



Clinical practice guidelines for the treatment of Ewing sarcoma (Spanish Sarcoma Research Group-GEIS)

Cristina Mata Fernández¹ · Ana Sebio² · Javier Orcajo Rincón³ · Javier Martín Broto⁴ · Antonio Martín Benlloch⁵ · David Marcilla Plaza⁶ · Antonio López Pousa² · Isidro Gracia Alegría⁷ · Martina Giuppi⁸ · Erica Collado Ballesteros⁹ · Daniel Bernabeu¹⁰ · Enrique de Alava¹¹ · Claudia Valverde Morales¹²

Received: 26 March 2024 / Accepted: 3 July 2024 / Published online: 19 August 2024
© The Author(s) 2024

Abstract

Ewing sarcoma is a small round-cell sarcoma characterized by gene fusion involving EWSR1 (or another TET family protein like FUS) and an ETS family transcription factor. The estimated incidence of this rare bone tumor, which occurs most frequently in adolescents and young adults, is 0.3 per 100,000/year. Although only 25% of patients with Ewing sarcoma are diagnosed with metastatic disease, historical series show that this is a systemic disease. Patient management requires multimodal therapies—including intensive chemotherapy—in addition to local treatments (surgery and/or radiotherapy). In the recurrent/refractory disease setting, different approaches involving systemic treatments and local therapies are also recommended as well as patient inclusion in clinical trials whenever possible. Because of the complexity of Ewing sarcoma diagnosis and treatment, it should be carried out in specialized centers and treatment plans should be designed upfront by a multidisciplinary tumor board. These guidelines provide recommendations for diagnosis, staging, and multimodal treatment of Ewing sarcoma.

Keywords Ewing sarcoma · Small round cell sarcomas · “Ewing like” sarcoma · Malignant bone tumours · EWSR1

Methodology

These guidelines were developed by a multidisciplinary panel of specialists from the different fields involved in the diagnosis and treatment of Ewing sarcoma (ES) in pediatric,

adolescent, and adult patients. A bibliographic search was conducted for published articles in the PubMed database and also common guidelines were consulted, including those of the National Comprehensive Cancer Network (NCCN) [1], European Society for Medical Oncology (ESMO)/European

✉ Cristina Mata Fernández
cristina.mata@salud.madrid.org

¹ Pediatric and Adolescent Oncohaematology Unit, Hospital Materno-Infantil Gregorio, Marañón Madrid, Spain

² Medical Oncology Department, Hospital Sant Pau, Barcelona, Spain

³ Nuclear Medicine Department, Hospital General Universitario Gregorio Marañón, Madrid, Spain

⁴ Medical Oncology Department, Fundación Jimenez Diaz University Hospital, University Hospital General de Villalba, and Instituto de Investigacion Sanitaria Fundacion Jimenez Diaz (IIS/FJD; UAM), Madrid, Spain

⁵ Section Spine Unit. Orthopaedic and Traumatology Department, Dr. Peset University Hospital, Valencia, Spain

⁶ Department of Pathology, Hospital Universitario Virgen del Rocío, Sevilla, Spain

⁷ Orthopaedic Oncology Unit, Orthopaedic and Traumatology Department, Hospital Sant Pau, Barcelona, Spain

⁸ Sarcoma patient, Madrid, Spain

⁹ Radiation Oncologist, Hospital Universitari i Politècnic La Fe, Valencia, Spain

¹⁰ Chief of Musculo-skeletal Radiology Section, Radiodiagnosis Service Hospital General Universitario La Paz, Madrid, Spain

¹¹ Department of Normal and Pathological Cytology and Histology, School of Medicine, University of Seville, Institute of Biomedicine of Sevilla, IBiS/Virgen del Rocío University Hospital /CSIC/University of Sevilla/CIBERONC, Seville, Spain

¹² Medical Oncology Department, Hospital Universitari Vall d'Hebrón, Barcelona, Spain

Reference Network for Cancers (EURACAN) [2]. In several telematic consensus meetings, experts gave presentations for subsequent discussion by the multidisciplinary panel, which adopted the Infectious Disease Society of America levels of evidence/grades of recommendation (Table 1) [3].

Introduction and epidemiology

These ES guidelines also cover the former entities known as Askin tumor and peripheral primitive neuroectodermal tumor (PNET). The ES group of small round-cell sarcomas are characterized by recurrent balanced translocations involving EWSR1 (or other TET family proteins like FUS) and an ETS family transcription factor. “Ewing-like” sarcomas or tumors lacking these characteristic fusions (including EWSR1/FUS non-ETS family members and CIC rearranged and BCOR—rearranged sarcomas), although currently similarly treated, have a different natural history and their management may differ, especially for CIC-rearranged tumors.

ES is the third most frequent malignant bone tumor in humans and the second most frequent after osteosarcoma in children. Median age at diagnosis is 15 years, and the male sex predominates (1.5:1) [4]. ES is practically non-existent in African and Afro-American populations, while the estimated incidence is 0.3 per 100,000/year in Caucasian adults.

Although the primary tumor is usually located in the bone (diaphysis of long bones like the femur, tibia or humerus pelvis, chest wall and spine), up to 10–20% of cases are extra-skeletal.

ES is a systemic disease. Although only 25% of patients have obvious metastases at diagnosis, in the past, 80–90% of

patients with apparently localized disease died when treatment was reduced to a local approach. Disease dissemination is predominantly hematogenous, being lung, bones, and bone marrow as the most common metastatic sites.

Clinical and radiological diagnosis

Persistent, localized bone pain (96% of cases), especially if asymmetric and causing the patient to wake at night, or a prolonged and unjustified limp, should raise suspicions and warrant further evaluation (evidence level grade: IV,A).

In conventional radiography, most ES appear as aggressive osteolytic lesions with a permeative pattern, most cases mixed lytic-sclerotic, or purely lytic. Around half of the lesions show complex periosteal reactions, such as multilaminated “onion-skin”, “sunburst,” and Codman’s triangle. Also common is a non-calcified soft tissue mass extending around the bone and displacing adjacent fatty lines.

Computed tomography (CT) can help detect bone destruction, periosteal reaction, and soft tissue mass in anatomically complex regions like the pelvis, spine, and ankle. Sclerotic bone may be present in up to 40% of cases, mostly related to osteonecrosis in central medullar areas or in flat bones.

Magnetic resonance imaging (MRI) is very sensitive in detecting bone and soft tissue tumor components and in defining tumor relationship to nearby nerves, vessels and fascial planes) [5]. Because skip lesions can be expected, MRI protocols must include proximal and distal joints to the affected bone (V,A). ES shows a non-specific signal pattern: hypointense in T1-weighted imaging (WI) and slightly hyperintense

Table 1 Levels of evidence and grades of recommendation (adapted from the Infectious Diseases Society of America-United States Public Health Service Grading System)

Levels of evidence	
I	Evidence from at least one large randomized, controlled trial of good methodological quality (low potential for bias) or meta-analyses of well conducted randomized trials without heterogeneity
II	Small randomized trials or large randomized trials with a suspicion of bias (lower methodological quality) or meta-analyses of such trials or trials with demonstrated heterogeneity
III	Prospective cohort studies
IV	Retrospective cohort studies or case–control studies
V	Studies without control group, case reports, expert opinions
Grades of recommendation	
A	Strong evidence for efficacy with a substantial clinical benefit, strongly recommended
B	Strong or moderate evidence for efficacy but with a limited clinical benefit, generally recommended
C	Insufficient evidence for efficacy or benefit does not outweigh the risk or the disadvantages (adverse events, costs,...) optional
D	Moderate evidence against efficacy or for adverse outcome, generally not recommended
E	Strong evidence against efficacy or for adverse outcome, never recommended

in T2-WI. The soft-tissue mass is almost always connected through cortical breaks to the bone lesion; however, because of rapid infiltrative spread, some cases show no cortical disruption, and the soft component appears to be wrapped around the bone (the wraparound sign). Contrast enhancement (CE), in both CT and MRI, is usually strong and diffuse. Diffusion-MRI showing a clear restriction pattern with very low apparent diffusion coefficient (ADC) values can help differentiate edema or necrosis from tumor cellularity [6]. The imaging appearance of extra-skeletal ES is non-specific, as happens with any other soft-tissue sarcoma.

Differential diagnosis should include osteomyelitis, osteosarcoma, lymphoma, leukemia, Langerhans cell histiocytosis, and metastatic neuroblastoma.

Percutaneous core needle biopsy using the coaxial trocar system, is the standard outpatient procedure, although an incisional biopsy may be necessary in a small minority of patients [7] (III,A). Biopsy must be directed to selected areas with cellularity or metabolic activity, as detected by CE-MRI, Doppler ultrasound (US), diffusion-MRI or positron emission tomography PET/CT [8] (IV,A). It is recommended to obtain multiple samples from the same biopsy. In most referral centers, surgeons and musculoskeletal radiologists together evaluate the optimal biopsy path [9] to be removed during the final surgery (V,C).

Pathology and molecular biology

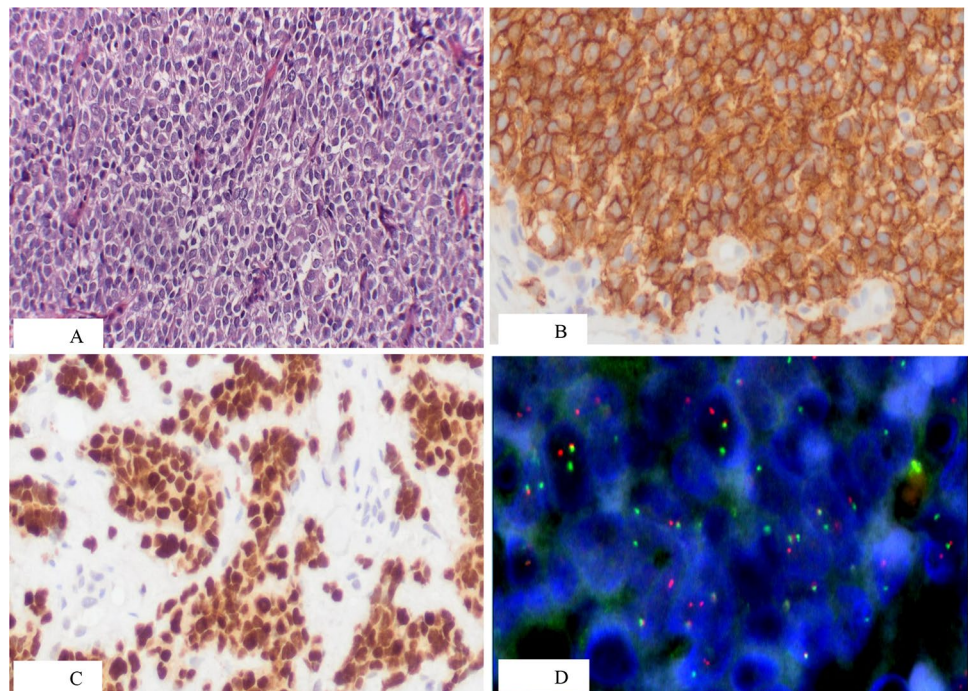
Although gross examination of untreated ES specimens is now uncommon because of the standard use of neoadjuvant treatment, a cut surface is soft, and grey-white color, and frequently includes areas of hemorrhage and necrosis [10].

Histologically, most cases are made up of solid sheets of uniform small round cells with round nuclei containing finely stippled chromatin, inconspicuous nucleoli, and scant clear or eosinophilic cytoplasm, with ill-defined cell borders [11] (Fig. 1A). Occasionally (in atypical ES), the tumor cells are larger, with more conspicuous nucleoli and irregular nuclear contours [12]. A minority of cases provide evidence of neuroectodermal differentiation with rosette formation.

Immunohistochemically, neoplastic cells show a membranous pattern of intense and diffuse CD99 expression (Fig. 1B) and NKX2.2 nuclear immunoreactivity (Fig. 1C). However, it is important to note that CD99 and NKX2.2 are not entirely specific for ES [13]. Nuclear FLI1 and ERG are often expressed in cases with the corresponding gene fusions [14].

Genetic confirmation is required for ES diagnosis. The most common ES translocations are t(11;22) (q24;q12), and. t(21;22) (q22;q12), resulting in the *EWSR1-FLI1* and *EWSR1-ERG* fusion transcripts and proteins (~85% and ~10% of cases, respectively); The remaining cases have alternative translocations that join *EWSR1* or *FUS* (which, along with *TAF15*, form the FET family) to other

Fig. 1 Histologic and molecular pattern of Ewing sarcoma. **A** Ewing Sarcoma. Conventional appearance with uniform small round cell and scanty cytoplasm. **B** Ewing Sarcoma. Strong and diffuse membranous reactivity for CD99. **C** Ewing Sarcoma. Intense and diffuse nuclear expression of NKX2.2. **D** Ewing Sarcoma. FISH analysis showing rearrangement of *EWSR1* (courtesy Dr. M. Biscuola)



ETS family members. All cases of ES harbor a FET-ETS fusion. The most commonly used diagnostic approach is fluorescence in-situ hybridization (FISH) using an EWSR1 break-apart probe (Fig. 1D). Molecular techniques, such as reverse-transcriptase polymerase chain reaction (RT-PCR) which detects both translocation partners, are more specific for ES, while next-generation sequencing (NGS) techniques appear to be moving to the forefront of molecular diagnostic approaches.

The differential diagnosis includes tumors with small round-cell morphology, mainly round cell sarcoma with EWSR1 non-ETS fusion, CIC-rearranged sarcoma, and sarcoma with a BCOR genetic alteration.

Described as a distinct subset of lesions carrying the same fusions, predominantly in the head and neck region, is adamantinoma-like ES, which often expresses pan-cytokeratin and markers of squamous differentiation. However, the relationship of this tumor type to classic ES is uncertain [15].

Definitive ES diagnosis should be performed (or reviewed) at a reference sarcoma center and based on sufficient biopsy material for conventional histology, immunohistochemistry, and molecular pathology [16] (I,A).

Staging and risk assessment

Staging is critical for treatment strategy planning and for prognostic assessment.

Initial laboratory investigations include blood count, serum biochemistry, and lactate dehydrogenase (LDH) testing.

CT scans show the highest sensitivity for lung metastasis detection. Extrapulmonary metastases may be ruled out by whole-body bone scans with ^{99m}Tc-hydroxydiphosphonate (HDP) bone scintigraphy (BS) or ¹⁸F-fluorodeoxyglucose FDG-PET/CT. Overall, PET/CT shows greater sensitivity (96%) and specificity (92%) for bone lytic metastasis detection [17] and it works especially well for nodal and soft tissue lesions.

Whole-body MRI may be an alternative to BS and PET/CT for bone marrow staging, especially when red bone marrow hyperplasia produces high tracer uptake [18]. Currently, most clinical guidelines recommend routine use of PET/CT for initial ES staging and surveillance (IV,A).

Bilateral iliac bone marrow biopsy is no longer standard in staging, because of the extremely low rate of bone marrow involvement in the absence of bone metastasis detection by BS or PET [19].

There is not standard staging classification system for ES and the American Joint Committee on Cancer (AJCC) bone cancer classification is not used by the most important international collaborative groups [20]. The most pragmatic

approach to assigning a treatment strategy is therefore based on risk factors.

Systematic reviews on prognostic factors for ES 5-year overall survival (OS) conclude that independent prognostic factors on diagnosis are age (< 15 years vs ≥ 15 years) [21], volume (< 200 mL vs ≥ 200 mL) [22], primary tumor location (appendicular vs axial), and disease extent (local vs lung metastasis vs extra-pulmonary metastasis) [23], and, after neoadjuvant treatment, histological response poor vs good (< 10% of viable tumor cells) [24]. While other studied variables, such as LDH [25], surgical margins, and local treatment modalities have been reported as prognostic factors, they are not all widely accepted as such.

Based on the above prognostic factors, the following risk groups have been proposed: standard-risk or good-risk (localized disease < 200 mL and, good histological response), High or poor risk (localized disease ≥ 200 mL or, poor histological response) [26]. Patients with metastasis constitute the group with the poorest prognosis, and outcomes are better for lung metastasis than for extrapulmonary metastasis) [27].

Treatment of localized disease on diagnosis

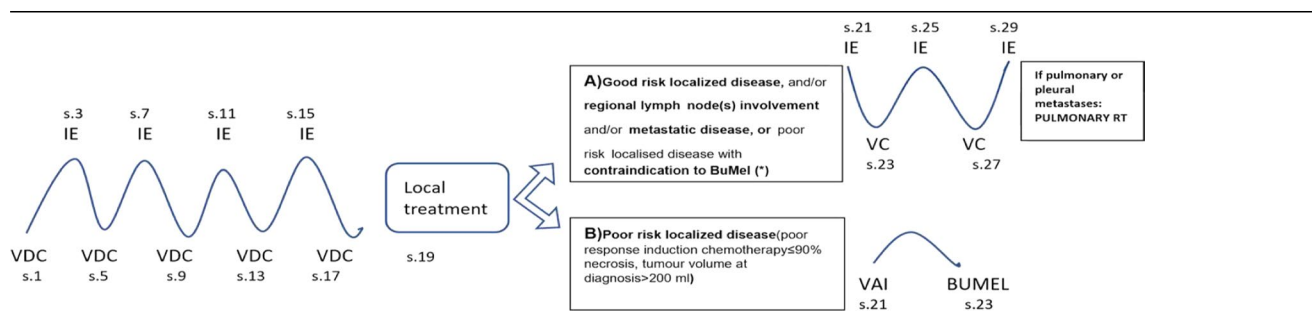
Systemic treatment

It is currently surmised that most patients with localized ES will have subclinical metastasis on diagnosis. Therefore, the accepted standard treatment for localized ES is multimodal, based on intensive multiagent chemotherapy (CTh) in addition to local treatment. After complete assessment (hematological, liver, cardiac, and renal function, and fertility preservation when desired), induction CTh is initiated (for volume reduction and subclinical metastatic disease control), followed by local treatment, and finally by consolidation CTh. OS is 70% and 50% at 5 and 10 years, respectively.

Almost all active protocols are based on combinations of the 5–6 most active drugs (doxorubicin, cyclophosphamide, vincristine, etoposide, ifosfamide, and dactinomycin).

Dose-dense regimens (every 2 weeks instead of the standard 3 weeks regimen) have been associated with a positive outcome in patients < 18 years [28]. The EuroEwing 2012 trial has also showed that an interval-compressed 2 weeks interval vincristine-doxorubicin-cyclophosphamide/ifosfamide-etoposide (VDC/IE) regimen is superior to a 3-week-interval vincristine-ifosfamide-doxorubicin-etoposide (VIDE) regimen [29] (I-A).

The consolidation use of high-dose (HD) CTh with stem-cell transplantation (SCT) for ES has been a topic of controversy in recent decades. A randomized study has demonstrated that, compared to seven cycles of maintenance chemotherapy, HD-CTh improves both event-free

Table 2 First line treatment plan. Adapted from Euroewing 2012

Agents and Dosage

Induction 9 cycles given at 14 day intervals on haematological recovery ($ANC \geq 0.75 \times 10^9/L$, platelets $\geq 75 \times 10^9/L$, with G-CSF support after every cycle)

VDC (1,5,9,13,17 weeks): Vincristine: 2 mg/m², d1 (IV push or short infusion)(max. single dose: 2 mg), Doxorubicin 37.5 mg/m²/d (IV infusion, 24 h) d1, d2 (75 mg/m²/cycle)

Cyclophosphamide 1200 mg/m² (IV infusion, 1 h) d1 plus MESNA and hydration*

IE (3,7,11,15 weeks) Ifosfamide 1800 mg/m²/d (IV infusion, 1 h) d 1–5 (9 g/m²/cycle) plus MESNA and hydration*, Etoposide 100 mg/m²/d (IV infusion, 2 h) d1–5 (500 mg/m²/cycle). (*¹ according to institutional guidelines)

Consolidation

(A) 5 cycles of alternating IE and VC at 14 day intervals on haematological recovery, same requirements)

IE (s.21,25,29) as in induction regimen; **VC (s.23,27)** as an induction therapy. G-CSF support. Maximum dosis calculated for a SA of 2 m²

(B) 1 cycle VAI plus BuMel:

VAI (s.21) Vincristine 1.5 mg/m² d1 (max. 2 mg), Actinomycin D 0.75 mg/m²/d (IV push) d1–2 (1.5 mg/m²/cycle) (max. single dose per day: 1.5 mg). Ifosfamide 3 g/m²/d (IV infusion, 1–3 h) d1–2 (6 g/m²/cycle) plus MESNA and hydration*

BuMel (s.23) Busulfan, IV 2 h infusion, (total of 4 doses from day -6 to day-3: Adults and children > 34 kg: 3.2 mg/kg/24 h, Children and adolescents: > 23–34 kg 3.8 mg/kg/24 h; 16–23 kg; 4.4 mg/kg/24 h; 9–< 16 kg: 4.8 mg/kg/24 h; < 9 kg: 4 mg/kg/24 h; Melphalan 140 mg/m² IV infusion over 30 min day -2 preinfusion. Hydration, G-CSF. Stem cell reinfusion on day 0. Clonazepam, and bone marrow transplantation support according to institutional guidelines

BuMel should be given on haematological recovery to $ANC 1.0 \times 10^9/L$, platelets $80 \times 10^9/L$

Mobilization and Harvesting: Recommended following the 9th induction cycle VDC. An option would be to do it from a steady state with G-CSF 10 mcgrs/kg/dia from the -4 day to collect a total of at least $3 \times 10^6/kg$ CD34+ cells. G-CSF must be continued daily collection has been completed

(*)Contraindication to BuMel: BuMel HDCT may interact with radiotherapy and must not compromise the ability to deliver an effective radiotherapy dose. Specific techniques and contraindications must be individualized. Failed harvesting is clearly a contraindication as well as other individual factors that have to be considered: Advanced age, comorbidities...

survival(EFS) and OS for patients aged < 50 years with localized high-risk ES (residual viable cells $\geq 10\%$) for patients undergoing surgery after induction chemotherapy alone or because of large tumor volume at diagnosis (≥ 200 mL) in unresected or initially resected tumors or resected tumors with preoperative irradiation but patients with a small unresected tumor were also eligible in case of poor clinical response to induction chemotherapy (< 50% radiologic reduction in soft tissue disease component); and no medical contraindication to treatment nor possibility of Busulfan interaction with radiotherapy [30, 31] (I,B).

Nevertheless, HD-CTh is not recommended for patients with metastasis to the pleura or to other sites, except within a clinical trial, as the benefit is unproven [32, 33].

No specific clinical trials have been conducted with adults aged > 40 years. Several single-center series and a retrospective analysis of the Surveillance, Epidemiology and End

Results (SEER) database show that survival rates approach pediatric survival rates when adults are treated with the same pediatric protocols [34–36] (II-B).

To summarize, our recommended approach for localized ES disease is based on EuroEwing 2012 Protocol (Table 2).

Response assessment

RECIST 1.1 is the standard oncology classification system for measuring response to treatment. Because ES bone lesions infrequently change in size, or show an asymmetric response to CTh, 3D-volumetric criteria [37] or reduced glycolytic activity outcomes [38] as evidenced by PET/CT, generally, correlate better with clinical

Pre- and post-treatment tumor glycolytic rate has been demonstrated to be a useful indirect measure of histological response to neoadjuvant CTh and progression-free survival

(PFS). Recent studies have shown that metabolic response ($> 55\%$ maximum standardized uptake value (SUVmax) reduction) is associated with 80% 3-year EFS [39], evaluable as early as nine days after starting treatment [40]. Thus, a PET-CT study is recommended for early neoadjuvant response assessment in ES (IV,A). Alternatives when PET-CT is not available are whole-body or local MRI with diffusion ADC, dynamic contrast enhancement and/or 3D-volume measures [41].

From the pathology perspective, response evaluation after neoadjuvant therapy, should be based on a full sagittal section and mapping of a whole representative slice of the tumor, as this allows evaluation of the relative proportions of viable and necrotic tumor [42].

Surgery

Recommended local treatment options include wide excision, definitive RT, or, in selected cases, amputation [43]. However, surgery is the treatment of choice if tumor resection is possible with negative margins, without significant morbidity, and with a reasonable functional outcome.

Data from retrospective analyses suggest that surgery yields better local control and survival than definitive RT in patients with localized disease [44]. Selection bias, however, likely accounts for at least some of those reported results, since larger, axial lesions with a higher rate of local failure and overall poorer prognosis are more frequently referred for RT. It is also becoming clear that in less accessible locations, like the pelvis, preoperative RT and surgical resection are associated with better histological response and OS rates.

Surgery has several advantages over RT, including avoided risk of secondary radiation induced sarcomas, the possibility of knowing the tumor necrosis percentage after induction CTh, and that it avoids bone growth retardation and deformities associated with irradiation of the immature child skeleton.

Since options have not been directly compared in a randomized trial, the choice of local control treatments should be individualized, as it depends on tumor location, and size, response to CTh, anticipated morbidity, the patient's age, and patient preferences. Patients and their treatment should be discussed by a multidisciplinary tumor board [45] (I,A).

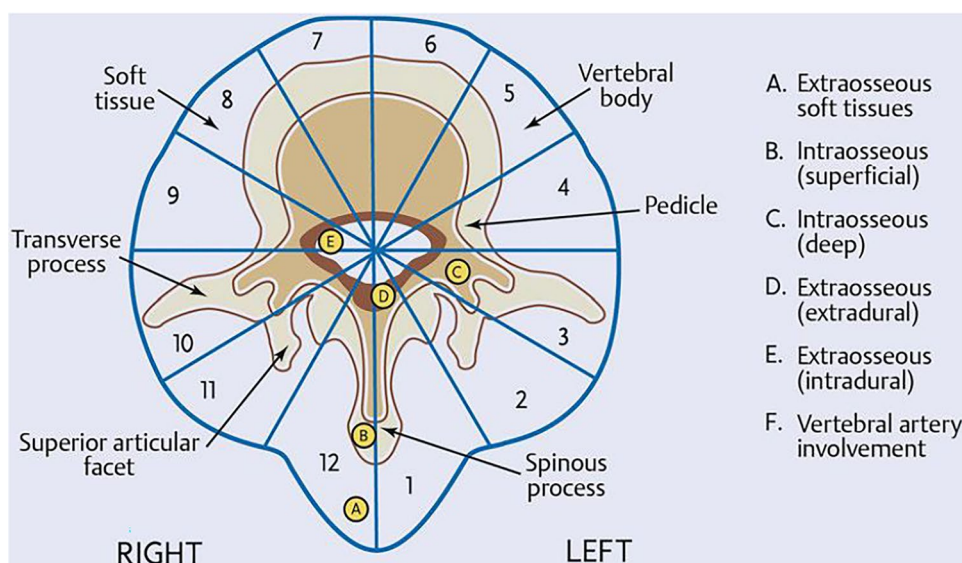
Surgical management of patients with limb ES is a major challenge. The main objectives of current orthopedic oncology are optimal tumor resection and a functional extremity, with greater OS of both, the patient and the tumor reconstruction [46].

The surgical goal in ES is a wide-margin en-bloc resection (i.e., extirpation of the tumor with a normal tissue cuff covering margins) [47]. This usually means removing 2 cm of normal tissue (1 cm if an anatomical barrier is present), and performing a bone osteotomy at a distance of 3–5 cm from the bone involvement level (II,A). Smaller margins in the bone may be acceptable after effective neoadjuvant treatment. Bone resections with joint preservation that use open physis cartilage as a margin are also oncologically acceptable, as favoring preservation of the joint.

Amputation is indicated when, in limb sparing surgery, tumor resection with wide margins is not feasible, or when the outcome in terms of functionality is not acceptable.

Surgery should include removal of all tissues originally affected by the tumor, i.e., not just the tissues remaining after

Fig. 2 WBB (Weinstein, Boriani, Biagini) Surgical Staging System. The transverse extension of the vertebral tumor is described with reference to 12 radiation zones (numbered 1 to 12 in a clockwise order) and to five concentric layers (A–E from the paravertebral extraosseous compartments to the dural involvement). The longitudinal extent of the tumor is recorded according to the levels involved



CTh shrinkage; when this is not possible, postoperative RT should be applied.

Regarding axial ES, there is no robust evidence in the literature for certain aspects of local management.

Neurological symptoms requiring laminectomy are common at presentation. However, in terms of neurological recovery, urgent decompressive surgery before an histological diagnosis is obtained may increase the risk of local recurrence, without providing a clear advantage over non-surgical treatment.

For patients with isolated (non-metastatic) spinal ES, surgical resection has been reported to be associated with significantly better OS than no surgery, after adjusting for age, RT, and extent of local tumor invasion [48, 49].

The additional challenge with axial tumors is that, at the moment of diagnosis, tumor margins have usually already exceeded the vertebral compartment with the involvement of paravertebral structures, and frequently, more than one vertebral segment. Thus, despite it is stated as wide-margin/ en-bloc -resection, spinal surgery is not radical, since the vertebral compartment must be opened to maintain the integrity of the uninvolved nervous structures [50] (Fig. 2).

As for implants, while carbon fiber-reinforced polyether etherketone (CFR-PEEK) and titanium implants have similar safety and efficacy profiles, CFR-PEEK has distinct advantages that make it a promising alternative for the treatment/outcome of spinal oncology patients, namely, that it decreases artefacts, improves early detection of local tumor recurrences, increases RT dose accuracy, and is also associated with lower complication rates.

For patients with primitive neuroectodermal tumor or extra-skeletal ES around the spine, the preferred treatment is multiagent CTh combined with wide- margin en-bloc excision and RT.

Finally, for unresectable spinal tumors, RT is the treatment of choice, as surgical debulking is not generally recommended.

Radiotherapy

RT as definitive local treatment RT is an effective local control option for patients for whom function-preserving surgery is not possible because of tumor location or extent, and for patients with unresectable primary tumors despite induction CTh. In the latter case, debulking surgery should be avoided, so the treatment of choice is definitive radiotherapy [51] (II,A), using doses in the range of 54–60 Gy.

RT as neo/adjuvant treatment RT should be started within 60 days after surgery [52] (II,A) and should be considered in the following situations:

- Bulky tumors in difficult sites (pelvis, sacrum, spine, and paraspinal and rib tumors associated with pleural effusion), when RT can be preoperative (45 Gy) or postoperative.
- Residual microscopic tumors (R1: 45–50.4 Gy), gross disease after surgery (R2: 54–55.8 Gy), and inadequate surgical margins (i.e., marginal or intralesional surgery).
- High-risk chest wall primary tumors with close or involved margins, initial pleural effusion, pleural infiltration, or intraoperative contamination of the pleural space. For which adjuvant hemithorax irradiation doses typically range from 15 to 18 Gy (by age) plus a boost to the initial tumor.
- When histological response to preoperative CTh is poor ($\leq 90\%$), even if surgical margins are negative (R0).

New RT technologies (such as stereotactic body RT (SBRT), intensity-modulated RT (IMRT), and proton therapy) potentially improve local control and toxicity outcomes. Recommended are conventional RT schedules of 1.8–2 Gy/fraction, once daily, rather than hyper-fractionated schedules.

Currently under evaluation are higher doses (64.8 Gy) for large tumors (> 8 cm, 100 mL) and lower doses (30–36 Gy) for small tumors (< 8 cm) that respond well to CTh.

Treatment of metastatic disease on diagnosis

Although patients with metastasis on diagnosis have a significantly poorer prognosis (5-year OS 15–40%), treatment must be administered with curative intent, first, because it is difficult to predict which patients can be cured, and second, because treatment may relieve pain and prolong PFS [53].

Systemic treatment

CTh regimens are broadly the same as for localized disease, except for certain considerations. Neither higher alkylator doses [54] nor HD-CTh with SCT [32, 33, 55] have shown any benefit for metastatic ES, so their use is not advisable except in clinical trials (I,C).

Local treatment

Treatment is individualized according to the primary tumor, and the number and location of metastases.

Primary site management Local treatment of the primary tumor follows the same principles as for localized disease, but radiotherapy is preferred to amputation especially in patients with extensive metastatic disease.

Pulmonary metastases Since patients with few pulmonary metastases, have a better prognosis, they may be good candidates for surgical managements [56]. Repeated pulmonary metastasis excisions have been associated with improved OS [57].

SBRT of residual lung metastases is also a viable option [58, 59].

Retrospective reports from large cooperative groups and single center series suggest that bilateral whole-lung irradiation (WLI) benefits patients with pulmonary metastases, even when the metastases have been resected or have completely disappeared with CTh [60]. Long-term lung toxicity is acceptable [61], while patients with metastases who undergo surgery or boost RT have slightly higher complications.

Bilateral WLI is recommended at the end of systemic treatment for good responders, specifically, 15–18 Gy in daily doses of 1.8–2 Gy plus a focal boost to 40–50 Gy for gross residual metastases (II,B).

Bone and soft tissue metastases While patients with isolated bone or soft tissue metastases have a poorer prognosis than patients with isolated lung metastases, 10% may survive over the long term.

RT in doses of 40–50 Gy and SBRT for metastatic lesions are well tolerated and may improve disease control and symptoms [62] (IV,C).

Our treatment approach for EW with pulmonary metastasis is multimodal therapy including CTh, local treatment of the primary tumor and local treatment of the residual lesions (except in disseminated metastatic disease at the time of local treatment) (III, B). Treatment of residual disease may be followed up by low-dose WLI even when nodules have disappeared.

Treatment on relapse

Local relapse

Local relapse occurs in approximately 5% of patients undergoing limb rescue surgery for ES in specialized centers. Treatment of a local relapse depends on the time of recurrence, the association with distant metastases, and also resectability using the same criteria as for initial

presentation. While prognosis for local ES relapse is usually poor, a non-significant impact on OS has also been reported [63].

RT and SBRT are options if the site has not previously been irradiated, and also as palliative care.

For local relapse, CTh is generally administered given the systemic nature of the disease and the high probability of developing metastasis if treatment is focused only on the local site (IV,B).

Systemic relapse/progression

Prognosis for ES following systemic relapse is poor, with 5-year OS reported as 8–30% [64]. Most recurrences occur within the first 2 years after initial therapy and most patients develop metastatic disease or metastatic and local disease [65].

Time since initial therapy is the most important prognostic factor as prognosis is more favorable for patients whose relapse occurs more than 18 months after initial diagnosis.

Several CTh regimens have been used for relapsed ES, including alkylating agents, camptothecin derivatives, and platinum agents. Evidence regarding those regimens comes from small retrospective series or early trials with limited numbers of patients. In the last decade, four of the most frequently used CTh regimens have been High Dose ifosfamide (HD-IFO) [66], gemcitabine-docetaxel (Gem-Doc) [67], irinotecan-temozolomide (Irn-Tmz), and topotecan-cyclophosphamide (Topo-Cyc) [68] (II,B). An ongoing international European phase III trial, rEECur, comparing those four CTh regimens as treatment for refractory/recurrent ES, reported in a first interim analysis that the poorest objective response and PFS was obtained for Gem-Doc [69] and, in subsequent interim analyses, that Irn-Tmz and Topo-Cyc were both inferior to HD-IFO in terms of response, PFS, and OS [70] (I,A). The HD-IFO arm is currently being compared to carboplatin-etoposide and to Lenvatinib plus HD-IFO, and new arms will be sequentially added and compared.

While several retrospective studies have suggested a potential survival benefit for HD-CTh followed by SCT [71], as yet no randomized trial data support this as standard practice (IV,C). Tyrosine kinase inhibitors although not approved for ES, have been evaluated. The single-arm phase II CABONE study evaluated cabozantinib in 39 patients with ES, and reported an objective response of 26% and median PFS of months [72] (III,B), and the non-comparative, randomized, double-blind, placebo-controlled phase II study of regorafenib, reported a median PFS of 11.4 weeks [73].

Other novel CTh combinations under investigation include irinotecan plus trabectedin, which, in a recent phase

Table 3 Proposed Follow-up (Adapted from NCCN/BSG)

	First 2 years	3–5 years	> 5 years	> 10 years
Clinical&PE	Every 2–3 months	Every 4–6 months	Annually	Annually (II-B)
Local Imaging. MRI. Add US or PET if metal artifacts	Every 2–3 months	Every 4–6 months	Based on clinical indications or annually	Based on clinical indications or annually(II-B)
Chest Imaging Chest Rx or CT	Every 2–3 m (mind alternate CT & CRX)	Based on clinical indications or every 4–6 m (mind alternate CT & CRX)	Based on clinical indications or annually	Based on clinical indications or annually (II-B)
Body Imaging Whole-body PET/ CT or bone scan	Based on clinical indications or every 6 months	Based on clinical indications or annually	Based on clinical indications	Based on clinical indications

Table 4 Checklist for patients with ES

Patients should be attended at centers belonging to a sarcoma network with a concrete expertise and multidisciplinary team
Children and adolescents should be referred to centers which in addition provide age-specific expertise
A clear treatment plan with objectives and stimated timelines should be discussed with the patient
Psychological support for patients and families is highly recommended. Psychologists dedicated to childhood and adolescence can provide expert assistance
Basal assessment (especially cardiological and endocrine) should be performed and other risk factors should be controlled in order to reduce toxicity burden of treatments
Oncofertility consultation should be assessed as soon as possible after the diagnosis
Encouraging of physical activity, adapted to patient situation, is highly recommended
Nutritional advice may give a greater sense of well-being and may help to control chemo and radio-therapy side effects
Basal work/ study activity should be assessed and patients should be refered to social workers as needed. Communication with school/university tutors and employers should be encouraged in order to facilitate a realistic plan for reintegration during and after treatment
Getting in touch with other patients and patients' family through patient associations may reduce isolation feeling and should be offered
Quick activation of palliative care is essential (when indicated)
All patients are entitled to request a second opinion from other oncologist(s)/team

I trial, showed encouraging activity and a good toxicity profile [74]. Finally, new molecules in monotherapy or combination therapy are currently under-evaluation in phase I trials.

Follow-up

In the first 2 years after the end of treatment, local follow-up with clinical and physical examination every 2–3 months is usually enough for extremities. However, a structured local imaging protocol may benefit survival [75] (IV,B), and local magnetic resonance imaging (MRI) is recommended every quarter during the first two years (IV,B).

A metabolic study using FDG-PET/-CT, which has 92% sensitivity and 93% specificity for the detection of recurring bone sarcoma [76], is indicated in two main scenarios: when radiological images are inconclusive for local recurrence, and when prosthetic or osteosynthetic material is present in the patient [77] (III,A). Metallic components, however, do

not cause significant artefacts that interfere with interpretation of FDG-PET-CT images.

Lung surveillance for metastases is usually by chest radiography or CT, and distant bone disease by PET/-CT or BS. For both bone and nodal surveillance, whole-body PET/-CT offers superior sensitivity, and is especially recommended in the case of new symptoms, abnormal imaging (primary tumors positive prior PET), and before any surgical decision-making [77] (IV,B). BS is reserved for cases when PET/-CT is not available (Table 3).

Note that the NCCN, ESMO and British Sarcoma Group (BSG) guidelines follow-up recommendations for high-grade sarcomas, including ES, vary (those in the BSG are less stringent). Recent evidence favors more individualized and less intensive follow-up regimes [78, 79].

Patient-centered care

A radiological suspicion of malignant primary tumor in patients with a bone or soft tissue lesion should be referred to a networked sarcoma center with specialized and multi-disciplinary expertise (IV,A). Children and adolescents, in particular, should be referred to centers with additional age-specific expertise.

Since ES occurs most frequently in children, adolescents, and young adults, communicating the diagnosis and its impact is particularly difficult. Psychological support for patients and family is highly recommended and clear communications between the patient and healthcare personnel is essential to avoid unfounded fears and concerns. Table 4 provides a short checklist for patients and physicians.

Treatment and follow-up visits should be scheduled taking into consideration quality of life, appropriate management of short- and long-term toxicities, early detection of relapses and secondary malignancies, and timely evaluation of co-morbidities and correlated risk patterns. Cardiological evaluation should include at least baseline and follow-up electrocardiogram (QTc interval and left ventricular ejection fraction).

Patient participation in clinical trials, although highly recommended, should be on an absolutely voluntary basis, and should be preceded by a family/patient discussion with the oncologist.

Funding No funds, grants, or other support was received.

Data Availability Not applicable.

Declarations

Conflict of interest The authors have no competing interests to declare that are relevant to the content of this article.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

1. https://www.nccn.org/professionals/physician_gls/pdf/bone.pdf. Accessed 20 Mar 2021.
2. Casali PG, Bielack S, Abecassis N, Aro HT, Bauer S, Biagini R, et al. Bone sarcomas: ESMO- PaedCan-EURACAN Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2018;29(Suppl):iv79–95.
3. Dykewicz CA. Summary of the guidelines for preventing opportunistic infections among hematopoietic stem cell transplant recipients. *Clin Infect Dis*. 2001;33(2):139–44.
4. Stiller CA, Trama A, Serraino D, Rossi S, Navarro C, Chirilaque MD, Casali PG. Descriptive epidemiology of sarcomas in Europe: report from the RARECARE project. *Eur J Cancer*. 2013;49(3):684–95. <https://doi.org/10.1016/j.ejca.2012.09.011>. (Epub 2012 Oct 15. PMID: 23079473).
5. Panicek DM, Gatsonis C, Rosenthal DI, Seeger LL, Huvois AG, Moore SG, Caudry DJ, Palmer WE, McNeil BJ. CT and MR imaging in the local staging of primary malignant musculoskeletal neoplasms: report of the Radiology Diagnostic Oncology Group. *Radiology*. 1997;202(1):237–46. <https://doi.org/10.1148/radiology.202.1.8988217>. (PMID: 8988217).
6. Clemente EJI, Navarro OM, Navallas M, Ladera E, Torner F, Sunol M, Garraus M, March JC, Barber I. Multiparametric MRI evaluation of bone sarcomas in children. *Insights Imaging*. 2022;13(1):1–18. <https://doi.org/10.1186/s13244-022-01177-9>.
7. Kubo T, Furuta T, Johan MP, Sakuda T, Ochi M, Adachi N. A meta-analysis supports core needle biopsy by radiologists for better histological diagnosis in soft tissue and bone sarcomas. *Medicine (United States)*. 2018. <https://doi.org/10.1097/MD.00000000000011567>.
8. Guimarães JB, Rigo L, Lewin F, Emerick A. The importance of PET/CT in the evaluation of patients with Ewing tumors. *Radiol Bras*. 2015;48(3):175–80. <https://doi.org/10.1590/0100-3984.2013.1887>. (PMID: 26185344; PMCID: PMC4492570).
9. Liu PT, Valadez SD, Chivers FS, Roberts CC, Beauchamp CP. Anatomically based guidelines for core needle biopsy of bone tumors: implications for limb-sparing surgery. *Radiographics*. 2007;27:189–206.
10. Grünewald TGP, Cidre-Aranaz F, Surdez D, Tomazou EM, de Álava E, Kovar H, Sorensen PH, Delattre O, Dirksen U. Ewing sarcoma. *Nat Rev Dis Primers*. 2018;4(1):5. <https://doi.org/10.1038/s41572-018-0003-x>. (PMID: 29977059).
11. Llombart-Bosch A, Machado I, Navarro S, Bertoni F, Bacchini P, Alberghini M, Karzeladze A, Savelov N, Petrov S, Alvarado-Cabrero I, Mihaila D, Terrier P, Lopez-Guerrero JA, Picci P. Histological heterogeneity of Ewing's sarcoma/PNET: an immunohistochemical analysis of 415 genetically confirmed cases with clinical support. *Virchows Arch*. 2009;455(5):397–411. <https://doi.org/10.1007/s00428-009-0842-7>. (Epub 2009 Oct 17 PMID: 19841938).
12. Machado I, Noguera R, Mateos EA, Calabuig-Fariñas S, López FI, Martínez A, Navarro S, Llombart-Bosch A. The many faces of atypical Ewing's sarcoma. A true entity mimicking sarcomas, carcinomas and lymphomas. *Virchows Arch*. 2011;458(3):281 (90 Koch R10.1007/s00428-010-1023-4. Epub 2010 Dec 23. PMID: 21181413).
13. Machado I, Yoshida A, Morales MGN, Abrahão-Machado LF, Navarro S, Cruz J, Lavernia J, Parafloriti A, Picci P, Llombart-Bosch A. Review with novel markers facilitates precise categorization of 41 cases of diagnostically challenging, “undifferentiated small round cell tumours”. A clinicopathologic, immunophenotypic and molecular analysis. *Ann Diagn Pathol*.

- 2018;34:1–12. <https://doi.org/10.1016/j.anndiagpath.2017.11.011>.
14. Wang WL, Patel NR, Caragea M, Hogendoorn PC, López-Terrada D, Hornick JL, Lazar AJ. Expression of ERG, an Ets family transcription factor, identifies ERG-rearranged Ewing sarcoma. *Mod Pathol*. 2012;25(10):1378–83. <https://doi.org/10.1038/modpathol.2012.97>. (Epub 2012 Jul 6 PMID: 22766791).
15. Bishop JA, Alaggio R, Zhang L, Seethala RR, Antonescu CR. Adamantinoma-like Ewing family tumors of the head and neck: a pitfall in the differential diagnosis of basaloid and myoepithelial carcinomas. *Am J Surg Pathol*. 2015;39(9):1267–74. <https://doi.org/10.1097/PAS.0000000000000460>. (PMID:26034869;PMCID:PMC4537687).
16. Casali PG, Bielack S, Abecassis N, Aro HT, Bauer S, Biagini R, et al. Bone sarcomas: ESMO- PaedCan-EURACAN Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2018;29(Suppl 4):iv79–95.
17. Treglia G, Salsano M, Stefanelli A, Mattoli MV, Giordano A, Bonomo L. Diagnostic accuracy of ¹⁸F-FDG-PET and PET/CT in patients with Ewing sarcoma family tumours: a systematic review and a meta-analysis. *Skeletal Radiol*. 2012;41(3):249–56.
18. Kalus S, Saifuddin A. Whole-body MRI vs bone scintigraphy in the staging of Ewing sarcoma of bone: a 12-year single-institution review. *Eur Radiol*. 2019;29(10):5700–8. <https://doi.org/10.1007/s00330-019-06132-9>. (Epub 2019 Mar 26 PMID: 30915559).
19. Cesari M, Righi A, Colangeli M, Gambarotti M, Spinnato P, Ferraro A, et al. Bone marrow biopsy in the initial staging of Ewing sarcoma: experience from a single institution. *Pediatr Blood Cancer*. 2019;66(6):e27653. <https://doi.org/10.1002/pbc.27653>. (Epub 2019 Feb 5. PMID: 30724024).
20. R.G. TNM staging of tumors of bone. PathologyOutlines.com website. <http://www.pathologyoutlines.com/topic/bonestaging.html>. Accessed 24 Feb 2022.
21. Cotterill SJ, Ahrens S, Paulussen M, Jürgens HF, Voûte PA, Gadner H, Craft AW. Prognostic factors in Ewing's tumor of bone: analysis of 975 patients from the European Intergroup Cooperative Ewing's Sarcoma Study Group. *J Clin Oncol*. 2000;18(17):3108–14. <https://doi.org/10.1200/JCO.2000.18.17.3108>. (PMID: 10963639).
22. Bacci G, Longhi A, Ferrari S, Mercuri M, Versari M, Bertoni F. Prognostic factors in non- metastatic Ewing's sarcoma tumor of bone: an analysis of 579 patients treated at a single institution with adjuvant or neoadjuvant chemotherapy between 1972 and 1998. *Acta Oncol*. 2006;45(4):469–75.
23. Bosma SE, Lancia C, Rueten-Budde AJ, et al. Easy-to-use clinical tool for survival estimation in Ewing sarcoma at diagnosis and after surgery. *Sci Rep*. 2019;9(1):11000.
24. Salzer-Kuntschik M, Brand G, Delling G. Determination of the degree of morphological regression following chemotherapy in malignant bone tumors. *Pathologe*. 1983;4(3):135–41.
25. Li S, Yang Q, Wang H, et al. Prognostic significance of serum lactate dehydrogenase levels in Ewing's sarcoma: a meta-analysis. *Mol Clin Oncol*. 2016;5(6):832–8.
26. Anderton J, Moroz V, Marec-Berard P, et al. International randomised controlled trial for the treatment of newly diagnosed EWING sarcoma family of tumours—EURO EWING 2012 Protocol. *Trials*. 2020;21(1):96.
27. Friedman DN, Chastain K, Chou JF, Moskowitz CS, Adsuar R, Wexler LH, Chou AJ, DeRosa A, Candela J, Magnan H, Pun S, Kahan T, Wolden SL, Meyers PA, Oeffinger KC. Morbidity and mortality after treatment of Ewing sarcoma: A single-institution experience. *Pediatr Blood Cancer*. 2017 Nov;64(11). <https://doi.org/10.1002/pbc.26562>. Epub 2017 Apr 18. PMID: 28417551
28. Womer RB, West DC, Krailo MD, et al. Randomized controlled trial of interval compressed chemotherapy for the treatment of localized Ewing sarcoma: a report from the Children's Oncology Group. *J Clin Oncol*. 2012;30:4148–54.
29. Brennan B, Kirton L, Marec-Bérard P, Gaspar N, Laurence V, Martín-Broto J, Sastre A, Gelderblom H, Owens C, Fenwick N, Strauss S, Moroz V, Whelan J, Wheatley K. Comparison of two chemotherapy regimens in patients with newly diagnosed Ewing sarcoma (EE2012): an open-label, randomised, phase 3 trial. *Lancet*. 2022;400(10362):1513–21. [https://doi.org/10.1016/S0140-6736\(22\)01790-1](https://doi.org/10.1016/S0140-6736(22)01790-1). (PMID: 36522207).
30. Ferrari S, Sundby Hall K, Luksch R, et al. Nonmetastatic Ewing family tumors. High dose chemotherapy with stem cell rescue in poor responder patients. Results of the Italian Sarcoma Group/Scandinavian Sarcoma Group III protocol. *Ann Oncol*. 2011;22:1221–7.
31. Whelan J, Le Deley MC, Dirksen U, Le Teuff G, Brennan B, Gaspar N, Hawkins DS, Amler S, Bauer S, Bielack S, Blay JY, Burdach S, Castex MP, Dilloo D, Eggert A, Gelderblom H, Gentet JC, Hartmann W, Hassenpflug WA, Hjorth L, Jimenez M, Klingebiel T, Kontny U, Kruseova J, Ladenstein R, Laurence V, Lervat C, Marec-Berard P, Marreaud S, Michon J, Morland B, Paulussen M, Ranft A, Reichardt P, van den Berg H, Wheatley K, Judson I, Lewis I, Craft A, Juergens H, Oberlin O; Euro-E.W.I.N.G.99 and EWING-2008 Investigators. High-Dose Chemotherapy and Blood Autologous Stem-Cell Rescue Compared With Standard Chemotherapy in Localized High-Risk Ewing Sarcoma: Results of Euro-E.W.I.N.G.99 and Ewing-2008. *J Clin Oncol*. 2018;36(31):JCO2018782516. <https://doi.org/10.1200/JCO.2018.78.2516>. Epub ahead of print. PMID: 30188789; PMCID: PMC6209090.
32. Dirksen U, Brennan B, Le Deley MC, Cozic N, van den Berg H, Bhadri V, Euro-E.W.I.N.G. 99andEwing2008Investigators, et al. High-Dose Chemotherapy Compared With Standard Chemotherapy and Lung Radiation in Ewing Sarcoma With Pulmonary Metastases: Results of the European Ewing Tumour Working Initiative of National Groups, 99 Trial and EWING 2008. *J Clin Oncol*. 2019;37(34):3192–202. <https://doi.org/10.1200/JCO.19.00915>. (Epub 2019 Sep 25. PMID: 31553693; PMCID: PMC6881099).
33. Koch R, Gelderblom H, Haveman L, et al. High dose treosulfan and melphalan as consolidation therapy versus standard therapy for high risk (metastatic)Ewing sarcoma. *J Clin Oncol*. 2022;40:2307.
34. Ahmed SK, Robinson SI, Okuno SH, et al. Adult Ewing sarcoma: survival and local control outcomes in 36 patients with metastatic disease. *Am J Clin Oncol*. 2014;37:423.
35. Bacci G, Baladelli A, Forni C, et al. Adjuvant and neoadjuvant chemotherapy for Ewing sarcoma family tumors in patients between 40 and 60: report of 35 cases and comparison of results with 586 younger patients treated with the same protocols in the same years. *Cancer*. 2007;109:780.
36. Pretz JL, Baryasuskas CM, George S, et al. Localized adult Ewing sarcoma: favourable outcomes with alternating vincristine, doxorubicin, cyclophosphamide, and ifosfamide, etoposide (VDC/IE)-based multimodality therapy. *Oncologists*. 2017;22:1265.
37. McCarville MB, Federico SM, Bishop MW, Pappo AS, Shulkin BL. Assessment of chemotherapy response in Ewing sarcoma. *Radiology*. 2016;281(2):647–8.
38. Raciborska A, Biliska K, Drabko K, Michalak E, Chaber R, Pogorzała M, Polczyńska K, Sobol G, Wiczorek M, Muszyńska-Roslan K, Rychłowska-Pruszyńska M, Rodriguez-Galindo C, Dziuk M. Response to chemotherapy estimates by FDG PET is an important prognostic factor in patients with Ewing sarcoma. *Clin Transl Oncol*. 2016;18(2):189–95. <https://doi.org/10.1007/s12094-015-1351-6>. (Epub 2015 Aug 7 PMID: 26250765).

39. Palmerini E, Colangeli M, Nanni C, Fanti S, Marchesi E, Paioli A, et al. The role of FDG PET/CT in patients treated with neoadjuvant chemotherapy for localized bone sarcomas. *Eur J Nucl Med Mol Imaging*. 2017;44(2):215–23.
40. Hyun OJ, Lubner BS, Leal JP, Wang H, Bolejack V, Schuetz SM, Schwartz LH, Helman LJ, Reinke D, Baker LH, Wahl RL. Response to early treatment evaluated with 18F-FDG PET and PERCIST 1.0 predicts survival in patients with Ewing sarcoma family of tumors treated with a monoclonal antibody to the insulin like growth factor 1 receptor. *J Nucl Med*. 2016;57(5):735–40. <https://doi.org/10.2967/jnumed.115.162412>. (Epub 2016 Jan 21. PMID: 26795289; PMCID: PMC5548359).
41. Asmar K, Saade C, Salman R, et al. The value of diffusion weighted imaging and apparent diffusion coefficient in primary Osteogenic and Ewing sarcomas for the monitoring of response to treatment: Initial experience. *European Journal of Radiology*. 2020;124:108855. <https://doi.org/10.1016/j.ejrad.2020.108855>. PMID: 32018075
42. Wardelmann E, Haas RL, Bovée JV, et al. Evaluation of response after neoadjuvant treatment in soft tissue sarcomas; the European Organization for Research and Treatment of Cancer-Soft Tissue and Bone Sarcoma Group (EORTC-STBSG) recommendations for pathological examination and reporting. *Eur J Cancer*. 2016;53:84–95.
43. Schuck A, Ahrens S, Paulussen M, et al. Local therapy in localized Ewing tumors: results of 1058 patients treated in the CESS 81, CESS 86, and EICESS 92 trials. *Int J Radiat Oncol Biol Phys*. 2003;55:168–77.
44. Schuck A, Ahrens S, Paulussen M, Kuhlen M, Köneemann S, Rube C, Winkelmann W, Kotz R, Dunst J, Willich N, Jürgens H. Local therapy in localized Ewing tumors: results of 1058 patients treated in the CESS 81, CESS 86, and EICESS 92 trials. *Int J Radiat Oncol Biol Phys*. 2003. [https://doi.org/10.1016/S0360-3016\(02\)03797-5](https://doi.org/10.1016/S0360-3016(02)03797-5).
45. Casali PG, Bielack S, Abecassis N, Aro HT, Bauer S, Biagini R, Bonvalot S, Boukovinas I, Bovee JVMG, Brennan B, Brodowicz T, Broto JM, Brugières L, Buonadonna A, De Álava E, Dei Tos AP, Del Muro XG, Dileo P, Dhooge C, Eriksson M, Fagioli F, Fedenko A, Ferraresi V, Ferrari A, Ferrari S, Frezza AM, Gaspar N, Gasperoni S, Gelderblom H, Gil T, Grignani G, Gronchi A, Haas RL, Hassan B, Hecker-Nolting S, Hohenberger P, Issels R, Joensuu H, Jones RL, Judson I, Jutte P, Kaal S, Kager L, Kasper B, Kopeckova K, Krákorová DA, Ladenstein R, Le Cesne A, Lugowska I, Merimsky O, Montemurro M, Morland B, Pantaleo MA, Piana R, Picci P, Piperno-Neumann S, Pousa AL, Reichardt P, Robinson MH, Rutkowski P, Safwat AA, Schöffski P, Sleijfer S, Stacchiotti S, Strauss SJ, Sundby Hall K, Unk M, Van Coevorden F, van der Graaf WTA, Whelan J, Wardelmann E, Zaikova O, Blay JY, ESMO Guidelines Committee, PaedCan and ERN EURACAN. Bone sarcomas: ESMO-PaedCan-EURACAN Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2018;29(Suppl 4):iv9–95. <https://doi.org/10.1093/annonc/mdy310>. (PMID: 30285218).
46. Sluga M, Windhager R, Lang S, Heinzl H, Krepler P, Mittermayer F, Dominkus M, Zoubek A, Kotz R. The role of surgery and resection margins in the treatment of Ewing's sarcoma. *Clin Orthop Relat Res*. 2001;392:394–9.
47. Alvarez-SanNicolas J, Gracia-Alegria I, Trullols-Tarrago L, et al. Prognostic factors and survival in Ewing's sarcoma treated by limb salvage surgery. *Clin Transl Oncol*. 2019;21:1374–82.
48. Miller BJ, Gao Y, Duchman KR. Does surgery or radiation provide the best overall survival in Ewing's sarcoma? A review of the National Cancer Data Base. *J Surg Oncol*. 2017;116(3):384–90. <https://doi.org/10.1002/jso.24652>. (Epub 2017 Apr 18 PMID: 28420037).
49. Yamazaki T, McLoughlin GS, Patel S, Rhines LD, Fournery DR. Feasibility and safety of en bloc resection for primary spine tumors: a systematic review by the Spine Oncology Study Group. *Spine (Phila Pa 1976)*. 2009;34(22 Suppl):S31–8. <https://doi.org/10.1097/BRS.0b013e3181b8b796>.
50. Boriani S, Weinstein JN, Biagini R. Primary bone tumors of the spine. Terminology and surgical staging. *Spine*. 1997;22:1036–44.
51. Donaldson SS. Ewing sarcoma: radiation dose and target volume. *Pediatr Blood Cancer*. 2004;42(5):471–6. <https://doi.org/10.1002/pbc.10472>. (PMID: 15049023).
52. Schuck A, Rube C, Köneemann S, Rube CE, Ahrens S, Paulussen M, Dunst J, Jürgens H, Willich N. Postoperative radiotherapy in the treatment of Ewing tumors: Influence of the interval between surgery and radiotherapy. *Strahlenther Onkol*. 2002. <https://doi.org/10.1007/s00066-002-0883-1>.
53. Haeusler J, Ranft A, Boelling T, Gosheger G, Braun-Munzinger G, Vieth V, Burdach S, vanden Berg H, Juergens H, Dirksen U. The value of local treatment in patients with primary, disseminated, multifocal Ewing sarcoma (PDMES). *Cancer*. 2010;116(2):443–50.
54. Granowetter L, Womer R, Devidas M, Krailo M, Wang C, Bernstein M, Marina N, Leavey P, Gebhardt M, Healey J, Shamberger RC, Goorin A, Miser J, Meyer J, Arndt CA, Sailer S, Marcus K, Perlman E, Dickman P, Grier HE. Dose-intensified compared with standard chemotherapy for nonmetastatic Ewing sarcoma family of tumors: a Children's Oncology Group Study. *J Clin Oncol*. 2009;27(15):2536–41. <https://doi.org/10.1200/JCO.2008.19.1478>. (Epub 2009 Apr 6. PMID: 19349548; PMCID: PMC2684856).
55. Meyers PA, Krailo MD, Ladanyi M, Chan KW, Sailer SL, Dickman PS, et al. High-dose melphalan, etoposide, total-body irradiation, and autologous stem-cell reconstitution as consolidation therapy for high-risk Ewing's sarcoma does not improve prognosis. *J Clin Oncol*. 2001;19(11):2812–20. <https://doi.org/10.1200/JCO.2001.19.11.2812>. (PMID: 11387352).
56. Reza J, Sammann A, Jin C, Horvai A, Hudnall M, Jablons DM, Jahan T, Kornak J, Mann MJ. Aggressive and minimally invasive surgery for pulmonary metastasis of sarcoma. *J Thorac Cardiovasc Surg*. 2014;147(4):1193–200. <https://doi.org/10.1016/j.jtcvs.2013.12.021>. (Epub 2014 Jan 2 PMID: 24507991).
57. Letourneau PA, Shackett B, Xiao L, Trent J, Tsao KJ, Lally K, Hayes-Jordan A. Resection of pulmonary metastases in pediatric patients with Ewing sarcoma improves survival. *J Pediatr Surg*. 2011;46(2):332–5.
58. Brown Lindsay C., Rachael A. Lester, Michael P. Grams, Michael G. Haddock, Kenneth R. Olivier, Carola A. S. Arndt, Peter S Rose, Nadia N Issa Laack. Stereotactic Body Radiotherapy for Metastatic and Recurrent Ewing Sarcoma and Osteosarcoma. *Sarcoma*. 2014;2014. <https://api.semanticscholar.org/CorpusID:15216102>
59. Baumann BC, Nagda SN, Kolker JD, et al. Efficacy and safety of stereotactic body radiation therapy for the treatment of pulmonary metastases from sarcoma: a potential alternative to resection. *J Surg Oncol*. 2016;114:65–9.
60. Bölling T, Schuck A, Paulussen M, Dirksen U, Ranft A, Köneemann S, Dunst J, Willich N, Jürgens H. Whole lung irradiation in patients with exclusively pulmonary metastases of Ewing tumors Toxicity analysis and treatment results of the EICESS-92 trial. *Strahlenther Onkol*. 2008;184(4):193–7. <https://doi.org/10.1007/s00066-008-1810-x>. (PMID: 18398583).
61. Casey DL, Alektiar KM, Gerber NK, Wolden SL. Whole lung irradiation for adults with pulmonary metastases from Ewing sarcoma. *Int J Radiat Oncol Biol Phys*. 2014;89(5):1069–75. <https://doi.org/10.1016/j.ijrobp.2014.04.036>. (Epub 2014 Jul 8 PMID: 25035210).
62. Liu AK, Stinauer M, Albano E, Greffe B, Tello T, Maloney K. Local control of metastatic sites with radiation therapy in

- metastatic Ewing sarcoma and rhabdomyosarcoma. *Pediatr Blood Cancer*. 2011;57(1):169–71. <https://doi.org/10.1002/pbc.23063>. (Epub 2011 Mar 24 PMID: 21557463).
63. Bosma SE, Ayua Fiocco M, Gelderblom H, Dijkstra PDS. Prognostic factors for survival in Ewing sarcoma: a systematic review. *Surg Oncol*. 2018;27(4):603–10.
 64. Barker LM, Pendergrass TW, Sanders JE, Hawkins DS. Survival after recurrence of Ewing's sarcoma family of tumors. *J Clin Oncol*. 2005;23(19):4354–62. <https://doi.org/10.1200/JCO.2005.05.105>. (Epub 2005 Mar 21 PMID: 15781881).
 65. Bacci G, Longhi A, Ferrari S, Mercuri M, Barbieri E, Bertoni F, Bacchini P, Picci P. Pattern of relapse in 290 patients with nonmetastatic Ewing's sarcoma family tumors treated at a single institution with adjuvant and neoadjuvant chemotherapy between 1972 and 1999. *Eur J Surg Oncol*. 2006;32(9):974–9. <https://doi.org/10.1016/j.ejso.2006.01.023>. (Epub 2006 Apr 18 PMID: 16621429).
 66. Ferrari S, et al. Response to high-dose ifosfamide in patients with advanced/recurrent Ewing sarcoma. *Pediatr Blood Cancer*. 2009;52(5):581–4.
 67. Rapkin L, Qayed M, Brill P, Martin M, Clark D, George BA, Olson TA, Wasilewski-Masker K, Alazraki A, Katzenstein HM. Gemcitabine and docetaxel (GEMDOX) for the treatment of relapsed and refractory pediatric sarcomas. *Pediatr Blood Cancer*. 2012;59(5):854–8. <https://doi.org/10.1002/pbc.24101>. (Epub 2012 Feb 2 PMID: 22302783).
 68. Farhat R, Raad R, Khoury NJ, Feghaly J, Eid T, Muwakkit S, Abboud M, El-Solh H, Saab R. Cyclophosphamide and topotecan as first-line salvage therapy in patients with relapsed Ewing sarcoma at a single institution. *J Pediatr Hematol Oncol*. 2013;35(5):356–60. <https://doi.org/10.1097/MPH.0b013e318270a343>. (PMID: 23042020).
 69. McCabe MG, et al. Results of the first interim assessment of rEECur, an international randomised controlled trial of chemotherapy for the treatment of recurrent and primary refractory Ewing sarcoma. *J Clin Oncol*. 2019;37(15_suppl):11007–11.
 70. McCabe MG, Kirton L, Khan M, Fenwick N, et al. Results of the second interim assessment of rEECur, an international randomised controlled trial of chemotherapy for the treatment of recurrent and primary refractory Ewing sarcoma (RR-ES). *J Clin Oncol*. 2020;38(15 suppl):11502.
 71. Rasper M, Jabar S, Ranft A, Jürgens H, Amler S, Dirksen U. The value of high-dose chemotherapy in patients with first relapsed Ewing sarcoma. *Pediatr Blood Cancer*. 2014;61(8):1382–6. <https://doi.org/10.1002/pbc.25042>. (Epub 2014 Apr 11 PMID: 24729428).
 72. Italiano A, Mir O, Mathoulin-Pelissier S, Penel N, Piperno-Neumann S, et al. Cabozantinib in patients with advanced Ewing sarcoma or osteosarcoma (CABONE): a multicentre, single-arm, phase 2 trial. *Lancet Oncol*. 2020;21(3):446–55. [https://doi.org/10.1016/S1470-2045\(19\)30825-3](https://doi.org/10.1016/S1470-2045(19)30825-3). (Epub 2020 Feb 17. PMID: 32078813; PMCID: PMC8763616).
 73. Duffaud F, Blay JY, Le Cesne A, Chevreau C, Boudou-Rouquette P, Kalbacher E, Penel N, Perrin C, Laurence V, Bompas E, Saada-Bouazid E, Delcambre C, Bertucci F, Cancel M, Schiffler C, Monard L, Bouvier C, Vidal V, Gaspar N, Chabaud S. Regorafenib in patients with advanced Ewing sarcoma: results of a non-comparative, randomised, double-blind, placebo-controlled, multicentre Phase II study. *Br J Cancer*. 2023;129(12):1940–8. <https://doi.org/10.1038/s41416-023-02413-9>. (Epub 2023 Nov 1. PMID: 37914801; PMCID: PMC10703915).
 74. Grohar P, Ballman KV, Heise R, Glod J, Wedekind MF, Mascarenhas L, Gedminas JM, DuBois SG, Maki RG, Crompton BD, Hayashi M, Peer CJ, Figg WD, Lindenberg ML, Gonzalez EM, Bagatell R, Laetsch TW, Widemann BC, Reinke DK, Chugh R. SARC037: Results of phase I study of trabectedin given as a 1-hour (h) infusion in combination with low dose irinotecan in relapsed/refractory Ewing sarcoma (ES). *J Clin Oncol*. 2023;41(16_suppl):11519.
 75. Heinemann M, Ranft A, Langer T, Jürgens H, Kreyer J, Vieth V, et al. Recurrence of Ewing sarcoma: Is detection by imaging follow-up protocol associated with survival advantage? *Pediatr Blood Cancer*. 2018;65(7):1–7.
 76. Liu F, Zhang Q, Zhu D, Liu F, Li Z, Li J, Wang B, Zhou D, Dong J. Performance of positron emission tomography and positron emission tomography/computed tomography using fluorine-18-fluorodeoxyglucose for the diagnosis, staging, and recurrence assessment of bone sarcoma: a systematic review and meta-analysis. *Medicine (Baltimore)*. 2015;94(36): e1462. <https://doi.org/10.1097/MD.0000000000001462>. (Erratum in: *Medicine (Baltimore)*. 2016 Jan;95(2):e187a. Liu, Fengxia [Added]. Erratum in: *Medicine (Baltimore)*. 2016 Jan 15;95(2):e187a. PMID: 26356700; PMCID: PMC4616630).
 77. Chang KJ, Kong CB, Cho WH, Jeon DG, Lee SY, Lim I, Lim SM. Usefulness of increased 18F- FDG uptake for detecting local recurrence in patients with extremity osteosarcoma treated with surgical resection and endoprosthetic replacement. *Skeletal Radiol*. 2015;44(4):529–37. <https://doi.org/10.1007/s00256-014-2063-7>. (Epub 2014 Nov 29 PMID: 25431093).
 78. Cipriano CA, Jang E, Tyler W. Sarcoma surveillance: a review of current evidence and guidelines. *J Am Acad Orthop Surg*. 2020;28(4):145–56.
 79. Goedhart LM, Leithner A, Ploegmakers JJW, Jutte PC. Follow-Up in Bone Sarcoma Care: A Cross-Sectional European Study. *Sarcoma*. 2020;2020:2040347. <https://doi.org/10.1155/2020/2040347>. PMID: 32675939; PMCID: PMC7350160.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.