Recombinant DNA: Basics and Advanced Applications

Code: 42895
ECTS Credits: 9

<table>
<thead>
<tr>
<th>Degree</th>
<th>Type</th>
<th>Year</th>
<th>Semester</th>
</tr>
</thead>
<tbody>
<tr>
<td>4313794 Biochemistry, Molecular Biology and Biomedicine</td>
<td>OT</td>
<td>0</td>
<td>A</td>
</tr>
</tbody>
</table>

**Contact**

Name: Alicia Roque Cordova
Email: Alicia.Roque@uab.cat

**Use of languages**

Principal working language: **catalan** (cat)

**Other comments on languages**

Language of most of the provided didactic material.

**Teachers**

Joaquín Ariño Carmona
Josep Antoni Biosca Vaqué
Inmaculada Ponte Marull
Antonio Casamayor Gracia
Jaume Piñol Ribas
Nerea Roher Armentia
Irantzu Pallarés Goitiz
Alicia Roque Cordova

**External teachers**

Jordi Moreno Romero
Marcus Buschbeck
Núria Sánchez Coll

**Prerequisites**

For graduates in Biochemistry, Biology, Biomedical Sciences, Genetics, Microbiology, Chemistry, Computer Science, Physics, Veterinary Medicine and Pharmacy

In any case, it is recommended to learn the basic techniques of recombinant DNA.

**Objectives and Contextualisation**
The main goal of the course is to provide an advanced and rigorous training about a diversity of recombinant DNA techniques, both basic and advanced. So, at the end of the module the student will have achieved a solid knowledge of different techniques involving the manipulation of recombinant DNA currently used in research laboratories as well as profits and limitations.

After completing this module, students will be able to:

1. Understand the methodological procedures and identify current instrumental tools based on recombinant DNA technology to address key issues in many research areas, such as the structure of DNA, the structure and function of chromatin, the evaluation of the expression and regulation, translation, and the subcellular localization of proteins, etc…

2. Design and conduct experiments using the most appropriate for each specific objective experimental recombinant DNA techniques.

3. Analyze and properly interpret and critically evaluate both, own and published in the scientific literature, experimental data.

4. Defining and understanding the specific techniques for specific organisms used as experimental models in research laboratories as well as profits and limitations.

**Skills**

- Analyse and correctly interpret the molecular mechanisms operating in living beings and identify their applications.
- Analyse research results to obtain new biotechnological or biomedical products to be transferred to society.
- Apply techniques for modifying living beings or parts of these in order to improve pharmaceutical and biotechnological processes and products or develop new products.
- Communicate and justify conclusions clearly and unambiguously to both specialist and non-specialist audiences.
- Continue the learning process, to a large extent autonomously.
- Develop critical reasoning within the subject area and in relation to the scientific or business context.
- Identify and propose scientific solutions to problems in molecular-level biological research and show understanding of the biochemical complexity of living beings.
- Integrate contents in biochemistry, molecular biology, biotechnology and biomedicine from a molecular perspective.
- Solve problems in new or little-known situations within broader (or multidisciplinary) contexts related to the field of study.
- Use and manage bibliography and IT resources related to biochemistry, molecular biology or biomedicine.
- Use scientific terminology to account for research results and present these orally and in writing.
- Work individually and in teams in a multidisciplinary context.

**Learning outcomes**

1. Analyse research results to obtain new biotechnological or biomedical products to be transferred to society.
2. Analyse, correctly interpret and critically assess both one's own experimental data and those published in the scientific literature.
3. Communicate and justify conclusions clearly and unambiguously to both specialist and non-specialist audiences.
4. Continue the learning process, to a large extent autonomously.
5. Decide on the most appropriate organism to use for each specific need.
6. Design and conduct experiments using the most appropriate experimental techniques of recombinant DNA for each particular objective.
7. Develop critical reasoning within the subject area and in relation to the scientific or business context.
8. Distinguish the bases of the most commonly used standard techniques in molecular biology.
9. Make the necessary changes to improve performance.
10. Solve problems in new or little-known situations within broader (or multidisciplinary) contexts related to the field of study.
11. Understand methodological procedures and the advantages and limitations of currently-used instruments for conducting research in this field (chromatin structure, gene expression and its regulation, mRNA processing, etc.).
12. Use and manage bibliography and IT resources related to biochemistry, molecular biology or biomedicine.
13. Use scientific terminology to account for research results and present these orally and in writing.
14. Work individually and in teams in a multidisciplinary context.

Content

The content of this module is as follows:

1) Introduction to basic molecular biology techniques.

1.1. Principles of gene cloning and DNA analysis.

- Amplification, labeling and detection of nucleic acids.
- Type of vectors, DNA molecular cloning strategies and gene libraries of DNA.
- DNA-directed mutagenesis.

1.2. Applications of gene cloning and DNA analysis for the study of gene expression.

- Techniques for studying gene expression based on DNA microarrays and massive sequencing of DNA (RT-PCR, Run On, microarrays and DNA footprinting, promoter analysis using reporter genes, mRNAs massive sequencing December, ChIP-Seq, GRO-Seq, etc.).

2) Characteristics of commonly used model organisms.

3) Techniques for the study of epigenetic mechanisms that regulate chromatin structure and its role in the replication, transcription and repair of eukaryotic DNA.

- Determination of heterochromatin and euchromatin regions (microscopic / sensitivity to nucleases / density gradients / chromatin precipitation curves, etc.).
- Modifications of histones (histone code). ChIP, ChIP-chip, ChIP-seq and others.
- Methods for characterizing high and low resolution nucleosome positioning and identification of sites of hypersensitivity to nucleases (End labeling, in vivo footprinting, LM-PCR, etc.).
- Methodology for the study of chromatin remodeling complexes.
- Techniques for studying the 3D organization of the genome (Hi-C).
- Applications of epigenetics in the analysis of genomic imprinting in plants.

4) Modification of genomes and gene silencing (RNA antisense techniques, KO, ribozymes, GM, using adjustable promoters, etc.).

5) Protein Expression. Characteristics of various protein expression systems (types of vectors, promoters, organizations, etc.). Study of the intracellular localization of proteins.

6) Detection of protein-protein interaction (double hybrid protein chips, FRET, etc.) and Interactomics.
7) Applications of recombinant DNA technology in industry and medicine (diagnosis, antibody engineering, metabolic, etc.).

8) Presentation and defense of a bibliographic work.

9) Case reports.

10) Practical sessions: 24h of laboratory work divided in 8 sessions of variable length (2-4h/session). Yeast two hybrid, PCR, RNA-seq analysis, chromatin immunoprecipitation, q-PCR, sequence databases and analysis tools.

11) Experts seminaries.

**Methodology**

Part of the teaching will be in the classroom and will comprise lectures, laboratory work and assistance in the defense of bibliographical works as detailed below. It is excluded the attendance at scheduled seminars, part of Module 2 Advanced seminars in Biochemistry and Molecular Biology.

Face class / Directed activities (60 h)

- Lectures and seminars 32 h
- Laboratory work 24 h
- Presentation and defense of bibliographical 4 h

Another part will be self-study by students, including the execution of tests and exercises throughout the course.

**Autonomous**

- Student Self Study: 67.5 h
- Theoretical and practical tests throughout the course (resolution of cases and problems): 24 h

The preparation by the students of a bibliographic work is the main activity supervised.

**Activities**

<table>
<thead>
<tr>
<th>Title</th>
<th>Hours</th>
<th>ECTS</th>
<th>Learning outcomes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Type: Directed</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Classroom lectures</td>
<td>32</td>
<td>1.28</td>
<td>2, 5, 7, 8, 11, 9, 4</td>
</tr>
<tr>
<td>Laboratory teaching</td>
<td>24</td>
<td>0.96</td>
<td>2, 6, 8, 11, 10, 3, 4, 14, 12</td>
</tr>
<tr>
<td>Supervised</td>
<td>4</td>
<td>0.16</td>
<td>1, 2, 7, 8, 11, 10, 4, 14, 12</td>
</tr>
</tbody>
</table>
### Type: Supervised

<table>
<thead>
<tr>
<th>Activity</th>
<th>Weighting</th>
<th>Hours</th>
<th>ECTS</th>
<th>Learning outcomes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Preparation of a bibliographic work</td>
<td>47</td>
<td>1.88</td>
<td>0.02</td>
<td>1, 2, 7, 8, 11, 10, 4, 14, 12, 13</td>
</tr>
</tbody>
</table>

### Type: Autonomous

<table>
<thead>
<tr>
<th>Activity</th>
<th>Weighting</th>
<th>Hours</th>
<th>ECTS</th>
<th>Learning outcomes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Autonomous learning</td>
<td>67.5</td>
<td>2.7</td>
<td>0.08</td>
<td>1, 2, 5, 7, 6, 8, 11, 10, 4, 14, 12</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Activity</th>
<th>Weighting</th>
<th>Hours</th>
<th>ECTS</th>
<th>Learning outcomes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Theoretical and practical tests throughout the course (resolution of cases and problems)</td>
<td>24</td>
<td>0.96</td>
<td>0.08</td>
<td>2, 5, 7, 6, 8, 11, 9, 10, 3, 4, 14, 12</td>
</tr>
</tbody>
</table>

### Evaluation

In order to pass the module the mark obtained by the weighted average of the different activities must be at least of 5 points.

**A retake exam will be available for those students who didn't get an average mark greater than 5 points.** To be eligible for the retake process, the student should have been previously evaluated in a set of activities equaling at least two thirds of the final score of the course or module. Thus, the student will be graded as "No Avaluable" if the weight in all conducted evaluation activities is less than 67% of the final score.

**Important:** If plagiarism is detected in any of the works submitted, the student will fail the whole module.

### Evaluation activities

<table>
<thead>
<tr>
<th>Title</th>
<th>Weighting</th>
<th>Hours</th>
<th>ECTS</th>
<th>Learning outcomes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Oral presentation of the bibliographic work</td>
<td>40%</td>
<td>0.5</td>
<td>0.02</td>
<td>1, 2, 5, 7, 6, 8, 11, 9, 10, 3, 4, 14, 12, 13</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Title</th>
<th>Weighting</th>
<th>Hours</th>
<th>ECTS</th>
<th>Learning outcomes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Practical sessions</td>
<td>30%</td>
<td>2</td>
<td>0.08</td>
<td>1, 2, 5, 6, 8, 11, 9, 10, 3, 4, 14</td>
</tr>
</tbody>
</table>

---

5
Resolution of exercises presented by teachers, throughout the course

<table>
<thead>
<tr>
<th>Percentage</th>
<th>Percentage Value</th>
<th>Exercises</th>
</tr>
</thead>
<tbody>
<tr>
<td>30 %</td>
<td>24</td>
<td>0.96</td>
</tr>
<tr>
<td></td>
<td>1, 2, 5, 7, 6, 8, 11, 10, 14, 12, 13</td>
<td></td>
</tr>
</tbody>
</table>

Bibliography


* Current Protocols in Molecular Biology
Ausubel et al.
J. Wiley, 2003

* Gene Cloning and DNA Analysis: An Introduction.
A. Brown.

* Lewin's GENES XI.

* Molecular Biotechnology: Principles and Applications of Recombinant DNA.
Bernard R. Glick, Jack J. Pasternak, Cheryl L. Patten.

* Next-Generation DNA Sequencing Informatics.
Stuart M. Brown

* Diverses revisions en revistes com: Current Opinion in Structural Biology, Trends in Biochemical Sciences, Trends in Biotechnology, Nature Biotechnology, Nature Methods, etc.