

## CONCLUSIONES

### OBJETIVO PRINCIPAL

-El timo humano expresa transcritos de todas las cadenas que conforman el receptor de acetilcolina, de todos los autoantígenos involucrados en esclerosis múltiple excepto MOG, y de los antígenos prostáticos PSA y PSM

-Los autoantígenos del sistema nervioso central no guardan una expresión restringida como se pensaba, excepto para MOG, que es el único antígeno realmente secuestrado en el SNC

-Los autoantígenos presentan tres patrones diferentes de expresión intratímica, a saber:

*Variable:* con marcadas diferencias interindividuales en los niveles de expresión

*Constitutivo:* paralelo a la expresión de genes constitutivos, como GAPDH

*Dependiente del desarrollo:* expresión relacionada con la edad

### OBJETIVOS SECUNDARIOS

#### OBJETIVO I

La expresión de los autoantígenos en el timo humano no guarda una relación directa con la edad, excepto para la cadena gamma del receptor fetal de acetilcolina.

#### OBJETIVO II

Las formas de expresión de los autoantígenos en el timo humano son similares a las formas de expresión en el órgano diana, excepto PLP, que se expresa como una variante mas corta de *splicing* alternativo. Esta conclusión es válida para nuestro sistema de amplificaciones, pero no se descarta que en otras regiones de los genes de los autoantígenos existan formas de *splicing* no detectadas por nuestros cebadores.

#### OBJETIVO III

Los experimentos de detección de antígenos en las diferentes fracciones celulares y en las microdisecciones tímicas indican que las células responsables de la transcripción de los autoantígenos en el timo son las células epiteliales, predominantemente de localización medular.



---

**BIBLIOGRAFÍA**

- Alam SM, Travers PJ, Wung JL, Nasholds W, Redpath S, Jameson SC, et al. T-cell receptor affinity and thymocyte positive selection. *Nature* 1996; 381: 616-620.
- Albert LJ, Inman RD. Molecular mimicry and autoimmunity. *N Engl J Med* 1999; 341: 2068-2074.
- Alexander R, Brady F, Ponniah S. Autoimmune prostatitis: evidence of T cell reactivity with normal prostatic proteins. *Urology* 1997; 50: 893-9.
- Alferink J, Aigner S, Reibke R, Hammerling GJ, Arnold B. Peripheral T-cell tolerance: the contribution of permissive T-cell migration into parenchymal tissues of the neonate. *Immunol Rev* 1999; 169: 255-61.
- Allore R, Friend W, O'Hanlon D, Neilson K, Baumal R, Dunn R, et al. Cloning and expression of the human S100 beta gene. *J Biol Chem* 1990; 265: 15537-43.
- Anderson AC, Nicholson LB, Legge KL, Turchin V, Zaghrouani H, Kuchroo VK. High frequency of autoreactive myelin proteolipid protein-specific T cells in the periphery of naive mice: mechanisms of selection of the self-reactive repertoire. *J Exp Med* 2000; 191: 761-70.
- Antonia SJ, Geiger T, Miller J, Flavell RA. Mechanisms of immune tolerance induction through the thymic expression of a peripheral tissue-specific protein. *Int Immunol* 1995; 7: 715-25.
- Archelos JJ, Storch MK, Hartung HP. The role of B cells and autoantibodies in multiple sclerosis. *Ann Neurol* 2000; 47: 694-706.
- Armengol MP, Juan M, Lucas-Martin A, Fernandez-Figueras MT, Jaraquemada D, Gallart T, et al. Thyroid autoimmune disease: Demonstration of thyroid antigen specific B cells and recombination-activating gene expression in chemokine containing active intrathyroidal germinal centers. *Am J Pathol* 2001.
- Baimbridge KG, Celio MR, Rogers JH. Calcium-binding proteins in the nervous system. *Trends Neurosci* 1992; 15: 303-8.
- Bajramovic JJ, Plomp AC, Goes A, Koevoets C, Newcombe J, Cuzner ML, et al. Presentation of alpha B-crystallin to T cells in active multiple sclerosis lesions: an early event following inflammatory demyelination. *J Immunol* 2000; 164: 4359-66.
- Balasa B, Deng C, Lee J, Bradley LM, Dalton DK, Christadoss P, et al. Interferon gamma (IFN-gamma) is necessary for the genesis of acetylcholine receptor-induced clinical experimental autoimmune myasthenia gravis in mice. *J Exp Med* 1997; 186: 385-91.

- Balasa B, Deng C, Lee J, Christadoss P, Sarvetnick N. The Th2 cytokine IL-4 is not required for the progression of antibody- dependent autoimmune myasthenia gravis. *J Immunol* 1998; 161: 2856-62.
- Barkas T, Gabriel JM, Mauron A, Hughes GJ, Roth B, Alliod C, et al. Monoclonal antibodies to the main immunogenic region of the nicotinic acetylcholine receptor bind to residues 61-76 of the alpha subunit. *J Biol Chem* 1988; 263: 5916-20.
- Beeson D, Morris A, Vincent A, Newsom-Davis J. The human muscle nicotinic acetylcholine receptor alpha-subunit exist as two isoforms: a novel exon. *Embo J* 1990; 9: 2101-6.
- Bell D, Young JW, Banchereau J. Dendritic cells. *Adv Immunol* 1999; 72: 255-324.
- Bellone M, Ostlie N, Lei S, Wu XD, Conti-Tronconi BM. The I-A<sup>bmi2</sup> mutation, which confers resistance to experimental myasthenia gravis, drastically affects the epitope repertoire of murine CD4<sup>+</sup> cells sensitized to nicotinic acetylcholine receptor. *J Immunol* 1991; 147: 1484-91.
- Bernard CC, Johns TG, Slavin A, Ichikawa M, Ewing C, Liu J, et al. Myelin oligodendrocyte glycoprotein: a novel candidate autoantigen in multiple sclerosis. *J Mol Med* 1997; 75: 77-88.
- Bernard CC, Kerlero de Rosbo N. Multiple sclerosis: an autoimmune disease of multifactorial etiology. *Curr Opin Immunol* 1992; 4: 760-5.
- Beroukhim R, Unwin N. Three-dimensional location of the main immunogenic region of the acetylcholine receptor. *Neuron* 1995; 15: 323-31.
- Bevan MJ. Cross-priming for a secondary cytotoxic response to minor H antigens with H-2 congenic cells which do not cross-react in the cytotoxic assay. *J Exp Med* 1976; 143: 1283-1288.
- Boison D, Bussow H, D. Du, Muller H, Stoffel W. Adhesive properties of proteolipid protein are responsible for the compaction of CNS myelin sheaths. *J Neurosci* 1995; 15: 5502-13.
- Boison D, Stoffel W. Disruption of the compacted myelin sheath of axons of the central nervous system in proteolipid protein-deficient mice. *Proc Natl Acad Sci USA* 1994; 91: 11709-13.
- Bornemann A, Kirchner T. Thymic myoid cell turnover in myasthenia gravis patients and in normal controls. *Virchows Arch* 1998; 432: 357-61.
- Brunner C, Lassmann H, Waehneltd T, Matthieu JM, Linington C. Differential ultrastructural localization of myelin basic protein, myelin/oligodendrocyte glycoprotein, and 2', 3'-cyclic nucleotide 3'-phosphodiesterase in the CNS of adult rats. *J Neurochem* 1989; 52: 296-304.

- Buckingham JM, Howard FM, Jr., Bernatz PE, Payne WS, Harrison EG, Jr., O'Brien PC, et al. The value of thymectomy in myasthenia gravis: a computer-assisted matched study. *Ann Surg* 1976; 184: 453-8.
- Burges J, Wray DW, Pizzighella S, Hall Z, Vincent A. A myasthenia gravis plasma immunoglobulin reduces miniature endplate potentials at human endplates in vitro. *Muscle Nerve* 1990; 13: 407-13.
- Campagnoni AT. Molecular biology of myelin proteins from the central nervous system. *J Neurochem* 1988; 51: 1-14.
- Cannella B, Raine CS. The adhesion molecule and cytokine profile of multiple sclerosis lesions. *Ann Neurol* 1995; 37: 424-35.
- Carbone F, Kurts C, Bennet S, Miller JF, WR H. Cross-presentation: a general mechanism for CTL immunity and tolerance. *Immunology Today* 1998; 19.
- Charukamnoetkanok P, Fukushima A, Whitcup SM, Gery I, Egwuagu CE. Expression of ocular autoantigens in the mouse thymus. *Curr Eye Res* 1998; 17: 788-92.
- Chen Y, Kuchroo VK, Inobe J, Hafler DA, Weiner HL. Regulatory T cell clones induced by oral tolerance: suppression of autoimmune encephalomyelitis. *Science* 1994; 265: 1237-1240.
- Chomczynski P, Sacchi N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal Biochem* 1987; 162: 156-9.
- Cleutjens K, van der Korput H, van Eekelen C, van Rooij H, Faber P, Trapman J. An androgen response element in a far upstream enhancer region is essential for high, androgen-regulated activity of the prostate -specific antigen promoter. *Mol Endocrinol* 1997; 11: 148-61.
- della Gaspera B, Pham-Dinh D, Roussel G, Nussbaum JL, Dautigny A. Membrane topology of the myelin/oligodendrocyte glycoprotein. *Eur J Biochem* 1998; 258: 478-84.
- Derbinski J, Schulte A, Kyewski B, Klein TC. Promiscuous gene expression in medullary thymic epithelial cells mirrors the peripheral self. *Nat Immunol* 2001.
- Diehl HJ, Schaich M, Budzinski RM, Stoffel W. Individual exons encode the integral membrane domains of human myelin proteolipid proteins. *Proc Natl Acad Sci USA* 1986; 83: 9807-11.
- Djabiri F, Caillat-Zucman S, Gajdos P, Jais JP, Gomez L, Khalil I, et al. Association of the AChR $\alpha$ -subunit gene (CHRNA), DQA1\*0101, and the DR3 haplotype in myasthenia gravis. Evidence for a three-gene disease model in a subgroup of patients. *J Autoimmun* 1997a; 10: 407-13.

- Djabiri F, Gajdos P, Eymard B, Gomez L, Bach JF, Garchon HJ. No evidence for an association of AChR beta-subunit gene (CHRNA1) with myasthenia gravis. *J Neuroimmunol* 1997b; 78: 86-9.
- Ebers GC, Sadovnick AD, Risch NJ. A genetic basis for familial aggregation in multiple sclerosis. Canadian Collaborative Study Group. *Nature* 1995; 377: 150-1.
- Egwuagu CE, Charukamnoetkanok P, Gery I. Thymic expression of autoantigens correlates with resistance to autoimmune disease. *J Immunol* 1997; 159: 3109-12.
- Fairchild PJ, Wildgoose R, Atherton E, Webb S, Wraith DC. An autoantigenic T cell epitope forms unstable complexes with class II MHC: a novel route for escape from tolerance induction. *Int Immunol* 1993; 5: 1151-58.
- Ferber I, Schonrich G, Schenkel J, Mellor AL, Hammerling GJ, Arnold B. Distinct levels of peripheral T cell tolerance induced by different doses of antigen. *Science* 1994; 263: 274-276.
- Foerster I. Controlling autorreactivity of CD4 T cells by local tolerance induction. *Dev Immunol* 1998; 6: 89-94.
- Folch J, Lees MB. Proteolipids, a new type of tissue lipoproteins, their isolation from brain. *J Biol Chem* 1951; 191: 807-817.
- Fowell D, Mason D. Evidence that the T cell repertoire of normal rats contains cells with the potential to cause diabetes. Characterization of the CD4+ T cell subset that inhibits this autoimmune potential. *J Exp Med* 1993; 177: 627-636.
- Fritz RB, Zhao ML. Thymic expression of myelin basic protein (MBP). Activation of MBP-specific T cells by thymic cells in the absence of exogenous MBP. *J Immunol* 1996; 157: 5249-53.
- Fuchs S, Neiro D, Tarrab-Hazdai R, Yaar G. Strain differences in the autoimmune response of mice to acetylcholine receptors. *Nature* 1976; 263: 329-30.
- Garchon HJ, Djabiri F, Viard JP, Gajdos P, Bach JF. Involvement of human muscle acetylcholine receptor alpha-subunit gene (CHRNA) in susceptibility to myasthenia gravis. *Proc Natl Acad Sci U S A* 1994; 91: 4668-72.
- Gardinier MV, Amiguet P, Linington C, Matthieu JM. Myelin/oligodendrocyte glycoprotein is a unique member of the immunoglobulin superfamily. *J Neurosci Res* 1992; 33: 177-187.
- Genain CP, Cannella B, Hauser SL, Raine CS. Identification of autoantibodies associated with myelin damage in multiple sclerosis. *Nat Med* 1999; 5: 170-5.
- Geuder KI, Marx A, Witzemann V, Schalke B, Toyka K, Kirchner T, et al. Pathogenetic significance of fetal-type acetylcholine receptors on thymic myoid cells in myasthenia gravis. *Dev Immunol* 1992; 2: 69-75.

- Greer JM, Sobel RA, Sette A, Southwood S, Lees MB, Kuchroo VK. Immunogenic and encephalitogenic epitope clusters of myelin proteolipid protein. *J Immunol* 1996; 156: 371-9.
- Gregorakis A, Holmes E, Murphy G. Prostate-specific membrane antigen: current and future utility. *Semin Urol Oncol* 1998; 16: 2-12.
- Gu D, Wogensen L, Calcutt NA, Xia C, Zhu S, Merlie JP, et al. Myasthenia gravis-like syndrome induced by expression of interferon gamma in the neuromuscular junction. *J Exp Med* 1995; 181: 547-57.
- Gu Y, Hall ZW. Characterization of acetylcholine receptor subunits in developing and in denervated mammalian muscle. *J Biol Chem* 1988; 263: 12878-85.
- Guyon T, Levasseur P, Truffault F, Cottin C, Gaud C, Berrih-Aknin S. Regulation of acetylcholine receptor alpha subunit variants in human myasthenia gravis. Quantification of steady-state levels of messenger RNA in muscle biopsy using the polymerase chain reaction. *J Clin Invest* 1994; 94: 16-24.
- Haase CG, Guggenmos J, Brehm U, Andersson M, Olsson T, Reindl M, et al. The fine specificity of the myelin oligodendrocyte glycoprotein autoantibody response in patients with multiple sclerosis and normal healthy controls. *J Neuroimmunol* 2001; 114: 220-5.
- Hafler DA, Weiner HL. Immunologic mechanisms and therapy in multiple sclerosis. *Immunol Rev* 1995; 144: 75-107.
- Hammerling GJ, Schonrich G, Ferber I, Arnold B. Peripheral tolerance as a multi-step mechanism. *Immunol Rev* 1993; 133: 93-104.
- Hanahan D. Peripheral-antigen-expressing cells in thymic medulla: factors in self-tolerance and autoimmunity. *Curr Opin Immunol* 1998; 10: 656-62.
- Hara H, Hayashi K, Ohta K, Itoh N, Ohta M. Nicotinic acetylcholine receptor mRNAs in myasthenic thymuses: association with intrathymic pathogenesis of myasthenia gravis. *Biochem Biophys Res Commun* 1993; 194: 1269-75.
- Hara Y, Ueno S, Uemichi T, Takahashi N, Yorifuji S, Fujii Y, et al. Neoplastic epithelial cells express alpha-subunit of muscle nicotinic acetylcholine receptor in thymomas from patients with myasthenia gravis. *FEBS Lett* 1991; 279: 137-40.
- Harrington CJ, Paez A, Hunkapiller T, Mannikko V, Brabb T, Ahearn M, et al. Differential tolerance is induced in T cells recognizing distinct epitopes of myelin basic protein. *Immunity* 1998; 8: 571-80.
- Hawke S, Matsuo H, Nicolle M, Malcherek G, Melms A, Willcox N. Autoimmune T cells in myasthenia gravis: heterogeneity and potential for specific immunotargeting. *Immunol Today* 1996; 17: 307-11.

- Haynes BF, Scearce RM, Lobach DF, Hensley LL. Phenotypic characterization and ontogeny of mesodermal-derived and endocrine epithelial components of the human thymic microenvironment. *J Exp Med* 1984; 159: 1149-68.
- Heath VL, Moore NC, Parnell SM, Mason D. Intrathymic expression of genes involved in organ-specific autoimmune diseases. *J Autoimmun* 1998; 11: 309-318.
- Heino M, Peterson P, Kudoh J, Nagamine K, Lagerstedt A, Ovod V, et al. Autoimmune regulator is expressed in the cells regulating immune tolerance in thymus medulla. *Biochem Biophys Res Commun* 1999; 257: 821-25.
- Hilton AA, Slavin AJ, Hilton DJ, Bernard CC. Characterization of cDNA and genomic clones encoding human myelin oligodendrocyte glycoprotein. *J Neurochem* 1995; 65: 309-18.
- Holz A, Bielekova B, Martin R, Oldstone MB. Myelin-associated oligodendrocytic basic protein: identification of an encephalitogenic epitope and association with multiple sclerosis. *J Immunol* 2000; 164: 1103-9.
- Horton RM, Manfredi AA, Conti-Tronconi BM. The 'embryonic' gamma subunit of the nicotinic acetylcholine receptor is expressed in adult extraocular muscle. *Neurology* 1993; 43: 983-6.
- Huseby ES, Goverman J. Tolerating the nervous system: a delicate balance. *J Exp Med* 2000; 191: 757-60.
- Huseby ES, Ohlen C, Goverman J. Cutting edge: myelin basic protein-specific cytotoxic T cell tolerance is maintained in vivo by a single dominant epitope in H-2k mice. *J Immunol* 1999; 163: 1115-8.
- Huseby ES, Sather B, Huseby PG, Goverman J. Age-dependent t cell tolerance and autoimmunity to myelin basic protein. *Immunity* 2001; 14: 471-81.
- Ichikawa M, Koh CS, Inaba Y, Seki C, Inoue A, Itoh M, et al. IgG subclass switching is associated with the severity of experimental autoimmune encephalomyelitis induced with myelin oligodendrocyte glycoprotein peptide in NOD mice. *Cell Immunol* 1999; 191: 97-104.
- Iglesias A, Bauer J, Litzemberger T, Schubart A, Linington C. T and B cell responses to Myelin Oligodendrocyte Glycoprotein in Experimental Autoimmune Encephalomyelitis and Multiple Sclerosis. *Glia* 2001; 36: 220-234.
- Israeli R, Miller W, Su L. Sensitive nested reverse transcription polymerase chain reaction detection of circulating prostate tumor cells: comparison of prostate-specific membrane antigen and prostate-specific antigen-based assays. *Cancer Res* 1994a; 54: 6306-10.



- Israeli R, Powell C, Corr J. Expression of the prostate-specific membrane antigen. *Cancer Res* 1994b; 54: 1807-11.
- Israeli R, Powell C, Fair W. Molecular cloning of a complementary DNA encoding a prostate-specific membrane antigen. *Cancer Res* 1993; 53: 227-30.
- Jensen R, Marshak D, Anderson C, Lukas T, Watterson D. Characterization of human brain S100 protein fraction: aminoacid sequence of S100 $\beta$ . *J Neurochem* 1985; 45: 700-6.
- Jolicoeur C, Hanahan D, Smith KM. T cell tolerance toward a transgenic beta-cell antigen and transcripcion of endogenous pancreatic genes in thymus. *Proc Natl Acad Sci U S A* 1994; 91: 6707-6711.
- Kaminski HJ, Fenstermaker RA, Abdul-Karim FW, Clayman J, Ruff RL. Acetylcholine receptor subunit gene expression in thymic tissue. *Muscle Nerve* 1993; 16: 1332-7.
- Kappler JW, Roehm N, Marrack P. T cell tolerance by clonal elimination in the thymus. *Cell* 1987; 49: 273-80.
- Karpuj M, Steinman L, Oksenberg JR. Multiple sclerosis: a polygenic disease involving epistatic interactions, germline rearrangements and enviromental effects. *Neurogenetics* 1997; 1: 21-8.
- Katzberg HD, Aziz T, Oger J. In myasthenia gravis cells from atrophic thymus secrete acetylcholine receptor antibodies. *Neurology* 2001; 56: 572-3.
- Kaul R, Shenoy M, Christadoss P. The role of major histocompatibility complex genes in myasthenia gravis and experimental autoimmune myasthenia gravis pathogenesis. *Adv Immunol* 1994; 4: 387-402.
- Kennedy MK, Tan LJ, Dal Canto MC, Tuohy VK, Lu ZJ, Trotter JL, et al. Inhibition of murine relapsing experimental autoimmune encephalomyelitis by immune tolerance to proteolipid protein and its encephalitogenic peptides. *J Immunol* 1990; 144: 909-15.
- Kerlero de Rosbo N, Hoffman M, Mendel I, Yust I, Kaye J, Bakimer R, et al. Predominance of the autoimmune response to myelin oligodendrocyte glycoprotein (MOG) in multiple sclerosis: reactivity to the extracellular domain of MOG is directed against three main regions. *Eur J Immunol* 1997; 27: 3059-69.
- Kirchner T, Hoppe F, Schalke B, Muller-Hermelink HK. Microenvironment of thymic myoid cells in myasthenia gravis. *Virchows Arch B Cell Pathol Incl Mol Pathol* 1988a; 54: 295-302.
- Kirchner T, Schalke B, Melms A, von Kugelgen T, Muller-Hermelink HK. Immunohistological patterns of non-neoplastic changes in the thymus in Myasthenia gravis. *Virchows Arch B Cell Pathol Incl Mol Pathol* 1986; 52: 237-57.

- Kirchner T, Tzartos S, Hoppe F, Schalke B, Wekerle H, Muller-Hermelink HK. Pathogenesis of myasthenia gravis. Acetylcholine receptor-related antigenic determinants in tumor-free thymuses and thymic epithelial tumors. *Am J Pathol* 1988b; 130: 268-80.
- Kisielow P, Bluthmann H, Staerz UD, Steinmetz M, von Boehmer H. Tolerance in T-cell-receptor transgenic mice involves deletion of nonmature CD4+8+ thymocytes. *Nature* 1988; 333: 742-6.
- Klein L, Klein T, Ruther U, Kyewski B. CD4 T cell tolerance to human C-reactive protein, an inducible serum protein, is mediated by medullary thymic epithelium. *J Exp Med* 1998; 188: 5-16.
- Klein L, Klugmann M, Nave KA, Tuohy VK, Kyewski B. Shaping of the autoreactive T-cell repertoire by a splice variant of self protein expressed in thymic epithelial cells. *Nat Med* 2000; 6: 56-61.
- Klein L, Kyewski B. "Promiscuous" expression of tissue antigens in the thymus: a key to T-cell tolerance and autoimmunity? *J Mol Med* 2000a; 78: 483-94.
- Klein L, Kyewski B. Self-antigen presentation by thymic stromal cells: a subtle division of labor. *Curr Opin Immunol* 2000b; 12: 179-86.
- Klein L, Roettinger B, Kyewski B. Sampling of complementing self-antigen pools by thymic stromal cells maximizes the scope of central T cell tolerance. *Eur J Immunol* 2001; 31: 2476-86.
- Klemenz R, Frohli E, Steiger RH, Schafer R, Aoyama A. Alpha B-crystallin is a small heat shock protein. *Proc Natl Acad Sci U S A* 1991; 88: 3652-6.
- Klugmann M, Schwab MH, Pulhofer A, Schneider A, Zimmermann F, Griffiths IR, et al. Assembly of CNS myelin in the absence of Proteolipid Protein. *Neuron* 1997; 18: 59-70.
- Kojima K, Berger T, Lassmann H, Hinze-Selch D, Zhang Y, Gehrman J, et al. Experimental autoimmune panencephalitis and uveoretinitis transferred to the Lewis rat by T lymphocytes specific for the S100 beta molecule, a calcium binding protein of astroglia. *J Exp Med* 1994; 180: 817-29.
- Kojima K, Reindl M, Lassmann H, Wekerle H, Linington C. The thymus and self-tolerance: co-existence of encephalitogenic S100 beta-specific T cells and their nominal autoantigen in the normal adult rat thymus. *Int Immunol* 1997; 9: 897-904.
- Kroepfl J, Viise L, Charron A, Linington C, Gardinier MV. Investigation of myelin/oligodendrocyte glycoprotein membrane topology. *J Neurochem* 1996; 67: 2219-22.

- Kuchroo VK, Martin C, Greer JM, Ju ST, Sobel RA, Dorf ME. Cytokines and adhesion molecules contribute to the ability of myelin proteolipid protein-specific T cell clones to mediate experimental allergic encephalomyelitis. *J Immunol* 1993; 151: 4371-.
- Kurts C, Kosaka H, Carbone FR, Miller JF, W.R. H. Class I-restricted-cross-presentation of exogenous self-antigens leads to deletion of autorreactive CD8(+) T cells. *J Exp Med* 1997; 186: 239-245.
- Kurts C, Sutherland RM, Davey G, Li M, Lew AM, Blanas E, et al. CD8 T cell ignorance or tolerance to islet antigens depends on antigen dose. *Proc Natl Acad Sci U S A* 1999; 96: 12703-12707.
- Lafaille JJ, Nagashima K, Katsuki M, Tonegawa S. High incidence of spontaneous autoimmune encephalomyelitis in immunodeficient anti-myelin basic protein T cell receptor transgenic mice. *Cell* 1994; 78: 399-408.
- Lanoue A, Bona C, von Boehmer H, Sarukhan A. Conditions that induce tolerance in mature CD4+ T cells. *J Exp Med* 1997; 185: 405-414.
- Laufer TM, DeKoning J, Markowitz JS, Lo D, Glimcher LH. Unopposed positive selection and autoreactivity in mice expressing class II MHC only on thymic cortex. *Nature* 1996; 383: 81-5.
- Lauriola L, Michetti F, Stolfi VM, Tallini G, Cocchia D. Detection by S-100 immunolabelling of interdigitating reticulum cells in human thymomas. *Virchows Arch B Cell Pathol Incl Mol Pathol* 1984; 45: 187-95.
- Le Douarin N, Corbel C, Bandeira A, Thomas-Vaslin V, Y. M, Coutinho A, et al. Evidence for a thymus-dependent form of tolerance that is not based on elimination or anergy of reactive T cells. *Immunol Rev* 1996; 149: 35-53.
- Lennon VA, Lindstrom J, Seybold ME. Experimental autoimmune myasthenia (EAMG): a model of myasthenia gravis in rats and guinea pigs. *J Exp Med* 1975; 141: 1365-75.
- Leprince C, Cohen-Kaminsky S, Berrih-Aknin S, Vernet-Der Garabedian B, Treton D, Galanaud P, et al. Thymic B cells from myasthenia gravis patients are activated B cells. Phenotypic and functional analysis. *J Immunol* 1990; 145: 2115-22.
- Lindstrom JM. Acetylcholine receptors and myasthenia. *Muscle Nerve* 2000; 23: 453-77.
- Linnington C, Webb S, Woodhams P. A novel myelin-associated glycoprotein defined by a mouse monoclonal antibody. *J Neuroimmunol* 1984; 6: 387-96.
- Link H, Olsson O, Sun J, Wang WZ, Andersson G, Ekre HP, et al. Acetylcholine receptor-reactive T and B cells in myasthenia gravis and controls. *J Clin Invest* 1991; 87: 2191-5.
- Link J, Soderstrom M, Ljungdahl A, Hojeberg B, Olsson T, Xu Z, et al. Organ-specific autoantigens induce interferon-gamma and interleukin-4 mRNA expression in

- mononuclear cells in multiple sclerosis and myasthenia gravis. *Neurology* 1994; 44: 728-34.
- Lintula S, Stenman U. The expression of prostate-specific membrane antigen in peripheral blood leukocytes. *J Urol* 1997; 157: 1969-72.
- Litzenburger T, Bluthmann H, Morales P, Pham-Dinh D, Dautigny A, Wekerle H, et al. Development of myelin oligodendrocyte glycoprotein autoreactive transgenic B lymphocytes: receptor editing in vivo after encounter of a self-antigen distinct from myelin oligodendrocyte glycoprotein. *J Immunol* 2000; 165: 5360-6.
- Liu H, MacKenzie-Graham AJ, Kim A, Voskuhl RR. Mice resistant to experimental autoimmune encephalomyelitis have increased thymic expression of myelin basic protein and increased MBP specific T cell tolerance. *J Neuroimmunol* 2001; 115: 118-126.
- Liu J, Marino MW, Wong G, Grail D, Dunn A, Bettadapura J, et al. TNF is a potent anti-inflammatory cytokine in autoimmune-mediated demyelination. *Nat Med* 1998; 4: 78-83.
- Liyanaige Y, Teo M, MacLennan C, Buckel A, Beeson D, Willcox N, et al. Expression of muscle proteins in thymomas of patients with myasthenia gravis. *Ann N Y Acad Sci* 1998; 841: 411-3.
- Lobos EA, Rudnick CH, Watson MS, Isenberg KE. Linkage disequilibrium study of RFLPs detected at the human muscle nicotinic acetylcholine receptor subunit genes. *Am J Hum Genet* 1989; 44: 522-33.
- Lucchinetti C, Bruck W, Parisi J, Scheithauer B, Rodriguez M, Lassmann H. Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination. *Ann Neurol* 2000; 47: 707-17.
- MacLennan CA, Beeson D, Willcox N, Vincent A, Newsom-Davis J. Muscle nicotinic acetylcholine receptor mRNA expression in hyperplastic and neoplastic myasthenia gravis thymus. *Ann N Y Acad Sci* 1998; 841: 407-10.
- Maitra A, Wistuba, II, Virmani AK, Sakaguchi M, Park I, Stucky A, et al. Enrichment of epithelial cells for molecular studies. *Nat Med* 1999; 5: 459-63.
- Manfredi AA, Protti MP, Dalton MW, Howard JF, Jr., Conti-Tronconi BM. T helper cell recognition of muscle acetylcholine receptor in myasthenia gravis. Epitopes on the gamma and delta subunits. *J Clin Invest* 1993; 92: 1055-67.
- Markovic-Plese S, Fukaura H, Zhang J, al-Sabbagh A, Southwood S, Sette A, et al. T cell recognition of immunodominant and cryptic proteolipid protein epitopes in humans. *J Immunol* 1995; 155: 982-92.

- Marten NW, Stohlman SA, Smith-Begolka W, Miller SD, Dimacali E, Yao Q, et al. Selection of CD8+ T cells with highly focused specificity during viral persistence in the central nervous system. *J Immunol* 1999; 162: 3905-14.
- Martens H, Goxe B, Geenen V. The thymic repertoire of neuroendocrine self-antigens: physiological implications in T-cell life and death. *Immunol Today* 1996; 17: 312-7.
- Martin R. Genetics of multiple sclerosis--how could disease-associated HLA-types contribute to pathogenesis? *J Neural Transm Suppl* 1997; 49: 177-94.
- Martin R, Gran B, Zhao Y, Markovic-Plese S, Bielekova B, Marques A, et al. Molecular mimicry and antigen-specific t cell responses in multiple sclerosis and chronic cns lyme disease. *J Autoimmun* 2001a; 16: 187-92.
- Martin R, Howell MD, Jaraquemada D, Flerlage M, Richert J, Brostoff S, et al. A myelin basic protein peptide is recognized by cytotoxic T cells in the context of four HLA-DR types associated with multiple sclerosis. *J Exp Med* 1991; 173: 19-24.
- Martin R, McFarland HF, McFarlin DE. Immunological aspects of demyelinating diseases. *Annu Rev Immunol* 1992; 10: 153-87.
- Martin R, Stürzebecher CS, McFarland EJ. Immunotherapy of multiple sclerosis: Where are we? Where should we go? *Nat Immunol* 2001b; 2: 785-88.
- Martino G, Hartung HP. Immunopathogenesis of multiple sclerosis: the role of T cells. *Curr Opin Neurol* 1999; 12: 309-21.
- Martinou JC, Falls DL, Fischbach GD, Merlie JP. Acetylcholine receptor-inducing activity stimulates expression of the epsilon-subunit gene of the muscle acetylcholine receptor. *Proc Natl Acad Sci U S A* 1991; 88: 7669-73.
- Marx A, Kirchner T, Hoppe F, O'Connor R, Schalke B, Tzartos S, et al. Proteins with epitopes of the acetylcholine receptor in epithelial cell cultures of thymomas in myasthenia gravis. *Am J Pathol* 1989; 134: 865-77.
- Marx A, Wilisch A, Schultz A, Gattenlohner S, Nenninger R, Muller-Hermelink HK. Pathogenesis of myasthenia gravis. *Virchows Arch* 1997; 430: 355-64.
- Mason D, Powrie F. Control of autoimmune pathology by regulatory T cells. *Curr Opin Immunol* 1998; 10: 649-655.
- Mathisen PM, Pease S, Garvey J, Hood L, Readhead C. Identification of an embryonic isoform of myelin basic protein that is expressed widely in the mouse embryo. *Proc Natl Acad Sci U S A* 1993; 90: 10125-9.
- Meinl E, Klinkert WE, Wekerle H. The thymus in myasthenia gravis: Changes typical for the human disease are absent in experimental autoimmune myasthenia gravis of the Lewis rat. *Am J Pathol* 1991; 139: 995-1008.

- Melms A, Chrestel S, Schalke BC, Wekerle H, Mauron A, Ballivet M, et al. Autoimmune T lymphocytes in myasthenia gravis. Determination of target epitopes using T lines and recombinant products of the mouse nicotinic acetylcholine receptor gene. *J Clin Invest* 1989; 83: 785-90.
- Melms A, Schalke BC, Kirchner T, Muller-Hermelink HK, Albert E, Wekerle H. Thymus in myasthenia gravis. Isolation of T-lymphocyte lines specific for the nicotinic acetylcholine receptor from thymuses of myasthenic patients. *J Clin Invest* 1988; 81: 902-8.
- Mihovilovic M, Denning S, Mai Y, Fisher CM, Whichard LP, Patel DD, et al. Thymocytes and cultured thymic epithelial cells express transcripts encoding alpha-3, alpha-5, and beta-4 subunits of neuronal nicotinic acetylcholine receptors. Preferential transcription of the alpha-3 and beta-4 genes by immature CD4+8+ thymocytes and evidence for response to nicotine in thymocytes. *Ann N Y Acad Sci* 1998; 841: 388-92.
- Mihovilovic M, Hulette C, Mittelstaedt J, Austin C, Roses AD. Nicotinic neuronal acetylcholine receptor alpha-3 subunit transcription in normal and myasthenic thymus. *Ann N Y Acad Sci* 1993; 681: 83-96.
- Mihovilovic M, Roses AD. Expression of alpha-3, alpha-5, and beta-4 neuronal acetylcholine receptor subunit transcripts in normal and myasthenia gravis thymus. Identification of thymocytes expressing the alpha-3 transcripts. *J Immunol* 1993; 151: 6517-24.
- Miller A, Lider O, Roberts AB, Sporn MB, Weiner HL. Suppressor T cells generated by oral tolerization to myelin basic protein suppress both in vitro and in vivo immune responses by the release of transforming growth factor beta after antigen-specific triggering. *Proc Natl Acad Sci U S A* 1992; 89: 421-425.
- Miller J, Kurts C, Allison J, Kosaka H, Carbone FR, Heath WR. Induction of peripheral CD8+ T cell tolerance by cross-presentation of self antigens. *Immunol Rev* 1998; 165: 267-277.
- Miller JF, Heath WR. Self-ignorance in the peripheral T-cell pool. *Immunol Rev* 1993; 133: 131-50.
- Mishina M, Takai T, Imoto K, Noda M, Takahashi T, Numa S, et al. Molecular distinction between fetal and adult forms of muscle acetylcholine receptor. *Nature* 1986; 321: 406-11.
- Missias AC, Chu GC, Klocke BJ, Sanes JR, Merlie JP. Maturation of the acetylcholine receptor in skeletal muscle: regulation of the AChR gamma-to-epsilon switch. *Dev Biol* 1996; 179: 223-38.

- Modigliani Y, Bandeira A, Coutinho A. A model for developmentally acquired thymus-dependent tolerance to central and peripheral antigens. *Immunol Rev* 1996; 149: 155-20.
- Moiola L, Galbiati F, Martino G, Amadio S, Brambilla E, Comi G, et al. IL-12 is involved in the induction of experimental autoimmune myasthenia gravis, an antibody-mediated disease. *Eur J Immunol* 1998; 28: 2487-97.
- Moiola L, Karachunski P, Protti MP, Howard JF, Jr., Conti-Tronconi BM. Epitopes on the beta subunit of human muscle acetylcholine receptor recognized by CD4+ cells of myasthenia gravis patients and healthy subjects. *J Clin Invest* 1994; 93: 1020-8.
- Mokhtarian F, Zhang Z, Shi Y, Gonzales E, Sobel RA. Molecular mimicry between a viral peptide and a myelin oligodendrocyte glycoprotein peptide induces autoimmune demyelinating disease in mice. *J Neuroimmunol* 1999; 95: 43-54.
- Moore BW. A soluble protein characteristic of the nervous system. *Biochem Biophys Res Commun* 1965; 19: 739-44.
- Muller-Hermelink HK, Marx A. Thymoma. *Curr Opin Oncol* 2000; 12: 426-33.
- Murakami M, Hosoi Y, Negishi T, Kamiya Y, Miyashita K, Yamada M, et al. Thymic hyperplasia in patients with Graves' disease. Identification of thyrotropin receptors in human thymus. *J Clin Invest* 1996; 98: 2228-34.
- Nagvekar N, Moody AM, Moss P, Roxanis I, Curnow J, Beeson D, et al. A pathogenetic role for the thymoma in myasthenia gravis. Autosensitization of IL-4- producing T cell clones recognizing extracellular acetylcholine receptor epitopes presented by minority class II isotypes. *J Clin Invest* 1998; 101: 2268-77.
- Nakano S, Engel AG. Myasthenia gravis: quantitative immunocytochemical analysis of inflammatory cells and detection of complement membrane attack complex at the end-plate in 30 patients. *Neurology* 1993; 43: 1167-72.
- Navaneetham D, Penn A, Howard J, Jr., Conti-Fine BM. Expression of the alpha 7 subunit of the nicotinic acetylcholine receptor in normal and myasthenic human thymuses. *Cell Mol Biol (Noisy-le-grand)* 1997; 43: 433-42.
- Nave KA, Lai C, Bloom FE, Milner RJ. Splice site selection in the proteolipid protein (PLP) gene transcript and primary structure of the DM-20 protein of the central nervous system myelin. *Proc Natl Acad Sci USA* 1986; 84: 5665-69.
- Nelson S, Conti-Tronconi BM. Adult thymus expresses an embryonic nicotinic acetylcholine receptor- like protein. *J Neuroimmunol* 1990; 29: 81-92.
- Noseworthy JH, Lucchinetti C, Rodriguez M, Weinshenker BG. Multiple sclerosis. *N Engl J Med* 2000; 343: 938-52.

- O'Connor KC, Bar-Or A, Hafler DA. The neuroimmunology of multiple sclerosis: possible roles of T and B lymphocytes in immunopathogenesis. *J Clin Immunol* 2001; 21: 81-92.
- Oehen SU, Ohashi PS, Burki K, Hengartner H, Zinkernagel RM, Aichele P. Escape of thymocytes and mature T cells from clonal deletion due to limiting tolerogen expression levels. *Cell Immunol* 1994; 158: 342-52.
- Ohashi PS, Oehen S, Buerki K, Pircher H, Ohashi CT, Odermatt B, et al. Ablation of "tolerance" and induction of diabetes by virus infection in viral antigen transgenic mice. *Cell* 1991; 65: 305-317.
- Okazaki K, Obata N, Inoue S, Hidaka H. S100 beta is a target protein of neurocalcin delta, an abundant isoform in glial cells. *Biochem J* 1995; 306: 551-5.
- Oldstone MB, Nerenberg M, Southern P, Price J, Lewicki H. Virus infection triggers insulin-dependent diabetes mellitus in a transgenic model: role of anti-self (virus) immune response. *Cell* 1991; 65: 319-331.
- Oshima M, Ashizawa T, Pollack M, Atassi M. Autoimmune T cell recognition of human acetylcholine receptor: The sites of T cell recognition in myasthenia gravis on the extracellular part of the  $\alpha$  subunit. *Eur J Immunol* 1990; 20: 2563-69.
- Ota K, Matsui M, Milford EL, Mackin GA, Weiner HL, Hafler DA. T-cell recognition of an immunodominant myelin basic protein epitope in multiple sclerosis. *Nature* 1990; 346: 183-7.
- Padberg F, Matsuda M, Fenk R, Patenge N, Kubuschok B, Hohlfeld R, et al. Myasthenia gravis: selective enrichment of antiacetylcholine receptor antibody production in untransformed human B cell cultures. *Eur J Immunol* 1999; 29: 3538-48.
- Patrick J, Lindstrom J. Autoimmune response to acetylcholine receptor. *Science* 1973; 180: 871-72.
- Pelfrey CM, Trotter JL, Tranquill LR, McFarland HF. Identification of a second T cell epitope of human proteolipid protein (residues 89-106) recognized by proliferative and cytolytic CD4+ T cells from multiple sclerosis patients. *J Neuroimmunol* 1994; 53: 153-61.
- Peterson P, Nagamine K, Scott HS, Heino M, Kudoh J, Shimizu N, et al. APECED: a monogenic autoimmune disease providing new clues to self-tolerance. *Immunol Today* 1998; 19: 384-86.
- Pham-Dinh D, Mattei MG, Nussbaum JL, Roussel G, Pontarotti P, Roeckel N, et al. Myelin/oligodendrocyte glycoprotein is a member of a subset of the immunoglobulin superfamily encoded within the major histocompatibility complex. *Proc Natl Acad Sci U S A* 1993; 90: 7990-4.



- Ponniah S, Arah I, Alexander R. PSA is a candidate self-antigen in autoimmune chronic prostatitis/chronic pelvic pain syndrome. *Prostate* 2000; 44: 49-54.
- Pribyl TM, Campagnoni C, Kampf K, Handley VW, Campagnoni AT. The major myelin protein genes are expressed in the human thymus. *J Neurosci Res* 1996; 45: 812-9.
- Pribyl TM, Campagnoni CW, Kampf K, Kashima T, Handley VW, McMahon J, et al. The human myelin basic protein gene is included within a 179-kilobase transcription unit: expression in the immune and central nervous systems. *Proc Natl Acad Sci U S A* 1993; 90: 10695-9.
- Protti MP, Manfredi AA, Howard JF, Jr., Conti-Tronconi BM. T cells in myasthenia gravis specific for embryonic acetylcholine receptor. *Neurology* 1991a; 41: 1809-14.
- Protti MP, Manfredi AA, Wu XD, Moiola L, Howard JF, Jr., Conti-Tronconi BM. Myasthenia gravis. T epitopes on the delta subunit of human muscle acetylcholine receptor. *J Immunol* 1991b; 146: 2253-61.
- Pugliese A, Brown D, Garza D, Murchison D, Zeller M, Redondo M, et al. Self-antigen-presenting cells expressing diabetes-associated autoantigens exist in both thymus and peripheral lymphoid organs. *J Clin Invest* 2001; 107: 555-64.
- Pugliese A, Zeller M, Fernandez A, Jr., Zalcborg LJ, Bartlett RJ, Ricordi C, et al. The insulin gene is transcribed in the human thymus and transcription levels correlated with allelic variation at the INS VNTR-IDD3 susceptibility locus for type 1 diabetes. *Nat Genet* 1997; 15: 293-7.
- Raine CS, Cannella B, Hauser SL, Genain CP. Demyelination in primate autoimmune encephalomyelitis and acute multiple sclerosis lesions: a case for antigen-specific antibody mediation. *Ann Neurol* 1999; 46: 144-60.
- Ramón y Cajal S. Contribucion al conocimiento de la neuroglia del cerebro humano. *Trab Lab Invest Biol (Madrid)* 1913.; 11: 255–315.
- Reindl M, Linington C, Brehm U, Egg R, Dilitz E, Deisenhammer F, et al. Antibodies against the myelin oligodendrocyte glycoprotein and the myelin basic protein in multiple sclerosis and other neurological diseases: a comparative study. *Brain* 1999; 122: 2047-56.
- Reinhardt C, Melms A. Elevated frequencies of natural killer T lymphocytes in myasthenia gravis. *Neurology* 1999; 52: 1485-7.
- Richman DP, Gomez CM, Berman PW, Burrell SA, Fitch FW, Amason BG. Monoclonal anti-acetylcholine receptor antibodies can cause experimental myasthenia. *Nature* 1980; 286: 738-39.

- Riemersma S, Vincent A, Beeson D, Newland C, Hawke S, Vernet-der Garabedian B, et al. Association of arthrogryposis multiplex congenita with maternal antibodies inhibiting fetal acetylcholine receptor function. *J Clin Invest* 1996; 98: 2358-63.
- Rivers TM, Sprunt DH, Berry GP. *J Exp Med* 1933; 58: 39-53.
- Roberts R, Lieber M, Bostwick D, Jacobsen S. A review of clinical and pathological prostatitis syndromes. *Urology* 1997; 49: 809-21.
- Rocha B, von Boehmer H. Peripheral selection of the T cell repertoire. *Science* 1991; 251: 1225-1228.
- Sakaguchi S. Regulatory T cells: key controllers of immunologic self-tolerance. *Cell* 2000; 101: 455-8.
- Sanes JR, Johnson YR, Kotzbauer PT, Mudd J, Hanley T, Martinou JC, et al. Selective expression of an acetylcholine receptor-lacZ transgene in synaptic nuclei of adult muscle fibers. *Development* 1991; 113: 1181-91.
- Schluep M, Willcox N, Vincent A, Dhoot GK, Newsom-Davis J. Acetylcholine receptors in human thymic myoid cells in situ: an immunohistological study. *Ann Neurol* 1987; 22: 212-22.
- Schmidt S. Candidate autoantigens in multiple sclerosis. *Mult Scler* 1999; 5: 147-60.
- Schmidt S, Linington C, Zipp F, Sotgiu S, de Waal Malefyt R, Wekerle H, et al. Multiple sclerosis: comparison of the human T-cell response to S100 beta and myelin basic protein reveals parallels to rat experimental autoimmune panencephalitis. *Brain* 1997; 120: 1437-45.
- Schonbeck S, Padberg F, Hohlfeld R, Wekerle H. Transplantation of thymic autoimmune microenvironment to severe combined immunodeficiency mice. *J Clin Invest* 1992; 90: 245-50.
- Schonrich G, Kalinke U, Momburg F, Malissen M, Schmitt-Verhulst AM, Malissen B, et al. Down-regulation of T cell receptors on self-reactive T cells as a novel mechanism for extrathymic tolerance induction. *Cell* 1991; 65: 293-304.
- Schwartz RH. A cell culture model for T lymphocyte clonal anergy. *Science* 1990; 248: 1349-1356.
- Sebzda E, Mariathasan S, Ohteki T, Jones R, Bachmann MF, Ohashi PS. Selection of the T cell repertoire. *Annu Rev Immunol* 1999; 17: 829-74.
- Sebzda E, Wallace VA, Mayer J, Yeung RS, Mak TW, Ohashi PS. Positive and negative thymocyte selection induced by different concentrations of a single peptide. *Science* 1994; 263: 1615-8.

- Seddon B, Mason D. Regulatory T cells in the control of autoimmunity: the essential role of TGF- $\beta$  and IL-4 in the prevention of autoimmune thyroiditis in rats by peripheral CD4<sup>+</sup>CD45RC<sup>-</sup> cells and CD4<sup>+</sup>CD8<sup>-</sup> thymocytes. *J Exp Med* 1999; 189: 279-288.
- Seddon B, Mason D. The third function of the thymus. *Immunol Today* 2000; 21: 95-9.
- Sempowski G, Thomasch J, Gooding M, Hale L, Edwards L, Ciafaloni E, et al. Effect of thymectomy on human peripheral blood T cell pools in myasthenia gravis. *J Immunol* 2001; 166: 2808-17.
- Shenoy M, Oshima M, Atassi M, Christadoss P. Suppression of experimental autoimmune myasthenia gravis by epitope-specific neonatal tolerance to synthetic region alpha 146-162 of acetylcholine receptor. *Clin Immunol Immunopathol* 1993; 66: 230-38.
- Smith KM, Olson DC, Hirose R, Hanahan D. Pancreatic gene expression in rare cells of thymic medulla: evidence for functional contribution to T cell tolerance. *Int Immunol* 1997; 9: 1355-65.
- Smith M, Biggar S, Hussain M. Prostate-specific antigen messenger RNA is expressed in non-prostate cells: implications for detection of micrometastases. *Cancer Res* 1995; 55: 2640-44.
- Soldan SS, Berti R, Salem N, Secchiero P, Flamand L, Calabresi PA, et al. Association of human herpes virus 6 (HHV-6) with multiple sclerosis: increased IgM response to HHV-6 early antigen and detection of serum HHV-6 DNA. *Nat Med* 1997; 3: 1394-7.
- Sommer N, Harcourt GC, Willcox N, Beeson D, Newsom-Davis J. Acetylcholine receptor-reactive T lymphocytes from healthy subjects and myasthenia gravis patients. *Neurology* 1991; 41: 1270-6.
- Sospedra M, Ferrer-Francesch X, Dominguez O, Juan M, Foz-Sala M, Pujol-Borrell R. Transcription of a broad range of self-antigens in human thymus suggests a role for central mechanisms in tolerance toward peripheral antigens. *J Immunol* 1998; 161: 5918-29.
- Spuler S, Sarropoulos A, Marx A, Hohlfeld R, Wekerle H. Thymoma-associated myasthenia gravis. Transplantation of thymoma and extrathymomal thymic tissue into SCID mice. *Am J Pathol* 1996; 148: 1359-65.
- Stefflerl A, Schubart A, Storch M, Amini A, Mather I, Lassmann H, et al. Butyrophilin, a milk protein, modulates the encephalitogenic T cell response to myelin oligodendrocyte glycoprotein in experimental autoimmune encephalomyelitis. *J Immunol* 2000; 165: 2859-65.
- Steinman L. Multiple sclerosis: a coordinated immunological attack against myelin in the central nervous system. *Cell* 1996; 85: 299-302.

- Storch MK, Piddlesden S, Haltia M, Iivanainen M, Morgan P, Lassmann H. Multiple sclerosis: in situ evidence for antibody- and complement- mediated demyelination. *Ann Neurol* 1998a; 43: 465-71.
- Storch MK, Stefferl A, Brehm U, Weissert R, Wallstrom E, Kerschensteiner M, et al. Autoimmunity to myelin oligodendrocyte glycoprotein in rats mimics the spectrum of multiple sclerosis pathology. *Brain Pathol* 1998b; 8: 681-94.
- Su S, Huang I, Fair W, Heston W. Alternatively spliced variants of prostate-specific membrane antigen RNA: ratio of expression as a potential measurement of progression. *Cancer Res* 1995; 55: 1441-45.
- Sun J, Link H, Olsson T, Xiao BG, Andersson G, Ekre HP, et al. T and B cell responses to myelin-oligodendrocyte glycoprotein in multiple sclerosis. *J Immunol* 1991a; 146: 1490-5.
- Sun JB, Olsson T, Wang WZ, Xiao BG, Kostulas V, Fredrikson S, et al. Autoreactive T and B cells responding to myelin proteolipid protein in multiple sclerosis and controls. *Eur J Immunol* 1991b; 21: 1461-8.
- Taguchi O, Kontani K, Ikeda H, Kezuka T, Takeuchi M, Takahashi H, et al. Tissue-specific suppressor T cells involved in self-tolerance are activated extrathymically by self-antigens. *Immunology* 1994; 82: 365-9.
- Talib S, Okarma TB, Lebkowski JS. Differential expression of human nicotinic acetylcholine receptor alpha subunit variants in muscle and non-muscle tissues. *Nucleic Acids Res* 1993; 21: 233-7.
- Targoni OS, Lehmann PV. Endogenous myelin basic protein inactivates the high avidity T cell repertoire. *J Exp Med* 1998; 187: 2055-2065.
- Trotter JL, Hickey WF, van der Veen RC, Sulze L. Peripheral blood mononuclear cells from multiple sclerosis patients recognize myelin proteolipid protein and selected peptides. *J Neuroimmunol* 1991; 33: 55-62.
- Tuohy VK, Lu Z, Sobel RA, Laursen RA, Lees MB. Identification of an encephalitogenic determinant of myelin proteolipid protein for SJL mice. *J Immunol* 1989; 142: 1523-7.
- Tzartos SJ, Barkas T, Cung MT, Kordossi A, Loutrari H, Marraud M, et al. The main immunogenic region of the acetylcholine receptor. Structure and role in myasthenia gravis. *Autoimmunity* 1991; 8: 259-70.
- Tzartos SJ, Kokla A, Walgrave SL, Conti-Tronconi BM. Localization of the main immunogenic region of human muscle acetylcholine receptor to residues 67-76 of the alpha subunit. *Proc Natl Acad Sci U S A* 1988; 85: 2899-903.

- Tzartos SJ, Remoundos M. Detection of antibodies directed against the cytoplasmic region of the human acetylcholine receptor in sera from myasthenia gravis patients. *Clin Exp Immunol* 1999; 116: 146-52.
- Tzartos SJ, Seybold ME, Lindstrom JM. Specificities of antibodies to acetylcholine receptors in sera from myasthenia gravis patients measured by monoclonal antibodies. *Proc Natl Acad Sci U S A* 1982; 79: 188-92.
- Vafiadis P, Bennett ST, Todd JA, Nadeau J, Grabs R, Goodyer CG, et al. Insulin expression in human thymus is modulated by INS VNTR alleles at the IDDM2 locus. *Nat Genet* 1997; 15: 289-92.
- van Meerwijk JP, Marguerat S, Lees RK, Germain RN, Fowlkes BJ, MacDonald HR. Quantitative impact of thymic clonal deletion on the T cell repertoire. *J Exp Med* 1997; 185: 377-383.
- van Noort JM, van Sechel AC, Bajramovic JJ, el Ouagmiri M, Polman CH, Lassmann H, et al. The small heat-shock protein alpha B-crystallin as candidate autoantigen in multiple sclerosis. *Nature* 1995; 375: 798-801.
- van Sechel AC, Bajramovic JJ, van Stipdonk MJ, Persoon-Deen C, Geutskens SB, van Noort JM. EBV-induced expression and HLA-DR-restricted presentation by human B cells of alpha B-crystallin, a candidate autoantigen in multiple sclerosis. *J Immunol* 1999; 162: 129-35.
- van Stipdonk MJ, Willems AA, Plomp AC, van Noort JM, Boog CJ. Tolerance controls encephalitogenicity of alphaB-crystallin in the Lewis rat. *J Neuroimmunol* 2000; 103: 103-11.
- Vernet-der Garabedian B, Lacokova M, Eymard B, Morel E, Faltin M, Zajac J, et al. Association of neonatal myasthenia gravis with antibodies against the fetal acetylcholine receptor. *J Clin Invest* 1994; 94: 555-9.
- Vincent A, Newland C, Brueton L, Beeson D, Riemersma S, Huson SM, et al. Arthrogryposis multiplex congenita with maternal autoantibodies specific for a fetal antigen. *Lancet* 1995; 346: 24-5.
- Vincent A, Newsom-Davis J. Acetylcholine receptor antibody characteristics in myasthenia gravis. I. Patients with generalized myasthenia or disease restricted to ocular muscles. *Clin Exp Immunol* 1982; 49: 257-65.
- Virchow R. Ueber das ausgebreitete Vorkommen einer dem Ner-venmark analogen substanz in den tierischen Geweben. *Virchows Arch Pathol Anat* 1854; 6: 562.
- von Herrath MG, Evans CF, Horwitz MS, Oldstone MB. Using transgenic mouse models to dissect the pathogenesis of virus- induced autoimmune disorders of the islets of Langerhans and the central nervous system. *Immunol Rev* 1996; 152: 111-43.

- Voskuhl RR. Myelin protein expression in lymphoid tissues: implications for peripheral tolerance. *Immunol Rev* 1998; 164: 81-92.
- Wakkach A, Guyon T, Bruand C, Tzartos S, Cohen-Kaminsky S, Berrih-Aknin S. Expression of acetylcholine receptor genes in human thymic epithelial cells: implications for myasthenia gravis. *J Immunol* 1996; 157: 3752-60.
- Wakkach A, Poeta S, Chastre E, Gespach C, Lecerf F, De La Porte S, et al. Establishment of a human thymic myoid cell line. Phenotypic and functional characteristics. *Am J Pathol* 1999; 155: 1229-40.
- Wang Z, Karachunski P, Howard FM, Jr., Conti-Fine BM. Myasthenia in SCID mice grafted with myasthenic patient lymphocytes. *Neurology* 1999; 52: 484-97.
- Webb S, Morris C, Sprent J. Extrathymic tolerance of mature T cells: clonal elimination as consequence of immunity. *Cell* 1990; 63: 1249-1256.
- Wekerle H. The viral triggering of autoimmune disease. *Nat Med* 1998; 4: 770-771.
- Wekerle H, Bradl M, Lington C, Kaab G, Kojima K. The shaping of the brain-specific T lymphocyte repertoire in the thymus. *Immunol Rev* 1996; 149: 231-43.
- Werdelin O, Cordes U, Jensen T. Aberrant expression of tissue-specific proteins in the thymus: a hypothesis for the development of central tolerance. *Scand J Immunol* 1998; 47: 95-100.
- Werner P, Pitt D, Raine CS. Multiple sclerosis: altered glutamate homeostasis in lesions correlates with oligodendrocyte and axonal damage. *Ann Neurol* 2001; 50: 169-80.
- Wheatley LM, Urso D, Tumas K, Maltzman J, Loh E, Levinson AI. Molecular evidence for the expression of nicotinic acetylcholine receptor alpha-chain in mouse thymus. *J Immunol* 1992; 148: 3105-9.
- Wilisch A, Gutsche S, Hoffacker V, Schultz A, Tzartos S, Nix W, et al. Association of acetylcholine receptor alpha-subunit gene expression in mixed thymoma with myasthenia gravis. *Neurology* 1999; 52: 1460-6.
- Willard HF, Riodan JR. Assignment of the gene for myelin proteolipid protein to the X-chromosome: implication for X-linked myelin disorders. *Science* 1985; 230: 940-42.
- Williams CB, Engle DL, Kersh GJ, Michael White J, Allen PM. A kinetic threshold between negative and positive selection based on the longevity of the T cell receptor-ligand complex. *J Exp Med* 1999; 189: 1531-44.
- Wucherpfennig KW, Strominger JL. Molecular mimicry in T cell-mediated autoimmunity: viral peptides activate human T clones specific for myelin basic protein. *Cell* 1995; 80: 695-705.

- 
- Xiao BG, Linington C, Link H. Antibodies to myelin-oligodendrocyte glycoprotein in cerebrospinal fluid from patients with multiple sclerosis and controls. *J Neuroimmunol* 1991; 31: 91-6.
- Zal T, Volkman A, Stockinger B. Mechanisms of tolerance induction in major histocompatibility complex class II-restricted T cells specific for a blood-borne self-antigen. *J Exp Med* 1994; 180: 2089-99.
- Zamvil SS, Steinman L. The T lymphocyte in experimental allergic encephalomyelitis. *Annu Rev Immunol* 1990; 8: 579-621.
- Zhang J, Xiao BG, Bakhiert M, van Der Meide P, Wigzell H, Link H, et al. Both CD4+ and CD8+ T cells are essential to induce experimental autoimmune myasthenia gravis. *J Exp Med* 1996; 184: 349-56.
- Zheng Y, Wheatley LM, Liu T, Levinson AI. Regulation of acetylcholine receptor alpha subunit mRNA expression in myasthenic thymus. *Ann N Y Acad Sci* 1998; 841: 393-6.
- Zheng Y, Wheatley LM, Liu T, Levinson AI. Acetylcholine receptor alpha subunit mRNA expression in human thymus: augmented expression in myasthenia gravis and upregulation by interferon-gamma. *Clin Immunol* 1999; 91: 170-7.
- Zimmer D, Cornwall E, Landar A, Song W. The S100 protein family: history, function, and expression. *Brain Res Bull* 1995; 37: 417-29.

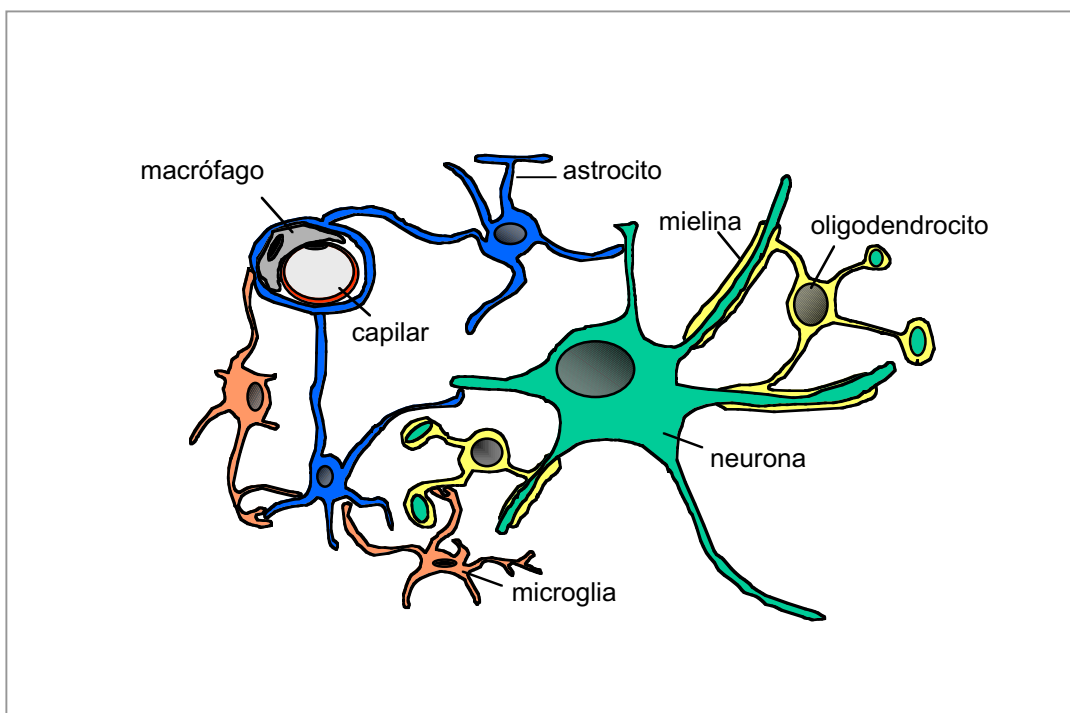




### **Las células del Sistema Nervioso Central**

En el año 1846, Virchow describió que en el cerebro, además de las neuronas, existía otro tipo celular, y pensando de que se trataba del tejido conectivo, lo llamó *neuroglia*. En 1913 Ramon y Cajal y Rio Hortega, utilizando técnicas de impregnación con metales, caracterizaron dos tipos celulares que forman la neuroglia, los astrocitos y oligodendrocitos, y los llamaron *macroglia* (Ramón y Cajal, 1913.). Años mas tarde, Rio Hortega describió un tercer tipo celular y lo llamó *microglia*, para diferenciarlo de la *macroglia*, los astrocitos y los oligodendrocitos. Las células gliales constituyen el principal tipo celular del sistema nervioso central (el 90% en los seres humanos) y son necesarias para el desarrollo y la correcta función neuronal.

Los **astrocitos**, transportan líquidos e iones hacia los capilares, y guían la migración celular durante la embriogénesis del sistema nervioso central. Desempeñan un papel estructural, además de formar la barrera hemato-encefálica rodeando a los endotelios capilares e impidiendo la circulación de las células sanguíneas en el sistema nervioso central (gráfico 13).



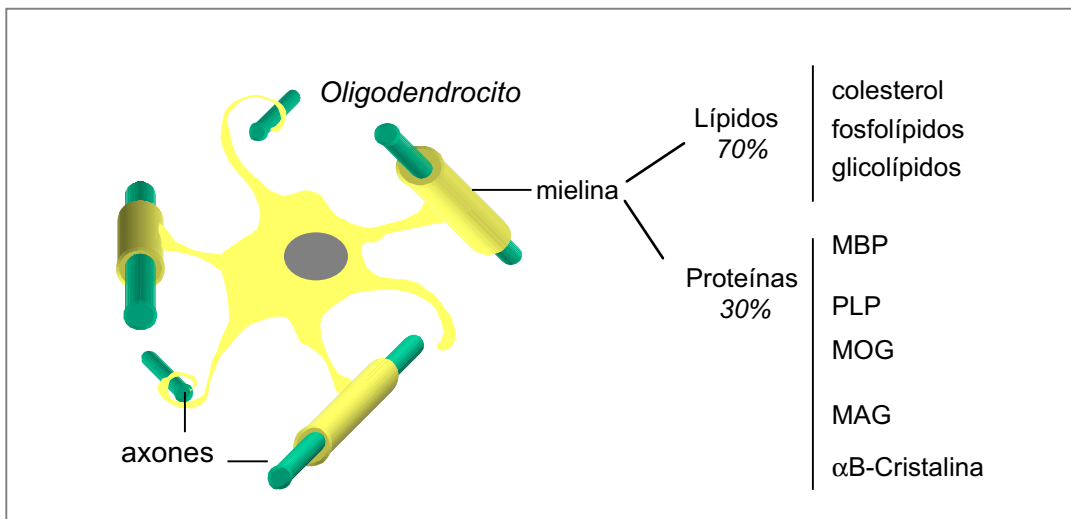
**Gráfico 13:** Representación de los tipos celulares en el sistema nervioso central. Los astrocitos son células estrelladas, con numerosas prolongaciones que les permitan contactar con todos los demás tipos celulares, y forman la barrera hemato-encefálica, debido a que rodean a las células endoteliales de los capilares sanguíneos. La microglia son los macrófagos residentes del sistema nervioso central. Tienen capacidad de fagocitar restos celulares y después de un daño neural son capaces de activarse y de proliferar mitóticamente. Los oligodendrocitos son las células formadoras de la mielina.

La **microglia**, son macrófagos especializados o células del sistema inmunológico en el sistema nervioso central (similares a las células dendríticas presentadoras de antígenos), debido a que tienen capacidad fagocítica y expresan moléculas de MHC de clase II.

Los **oligodendrocitos**, son las células formadoras de la mielina. La vaina de mielina debe su nombre a Virchow (Virchow, 1854). Es una estructura espiral constituida por extensiones de la membrana plasmática de los oligodendrocitos en el sistema nervioso central, la cual se dispone de manera concéntrica alrededor de los axones, constituyendo una vaina multilaminar (gráfico 14).

Se compone de un 70% de lípidos, de un 30% de proteínas y de un bajo contenido en agua. Esta proporción es inversa a la composición normal de la membrana plasmática, pero es acorde con la función de conducción del impulso nervioso, a alta velocidad y a largas distancias.

Todos los constituyentes de la mielina son sintetizados por el oligodendrocito. Los lípidos que constituyen la mielina son colesterol, fosfolípidos y glicolípidos en una proporción aproximada de 4:3:2. Los componentes proteicos mas importantes de la mielina son la proteína básica de mielina (MBP) y la proteína proteolipídica (PLP), que son proteínas de bajo peso molecular y representan el 80% de las proteínas mielínicas (Campagnoni, 1988). La mielina también contiene otros componentes proteicos minoritarios, como la glicoproteína asociada a la mielina (MAG), y la glicoproteína mielínica de los oligodendrocitos (MOG).



**Gráfico 14:** Componentes de la vaina de mielina del sistema nervioso central

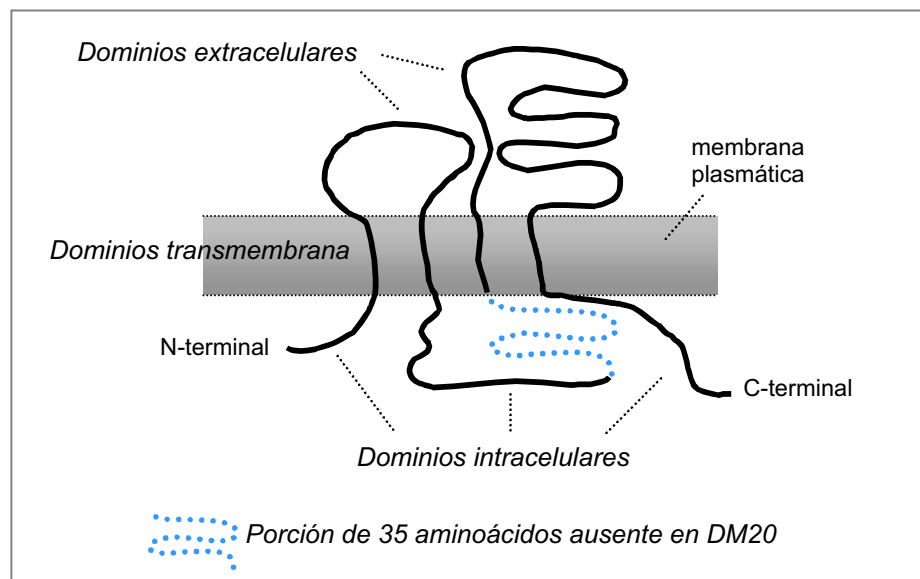
## Antígenos mielínicos

### Proteína Proteolípídica (PLP)

La PLP es uno de los principales componentes proteicos de la mielina, ya que representa casi el 50% de las mismas. Recibe el nombre de proteína proteolípídica debido a que en el año 1951, Folch extrajo gran cantidad de proteínas del cerebro empleando mezclas de solventes orgánicos y pensando que se trataba de complejos proteolípídicos, las denominó “*proteolípidos*” para diferenciarlas de otras lipoproteínas solubles en agua (Folch and Lees, 1951).

La PLP del sistema nervioso central existe en dos isoformas: la isoforma predominante llamada PLP *full-length* (de 25-kD) y la isoforma corta, llamada DM20 (de 20-kD). Ambas isoformas son codificadas por el mismo gen de 15 Kb (Nave et al., 1986) localizado en el cromosoma X en la posición Xq22 en humanos, y en el área H2C en ratones (Willard and Riodan, 1985). El gen comprende 7 exones, que codifican el mRNA de la forma full-length de PLP, mientras que la isoforma DM-20, es el resultado de *splicing* alternativo del mRNA, que pierde parte del exón 3 (exón 3B) y resulta en la delección de 35 aminoácidos de la secuencia proteica.

Existe un 100% de homología en la secuencia peptídica de PLP de ratón y la humana (Diehl et al., 1986). La estructura secundaria de la proteína comprende dos dominios extracitoplasmáticos, cuatro dominios de transmembrana, y tres dominios intracitoplasmáticos que incluyen los extremos N y C-terminal (gráfico 15).



**Gráfico 15:** Representación de la estructura secundaria de PLP

La función de PLP en la vaina de mielina es la de estabilizar las uniones de la membrana (Boison et al., 1995; Klugmann et al., 1997). En los ratones knock-out de PLP, que no expresan PLP ni DM20, se observa que si bien los oligodendrocitos son capaces de mielinizar a los axones, la vaina de mielina carece de estabilidad (Boison and Stoffel, 1994).

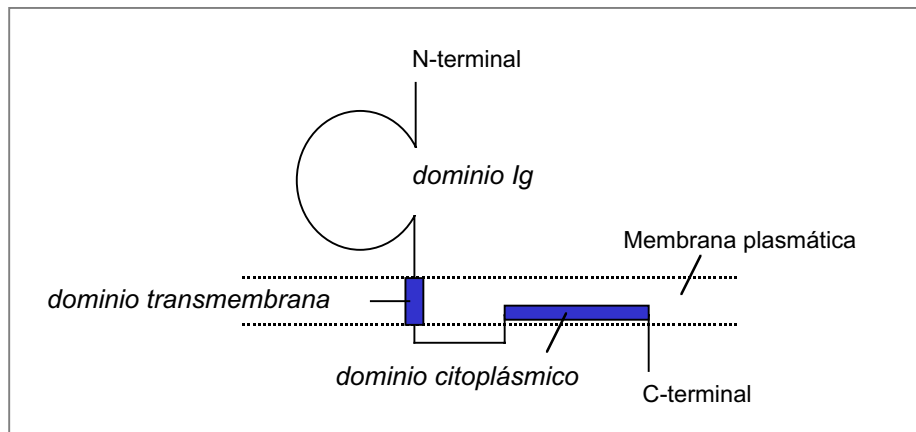
### ***Glicoproteína mielínica de los oligodendrocitos (MOG)***

MOG, es una proteína específica de los oligodendrocitos localizada en la superficie exterior de la vaina de mielina (Brunner et al., 1989). Es un componente minoritario de las proteínas mielínicas, ya que representa solo el 0,01-0,05 % en peso de las proteínas de membrana (Linnington et al., 1984).

MOG es una proteína de 218 aminoácidos que pertenece a la superfamilia de las inmunoglobulinas (Gardinier et al., 1992) y está codificada dentro de la región distal del MHC (Pham-Dinh et al., 1993). El gen MOG humano tiene un tamaño de 11.1 Kb y se organiza en 10 exones. Los transcritos dominantes de MOG, sin embargo, están codificados por 8 exones homólogos (Hilton et al., 1995). La proteína resultante es una glicoproteína de membrana de tipo I (ver gráfico 16), de 26-28 kD, que presenta un dominio Ig en el extremo N-terminal, un dominio transmembrana y un tercer dominio integrado con la cara interna de la membrana plasmática (della Gaspera et al., 1998; Kroepfl et al., 1996).

La función de MOG no se conoce, pero dada la localización extracelular de su dominio N-terminal en la vaina de mielina (Brunner et al., 1989), algunos autores sostienen que podría actuar como receptor, mientras que otros piensan que desempeña un papel estructural en el mantenimiento de la mielina, a través de interacciones con las proteínas de la matrix extracelular. El extremo C-terminal también parece ser funcionalmente importante en interacciones intracelulares, ya que es un dominio altamente conservado en diferentes especies (Gardinier et al., 1992; Pham-Dinh et al., 1993).

Desde el punto de vista inmunológico, MOG es un importante autoantígeno en las enfermedades desmielinizantes del sistema nervioso central. En el modelo animal de esclerosis múltiple, EAE, la inoculación de péptidos de MOG provoca enfermedad desmielinizante, como consecuencia de la respuesta inmune humoral (Bernard et al., 1997; Ichikawa et al., 1999). Además, en ausencia de anticuerpos específicos contra MOG, la patología en los modelos animales es solamente inflamatoria, mientras que, por el contrario, la inyección de un anticuerpo monoclonal específico contra MOG al inicio de la enfermedad produce una extensiva desmielinización, aumenta la respuesta inflamatoria y la severidad de la enfermedad (Iglesias et al., 2001).



**Gráfico 16:** Representación de la estructura secundaria de MOG

### **$\alpha$ B Cristalina**

$\alpha$ -Cristalina es un miembro de la familia de proteínas pequeñas del shock térmico (*sHSP*) (Klemenz et al., 1991), y constituye un agregado de dos polipéptidos,  $\alpha$ A- y  $\alpha$ B-Cristalina, de 20-kDa cada uno, que comparten un 55% de homología y que forman agregados heterogéneos solubles, en proporción 3:1.  $\alpha$ -Cristalina es la principal proteína soluble del cristalino del ojo, y tiene una importante función en el establecimiento del índice de refracción del citoplasma. Sin embargo, sus funciones celulares no se restringen al cristalino debido a su expresión ubicua, la actividad de autokinasa, la asociación con enfermedades neurodegenerativas y la capacidad de proteger contra el golpe térmico impidiendo la agregación proteica. De las dos subunidades que la constituyen,  $\alpha$ A-Cristalina se encuentra algo más restringida al cristalino, en tanto que  $\alpha$ B-Cristalina es más ubicua y su expresión se puede inducir por stress.

$\alpha$ B-Cristalina se ha encontrado expresada en altas concentraciones en el citoplasma de los oligodendrocitos y de los astrocitos en las lesiones desmielinizantes en la esclerosis múltiple, y se la considera por ello un antígeno mielínico. Además, en las lesiones activas de la esclerosis múltiple, caracterizadas por desmielinización y presencia de infiltrado inflamatorio, se puede detectar  $\alpha$ B-Cristalina entre las proteínas fagocitadas por los macrófagos, lo cual sugiere que la  $\alpha$ B-Cristalina es probablemente procesada y presentada como antígeno al sistema inmune celular (Bajramovic et al., 2000).

**Antígenos no mielínicos****S100  $\beta$** 

S100 $\beta$  es una proteína que une el calcio y que se encuentra abundantemente expresada en los astrocitos (Baimbridge et al., 1992). Forma parte de una familia de proteínas ligadoras de calcio constituida aproximadamente por 16 miembros, cada uno de los cuales presenta un patrón de expresión específico de tejido. Aunque la distribución de esta familia de proteínas no se encuentra restringida al SNC algunos miembros como S100 $\alpha$  y S100 $\beta$  actúan en el SNC como proteínas moduladoras del calcio intracelular y uniendo estímulos extracelulares con respuestas intracelulares a través de la interacción con otras proteínas citoplasmáticas (Zimmer et al., 1995), como por ejemplo neurocalcin delta (Okazaki et al., 1995).

S100 $\beta$  está codificada en el cromosoma 21. Presenta 3 exones y dos intrones (Allore et al., 1990) y los exones 2 y 3 codifican dos dominios independientes de unión al calcio. Aunque el significado funcional de esta proteína no está claro, se ha comprobado que contribuye al funcionamiento normal del hipocampo. Es una proteína pequeña y soluble, de 91 aminoácidos (Jensen et al., 1985), que representa un 0,4% del total de las proteínas solubles del SNC (Moore, 1965). Su expresión en el SNC está restringida a los astrocitos, quienes además secretan una forma biológicamente activa de S100 $\beta$  que puede ser detectada en el líquido cefaloraquídeo. En el sistema nervioso periférico se encuentra expresada en las células de Schwann, y en las células de Müller en la retina.

## ***Antígenos prostáticos***

### ***Antígeno Prostático específico (PSA)***

El PSA es una glicoproteína de 240 aminoácidos, con un peso molecular de 28-30 KDa. Se sintetiza en el epitelio prostático y se secreta como un constituyente normal del fluido seminal. Pequeñas cantidades son secretadas a la sangre, y por ello es posible detectarlo en suero.

Su función biológica es enzimática, es una serinoproteasa y la mayor proporción de PSA circulante en el suero se encuentra unida a inhibidores de proteasas endógenos, como la  $\alpha$ -1 antitripsina y la  $\alpha$ -2 macroglobulina. El PSA se expresa en altos niveles en el tejido prostático pero en bajos niveles en otros tejidos.

El gen que codifica para PSA tiene 5 exones. Su expresión está regulada por andrógenos: la transcripción del PSA comienza cuando el receptor de andrógenos, activado por la unión de su ligando, se une a la región promotora del gen PSA (Cleutjens et al., 1997)

Líneas celulares de origen no prostático, células sanguíneas y células de la médula ósea pueden expresar bajas cantidades de RNA de PSA (Smith et al., 1995).

### ***Antígeno Prostático Específico de Membrana (PSM)***

El PSM es una glicoproteína de transmembrana de tipo II, de 84,3 KDa constituida por 750 aminoácidos que presenta tres dominios estructurales: un dominio C-terminal extracelular (de 707 aminoácidos), un dominio transmembrana (de 24 aminoácidos) y un dominio N-terminal intracelular (de 19 aminoácidos) (Gregorakis et al., 1998). La secuencia presenta un 54% de homología con el mRNA del receptor de transferrina humano (Israeli et al., 1993).

El gen que la codifica se localiza en el cromosoma 11, el DNA genómico tiene un tamaño de 93525 pb y se organiza en 19 exones y 18 intrones. Su expresión está aumentada por bFGF, TGF- $\alpha$  y EGF (Israeli et al., 1994a) y disminuida por andrógenos como la 5 $\alpha$ -DHT (Israeli et al., 1994b).

Su función biológica es enzimática, es una  $\alpha$ -dipeptidasa expresada en la superficie de las células epiteliales prostáticas (Gregorakis et al., 1998). Se utiliza como marcador específico del epitelio prostático ya que las células basales son negativas para PSM.

También se ha detectado expresión de PSM fuera de la próstata, en la mucosa duodenal, los túbulos renales y en linfocitos de sangre periférica (Lintula and Stenman, 1997).

En tejido prostático normal existe además una variante mas corta resultante de splicing alternativo, llamada PSM'. PSM' ha perdido 266 nucleótidos del extremo 5' del cDNA que codifican para el codón de iniciación de la traducción y codones que codifican para 57

aminoácidos que forman parte del dominio de transmembrana de PSM. La proteína resultante es una proteína de 693 aminoácidos, con un peso molecular de 78 KDa, de localización citosólica (Su et al., 1995).

Aun no está clara la consecuencia biológica de estas variantes de *splicing* en la próstata, pero se ha demostrado que en los tumores primarios de próstata, el PSM es la forma predominante, mientras que en la próstata normal, la forma corta PSM' es la predominante (Su et al., 1995).



## PUBLICACIONES

Los resultados descritos en esta memoria están siendo preparados para su publicación:

Bruno R, Tolosa E, Sospedra M, Francesc-Ferrer X, Melms A, Foz-Sala M, Coll-Cantí J and Pujol-Borrell R.

*Transcription of Nicotinic Acetylcholine Receptor subunits in human thymus: three patterns of expression emerge.*

Bruno R, Sospedra M, Sabater L, Francesc-Ferrer X, Escudero D, Martínez-Cáceres E and Pujol-Borrell R.

*Multiple Sclerosis candidate autoantigens except MOG are transcribed in human thymus.*

Otras publicaciones relacionadas con esta tesis:

Sospedra M, Ferrer-Francesch X, Dominguez O, Juan M, Foz-Sala M, Pujol-Borrell R.

Transcription of a broad range of self-antigens in human thymus suggests a role for central mechanisms in tolerance toward peripheral antigens. *J Immunol* 1998; 161: 5918-29.