipients: a systematic review of randomized controlled trials. *Transplant Rev* 2021;35: 100587. <https://doi.org/10.1016/j.trre.2020.100587>.
- [17] Kumar L, Murray-Krezan C, Singh N, Brennan DC, Rakita RM, Dasgupta S, et al. A systematic review and Meta-analysis of optimized CMV preemptive therapy and antiviral prophylaxis for CMV disease prevention in CMV high-risk (D+R-) kidney transplant recipients. *Transplant Direct* 2023;9:e1514. <https://doi.org/10.1097/TXD.00000000000001514>.
- [18] Blom KB, Birkeland GK, Midtvedt K, Jenssen TG, Reisæter AV, Rollag H, et al. Cytomegalovirus high-risk kidney transplant recipients show no difference in long-term outcomes following preemptive versus prophylactic management. *Transplantation* 2023;107:1846–53. <https://doi.org/10.1097/TP.0000000000004615>.
- [19] Limaye AP, Budde K, Humar A, Vincenti F, Kuypers DRJ, Carroll RP, et al. Letermovir vs Valganciclovir for prophylaxis of cytomegalovirus in high-risk kidney transplant recipients. *JAMA* 2023;330:33. <https://doi.org/10.1001/jama.2023.9106>.
- [20] Humar A, Lebranchu Y, Vincenti F, Blumberg EA, Punch JD, Limaye AP, et al. The efficacy and safety of 200 days Valganciclovir cytomegalovirus prophylaxis in high-risk kidney transplant recipients. *Am J Transplant* 2010;10:1228–37. <https://doi.org/10.1111/j.1600-6143.2010.03074.x>.
- [21] Humar A, Limaye AP, Blumberg EA, Hauser IA, Vincenti F, Jardine AG, et al. Extended Valganciclovir prophylaxis in D+/R– kidney transplant recipients is associated with long-term reduction in cytomegalovirus disease: two-year results of the IMPACT study. *Transplantation* 2010;90:1427–31. <https://doi.org/10.1097/TP.0b013e3181ff1493>.
- [22] Kumar D, Chernenko S, Moussa G, Cobos I, Manuel O, Preiksaitis J, et al. Cell-mediated immunity to predict cytomegalovirus disease in high-risk solid organ transplant recipients. *Am J Transplant* 2009;9:1214–22. <https://doi.org/10.1111/j.1600-6143.2009.02618.x>.
- [23] Kumar D, Chin-Hong P, Kayler L, Wojciechowski D, Limaye AP, Osama Gaber A, et al. A prospective multicenter observational study of cell-mediated immunity as a predictor for cytomegalovirus infection in kidney transplant recipients. *Am J Transplant* 2019;19:2505–16. <https://doi.org/10.1111/ajt.15315>.
- [24] Manuel O, Husain S, Kumar D, Zayas C, Mawhorter S, Levi ME, et al. Assessment of cytomegalovirus-specific cell-mediated immunity for the prediction of cytomegalovirus disease in high-risk solid-organ transplant recipients: a multicenter cohort study. *Clin Infect Dis* 2013;56:817–24. <https://doi.org/10.1093/cid/cis993>.
- [25] Deborska-Materkowska D, Perkowska-Ptasinska A, Sadowska A, Gozdowska J, Ciszek M, Serwanska-Swietek M, et al. Diagnostic utility of monitoring cytomegalovirus-specific immunity by QuantiFERON-cytomegalovirus assay in kidney transplant recipients. *BMC Infect Dis* 2018;18:179. <https://doi.org/10.1186/s12879-018-3075-z>.
- [26] Ruan Y, Guo W, Liang S, Xu Z, Niu T. Diagnostic performance of cytomegalovirus (CMV) immune monitoring with ELISPOT and QuantiFERON-CMV assay in kidney transplantation: a PRISMA-compliant article. *Medicine* 2019;98:e15228. <https://doi.org/10.1097/MD.00000000000015228>.
- [27] Manuel O, Laager M, Hirzel C, Neofytos D, Walti LN, Hoenger G, et al. Immune monitoring-guided vs fixed duration of antiviral prophylaxis against cytomegalovirus in solid-organ transplant recipients. A multicenter, randomized Clinical Trial. *Clin Infect Dis* 2023. <https://doi.org/10.1093/cid/ciad575>.
- [28] Abate D, Saldan A, Mengoli C, Fiscon M, Silvestre C, Fallico L, et al. Comparison of cytomegalovirus (CMV) enzyme-linked immunosorbent spot and CMV quantiferon gamma interferon-releasing assays in assessing risk of CMV infection in kidney transplant recipients. *J Clin Microbiol* 2013;51:2501–7. <https://doi.org/10.1128/JCM.00563-13>.
- [29] San-Juan R, Navarro D, García-Reyne A, Montejó M, Muñoz P, Carratala J, et al. Effect of delaying prophylaxis against CMV in D+/R– solid organ transplant recipients in the development of CMV-specific cellular immunity and occurrence of late CMV disease. *J Inf Secur* 2015;71:561–70. <https://doi.org/10.1016/j.jinf.2015.06.013>.
- [30] Witzke O, Hauser IA, Bartels M, Wolf G, Wolters H, Nitschke M. Valganciclovir prophylaxis versus preemptive therapy in cytomegalovirus-positive renal allograft recipients: 1-year results of a randomized clinical trial. *Transplantation* 2012;93:61–8. <https://doi.org/10.1097/TP.0b013e318238dab3>.
- [31] Witzke O, Nitschke M, Bartels M, Wolters H, Wolf G, Reinke P, et al. Valganciclovir prophylaxis versus preemptive therapy in cytomegalovirus-positive renal allograft recipients: long-term results after 7 years of a randomized clinical trial. *Transplantation* 2018;102:876–82. <https://doi.org/10.1097/TP.0000000000002024>.

- [32] Jarque M, Crespo E, Melilli E, Gutiérrez A, Moreso F, Guirado L, et al. Cellular immunity to predict the risk of cytomegalovirus infection in kidney transplantation: a prospective, interventional. Multicenter Clin Trial Clin Infect Dis 2020;71:2375–85. <https://doi.org/10.1093/cid/ciz1209>.
- [33] Bestard O, Lucia M, Crespo E, Van Liempt B, Palacio D, Melilli E, et al. Pretransplant immediately early-1-specific T cell responses provide protection for CMV infection after kidney transplantation. Am J Transplant 2013;13:1793–805. <https://doi.org/10.1111/ajt.12256>.
- [34] Fernández-Ruiz M, Giménez E, Vinuesa V, Ruiz-Merlo T, Parra P, Amat P, et al. Regular monitoring of cytomegalovirus-specific cell-mediated immunity in intermediate-risk kidney transplant recipients: predictive value of the immediate post-transplant assessment. Clin Microbiol Infect 2019;25:381.e1–381.e10. <https://doi.org/10.1016/j.cmi.2018.05.010>.
- [35] Cantisán S, Rodelo-Haas C, Páez-Vega A, Nieto A, Vaquero JM, Poyato A, et al. Factors related to the development of CMV-specific CD8+ T cell response in CMV-seropositive solid organ transplant candidates. Am J Transplant 2015;15:715–22. <https://doi.org/10.1111/ajt.13012>.
- [36] Cantisán S, Lara R, Montejo M, Redel J, Rodríguez-Benot A, Gutiérrez-Aroca J, et al. Pretransplant interferon- γ secretion by CMV-specific CD8+ T cells informs the risk of CMV replication after transplantation. Am J Transplant 2013;13:738–45. <https://doi.org/10.1111/ajt.12049>.
- [37] López-Oliva MO, Martínez V, Buitrago A, Jiménez C, Rivas B, Escuin F, et al. Pretransplant CD8 T-cell response to IE-1 discriminates seropositive kidney recipients at risk of developing CMV infection posttransplant. Transplantation 2014;97:839–45. <https://doi.org/10.1097/TP.000000000000438025.96334.eb>.
- [38] Santos CAQ, Brennan DC, Fraser VJ, Olsen MA. Delayed-onset cytomegalovirus disease coded during hospital readmission after kidney transplantation. Transplantation 2014;98:187–94. <https://doi.org/10.1097/TP.0000000000000030>.
- [39] Lee Y-M, Kim YH, Han DJ, Park S-K, Park JS, Sung H, et al. Cytomegalovirus infection after acute rejection therapy in seropositive kidney transplant recipients. Transpl Infect Dis 2014;16:397–402. <https://doi.org/10.1111/tid.12227>.
- [40] Schachtnar T, Stein M, Reinke P. ABO desensitization affects cellular immunity and infection control after renal transplantation. Transpl Int 2015;28:1179–94. <https://doi.org/10.1111/tri.12616>.
- [41] Song SH, Lee J, Kim BS, Kim S, Lee JG, Jeong HJ, et al. Successful launch of an ABO-incompatible kidney transplantation program to overcome the shortage of compatible living donors: experience at a single center. Clin Nephrol 2017;88:117–23. <https://doi.org/10.5414/CN109114>.
- [42] Kute VB, Pathak V, Ray DS, Bhalla AK, Godara SM, Narayanan S, et al. A multicenter retrospective cohort study on management protocols and clinical outcomes after ABO-incompatible kidney transplantation in India. Transplantation 2023. <https://doi.org/10.1097/TP.0000000000004789>.
- [43] Páez-Vega A, Gutiérrez-Benot B, Facundo C, Redondo-Pachón D, Suárez M, et al. Immunoguided discontinuation of prophylaxis for cytomegalovirus disease in kidney transplant recipients treated with antithymocyte globulin: a randomized clinical trial. Clin Infect Dis 2022;74:757–65. <https://doi.org/10.1093/cid/ciab574>.
- [44] Singh N, Winston DJ, Razonable RR, Lyon GM, Silveira FP, Wagener MM, et al. Effect of preemptive therapy vs antiviral prophylaxis on cytomegalovirus disease in seronegative liver transplant recipients with seropositive donors: a randomized clinical trial. JAMA 2020;323:1378–87. <https://doi.org/10.1001/jama.2020.3138>.
- [45] Kumar L, Dasgupta S, Murray-Krezan C, Singh N, Rakita RM, Fisher CE, et al. Association of CMV DNAemia with long-term mortality in a randomized trial of preemptive therapy (PET) and antiviral prophylaxis (AP) for prevention of CMV disease in high-risk donor seropositive, recipient seronegative (D+R-) liver transplant recipients. Clin Infect Dis 2023. <https://doi.org/10.1093/cid/ciad643>.
- [46] Mumtaz K, Faisal N, Husain S, Morillo A, Renner EL, Shah PS. Universal prophylaxis or preemptive strategy for cytomegalovirus disease after liver transplantation: a systematic review and Meta-analysis. Am J Transplant 2015;15:472–81. <https://doi.org/10.1111/ajt.13044>.
- [47] Florescu DF, Qiu F, Schmidt CM, Kalil AC. A direct and indirect comparison Meta-analysis on the efficacy of cytomegalovirus preventive strategies in solid organ transplant. Clin Infect Dis 2014;58:785–803. <https://doi.org/10.1093/cid/cit945>.
- [48] Yadav DK, Adhikari VP, Yadav RK, Singh A, Huang X, Zhang Q, et al. Antiviral prophylaxis or preemptive therapy for cytomegalovirus after liver transplantation? a systematic review and meta-analysis. Front Immunol 2022;13:953210. <https://doi.org/10.3389/fimmu.2022.953210>.
- [49] Campos-Varela I, Blumberg EA, Giorgio P, Kotton CN, Saliba F, Wey EQ, et al. What is the optimal antimicrobial prophylaxis to prevent postoperative infectious complications after liver transplantation? A systematic review of the literature and expert panel recommendations. Clin Transpl 2022;36. <https://doi.org/10.1111/ctr.14631>.
- [50] Gliga S, Fiedler M, Dornieden T, Achterfeld A, Paul A, Horn PA, et al. Comparison of three cellular assays to predict the course of CMV infection in liver transplant recipients. Vaccines (Basel) 2021;9. <https://doi.org/10.3390/vaccines9020088>.
- [51] Bhugra A, Khodare A, Agarwal R, Pamecha V, Gupta E. Role of cytomegalovirus specific cell-mediated immunity in the monitoring of cytomegalovirus infection among living donor liver transplantation adult recipients: a single-center experience. Transpl Infect Dis 2023;25:e14011. <https://doi.org/10.1111/tid.14011>.
- [52] Páez-Vega A, Poyato A, Rodriguez-Benot A, Guirado L, Fortún J, Len O, et al. Analysis of spontaneous resolution of cytomegalovirus replication after transplantation in CMV-seropositive patients with pretransplant CD8+IFNG+ response. Antiviral Res 2018;155:97–105. <https://doi.org/10.1016/j.antiviral.2018.05.006>.
- [53] Manuel O, Kralidis G, Mueller NJ, Hirsch HH, Garzoni C, van Delden C, et al. Impact of antiviral preventive strategies on the incidence and outcomes of cytomegalovirus disease in solid organ transplant recipients. Am J Transplant 2013;13:2402–10. <https://doi.org/10.1111/ajt.12388>.
- [54] L’Huillier AG, Ferreira VH, Ku T, Bahinskaya I, Kumar D, Humar A. Improving our mechanistic understanding of the indirect effects of CMV infection in transplant recipients. Am J Transplant 2019;19:2495–504. <https://doi.org/10.1111/ajt.15371>.
- [55] Imlay H, Dumitriu Carcoana AO, Fisher CE, Wong B, Rakita RM, Fishbein DP, et al. Impact of valganciclovir prophylaxis duration on cytomegalovirus disease in high-risk donor seropositive/recipient seronegative heart transplant recipients. Transpl Infect Dis 2020;22. <https://doi.org/10.1111/tid.13255>.
- [56] Schachtnar T, Stein M, Reinke P. CMV-specific T cell monitoring offers superior risk stratification of CMV-seronegative kidney transplant recipients of a CMV-seropositive donor. Transplantation 2017;101:e315–25. <https://doi.org/10.1097/TP.00000000000001825>.
- [57] Gardiner BJ, Bailey JP, Percival MA, Morgan BA, Warner VM, Lee SJ, et al. Incidence and severity of cytomegalovirus infection in seropositive heart transplant recipients. Clin Transpl 2023;37. <https://doi.org/10.1111/ctr.14982>.
- [58] Lerman JB, Green CL, Molina MR, Maharaj V, Ortega-Legaspi JM, Sen S, et al. Multicenter study of universal prophylaxis versus pre-emptive therapy for patients at intermediate risk (R+) for CMV following heart transplantation. Clin Transpl 2023;37:e15065. <https://doi.org/10.1111/ctr.15065>.
- [59] Chierighini A, Potena L, Borgese L, Gibertoni D, Squarzoni D, Turello G, et al. Monitoring of cytomegalovirus (CMV)-specific cell-mediated immunity in heart transplant recipients: clinical utility of the Quantiferon-CMV assay for Management of Posttransplant CMV infection. J Clin Microbiol 2018;56. <https://doi.org/10.1128/JCM.01040-17>.
- [60] Palmer SM, Limaye AP, Banks M, Gallup D, Chapman J, Lawrence EC, et al. Extended valganciclovir prophylaxis to prevent cytomegalovirus after lung transplantation: a randomized, controlled trial. Ann Intern Med 2010;152:761–9. <https://doi.org/10.7326/0003-4819-152-12-201006150-00003>.
- [61] Jakobsch P, Zweytkow B, Kerschner H, Hoda AM, Keplinger M, Lang G, et al. Cytomegalovirus prevention in high-risk lung transplant recipients: comparison of 3- vs 12-month valganciclovir therapy. J Heart Lung Transplant 2009;28:670–5. <https://doi.org/10.1016/j.healun.2009.03.012>.
- [62] Finlen Copeland CA, Davis WA, Snyder LD, Banks M, Avery R, Davis RD, et al. Long-term efficacy and safety of 12 months of valganciclovir prophylaxis compared with 3 months after lung transplantation: a single-center, long-term follow-up analysis from a randomized, controlled cytomegalovirus prevention trial. J Heart Lung Transplant 2011;30:990–6. <https://doi.org/10.1016/j.healun.2011.02.017>.
- [63] Schoeppler KE, Lyu DM, Grazia TJ, Crossno JT, Vandervest KM, Zamora MR. Late-onset cytomegalovirus (CMV) in lung transplant recipients: can CMV Serostatus guide the duration of prophylaxis? Am J Transplant 2013;13:376–82. <https://doi.org/10.1111/j.1600-6143.2012.04339.x>.
- [64] Paraskeva M, Bailey M, Levvey BJ, Griffiths AP, Kotsimbos TC, Williams TP, et al. Cytomegalovirus replication within the lung allograft is associated with bronchiolitis obliterans syndrome. Am J Transplant 2011;11:2190–6. <https://doi.org/10.1111/j.1600-6143.2011.03663.x>.
- [65] Westall GP, Cristiano Y, Levvey BJ, Whitford H, Paraskeva MA, Paul E, et al. A randomized study of Quantiferon CMV-directed versus fixed-duration Valganciclovir prophylaxis to reduce late CMV after lung transplantation. Transplantation 2019;103:1005–13. <https://doi.org/10.1097/TP.0000000000002454>.
- [66] Gardiner BJ, Lee SJ, Robertson AN, Cristiano Y, Snell GI, Morrissey CO, et al. Real-world experience of Quantiferon-CMV directed prophylaxis in lung transplant recipients. J Heart Lung Transplant 2022;41:1258–67. <https://doi.org/10.1016/j.healun.2022.05.004>.
- [67] Monforte V, Lopez C, Santos F, Zurbano F, de la Torre M, Sole A, et al. A multicenter study of Valganciclovir prophylaxis up to day 120 in CMV-seropositive lung transplant recipients. Am J Transplant 2009;9:1134–41. <https://doi.org/10.1111/j.1600-6143.2009.02574.x>.
- [68] Monforte V, Sintes H, López-Gallo C, Delgado M, Santos F, Zurbano F, et al. Risk factors, survival, and impact of prophylaxis length in cytomegalovirus-seropositive lung transplant recipients: a prospective, observational, multicenter study. Transpl Infect Dis 2017;19. <https://doi.org/10.1111/tid.12694>.
- [69] Donadeu L, Revilla-López E, Jarque M, Crespo E, Torija A, Bravo C, et al. CMV-specific cell-mediated immunity predicts a high level of CMV replication after prophylaxis withdrawal in lung transplant recipients. J Infect Dis 2021;224:526–31. <https://doi.org/10.1093/infdis/jia727>.
- [70] Monforte V, Sintes H, Ussetti P, Castejón R, Pérez VL, Laporta R, et al. Assessment of Quantiferon®-CMV and Immuknow® assays in CMV-seropositive lung transplant recipients to stratify risk of CMV infection. Arch Bronconeumol 2022;58:614–7. <https://doi.org/10.1016/j.arbres.2021.10.002>.
- [71] Chen-Yoshikawa TF. ABO blood type incompatible lung transplantation. J Thorac Dis 2023;15:3437–42. <https://doi.org/10.21037/jtd-23-48>.
- [72] Ambrose T, Sharkey LM, Louis-Auguste J, Rutter CS, Duncan S, English S, et al. Cytomegalovirus infection and rates of antiviral resistance following intestinal and multivisceral transplantation. Transplant Proc 2016;48:492–6. <https://doi.org/10.1016/j.transproceed.2015.09.070>.
- [73] Nagai S, Mangus RS, Anderson E, Ekser B, Kubal CA, Fridell JA, et al. Cytomegalovirus infection after intestinal/multivisceral transplantation: a single-

- center experience with 210 cases. *Transplantation* 2016;100:451–60. <https://doi.org/10.1097/TP.00000000000000832>.
- [74] Knoll BM, Hammond SP, Koo S, Issa NC, Tullius SG, Baden LR, et al. Infections following facial composite tissue Allotransplantation—single center experience and review of the literature. *Am J Transplant* 2013;13:770–9. <https://doi.org/10.1111/ajt.12013>.
- [75] Fallatah SM, Marquez MA, Bazerbachi F, Schiff JR, Cattral MS, McGilvray ID, et al. Cytomegalovirus infection post-pancreas-kidney transplantation – results of antiviral prophylaxis in high-risk patients. *Clin Transpl* 2013;27:503–9. <https://doi.org/10.1111/ctr.12138>.
- [76] Luan FL, Stuckey LJ, Park JM, Kaul D, Cibrik D, Ojo A. Six-month prophylaxis is cost effective in transplant patients at high risk for cytomegalovirus infection. *J Am Soc Nephrol* 2009;20:2449–58. <https://doi.org/10.1681/ASN.2008111166>.
- [77] López-Medrano F, Rueda B, Lizasoain M, Juan RS, Folgueira D, Andrés A, et al. Preemptive therapy is not adequate for prevention of cytomegalovirus disease in pancreas-kidney transplant recipients. *Transpl Infect Dis* 2009;11:400–4. <https://doi.org/10.1111/j.1399-3062.2009.00416.x>.
- [78] Lisboa LF, Kumar D, Wilson LE, Humar A. Clinical utility of cytomegalovirus cell-mediated immunity in transplant recipients with cytomegalovirus viremia. *Transplantation* 2012;93:195–200. <https://doi.org/10.1097/TP.0b013e31823c1cd4>.
- [79] Andreani M, Albano L, Benzaken S, Cassuto E, Jeribi A, Caramella A, et al. Monitoring of CMV-specific cell-mediated immunity in kidney transplant recipients with a high risk of CMV disease (D+R-): a case series. *Transplant Proc* 2020;52:204–11. <https://doi.org/10.1016/j.transproced.2019.11.002>.
- [80] Doss KM, Kling CE, Heldman MR, Singh N, Wagener M, Rakita RM, et al. Real-world effectiveness of preemptive therapy (PET) for cytomegalovirus (CMV) disease prevention in CMV high-risk donor seropositive/recipient seronegative (D+R-) liver transplant recipients (LTxR). *Transpl Infect Dis* 2023;25. <https://doi.org/10.1111/tid.14015>.
- [81] Singh N, Winston DJ, Razonable RR, Lyon GM, Silveira FP, Wagener MM, et al. Cost-effectiveness of preemptive therapy versus prophylaxis in a randomized clinical trial for the prevention of cytomegalovirus disease in seronegative liver transplant recipients with seropositive donors. *Clin Infect Dis* 2021;73:e2739–45. <https://doi.org/10.1093/cid/ciaa1051>.
- [82] Paez-Vega A, Cantisan S, Vaquero JM, Vidal E, Luque-Pineda A, Lobo-Acosta MA, et al. Efficacy and safety of the combination of reduced duration prophylaxis followed by immuno-guided prophylaxis to prevent cytomegalovirus disease in lung transplant recipients (CYTOCOR STUDY): an open-label, randomised, non-inferiority clinical trial. *BMJ Open* 2019;9:e030648. <https://doi.org/10.1136/bmjopen-2019-030648>.
- [83] Le Page AK, Jager MM, Kotton CN, Simoons-Smit A, Rawlinson WD. International survey of cytomegalovirus Management in Solid Organ Transplantation after the publication of consensus guidelines. *Transplantation* 2013;95:1455–60. <https://doi.org/10.1097/TP.0b013e31828ee12e>.
- [84] Lee JH, Lee SW, Hwang SD, Song JH. Efficacy and safety according to the dose of Valganciclovir for cytomegalovirus prophylaxis in transplantation: network meta-analysis using recent data. *Transplant Proc* 2021;53:1945–50. <https://doi.org/10.1016/j.transproced.2021.05.006>.
- [85] Bixby AL, Fitzgerald L, Park JM, Kaul D, Tischer S. Comparison of standard versus low-dose valganciclovir regimens for cytomegalovirus prophylaxis in high-risk liver transplant recipients. *Transpl Infect Dis* 2021;23:e13713. <https://doi.org/10.1111/tid.13713>.
- [86] Stevens DR, Sawinski D, Blumberg E, Galanakis N, Bloom RD, Trofe-Clark J. Increased risk of breakthrough infection among cytomegalovirus donor-positive/recipient-negative kidney transplant recipients receiving lower-dose valganciclovir prophylaxis. *Transpl Infect Dis* 2015;17:163–73. <https://doi.org/10.1111/tid.12349>.
- [87] Tedesco-Silva H, Felipe C, Ferreira A, Cristelli M, Oliveira N, Sandes-Freitas T, et al. Reduced incidence of cytomegalovirus infection in kidney transplant recipients receiving Everolimus and reduced tacrolimus doses. *Am J Transplant* 2015;15:2655–64. <https://doi.org/10.1111/ajt.13327>.
- [88] Berger SP, Sommerer C, Witzke O, Tedesco H, Chadban S, Mulgaonkar S, et al. Two-year outcomes in de novo renal transplant recipients receiving everolimus-facilitated calcineurin inhibitor reduction regimen from the TRANSFORM study. *Am J Transplant* 2019;19:3018–34. <https://doi.org/10.1111/ajt.15480>.
- [89] Cervera C, Cofan F, Hernandez C, Soy D, Marcos MA, Sanclemente G, et al. Effect of mammalian target of rapamycin inhibitors on cytomegalovirus infection in kidney transplant recipients receiving polyclonal antilymphocyte globulins: a propensity score-matching analysis. *Transpl Int* 2016;29:1216–25. <https://doi.org/10.1111/tri.12848>.
- [90] Kaminski H, Kamar N, Thaunat O, Bouvier N, Caillard S, Garrigue I, et al. Incidence of cytomegalovirus infection in seropositive kidney transplant recipients treated with everolimus: a randomized, open-label, multicenter phase 4 trial. *Am J Transplant* 2022;22:1430–41. <https://doi.org/10.1111/ajt.16946>.
- [91] Viana LA, Cristelli MP, Basso G, Santos DW, Dantos MTC, Dreigas YC, et al. Conversion to mTOR inhibitor to reduce the incidence of cytomegalovirus recurrence in kidney transplant recipients receiving preemptive treatment: a prospective. Randomiz Trial *Transpl* 2023;107:1835–45. <https://doi.org/10.1097/TP.00000000000004559>.
- [92] Eisen HJ, Kobashigawa J, Starling RC, Pauly DF, Kfoury A, Ross H, et al. Everolimus versus mycophenolate Mofetil in heart transplantation: a randomized. Multicenter Trial *Am J Transpl* 2013;13:1203–16. <https://doi.org/10.1111/ajt.12181>.
- [93] Kobashigawa J, Ross H, Bara C, Delgado JF, Dengler T, Lehmkohl HB, et al. Everolimus is associated with a reduced incidence of cytomegalovirus infection following de novo cardiac transplantation. *Transpl Infect Dis* 2013;15:150–62. <https://doi.org/10.1111/tid.12007>.
- [94] Ueyama H, Kuno T, Takagi H, Alvarez P, Asleh R, Briasoulis A. Maintenance immunosuppression in heart transplantation: insights from network meta-analysis of various immunosuppression regimens. *Heart Fail Rev* 2022;27:869–77. <https://doi.org/10.1007/s10741-020-09967-3>.
- [95] Ghassemieh B, Ahya VN, Baz MA, Valentine VG, Arcasoy SM, Love RB, et al. Decreased incidence of cytomegalovirus infection with sirolimus in a post hoc randomized, multicenter study in lung transplantation. *J Heart Lung Transplant* 2013;32:701–6. <https://doi.org/10.1016/j.healun.2013.04.010>.
- [96] Kaminski H, Marselles G, Yared N, Nokin M-J, Pitard V, Zouine A, et al. mTOR inhibitors prevent CMV infection through the restoration of functional $\alpha\beta$ and $\gamma\delta$ T cells in kidney transplantation. *J Am Soc Nephrol* 2022;33:121–37. <https://doi.org/10.1681/ASN.2020121753>.
- [97] Florescu DF, Kalil AC, Qiu F, Schmidt CM, Sandkovsky U. What is the impact of hypogammaglobulinemia on the rate of infections and survival in solid organ transplantation? A meta-analysis. *Am J Transplant* 2013;13:2601–10. <https://doi.org/10.1111/ajt.12401>.
- [98] Immohr MB, Akhyari P, Böttger C, Mehdianni A, Dalyanoglu H, Westenfeld R, et al. Cytomegalovirus mismatch after heart transplantation: impact of antiviral prophylaxis and intravenous hyperimmune globulin. *Immun Inflamm Dis* 2021;9:1554–62. <https://doi.org/10.1002/iid.3.508>.
- [99] Zamora MR. Use of cytomegalovirus immune globulin and ganciclovir for the prevention of cytomegalovirus disease in lung transplantation. *Transpl Infect Dis* 2001;3:49–56. <https://doi.org/10.1034/j.1399-3062.2001.00010.x>.
- [100] Carbone J, Gallego A, Fernandez Yáñez J, Sousa I, Sarmiento E. Potential immunomodulatory role of specific Anticytomegalovirus intravenous immunoglobulin in heart recipients. *Transplant Proc* 2016;48:3027–9. <https://doi.org/10.1016/j.transproced.2016.07.039>.
- [101] Solidoro P, Patrucco F, Albera C, Pennazio V, Guerrera F, Boffini M, et al. Keep on tailoring CMV management in lung transplantation: 24 versus 12-month CMV hyperimmune globulins regimen effects in combined universal prophylaxis. *Panminerva Med* 2022;64:438–41. <https://doi.org/10.23736/S0031-0808.22.04736-X>.
- [102] Augusto J-F, Garnier A-S, Demiselle J, Langs V, Picquet J, Legall R, et al. Hypogammaglobulinemia and risk of severe infection in kidney transplant recipients. *Transpl Infect Dis* 2016;18:741–51. <https://doi.org/10.1111/tid.12593>.
- [103] Boddana P, Webb LH, Unsworth J, Brealey M, Bingham C, Harper SJ. Hypogammaglobulinemia and bronchiectasis in mycophenolate mofetil-treated renal transplant recipients: an emerging clinical phenomenon? *Clin Transpl* 2011;25:417–9. <https://doi.org/10.1111/j.1399-0012.2010.01255.x>.
- [104] Bonaros N, Mayer B, Schachner T, Laufer G, Kocher A. CMV-hyperimmune globulin for preventing cytomegalovirus infection and disease in solid organ transplant recipients: a meta-analysis. *Clin Transpl* 2008;22:89–97. <https://doi.org/10.1111/j.1399-0012.2007.00750.x>.
- [105] Yamani MH, Avery RK, Mawhorter SD, Young JB, Ratliff NB, Hobbs RE, et al. Hypogammaglobulinemia following cardiac transplantation: a link between rejection and infection. *J Heart Lung Transplant* 2001;20:425–30. [https://doi.org/10.1016/s1053-2498\(00\)00331-4](https://doi.org/10.1016/s1053-2498(00)00331-4).
- [106] Yamani MH, Avery R, Mawhorter S, Young JB, McNeill A, Cook DJ, et al. Hypogammaglobulinemia after heart transplantation: impact of pre-emptive use of immunoglobulin replacement (CytoGam) on infection and rejection outcomes. *Transpl Infect Dis* 2001;3:40–3. <https://doi.org/10.1034/j.1399-3062.2001.00008.x>.
- [107] Yamani MH, Avery R, Mawhorter SD, McNeill A, Cook D, Ratliff NB, et al. The impact of CytoGam on cardiac transplant recipients with moderate hypogammaglobulinemia: a randomized single-center study. *J Heart Lung Transplant* 2005;24:1766–9. <https://doi.org/10.1016/j.healun.2004.11.016>.
- [108] Sarmiento E, Diez P, Arraya M, Jaramillo M, Calahorra L, Fernandez-Yáñez J, et al. Early intravenous immunoglobulin replacement in hypogammaglobulinemic heart transplant recipients: results of a clinical trial. *Transpl Infect Dis* 2016;18:832–43. <https://doi.org/10.1111/tid.12610>.
- [109] Gardiner BJ, Nierenberg NE, Chow JK, Ruthazer R, Kent DM, Snydman DR. Absolute lymphocyte count: a predictor of recurrent cytomegalovirus disease in solid organ transplant recipients. *Clin Infect Dis* 2018;67:1395–402. <https://doi.org/10.1093/cid/ciy295>.
- [110] Schoeberl A-K, Zuckermann A, Kaider A, Aliaabadi-Zuckermann A, Uyanik-Uenal K, Laufer G, et al. Absolute lymphocyte count as a marker for cytomegalovirus infection after heart transplantation. *Transplantation* 2023;107:748–52. <https://doi.org/10.1097/TP.00000000000004360>.
- [111] Meesing A, Razonable RR. Absolute lymphocyte count thresholds: a simple, readily available tool to predict the risk of cytomegalovirus infection after transplantation. *Open Forum Infect Dis* 2018;5. <https://doi.org/10.1093/ofid/ofy230>.
- [112] Fernández-Ruiz M, Silva JT, López-Medrano F, Allende LM, San Juan R, Cambra F, et al. Post-transplant monitoring of NK cell counts as a simple approach to predict the occurrence of opportunistic infection in liver transplant recipients. *Transpl Infect Dis* 2016;18:552–65. <https://doi.org/10.1111/tid.12564>.
- [113] Fernández-Ruiz M, López-Medrano F, Allende LM, Andrés A, García-Reyne A, Lumbrieras C, et al. Kinetics of peripheral blood lymphocyte subpopulations predicts the occurrence of opportunistic infection after kidney transplantation. *Transpl Int* 2014;27:674–85. <https://doi.org/10.1111/tri.12321>.
- [114] Calarota SA, Chiesa A, De Silvestri A, Morosini M, Oggionni T, Marone P, et al. T-lymphocyte subsets in lung transplant recipients: association between nadir CD4

- T-cell count and viral infections after transplantation. *J Clin Virol* 2015;69:110–6. <https://doi.org/10.1016/j.jcv.2015.06.078>.
- [115] San-Juan R, Fernández-Ruiz M, Ruiz-Ruigómez M, López-Medrano F, Ruiz-Merlo T, Andrés A, et al. A new clinical and Immunovirological score for predicting the risk of late severe infection in solid organ transplant recipients: the CLIV score. *J Infect Dis* 2020;222:479–87. <https://doi.org/10.1093/infdis/jiaa090>.
- [116] Fernández-Ruiz M, Serón D, Alonso Á, Lora D, Hernández D, González E, et al. Derivation and external validation of the SIMPLICITY score as a simple immune-based risk score to predict infection in kidney transplant recipients. *Kidney Int* 2020;98:1031–43. <https://doi.org/10.1016/j.kint.2020.04.054>.
- [117] Sarmiento E, Navarro J, Fernandez-Yáñez J, Palomo J, Muñoz P, Carbone J. Evaluation of an immunological score to assess the risk of severe infection in heart recipients. *Transpl Infect Dis* 2014;16:802–12. <https://doi.org/10.1111/tid.12284>.
- [118] Doron S, Ruthazer R, Werner BG, Rabson A, Snydman DR. Hypogammaglobulinemia in liver transplant recipients: incidence, timing, risk factors, and outcomes. *Transplantation* 2006;81:697–703. <https://doi.org/10.1097/01.tp.0000180531.66518.9e>.
- [119] Ohsumi A, Chen F, Yamada T, Sato M, Aoyama A, Bando T, et al. Effect of hypogammaglobulinemia after lung transplantation: a single-institution study. *Eur J Cardiothorac Surg* 2014;45:e61–7. <https://doi.org/10.1093/ejcts/ezt583>.
- [120] Mozer-Glassberg Y, Shamir R, Steinberg R, Kadmon G, Har-Lev E, Mor E, et al. Hypogammaglobulinemia in the early period after liver transplantation in children. *Clin Transpl* 2013;27:E289–94. <https://doi.org/10.1111/ctr.12116>.
- [121] Fernández-Ruiz M, López-Medrano F, San-Juan R, Aguado JM. Post-transplant hypogammaglobulinemia and risk of infection after kidney transplantation: magnitude matters. *Transpl Infect Dis* 2017;19. <https://doi.org/10.1111/tid.12628>.
- [122] Sarmiento E, Jimenez M, di Natale M, Rodriguez-Ferrero M, Anaya F, Lopez-Hoyos M, et al. Secondary antibody deficiency is associated with development of infection in kidney transplantation: results of a multicenter study. *Transpl Infect Dis* 2021;23:e13494. <https://doi.org/10.1111/tid.13494>.
- [123] Fishman JA, Gans H. AST Infectious diseases Community of Practice. Pneumocystis jiroveci in solid organ transplantation: guidelines from the American Society of Transplantation Infectious Diseases Community of Practice. *Clin Transpl* 2019;33:e13587. <https://doi.org/10.1111/ctr.13587>.
- [124] Muñoz P, Giannella M, Alcalá L, Sarmiento E, Fernandez-Yáñez J, Palomo J, et al. *Clostridium difficile*-associated diarrhea in heart transplant recipients: is hypogammaglobulinemia the answer? *J Heart Lung Transplant* 2007;26:907–14. <https://doi.org/10.1016/j.healun.2007.07.010>.
- [125] Mawhorter S, Yamani MH. Hypogammaglobulinemia and infection risk in solid organ transplant recipients. *Curr Opin Organ Transplant* 2008;13:581–5. <https://doi.org/10.1097/MOT.0b013e3283186b6c>.
- [126] Sarmiento E, Jaramillo M, Calahorra L, Fernandez-Yáñez J, Gomez-Sánchez M, Crespo-Leiro MG, et al. Evaluation of humoral immunity profiles to identify heart recipients at risk for development of severe infections: a multicenter prospective study. *J Heart Lung Transplant* 2017;36:529–39. <https://doi.org/10.1016/j.healun.2016.10.004>.
- [127] Fujino T, Kumai Y, Nitta D, Holzhauser L, Nguyen A, Lourenco L, et al. Hypogammaglobulinemia following heart transplantation: prevalence, predictors, and clinical importance. *Clin Transpl* 2020;34:e14087. <https://doi.org/10.1111/ctr.14087>.
- [128] Lee H, Oh E-J. Laboratory diagnostic testing for cytomegalovirus infection in solid organ transplant patients. *Korean J Transpl* 2022;36:15–28. <https://doi.org/10.4285/kjt.22.0001>.
- [129] Fernández-Ruiz M, García-Ríos E, Redondo N, Rodríguez-Goncer I, Ruiz-Merlo T, Parra P, et al. Post-transplant dynamics and clinical significance of CMV-specific neutralizing antibodies in kidney transplant recipients treated with T-cell-depleting agents. *J Infect Dis* 2023. <https://doi.org/10.1093/infdis/jiad411>.
- [130] Zangerer N, Oxenius A. T cell immunity to cytomegalovirus infection. *Curr Opin Immunol* 2022;77:102185. <https://doi.org/10.1016/j.co.2022.102185>.
- [131] Wills MR, Carmichael AJ, Mynard K, Jin X, Weekes MP, Plachter B, et al. The human cytotoxic T-lymphocyte (CTL) response to cytomegalovirus is dominated by structural protein pp65: frequency, specificity, and T-cell receptor usage of pp65-specific CTL. *J Virol* 1996;70:7569–79. <https://doi.org/10.1128/JV.70.11.7569-7579.1996>.
- [132] Rogers R, Saharia K, Chandorkar A, Weiss ZF, Vieira K, Koo S, et al. Clinical experience with a novel assay measuring cytomegalovirus (CMV)-specific CD4+ and CD8+ T-cell immunity by flow cytometry and intracellular cytokine staining to predict clinically significant CMV events. *BMC Infect Dis* 2020;20:58. <https://doi.org/10.1186/s12879-020-4787-4>.
- [133] Gerna G, Lilleri D, Fornara C, Comolli G, Lozza L, Campana C, et al. Monitoring of human cytomegalovirus-specific CD4 and CD8 T-cell immunity in patients receiving solid organ transplantation. *Am J Transplant* 2006;6:2356–64. <https://doi.org/10.1111/j.1600-6143.2006.01488.x>.
- [134] Fernández-Ruiz M, Rodríguez-Goncer I, Parra P, Ruiz-Merlo T, Corbella L, López-Medrano F, et al. Monitoring of CMV-specific cell-mediated immunity with a commercial ELISA-based interferon- γ release assay in kidney transplant recipients treated with antithymocyte globulin. *Am J Transplant* 2020;20:2070–80. <https://doi.org/10.1111/ajt.15793>.
- [135] Abate D, Saldan A, Fiscon M, Cofano S, Paciolla A, Furian L, et al. Evaluation of cytomegalovirus (CMV)-specific T cell immune reconstitution revealed that baseline antiviral immunity, prophylaxis, or preemptive therapy but not antithymocyte globulin treatment contribute to CMV-specific T cell reconstitution in kidney transplant recipients. *J Infect Dis* 2010;202:585–94. <https://doi.org/10.1086/654931>.
- [136] Weseslindtner L, Kerschner H, Steinacher D, Nachbagauer R, Kundi M, Jaksch P, et al. Prospective analysis of human cytomegalovirus DNAemia and specific CD8+ T cell responses in lung transplant recipients. *Am J Transplant* 2012;12:2172–80. <https://doi.org/10.1111/j.1600-6143.2012.04076.x>.
- [137] Sund F, Lidehäll A-K, Claesson K, Foss A, Tötterman TH, Korsgren O, et al. CMV-specific T-cell immunity, viral load, and clinical outcome in seropositive renal transplant recipients: a pilot study. *Clin Transpl* 2010;24:401–9. <https://doi.org/10.1111/j.1399-0012.2009.00976.x>.
- [138] Gerna G, Lilleri D, Chiesa A, Zelini P, Furione M, Comolli G, et al. Virologic and immunologic monitoring of cytomegalovirus to guide preemptive therapy in solid-organ transplantation. *Am J Transplant* 2011;11:2463–71. <https://doi.org/10.1111/j.1600-6143.2011.03636.x>.
- [139] Egli A, Binet I, Binggeli S, Jäger C, Dumoulin A, Schaub S, et al. Cytomegalovirus-specific T-cell responses and viral replication in kidney transplant recipients. *J Transl Med* 2008;6:29. <https://doi.org/10.1186/1479-5876-6-29>.
- [140] Sester M, Sester U, Gärtner B, Heine G, Girndt M, Mueller-Lantzsch N, et al. Levels of virus-specific CD4 T cells correlate with cytomegalovirus control and predict virus-induced disease after renal transplantation. *Transplantation* 2001;71:1287–94. <https://doi.org/10.1097/0000000000105150-00018>.
- [141] Pongsakornkullachart K, Chayakulkeeree M, Vongwiwatana A, Kantakkamalakul W, Skulratanasak P, Phoompong P. Quantiferon-Cytomegalovirus assay for prediction of cytomegalovirus viremia in kidney transplant recipients: study from high cytomegalovirus Seroprevalence country. *Front Cell Infect Microbiol* 2022;12:893232. <https://doi.org/10.3389/fcimb.2022.893232>.
- [142] Pipeling MR, John ER, Orens JB, Lechtzin N, McDyer JF. Primary cytomegalovirus phosphoprotein 65-specific CD8+ T-cell responses and T-bet levels predict immune control during early chronic infection in lung transplant recipients. *J Infect Dis* 2011;204:1663–71. <https://doi.org/10.1093/infdis/jir624>.
- [143] Kumar D, Mian M, Singer L, Humar A. An interventional study using cell-mediated immunity to personalize therapy for cytomegalovirus infection after transplantation. *Am J Transplant* 2017;17:2468–73. <https://doi.org/10.1111/ajt.14347>.
- [144] Lúcia M, Crespo E, Melilli E, Cruzado JM, Luque S, Llaudó I, et al. Preformed frequencies of cytomegalovirus (CMV)-specific memory T and B cells identify protected CMV-sensitized individuals among seronegative kidney transplant recipients. *Clin Infect Dis* 2014;59:1537–45. <https://doi.org/10.1093/cid/ciu589>.
- [145] Hu X, Wang H-Y, Otero CE, Jenks JA, Permar SR. Lessons from acquired natural immunity and clinical trials to inform next-generation human cytomegalovirus vaccine development. *Annu Rev Virol* 2022;9:491–520. <https://doi.org/10.1146/annurev-virology-100220-010653>.
- [146] Griffiths PD, Stanton A, MacCarron E, Smith C, Osman M, Harber M, et al. Cytomegalovirus glycoprotein-B vaccine with MF59 adjuvant in transplant recipients: a phase 2 randomised placebo-controlled trial. *Lancet* 2011;377:1256–63. [https://doi.org/10.1016/S0140-6736\(11\)60136-0](https://doi.org/10.1016/S0140-6736(11)60136-0).
- [147] Rzymski P, Szuster-Ciesielska A, Dzieciątkowski T, Gwenzli W, Fal A. mRNA vaccines: the future of prevention of viral infections? *J Med Virol* 2023;95:e28572. <https://doi.org/10.1002/jmv.28572>.
- [148] Humar A, Siegal D, Moussa G, Kumar D. A prospective assessment of valganciclovir for the treatment of cytomegalovirus infection and disease in transplant recipients. *J Infect Dis* 2005;192:1154–7. <https://doi.org/10.1086/444398>.
- [149] Len O, Gavaldà J, Aguado JM, Borrell N, Cervera C, Cisneros JM, et al. Valganciclovir as treatment for cytomegalovirus disease in solid organ transplant recipients. *Clin Infect Dis* 2008;46:20–7. <https://doi.org/10.1086/523590>.
- [150] Caldés A, Gil-Vernet S, Armendariz Y, Colom H, Pou L, Niubó J, et al. Sequential treatment of cytomegalovirus infection or disease with a short course of intravenous ganciclovir followed by oral valganciclovir: efficacy, safety, and pharmacokinetics. *Transpl Infect Dis* 2009;12:204–12. <https://doi.org/10.1111/j.1399-3062.2009.00481.x>.
- [151] Asberg A, Humar A, Rollag H, Jardine AG, Mouas H, Pescovitz MD, et al. Oral valganciclovir is noninferior to intravenous ganciclovir for the treatment of cytomegalovirus disease in solid organ transplant recipients. *Am J Transplant* 2007;7:2106–13. <https://doi.org/10.1111/j.1600-6143.2007.01910.x>.
- [152] Luan FL, Chopra P, Park J, Norman S, Cibrik D, Ojo A. Efficacy of valganciclovir in the treatment of cytomegalovirus disease in kidney and pancreas transplant recipients. *Transplant Proc* 2006;38:3673–5. <https://doi.org/10.1016/j.transproceed.2006.10.105>.
- [153] Asberg A, Humar A, Jardine AG, Rollag H, Pescovitz MD, Mouas H, et al. Long-term outcomes of CMV disease treatment with valganciclovir versus IV ganciclovir in solid organ transplant recipients. *Am J Transplant* 2009;9:1205–13. <https://doi.org/10.1111/j.1600-6143.2009.02617.x>.
- [154] Czock D, Scholle C, Rasche FM, Schaarschmidt D, Keller F. Pharmacokinetics of valganciclovir and ganciclovir in renal impairment. *Clin Pharmacol Ther* 2002;72:142–50. <https://doi.org/10.1067/mcp.2002.126306>.
- [155] Lalagkas PN, Iliou J, Rigo R, Miaronis M, Fernández-Alarcon B, Bestard O, et al. Comparison of three renal function formulas for ganciclovir/Valganciclovir dose individualization in CMV-infected solid organ transplantation patients using a population approach. *Clin Pharmacokinet* 2023;62:861–80. <https://doi.org/10.1007/s40262-023-1237-3>.
- [156] Linder KA, Kovacs C, Mullane KM, Wolfe C, Clark NM, La Hoz RM, et al. Letermovir treatment of cytomegalovirus infection or disease in solid organ and

- hematopoietic cell transplant recipients. *Transpl Infect Dis* 2021;23:e13687. <https://doi.org/10.1111/tid.13687>.
- [157] Marty FM, Winston DJ, Chemaly RF, Mullane KM, Shore TB, Papanicolaou GA, et al. A randomized, double-blind, placebo-controlled phase 3 trial of Oral Brincidofovir for cytomegalovirus prophylaxis in allogeneic hematopoietic cell transplantation. *Biol Blood Marrow Transplant* 2019;25:369–81. <https://doi.org/10.1016/j.bbmt.2018.09.038>.
- [158] Faure E, Galperine T, Cannesson O, Alain S, Gneimmi V, Goeminne C, et al. Case report: Brincidofovir-induced reversible severe acute kidney injury in 2 solid-organ transplant for treatment of cytomegalovirus infection. *Medicine* 2016;95: e5226. <https://doi.org/10.1097/MD.00000000000005226>.
- [159] Frange P, Leruez-Ville M. Maribavir, brincidofovir and letermovir: efficacy and safety of new antiviral drugs for treating cytomegalovirus infections. *Med Mal Infect* 2018;48:495–502. <https://doi.org/10.1016/j.medmal.2018.03.006>.
- [160] Kerr ER, Bidanset DJ, Hartline CB, Yan Z, Zemlicka J, Quenelle DC. Oral activity of a methylenecyclopropane analog, cyclopavopavir, in animal models for cytomegalovirus infections. *Antimicrob Agents Chemother* 2004;48:4745–53. <https://doi.org/10.1128/AAC.48.12.4745-4753.2004>.
- [161] Toth K, Hussein ITM, Tollefson AE, Ying B, Spencer JE, Eagar J, et al. Filociclovir is a potent *in vitro* and *in vivo* inhibitor of human adenoviruses. *Antimicrob Agents Chemother* 2020;64. <https://doi.org/10.1128/AAC.01299-20>.
- [162] Roushaw NG, Hurwitz SJ, Hart M, Beck A, Anderson EJ, Deye G, et al. Phase Ib trial to evaluate the safety and pharmacokinetics of multiple ascending doses of Filociclovir (MBX-400, Cyclopavopavir) in healthy volunteers. *Antimicrob Agents Chemother* 2019;63. <https://doi.org/10.1128/AAC.00717-19>.
- [163] Nuévalos M, García-Ríos E, Mancebo FJ, Martín-Martín C, Pérez-Romero P. Novel monoclonal antibody-based therapies: implications for the treatment and prevention of HCMV disease. *Trends Microbiol* 2023;31:480–97. <https://doi.org/10.1016/j.tim.2022.12.003>.
- [164] Shanahan A, Malani PN, Kaul DR. Relapsing cytomegalovirus infection in solid organ transplant recipients. *Transpl Infect Dis* 2009;11:513–8. <https://doi.org/10.1111/j.1399-3062.2009.00443.x>.
- [165] Eid AJ, Arthurs SK, Deziel PJ, Wilhelm MP, Razonable RR. Clinical predictors of relapse after treatment of primary gastrointestinal cytomegalovirus disease in solid organ transplant recipients. *Am J Transplant* 2010;10:157–61. <https://doi.org/10.1111/j.1600-6143.2009.02861.x>.
- [166] Natori Y, Alghamdi A, Tazari M, Miller V, Husain S, Komatsu T, et al. Use of viral load as a surrogate marker in clinical studies of cytomegalovirus in solid organ transplantation: a systematic review and meta-analysis. *Clin Infect Dis* 2018;66: 617–31. <https://doi.org/10.1093/cid/cix793>.
- [167] Gardiner BJ, Chow JK, Price LL, Nierenberg NE, Kent DM, Snydman DR. Role of secondary prophylaxis with Valganciclovir in the prevention of recurrent cytomegalovirus disease in solid organ transplant recipients. *Clin Infect Dis* 2017; 65:2000–7. <https://doi.org/10.1093/cid/cix696>.
- [168] Sullivan T, Brodzinski A, Patel G, Uprikar S. The role of secondary cytomegalovirus prophylaxis for kidney and liver transplant recipients. *Transplantation* 2015;99:855–9. <https://doi.org/10.1097/TP.0000000000000386>.
- [169] Natori Y, Humar A, Husain S, Rotstein C, Renner E, Singer L, et al. Recurrence of CMV infection and the effect of prolonged antivirals in organ transplant recipients. *Transplantation* 2017;101:1449–54. <https://doi.org/10.1097/TP.0000000000001338>.
- [170] Gardiner BJ, Nierenberg NE, Chow JK, Ruthazer R, Kent DM, Snydman DR. Absolute lymphocyte count: a predictor of recurrent cytomegalovirus disease in solid organ transplant recipients. *Clin Infect Dis* 2018;67:1395–402. <https://doi.org/10.1093/cid/ciy295>.
- [171] Helanterä I, Lautenschlager I, Koskinen P. The risk of cytomegalovirus recurrence after kidney transplantation. *Transpl Int* 2011;24:1170–8. <https://doi.org/10.1111/j.1432-2277.2011.01321.x>.
- [172] Ljungman P, Chemaly RF, Khawaya F, Alain S, Avery R, Badshah C, et al. Consensus Definitions of Cytomegalovirus (CMV) Infection and Disease in Transplant Patients Including Resistant and Refractory CMV for Use in Clinical Trials: 2024 Update From the Transplant Associated Virus Infections Forum. *Clin Infect Dis* 2024. <https://doi.org/10.1093/cid/ciae321>.
- [173] Kleiboeker SB. Prevalence of cytomegalovirus antiviral drug resistance in transplant recipients. *Antiviral Res* 2023;215:105623. <https://doi.org/10.1016/j.antiviral.2023.105623>.
- [174] Grossi PA, Kamar N, Saliba F, Baldanti F, Aguado JM, Gottlieb J, et al. Cytomegalovirus Management in Solid Organ Transplant Recipients: a pre-COVID-19 survey from the working Group of the European Society for organ transplantation. *Transpl Int* 2022;35. <https://doi.org/10.3389/ti.2022.10332>.
- [175] Lurain NS, Chou S. Antiviral drug resistance of human cytomegalovirus. *Clin Microbiol Rev* 2010;23:689–712. <https://doi.org/10.1128/CMR.00009-10>.
- [176] Fisher CE, Knudsen JL, Lease ED, Jerome KR, Rakita RM, Boeckh M, et al. Risk factors and outcomes of ganciclovir-resistant cytomegalovirus infection in solid organ transplant recipients. *Clin Infect Dis* 2017;65:57–63. <https://doi.org/10.1093/cid/cix259>.
- [177] Tamzali Y, Pourcher V, Azoyan L, Ouali N, Barrou B, Conti F, et al. Factors associated with genotypic resistance and outcome among solid organ transplant recipients with refractory cytomegalovirus infection. *Transpl Int* 2023;36. <https://doi.org/10.3389/ti.2023.11295>.
- [178] Khawaja F, Spallone A, Kotton CN, Chemaly RF. Cytomegalovirus infection in transplant recipients: newly approved additions to our armamentarium. *Clin Microbiol Infect* 2023;29:44–50. <https://doi.org/10.1016/j.cmi.2022.07.001>.
- [179] Chou S. Phenotypic diversity of cytomegalovirus DNA polymerase gene variants observed after antiviral therapy. *J Clin Virol* 2011;50:287–91. <https://doi.org/10.1016/j.jcv.2011.01.004>.
- [180] Piret J, Boivin G. Clinical development of letermovir and maribavir: overview of human cytomegalovirus drug resistance. *Antiviral Res* 2019;163:91–105. <https://doi.org/10.1016/j.antiviral.2019.01.011>.
- [181] Hofmann E, Sidler D, Dahdal S, Bittel P, Suter-Riniker F, Manuel O, et al. Emergence of letermovir resistance in solid organ transplant recipients with ganciclovir resistant cytomegalovirus infection: a case series and review of the literature. *Transpl Infect Dis* 2021;23:e13515. <https://doi.org/10.1111/tid.13515>.
- [182] Chou S. Advances in the genotypic diagnosis of cytomegalovirus antiviral drug resistance. *Antiviral Res* 2020;176:104711. <https://doi.org/10.1016/j.antiviral.2020.104711>.
- [183] López-Aladid R, Guiu A, Sanclemente G, López-Medrano F, Cofán F, Mosquera MM, et al. Detection of cytomegalovirus drug resistance mutations in solid organ transplant recipients with suspected resistance. *J Clin Virol* 2017;90: 57–63. <https://doi.org/10.1016/j.jcv.2017.03.014>.
- [184] Kotton CN, Kamar N. New insights on CMV Management in Solid Organ Transplant Patients: prevention, treatment, and Management of Resistant/ refractory disease. *Infect Dis Ther* 2023;12:333–42. <https://doi.org/10.1007/s40121-022-00746-1>.
- [185] Jo H, Kwon DE, Han SH, Min SY, Hong Y-M, Lim BJ, et al. De novo genotypic heterogeneity in the UL56 region in cytomegalovirus-infected tissues: implications for primary Letermovir resistance. *J Infect Dis* 2020;221:1480–7. <https://doi.org/10.1093/infdis/jiz642>.
- [186] Sahoo MK, Leterova MI, Yamamoto F, Waggoner JJ, Chou S, Holmes SP, et al. Detection of cytomegalovirus drug resistance mutations by next-generation sequencing. *J Clin Microbiol* 2013;51:3700–10. <https://doi.org/10.1128/JCM.01605-13>.
- [187] Santos Bravo M, Plaust N, Sánchez-Palomino S, Rodríguez C, Navarro Gabriel M, Mosquera MM, et al. Genotypic and phenotypic study of antiviral resistance mutations in refractory cytomegalovirus infection. *J Infect Dis* 2022;226: 1528–36. <https://doi.org/10.1093/infdis/jiac349>.
- [188] Chou S, Boivin G, Ives J, Elston R. Phenotypic evaluation of previously uncharacterized cytomegalovirus DNA polymerase sequence variants detected in a valganciclovir treatment trial. *J Infect Dis* 2014;209:1219–26. <https://doi.org/10.1093/infdis/jit654>.
- [189] Gracia-Ahufinger I, Gutiérrez-Aroca J, Cordero E, Vidal E, Cantisán S, del Castillo D, et al. Use of high-dose ganciclovir for the treatment of cytomegalovirus replication in solid organ transplant patients with ganciclovir resistance-inducing mutations. *Transplantation* 2013;95:1015–20. <https://doi.org/10.1097/TP.0b013e31828555ac>.
- [190] Avery RK, Arav-Boger R, Marr KA, Kraus E, Shoham S, Lees L, et al. Outcomes in transplant recipients treated with Foscarnet for ganciclovir-resistant or refractory cytomegalovirus infection. *Transplantation* 2016;100:e74–80. <https://doi.org/10.1097/TP.0000000000001418>.
- [191] Eid AJ, Arthurs SK, Deziel PJ, Wilhelm MP, Razonable RR. Emergence of drug-resistant cytomegalovirus in the era of valganciclovir prophylaxis: therapeutic implications and outcomes. *Clin Transpl* 2008;22:162–70. <https://doi.org/10.1111/j.1399-0012.2007.00761.x>.
- [192] Papanicolaou GA, Silveira FP, Langston AA, Pereira MR, Avery RK, Uknis M, et al. Maribavir for refractory or resistant cytomegalovirus infections in hematopoietic-cell or solid-organ transplant recipients: a randomized, dose-ranging, double-blind, phase 2 study. *Clin Infect Dis* 2019;68:1255–64. <https://doi.org/10.1093/cid/ciy706>.
- [193] Avery RK, Alain S, Alexander BD, Blumberg EA, Chemaly RF, Cordonnier C, et al. Maribavir for refractory cytomegalovirus infections with or without resistance post-transplant: results from a phase 3 randomized clinical trial. *Clin Infect Dis* 2022;75:690–701. <https://doi.org/10.1093/cid/ciab988>.
- [194] Chou S, Song K, Wu J, Bo T, Crumpacker C. Drug resistance mutations and associated phenotypes detected in clinical trials of Maribavir for treatment of cytomegalovirus infection. *J Infect Dis* 2022;226:576–84. <https://doi.org/10.1093/infdis/jiaa462>.
- [195] Carbone J, Palomo J, Fernandez-Yáñez J, Sarmiento E. Subcutaneous immunoglobulin replacement therapy in a heart transplant recipient with severe recurrent infections. *Heart Lung Vessel* 2015;7:256–9.
- [196] Carbone J, Sarmiento E, Del Pozo N, Rodríguez-Molina JJ, Navarro J, Fernández-Yáñez J, et al. Restoration of humoral immunity after intravenous immunoglobulin replacement therapy in heart recipients with post-transplant antibody deficiency and severe infections. *Clin Transpl* 2012;26:E277–83. <https://doi.org/10.1111/j.1399-0012.2012.01653.x>.
- [197] Mylonakis E, Kallas WM, Fishman JA. Combination antiviral therapy for ganciclovir-resistant cytomegalovirus infection in solid-organ transplant recipients. *Clin Infect Dis* 2002;34:1337–41. <https://doi.org/10.1086/340101>.
- [198] Sarmiento E, Fernández-Yáñez J, Muñoz P, Palomo J, Rodríguez-Molina JJ, Bermejo J, et al. Hypogammaglobulinemia after heart transplantation: use of intravenous immunoglobulin replacement therapy in relapsing CMV disease. *Int Immunopharmacol* 2005;5:97–101. <https://doi.org/10.1016/j.intimp.2004.09.006>.
- [199] Santhanakrishnan K, Yonan N, Iyer K, Callan P, Al-Aloul M, Venkateswaran R. Management of ganciclovir resistance cytomegalovirus infection with CMV hyperimmune globulin and leflunomide in seven cardiothoracic transplant recipients and literature review. *Transpl Infect Dis* 2022;24:e13733. <https://doi.org/10.1111/tid.13733>.

- [200] Heslop HE, Slobod KS, Pule MA, Hale GA, Rousseau A, Smith CA, et al. Long-term outcome of EBV-specific T-cell infusions to prevent or treat EBV-related lymphoproliferative disease in transplant recipients. *Blood* 2010;115:925–35. <https://doi.org/10.1182/blood-2009-08-239186>.
- [201] Blyth E, Clancy L, Simms R, Ma CKK, Burgess J, Deo S, et al. Donor-derived CMV-specific T cells reduce the requirement for CMV-directed pharmacotherapy after allogeneic stem cell transplantation. *Blood* 2013;121:3745–58. <https://doi.org/10.1182/blood-2012-08-448977>.
- [202] Grau-Vorster M, López-Montañés M, Cantó E, Vives J, Oliver-Vila I, Barba P, et al. Characterization of a cytomegalovirus-specific T lymphocyte product obtained through a rapid and scalable production process for use in adoptive immunotherapy. *Front Immunol* 2020;11:271. <https://doi.org/10.3389/fimmu.2020.00271>.
- [203] Fuji S, Einsele H, Kapp M. Cytomegalovirus disease in hematopoietic stem cell transplant patients: current and future therapeutic options. *Curr Opin Infect Dis* 2017;30:372–6. <https://doi.org/10.1097/QCO.0000000000000375>.
- [204] Tzannou I, Papadopoulou A, Naik S, Leung K, Martinez CA, Ramos CA, et al. Off-the-shelf virus-specific T cells to treat BK virus, human herpesvirus 6, cytomegalovirus, Epstein-Barr virus, and adenovirus infections after allogeneic hematopoietic stem-cell transplantation. *J Clin Oncol* 2017;35:3547–57. <https://doi.org/10.1200/JCO.2017.73.0655>.
- [205] Ouellette CP. Adoptive immunotherapy for prophylaxis and treatment of cytomegalovirus infection. *Viruses* 2022;14:2370. <https://doi.org/10.3390/v14112370>.
- [206] Haque T, Wilkie GM, Taylor C, Amlot PL, Murad P, Iley A, et al. Treatment of Epstein-Barr-virus-positive post-transplantation lymphoproliferative disease with partly HLA-matched allogeneic cytotoxic T cells. *Lancet* 2002;360:436–42. [https://doi.org/10.1016/S0140-6736\(02\)09672-1](https://doi.org/10.1016/S0140-6736(02)09672-1).
- [207] Haque T, Wilkie GM, Jones MM, Higgins CD, Urquhart G, Wingate P, et al. Allogeneic cytotoxic T-cell therapy for EBV-positive posttransplantation lymphoproliferative disease: results of a phase 2 multicenter clinical trial. *Blood* 2007;110:1123–31. <https://doi.org/10.1182/blood-2006-12-063008>.
- [208] Macesic N, Langsford D, Nicholls K, Hughes P, Gottlieb DJ, Clancy L, et al. Adoptive T cell immunotherapy for treatment of ganciclovir-resistant cytomegalovirus disease in a renal transplant recipient. *Am J Transplant* 2015;15: 827–32. <https://doi.org/10.1111/ajt.13023>.
- [209] Holmes-Liew C-L, Holmes M, Beagley L, Hopkins P, Chambers D, Smith C, et al. Adoptive T-cell immunotherapy for ganciclovir-resistant CMV disease after lung transplantation. *Clin Transl Immunol* 2015;4:e35. <https://doi.org/10.1038/cti.2015.5>.
- [210] Gupta MP, Koenig LR, Doubrovina E, Hasan A, Dahi PB, O'Reilly RJ, et al. Ocular outcomes after treatment of cytomegalovirus retinitis using adoptive immunotherapy with cytomegalovirus-specific cytotoxic T lymphocytes. *Ophthalmol Retina* 2021;5:838–49. <https://doi.org/10.1016/j.oret.2021.04.009>.
- [211] Smith C, Beagley L, Rehan S, Neller MA, Crooks P, Solomon M, et al. Autologous adoptive T-cell therapy for recurrent or drug-resistant cytomegalovirus complications in solid organ transplant recipients: a single-arm open-label phase I clinical trial. *Clin Infect Dis* 2019;68:632–40. <https://doi.org/10.1093/cid/ciy549>.
- [212] Amini L, Wagner DL, Rössler U, Zarrinrad G, Wagner LF, Vollmer T, et al. CRISPR-Cas9-edited tacrolimus-resistant antiviral T cells for advanced adoptive immunotherapy in transplant recipients. *Mol Ther* 2021;29:32–46. <https://doi.org/10.1016/j.ymthe.2020.09.011>.