Recombinant DNA: Basics and Advanced Applications

Code: 42895
ECTS Credits: 9

<table>
<thead>
<tr>
<th>Degree</th>
<th>Type</th>
<th>Year</th>
<th>Semester</th>
</tr>
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<tbody>
<tr>
<td>4313794 Biochemistry, Molecular Biology and Biomedicine</td>
<td>OT</td>
<td>0</td>
<td>A</td>
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</tbody>
</table>

Contact

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Email: Antonio.Casamayor@uab.cat

Other comments on languages

Language of most of the provided didactic material.

Use of languages

Principal working language: catalan (cat)

Teachers

Joaquín Ariño Carmona
Josep Antoni Biosca Vaqué
Inmaculada Ponte Marull
Salvador Ventura Zamora
Jaume Piñol Ribas
Irantzu Pallarés Goitz
Alicia Roque Cordova

External teachers

Dra. Nerea Roher Armentia
Dra. Núria Sánchez Coll

Prerequisites

For graduates in Biochemistry, Biology, Biomedical Sciences, Genetics, Microbiology, Chemistry, Computer Science, Physics, Veterinary Medicine and Pharmacy

In any case, it is recommended to learn the basic techniques of recombinant DNA.

Objectives and Contextualisation

The main goal of the course is to provide an advanced and rigorous training about a diversity of recombinant DNA techniques, both basic and advanced. So, at the end of the module the student will have achieved a solid knowledge of different techniques involving the manipulation of recombinant DNA currently used in research laboratories as well as profits and limitations.

After completing this module, students will be able to:
1. Understand the methodological procedures and identify current instrumental tools based on recombinant DNA technology to address key issues in many research areas, such as the structure of DNA, the structure and function of chromatin, the evaluation of the expression and regulation, translation, and the subcellular localization of proteins, etc...

2. Design and conduct experiments using the most appropriate for each specific objective experimental recombinant DNA techniques.

3. Analyze and properly interpret and critically evaluate both, own and published in the scientific literature, experimental data.

4. Defining and understanding the specific techniques for specific organisms used as experimental models in research laboratories as well as profits and limitations.

Skills

- Analyse and correctly interpret the molecular mechanisms operating in living beings and identify their applications.
- Analyse research results to obtain new biotechnological or biomedical products to be transferred to society.
- Apply techniques for modifying living beings or parts of these in order to improve pharmaceutical and biotechnological processes and products or develop new products.
- Communicate and justify conclusions clearly and unambiguously to both specialist and non-specialist audiences.
- Continue the learning process, to a large extent autonomously.
- Develop critical reasoning within the subject area and in relation to the scientific or business context.
- Identify and propose scientific solutions to problems in molecular-level biological research and show understanding of the biochemical complexity of living beings.
- Integrate contents in biochemistry, molecular biology, biotechnology and biomedicine from a molecular perspective.
- Solve problems in new or little-known situations within broader (or multidisciplinary) contexts related to the field of study.
- Use and manage bibliography and IT resources related to biochemistry, molecular biology or biomedicine.
- Use scientific terminology to account for research results and present these orally and in writing.
- Work individually and in teams in a multidisciplinary context.

Learning outcomes

1. Analyse, correctly interpret and critically assess both one's own experimental data and those published in the scientific literature.
2. Analyse research results to obtain new biotechnological or biomedical products to be transferred to society.
3. Communicate and justify conclusions clearly and unambiguously to both specialist and non-specialist audiences.
4. Continue the learning process, to a large extent autonomously.
5. Decide on the most appropriate organism to use for each specific need.
6. Design and conduct experiments using the most appropriate experimental techniques of recombinant DNA for each particular objective.
7. Develop critical reasoning within the subject area and in relation to the scientific or business context.
8. Distinguish the bases of the most commonly used standard techniques in molecular biology.
9. Make the necessary changes to improve performance.
10. Solve problems in new or little-known situations within broader (or multidisciplinary) contexts related to the field of study.
11. Understand methodological procedures and the advantages and limitations of currently-used instruments for conducting research in this field (chromatin structure, gene expression and its regulation, mRNA processing, etc.).
12. Use and manage bibliography and IT resources related to biochemistry, molecular biology or biomedicine.
13. Use scientific terminology to account for research results and present these orally and in writing.
14. Work individually and in teams in a multidisciplinary context.

Content

The content of this module is as follows:

1) Introduction to basic molecular biology techniques.

1.1. Principles of gene cloning and DNA analysis.

- Amplification, labeling and detection of nucleic acids.
- Type of vectors, DNA molecular cloning strategies and gene libraries of DNA.
- DNA-directed mutagenesis.

1.2. Applications of gene cloning and DNA analysis for the study of gene expression.

- Techniques for studying gene expression based on DNA microarrays and massive sequencing of DNA (RT-PCR, Run On, microarrays and DNA footprinting, promoter analysis using reporter genes, mRNAs massive sequencing December, ChIP-Seq, GRO-Seq, etc.).

2) Characteristics of commonly used model organisms.

3) Techniques for the study of epigenetic mechanisms that regulate chromatin structure and its role in the replication, transcription and repair of eukaryotic DNA.

- Determination of heterochromatin and euchromatin regions (microscopic / sensitivity to nucleases / density gradients / chromatin precipitation curves, etc.).
- Modifications of histones (histone code). ChIP, ChIP-chip, ChIP-seq and others.
- Methods for characterizing high and low resolution nucleosome positioning and identification of sites of hypersensitivity to nucleases (End labeling, in vivo footprinting, LM-PCR, etc.).
- Methodology for the study of chromatin remodeling complexes.

4) Modification of genomes and gene silencing (RNA antisense techniques, KO, ribozymes, GM, using adjustable promoters, etc.).

5) Protein Expression. Characteristics of various protein expression systems (types of vectors, promoters, organizations, etc.). Study of the intracellular localization of proteins.

6) Detection of protein-protein interaction (double hybrid protein chips, FRET, etc.) and Interactomics.

7) Applications of recombinant DNA technology in industry and medicine (diagnosis, antibody engineering, metabolic, etc.).

8) Presentation and defense of a bibliographic work.

Methodology

Part of the teaching will be in the classroom and will comprise lectures, laboratory work and assistance in the defense of bibliographical works as detailed below. It is excluded the attendance at scheduled seminars, part of Module 2 Advanced seminars in Biochemistry and Molecular Biology.
Face class / Dirigidas (52 h)
- Lectures and seminars 32 h
- Laboratory work 16 h
- Presentation and defense of bibliographical 4 pm

Another part will be self-study by students, including the execution of tests and exercises throughout the course.

Autonomous (120 h)
- Student Self Study: 93 pm
- Theoretical and practical tests throughout the course (resolution of cases and problems): 25h

The preparation by the students of a bibliographic work is the main activity supervised.

### Activities

<table>
<thead>
<tr>
<th>Title</th>
<th>Hours</th>
<th>ECTS</th>
<th>Learning outcomes</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Type: Directed</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Classroom lectures</td>
<td>32</td>
<td>1.28</td>
<td>1, 5, 7, 8, 11, 9, 4</td>
</tr>
<tr>
<td>Laboratory teaching</td>
<td>16</td>
<td>0.64</td>
<td>1, 6, 8, 11, 10, 3, 4, 14, 12</td>
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<tr>
<td>Supervised</td>
<td>4</td>
<td>0.16</td>
<td>2, 1, 7, 8, 11, 10, 4, 14, 12, 13</td>
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<tr>
<td><strong>Type: Supervised</strong></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Preparation of a bibliographic work</td>
<td>55</td>
<td>2.2</td>
<td>2, 1, 7, 8, 11, 10, 4, 14, 12, 13</td>
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<tr>
<td><strong>Type: Autonomous</strong></td>
<td></td>
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<tr>
<td>Autonomous learning</td>
<td>67.5</td>
<td>2.7</td>
<td>2, 1, 5, 7, 6, 8, 11, 10, 4, 14, 12</td>
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<tr>
<td>Theoretical and practical tests throughout the course (resolution of cases and problems)</td>
<td>25</td>
<td>1</td>
<td>1, 5, 7, 6, 8, 11, 9, 10, 3, 4, 14, 12</td>
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### Evaluation

The evaluation system described is subject to an attendance of at least 80% of the sessions.
Attendance of at least 80% provides the maximum obtained mark (100 %).

It will be considered "not evaluable" when the number of tests / assignments / activities assessed made by the student do not allow to achieve a minimum overall score of 5.0, assuming that all tests had obtained the highest rating. Thus, assuming a total of 36 theoretical sessions, attending to 29 or more sessions will represent 100% of the score obtained. Attendance to 14 sessions represents 50% of the mark. Attendance to less than 14 sessions will be considered as "non-assessable"

### Evaluation activities

<table>
<thead>
<tr>
<th>Title</th>
<th>Weighting</th>
<th>Hours</th>
<th>ECTS</th>
<th>Learning outcomes</th>
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<tbody>
<tr>
<td>Completion of laboratory sessions</td>
<td>10%</td>
<td>1</td>
<td>0.04</td>
<td>2, 1, 5, 6, 8, 11, 9, 10, 3, 4, 14</td>
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<td>Exercises eventually presented by teachers and theoretical and practical tests throughout the course</td>
<td>40%</td>
<td>0.5</td>
<td>0.02</td>
<td>2, 1, 5, 7, 6, 8, 11, 9, 10, 3, 4, 14, 12, 13</td>
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<td>Oral presentation of the bibliographic work</td>
<td>40%</td>
<td>24</td>
<td>0.96</td>
<td>2, 1, 5, 7, 6, 8, 11, 10, 14, 12, 13</td>
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</tbody>
</table>

### Bibliography


* Current Protocols in Molecular Biology

Ausubel et al.

J. Willey, 2003

* Gene Cloning and DNA Analysis: An Introduction.
Recombinant DNA: Basics and Advanced Applications 2015 - 2016

A. Brown.


* Lewin's GENES XI.


* Molecular Biotechnology: Principles and Applications of Recombinant DNA.

Bernard R. Glick, Jack J. Pasternak, Cheryl L. Patten.


* Next-Generation DNA Sequencing Informatics.

Stuart M. Brown


* Diverses revisions en revistes com: Current Opinion in Structural Biology, Trends in Biochemical Sciences, Trends in Biotechnology, Nature Biotechnology, Nature Methods, etc.