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Faculty of Biosciences  
Department of Genetics and Microbiology  
Genome Instability and DNA Repair group

**GENETIC AND PROTEOMIC STUDY OF *ERCC4/XPF*  
IN DNA REPAIR AND HUMAN DISEASES**

**DOCTORAL THESIS**

**Maria Marín Vilar**





Universitat Autònoma de Barcelona

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## GENETIC AND PROTEOMIC STUDY OF *ERCC4/XPF* IN DNA REPAIR AND HUMAN DISEASES

Dissertation respectfully submitted by

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To Universitat Autònoma de Barcelona in partial fulfilment of the requirements for the degree of Doctor of Philosophy, as per the Doctorate Program in Genetics with the International Degree Mention.

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***“There is a driving force more powerful than steam, electricity and nuclear power:  
the will.”***

*Albert Einstein*



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*“La ciencia avanza de verdad sólo cuando desobedece, sólo cuando es rebelde.”*  
(Delegada zapatista de los Caracoles, 2a edición ConCiencias por la Humanidad, 2017)

*“Instruïu-vos, perquè necessitarem tota la nostra intel·ligència. Emocioneu-vos, perquè necessitarem tot el nostre entusiasme. I organitzeu-vos, perquè necessitarem tota la nostra força.”* (Antonio Gramsci, *L'ordine Nuovo*, 1919)

## ABBREVIATIONS

<b>6-4 PPs</b>	<i>6-4 photoproducts</i>
<b>aa</b>	<i>Amino-acid</i>
<b>a-EJ</b>	<i>Alternative end joining</i>
<b>ATM</b>	<i>Ataxia-telangiectasia</i>
<b>ATR</b>	<i>ATM and Rad3-related</i>
<b>ATRIP</b>	<i>ATR interacting protein</i>
<b>BER</b>	<i>base excision repair</i>
<b>BLM</b>	<i>Bloom syndrome RecQ-like helicase</i>
<b>bp</b>	<i>Base pair</i>
<b>CBP</b>	<i>Calmodulin binding protein</i>
<b>C. elegans</b>	<i>Caenorhabditis elegans</i>
<b>C-NHEJ</b>	<i>Canonical nonhomologous end joining</i>
<b>CDK</b>	<i>Cyclin-dependent kinase</i>
<b>CoIP</b>	<i>Coimmunoprecipitation</i>
<b>CPDs</b>	<i>Cyclobutane-pyrimidine dimers</i>
<b>CRISPR</b>	<i>Clustered Regularly Interspaced Short Palindromic Repeats</i>
<b>CS</b>	<i>Cockayne Syndrome</i>
<b>CSA</b>	<i>Cockayne syndrome WD repeat protein A</i>
<b>CSB</b>	<i>Cockayne syndrome protein B</i>

<b>DEB</b>	<i>Diepoxybutane</i>
<b>DDR</b>	<i>DNA damage repair</i>
<b>DMEM</b>	<i>Dulbecco's Modified Eagle Medium</i>
<b>DTT</b>	<i>Dithiothreitol</i>
<b>DUBs</b>	<i>Deubiquitylases</i>
<b>D. melanogaster</b>	<i>Drosophila melanogaster</i>
<b>DNA2</b>	<i>DNA replication helicase/nuclease 2</i>
<b>DNA-PK</b>	<i>DNA-dependent protein kinase</i>
<b>DSB</b>	<i>Double strand break</i>
<b>dsDNA</b>	<i>Double strand DNA</i>
<b>E. coli</b>	<i>Escherichia coli</i>
<b>EdU</b>	<i>5-ethynyl-2'-deoxyuridine</i>
<b>EMA</b>	<i>Ethidium Monoazide Bromide</i>
<b>EU</b>	<i>5-ethynyluridine</i>
<b>EV</b>	<i>Empty vector</i>
<b>FA</b>	<i>Fanconi anemia</i>
<b>FAN1</b>	<i>Fanconi Associated Nuclease 1</i>
<b>FBOC</b>	<i>Familiar breast and ovarian cancer</i>
<b>FBS</b>	<i>Fetal Bovine Serum</i>
<b>GFP</b>	<i>Green fluorescence protein</i>
<b>GG-NER</b>	<i>Global genome repair of NER</i>
<b>GGR</b>	<i>Global genome repair</i>

<b>HEK293T</b>	<i>Human embryonic kidney 293T</i>
<b>HhH</b>	<i>Helix-hairpin-helix</i>
<b>HJ</b>	<i>Holliday junction</i>
<b>HMGN1</b>	<i>High mobility group nucleosome-binding domain-containing protein 1</i>
<b>HR</b>	<i>Homologous recombination</i>
<b>ICL</b>	<i>Interstrand crosslink</i>
<b>ICLR</b>	<i>Interstrand crosslink repair</i>
<b>Lig1</b>	<i>Ligase I</i>
<b>Lig3</b>	<i>Ligase III</i>
<b>MH</b>	<i>Microhomologies</i>
<b>MMC</b>	<i>Mitomycin C</i>
<b>MMEJ</b>	<i>Microhomology-mediated end joining</i>
<b>MMR</b>	<i>Mismatch repair</i>
<b>MN</b>	<i>Micronucleus</i>
<b>NBS1</b>	<i>Nijmegen Breakage Syndrome 1</i>
<b>NER</b>	<i>Nucleotide-excision repair</i>
<b>NHEJ</b>	<i>Nonhomologous end joining</i>
<b>PAM</b>	<i>Protospacer Adjacent Motif</i>
<b>PARP1</b>	<i>Poly (ADP-ribose) polymerase 1</i>
<b>PCNA</b>	<i>Proliferating cell nuclear antigen</i>
<b>PEI</b>	<i>Polyethylenimine</i>

<b>PI</b>	<i>5-ethynyluridine</i>
<b>Pol θ</b>	<i>Polymerase theta</i>
<b>Polη</b>	<i>Polymerase eta</i>
<b>RIR</b>	<i>Replication-independent repair</i>
<b>RPA</b>	<i>Replication protein A</i>
<b>SBP</b>	<i>Streptavidine binding protein</i>
<b>S. cerevisiae</b>	<i>Saccharomyces cerevisiae</i>
<b>SILAC</b>	<i>Stable isotope labeling by amino acids in cell culture</i>
<b>S. pombe</b>	<i>Schizosaccharomyces pombe</i>
<b>SSA</b>	<i>Single strand annealing</i>
<b>ssDNA</b>	<i>Single strand DNA</i>
<b>SD</b>	<i>Standard deviation</i>
<b>SDSA</b>	<i>Synthesis dependent strand annealing</i>
<b>SEM</b>	<i>Standard error of the mean</i>
<b>TALEN</b>	<i>Transcription activator-like effector nuclease</i>
<b>TAP</b>	<i>Tandem affinity purification</i>
<b>TC-NER</b>	<i>Transcription-couple repair of NER</i>
<b>TCR</b>	<i>Transcription-coupled repair</i>
<b>T-loop</b>	<i>Telomeric loop</i>
<b>TLS</b>	<i>Translesion synthesis</i>
<b>TFI IH</b>	<i>Transcription initiation factor IIH</i>
<b>TOP1</b>	<i>Topoisomerase I</i>

<b>UBZ</b>	<i>Ubiquitin-binding</i>
<b>USP7</b>	<i>Ubiquitin-specific-processing protease 7</i>
<b>USP11</b>	<i>Ubiquitin Carboxyl-terminal hydrolase 11</i>
<b>UV</b>	<i>Ultraviolet</i>
<b>UV-DDB</b>	<i>UV radiation-DNA damage-binding protein</i>
<b>UVSSA</b>	<i>UV-stimulated scaffold protein A</i>
<b>vs</b>	<i>Versus</i>
<b>WB</b>	<i>Western blot</i>
<b>WCE</b>	<i>Whole cell extract</i>
<b>XAB2</b>	<i>XPA-binding protein 2</i>
<b>XFE</b>	<i>Segmental Progeria</i>
<b>X. laevis</b>	<i>Xenopus laevis</i>
<b>XP</b>	<i>Xeroderma Pigmentosum</i>
<b>XP-F</b>	<i>Xeroderma Pigmentosum complementation group F</i>



## Abstract

A change in an organism's DNA can affect all the aspects of its life, until the point of compromising it. To overcome this, cells have evolved sophisticated machineries of DNA damage repair. Thus, our DNA contains the necessary information to produce proteins that participate in damage recognition, binding, excision and in the reestablishment of correct genetic information. One of these proteins is the XPF endonuclease, which is the catalytic subunit of the stable heterodimer XPF-ERCC1, able to incise at the 5' side at different DNA damages. This essential protein is encoded by *ERCC4* gene and participates in multiple genome maintenance pathways including nucleotide excision repair (NER), interstrand crosslink (ICL) repair, double strand break (DSB) repair pathways such as microhomology-mediated end joining (MMEJ) and single strand annealing (SSA). XPF has also been suggested to have possible backup roles in repairing oxidative damage and in telomere maintenance besides to have a role in the response of cancer cells to chemotherapy. Considering its wide involvement in multiple DNA repair pathways, it is not surprising that *ERCC4* mutations are associated to a range of human diseases including Xeroderma Pigmentosum (XP), Segmental Progeria (XFE), Fanconi Anemia (FA), Cockayne Syndrome (CS) and several cases combined diseases of Xeroderma and Cockayne syndromes (XPCSCD). A better understanding of (i) the correlation between the pathogenic mutations and patients' phenotype and (ii) the essential DNA repair mechanisms is expected to promote a faster development of possible treatments. Here we report a detailed overview of functional studies performed with a set of cells containing pathogenic XPF mutations in a genetically homogeneous background. The selected XPF mutants, located in different domains of the protein, and the resulting human syndromes, were the following: XPF<sup>R153P</sup> (XFE), XPF<sup>I225M</sup> (XP), XPF<sup>L230P</sup> (FA), XPF<sup>C236R</sup> (CS), XPF<sup>R589W</sup> (XP/XPCSCD), XPF<sup>R689S</sup> (FA), XPF<sup>R799W</sup> (XP/XFE-CS). Detailed functional studies include the analyses of NER pathway (UVC sensitivity, UDS and RRS) and ICLR pathway (ICL sensitivity, ICL-induced G2/M arrest and

ICL-induced chromosome fragility). Our results emphasize the importance of other factors, beyond protein position of the variant, such as protein levels, cell localization and the molecular interactions, in order to associate any XPF mutant to a clinical phenotype. In this framework, and to increase our knowledge about XPF interactions that can regulate XPF functions in the distinct DNA repair pathways, we investigated the XPF interactome. The implementation of the most advanced proteomic techniques including tandem affinity purification, co-immunoprecipitation and SILAC, coupled to mass spectrometry led us to identify a new XPF interactor involved in genome maintenance: USP11, a deubiquitinase that is known to regulate the activity of functionally related proteins such as BRCA2 or XPC. Here we prove XPF-USP11 interaction occurs irrespective of DNA damage and is DNA-independent. We also demonstrate that USP11 regulates DSB repair by SSA and possibly NER but is not involved in HR or ICLR.

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# **Introduction**



# I. INTRODUCTION

## **I.1- GENOMIC INSTABILITY AND DNA REPAIR**

The maintenance of genome integrity is crucial for the survival of cells because it ensures the faithful transmission of genetic information to the progeny and safeguards the proper functioning and survival of all the organisms. DNA is by far the most prone subcellular molecule to be threatened by lesions and yet the one that can deal better with them due to multiple DNA repair systems. The different DNA repair pathways are essential to identify DNA damage and correct it, therefore investing in genome maintenance (Hoeijmakers 2009).

Human cells have around 24,000 genes confined in 3 billion base pairs (bp). It is estimated that DNA damage occurs at a rate of 10,000 to 100,000 molecular lesions per cell per day (Vermeij et al. 2014). Thanks to the DNA repair pathways most of these lesions can be fixed but permanent changes in the DNA may occur and they are defined as mutations. If these unrepaired lesions affect critical genes, the integrity of the genome will be compromised and hence the cells' stability.

Genome instability can be induced by different DNA damage sources: exogenous physical and chemical agents and endogenous chemical genotoxic agents which are products of metabolism (Hoeijmakers 2009). Accumulation of cellular damage can lead to two different processes: ageing and cancer (Lopez-Otin et al. 2013). Ageing is produced when damage interferes with the vital processes of the cell and drives it to apoptosis or senescence, while the hallmark of cancer is the accumulation of damage that confers aberrant proliferation advantages to certain cells which become immortal.

DNA repair mechanisms have evolved to detect the damage and remove it, therefore ensuring the integrity of the genome by several multiple protein pathways such as base-

excision repair (BER), nucleotide-excision repair (NER), transcription-coupled repair (TCR), nonhomologous end joining (NHEJ), homologous recombination (HR) and interstrand cross-link (ICL) repair. This wide machinery evidences the importance of investment in genome maintenance and cell survival.

## I.2- XPF PROTEIN

XPF is a protein encoded by *ERCC4* gene and also receives the alias of FANCQ. It was identified as the defective gene in Xeroderma Pigmentosum complementation group F (XP-F) (Sijbers et al. 1996) and its cDNA complemented the human XPF cells and the deficient rodent NER cells *Ercc4* and *Ercc11* (Yagi et al. 1998). The human *ERCC4* is located in 16p13.1-p13.2 and encodes for a 916 amino-acid (aa) protein (Brookman et al. 1996) which forms a stable heterodimer with ERCC1 in order to constitute a structure-specific endonuclease that incises in the 5' side of DNA damage. This dependent stabilization is reciprocal; the heterodimer is found to be formed stoichiometrically 1:1 and it is essential for life, since the complete inactivation of the *ERCC4* gene in humans seems to be incompatible with postnatal survival (Osorio et al. 2013; Tian et al. 2004).

XPF is organized in three different domains: a N-terminal helicase domain (residues 15-647), a central nuclease domain (residues 667-824) and a C-terminal helix-hairpin-helix (HhH) domain (residues 848-916) (Klein Douwel et al. 2017) (Figure 1).



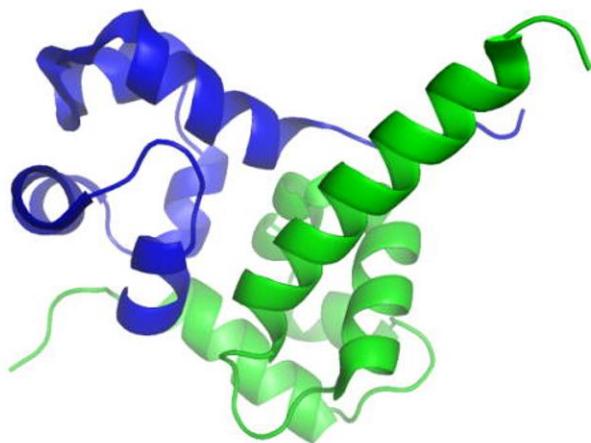
**Figure 1: Domain organization of XPF and ERCC1** (adapted from (McNeil and Melton 2012)).

The N-terminal helicase domain is homologous to superfamily II helicases and contributes to DNA binding activity, the nuclease domain of XPF is responsible for the catalytic activity of the protein (Tsodikov et al. 2005), and the C-terminal HhH domain works as a scaffold for the correct folding of ERCC1 to form a functional endonuclease (Tripsianes et al. 2005) besides to be also involved in DNA substrate binding. Residues confined in 224-297 from ERCC1 protein and 825-916 from XPF are the regions responsible for the stable heterodimer formation (de Laat et al. 1998).

From the two components of the heterodimer, XPF specifically recognizes single strand DNA (ssDNA) through the N-terminal helicase domain, while ERCC1 rather binds to dsDNA through its hairpin region. This different substrate specificity allows the heterodimer to bind to ss/double strand DNA (dsDNA) substrate and dictates the incision position during the DNA repair (Das et al. 2017).

Previously, a time-consistent model proposed that the two HhH domains of the two proteins were binding to ssDNA and the central domain of ERCC1 bound ssDNA/dsDNA junctions with a preference for 5' single-stranded overhang (Tsodikov et al. 2005). Notwithstanding, it has been recently elucidated that ERCC1 specifically recognizes dsDNA through its HhH domain, and that the C-terminal heterodimer complex binds more tightly to the sites with a ss-dsDNA junction, such as bubble and splayed arm substrates than to either dsDNA or ssDNA alone. In fact, the two motifs of the HhH domain form a cavity where a guanine base can be bound. Apparently, there is a sequence dependence in the recognition of the damaged DNA, being guanine the most prone to be recognized, followed by cytosine and thymidine, and leaving adenine as the less recognized one.

The two HhH domains of XPF and ERCC1 have been solved by crystallography and the dimer structure consists of a wide network of hydrophobic interactions between the two proteins that make them to mutually stabilize one another (Croteau et al. 2008) (Figure 2).

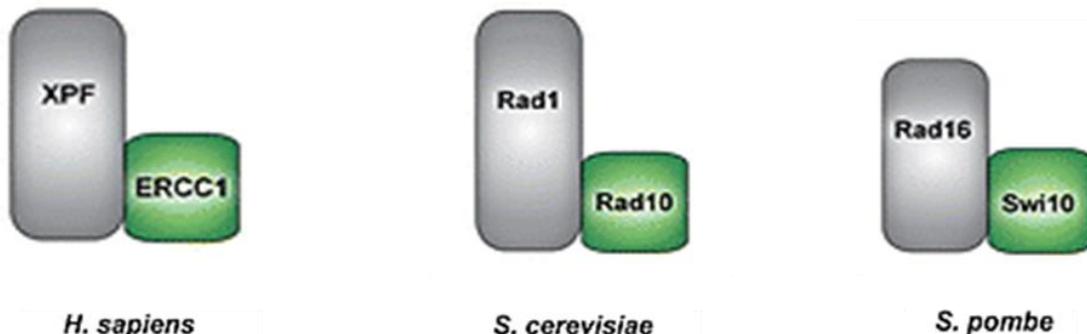


**Figure 2: Crystal structure of the human C-terminal HhH<sub>2</sub> domain complex of XPF (blue) and ERCC1 (green)** (adapted from (Croteau et al. 2008)).

Considering this ss-dsDNA junction, XPF would bind to the 5' site of the ssDNA and this ssDNA would be linked by the 3' to the dsDNA that ERCC1 would recognize. Putting this all together, an important idea to emphasize would be that the XPF-ERCC1 would be positioned according to the DNA structure and in a sequence dependent way, which would moreover dictate the cleavage efficiency of the endonuclease heterodimer (Das et al. 2017).

### I.2.1 EVOLUTION OF XPF PROTEIN

XPF protein seems to be highly conserved during evolution; its structure reveals similarities in archaeal (Rouillon and White 2011) and in eukarya (Barve et al. 2013), proving its essential role for the cell maintenance. Mammalian XPF shares a degree of homology with Rad1, a protein of 1,100 aa from *Saccharomyces cerevisiae* (*S. cerevisiae*); Rad16, a protein of 892 aa from *Schizosaccharomyces Pombe* (*S.pombe*) and MEI-9, a 946 aa *Drosophila melanogaster* (*D. melanogaster*) protein that ranges from 28% homology with Rad1 to 40% of MEI-9 (Brookman et al. 1996) (Figure 3).



**Figure 3: Heterodimeric associations of mammalian XPF and its homologous in yeast** (adapted from (Ciccia et al. 2008)).

Rad1 and Rad16 are essential components in yeast. Rad1 activity requires to be bound in a heterodimer to Rad10 by its C-terminal regions (809-997 aa of Rad1 and 90-210 aa of Rad10). This complex would be the homologous to the mammalian XPF-ERCC1 and it shows ssDNA endonuclease activity, cleaving at 3' of the ssDNA at the junction with dsDNA, with a clear preference for splayed-arm DNA substrates (Ciccia et al. 2008). This complex is an essential component of NER in yeast, it interacts with Rad14, the homologous of mammalian XPA protein from NER (de Laat et al. 1998), and it has also been proved to be involved in the removal of the two non-homologous 3'-ended ssDNAs that are formed as intermediates in single strand annealing (SSA) (Prakash and Prakash 2000). Despite all these similarities, the XPF interaction site of ERCC1 is different from the Rad1-binding site of Rad10. XPF interaction site in ERCC1 comprises aa 224-297, which would be out of the homologous region in Rad10; the correspondent region of XPF-binding in Rad10 comprises aa 98-214. This shows an extended C-terminal region in mammalian ERCC1, which is composed by the double HhH motif (residues 236-289). This HhH motifs have been found in many DNA break processing enzymes like in UvrC NER protein of *Escherichia coli* (*E. coli*) or *S. pombe* and contribute to DNA binding (de Laat et al. 1998).

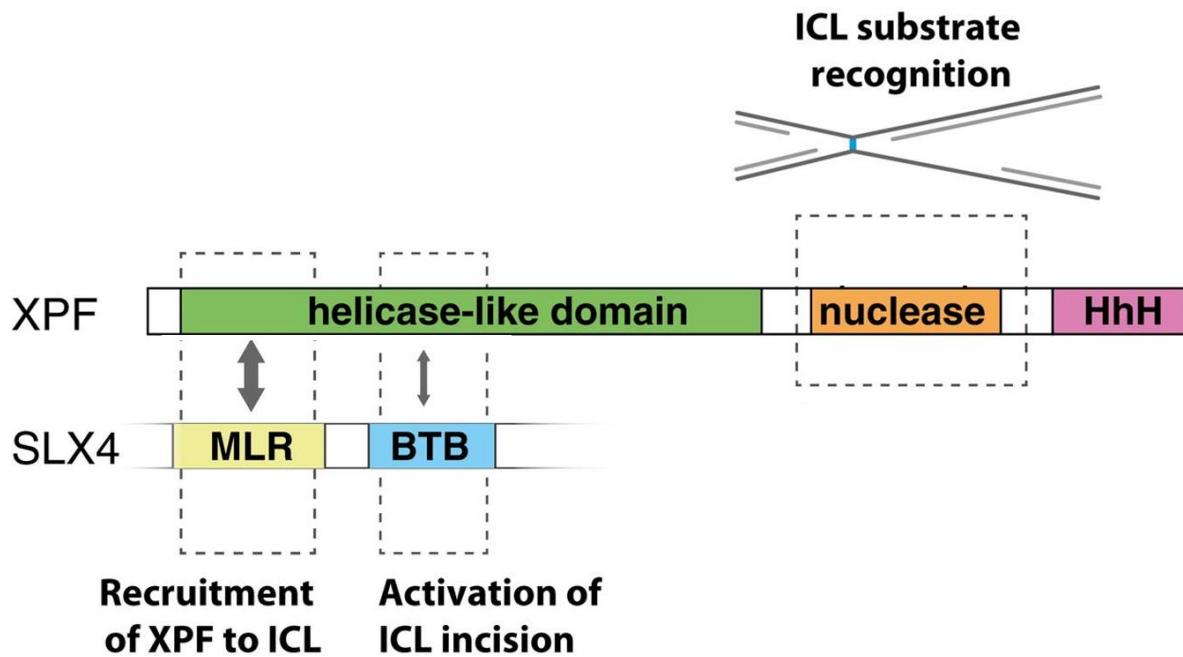
Rad16 is the ortholog of XPF in *S. pombe* and it forms a complex with Swi10/Rad23 (ERCC1). Rad16 promotes recombination repair of broken replication forks without using sister chromatids repair, but ectopic donor sequences, thus contributing to genome stability.

Rad1 and Rad16 seem to not have any role in meiosis while Mei-9, the XPF NER-homologue in *D. melanogaster* is also essential for resolution of meiotic chiasmata. In *Caenorhabditis elegans* (*C. elegans*), XPF shares roles with other nucleases involved in meiosis such as Mus81 and SLX-1 (Mastro and Forsburg 2014).

### I.2.2 XPF INTERACTIONS

Besides the already explained interaction of XPF with ERCC1, the nuclease heterodimer is able to establish temporal interactions with other proteins to participate in the different damage repair pathways in which it is seen to be involved. XPF interactome had not been deeply investigated but some important interactor partners are already unrevealed and, considering the relevance they can have to understand the different pathways where XPF works, potentially more will arise.

One of these important interactions is established with SLX4, a protein involved in repairing the damage produced by ICL causing agents, topoisomerase I (TOP1) inhibitors and in Holliday junction (HJ) resolution. This protein works as a platform for several nucleases in ICL repair (ICLR). Two interaction sites are described between XPF and SLX4 (Figure 4): the BTB domain and the MUS312/MEI9 interaction-like, or MLR domain, one ensuring the recruitment of XPF to the damage sites and another one enhancing its nuclease activity (Y. Kim 2014). The BTB domain of SLX4 is not essential for the SLX4 and XPF interaction, the binding is established through the SLX4 N-terminal segment MLR domain (Y. Kim et al. 2013). Recent studies performed in *Xenopus laevis* (*X. laevis*) suggest the interaction takes place most likely between the MLR domain and the 230 leucine of human XPF aa sequence but there is a second interaction site in BTB domain of SLX4 and the residues 323-326 of XPF, which is transient but important to promote XPF nuclease activity (Klein Douwel et al. 2017).



**Figure 4: Model of XPF-SLX4 domains interactions for ICLR** (adapted from (Klein Douwel et al. 2017)).

SLX4 is a SUMO E3 ligase able to SUMOylate itself and XPF, although this post translational modification does not seem to interfere in any of these two proteins ICLR ability (Guervilly et al. 2015).

The replication protein A (RPA) is the major protein that binds to ssDNA in eukaryotic cells. It prevents the ssDNA from winding back on itself or forming secondary structures during replication. RPA recruitment to ssDNA stimulates XPF-ERCC1 activity and selectively permits the endonuclease to incise at 5' of the fork junction when there is a nascent leading strand (Matsunaga et al. 1996). Therefore, RPA is considered an XPF interactor partner (Abdullah et al. 2017). In fact, a direct interaction between XPF and RPA has been proved in the presence of DNA bubble substrates (Bessho et al. 1997) which explains how RPA orientates properly XPF-ERCC1 on the DNA substrate.

The XPF-ERCC1 heterodimer physically interacts with XPA, another protein from NER pathway, but in this case it does it through ERCC1 instead of XPF subunit. The central

domain of ERCC1 is structurally homologous to the XPF nuclease domain but it differs with it in the presence of a grooved lined with basic aromatic residues that interact with XPA protein. Consequently, ERCC1, and therefore, XPF heterodimer, is connected to the NER machinery through XPA (Su et al. 2012).

Due to the wide range of processes where XPF is involved, it is reasonable that more interacting partners will come out.

### **I.3- XPF ROLE IN GENOMIC INSTABILITY SYNDROMES**

The XPF-ERCC1 heterodimer participates in multiple genome maintenance pathways, such as NER (Sijbers et al. 1996), ICLR (Damia et al. 1996), an alternative end joining (a-EJ) also known as microhomology-mediated end joining (MMEJ) (Ahmad et al. 2008), the SSA branch of double strand breaks (DSB) repair (Ahmad et al. 2008) and telomere maintenance (Zhu et al. 2003). Moreover, XPF has been found to have possible backup roles in repairing oxidative damage and DNA breaks with damaged ends (Scharer 2017). Considering its wide involvement in DNA repair machinery, it is not surprising the range of human diseases which are associated with mutations in XPF-ERCC1 heterodimer.

#### **I.3.1 NUCLEOTIDE EXCISION REPAIR AND PHOTOSENSITIVE SYNDROMES**

NER is one of the most versatile DNA damage repair pathways. It is involved in the removal of lesions caused by ultraviolet (UV) radiation like cyclobutane-pyrimidine dimers (CPDs) and 6-4 photoproducts (6-4 PPs), several natural and induced bulky chemical adducts, intrastrand crosslinks caused by some drugs and ROS-generated cyclopurines. All of these lesions distort the DNA double helix and must be removed to allow a proper DNA replication (Marteijn et al. 2014).

NER is divided in two subpathways (Figure 5): global genome repair (GGR) and TCR, which differ in the damage recognition process but share the same mechanism to incise at both sides of the lesion, repair and ligate the DNA gap.

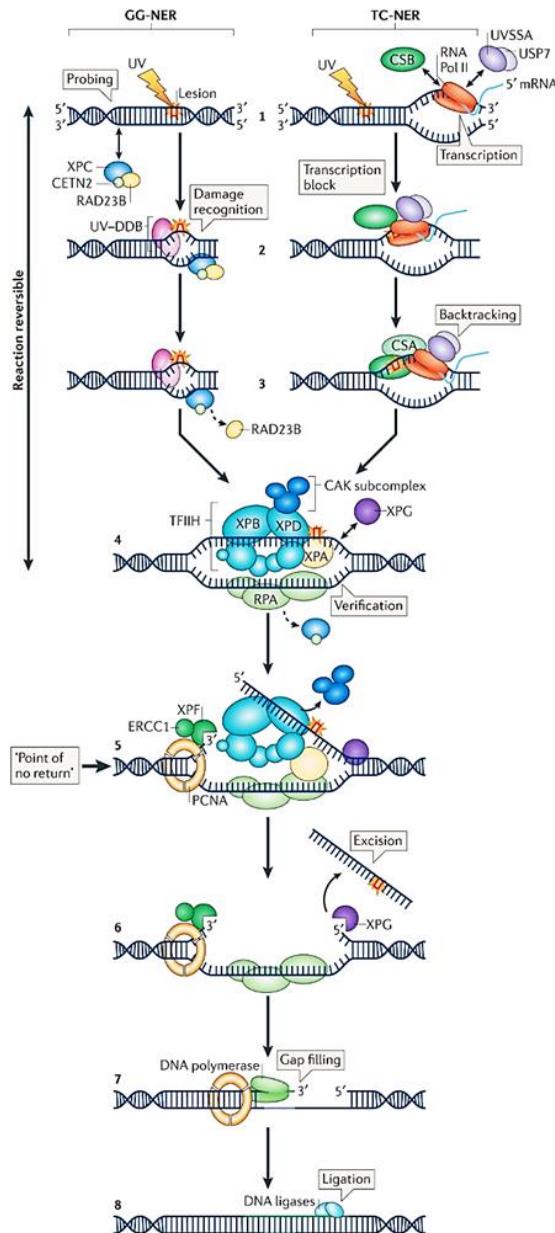


Figure 5: NER pathway (Marteijn et al. 2014).

In GGR subpathway, the entire genome is probed by the protein sensor XPC in association with RAD23B and CETN2 to find any helix distortion (Sugasawa et al. 1998). As some of the UV induced lesions just mildly destabilize the DNA helix, the XPC sensor is unable to detect them and requires the help of the UV radiation-DNA damage-binding protein (UV-DDB) complex, which comprises DDB1 and DDB2 (Chu and Chang 1988). In contrast, TCR subpathway is activated when the UV-induced lesions arrest the transcriptional machinery; the RNA Pol II stalls at the damaged site and recruits the proteins Cockayne syndrome WD repeat protein A (CSA or ERCC8) and the Cockayne syndrome protein B (CSB or ERCC6) (Fousteri et al. 2006). Both proteins are required for the recruitment of NER factors and some TCR-specific proteins such as UV-stimulated scaffold protein A (UVSSA), ubiquitin-specific-processing protease 7 (USP7) (X. Zhang et al. 2012), XPA-binding protein 2 (XAB2) (Kuraoka et al. 2008) and high mobility group nucleosome-binding domain-containing protein 1 (HMGN1) (Fousteri et al. 2006). It is thought that CSB is the protein involved in the backtracking that the RNA Pol II experiences when it is stalled at the lesion site, leaving it accessible for the rest of the complexes to be recruited for the damage repair (Sigurdsson et al. 2010).

Once the damage is recognized, the two molecular subpathways converge into a single one and the transcription initiation factor IIH (TFIIH or XPB) complex is recruited. The two helicases of this protein complex, XPB and XPD, extend the open DNA around the lesion in opposite directions. XPA protein binds to them and is able to detect distortions in the ssDNA, contributing to the damage verification. Following damage verification, the unwound DNA strand gets covered by ssDNA binding RPA and XPA recruits XPF-ERCC1 heterodimer by binding to ERCC1 (Tsodikov et al. 2007; Volker et al. 2001). This endonuclease complex, whose specificity is stimulated by RPA (Matsunaga et al. 1996), binds to the damaged strand to create an incision 5' to the lesion. Right after the lesion the trimeric proliferating cell nuclear antigen (PCNA) ring is loaded (Hutton et al. 2010) and recruits the polymerases which will fill the gap from the excision. The 5' incision sets the machinery in a “point of no return”, as the damaged DNA strand is already cleaved by one site and the process must move onwards to avoid the creation of damaging intermediates. Next step is the XPG 3' incision to the lesion which leaves a gap in the damaged strand of 22-30 nucleotides. The

gap filling is provided by the DNA Pol  $\delta$ , DNA Pol  $\varepsilon$  or DNA Pol  $\kappa$  and the process ends with the sealing of the nick by the DNA ligase 1 or DNA ligase 3 (A. R. Lehmann 2011).

### I.3.1.1 XERODERMA PIGMENTOSUM (XP)

*ERCC4* is also known as *XPF* because it was identified as the defective gene in complementation group F of Xeroderma Pigmentosum (XP) (Sijbers et al. 1996), an autosomal recessive syndrome with 100% penetrance characterized by extreme photosensitivity and a 10,000-fold increased risk of skin cancers due to failure to repair DNA lesions produced by UV light (Gregg et al. 2011). Ocular abnormalities and increased risk of cancers of the oral cavity are also very common among the patients (A. R. Lehmann et al. 2011). Despite XP is more frequently diagnosed in Asian patients (Imoto et al. 2013), there are few Caucasians cases with mild UV sensitivity and no skin cancer but presenting acute neurological syndromes such as recessive cerebellar ataxia and chorea (Carre et al. 2017). There have been described eight XP complementation groups (XP-A to XP-G and XPV); proteins from XP-A to XP-G are involved in the repair of the UV lesions while XPV is involved in DNA replication bypassing the UV-damage. XP patients of group F have mild XP symptoms and a reduced level of nuclear XPF indicating that its mutations promote mislocalization of the heterodimer XPF-ERCC1 to the cytoplasm of cells (Ahmad et al. 2010) and lead to insufficient levels of the endonuclease to complete NER functions.

### I.3.1.2 SEGMENTAL PROGERIA CASES (XFE)

Few cases described patients characterized by a progeria-like phenotype produced by a characteristic failure of the mutant XPF protein to properly translocate to the nucleus, likely through aggregation of the protein in the cytoplasm, and being avoided to be recruited to sites of active NER. The very low levels of nuclear XPF were apparently insufficient to support either NER or ICL. The patients presented severe photosensitivity, neurological and musculoskeletal abnormalities and hematopoietic symptoms (Mori et al. 2018; Niedernhofer et al. 2006).

### **I.3.1.3 COCKAYNE SYNDROME (CS)**

Some XPF mutations can produce Cockayne Syndrome (CS), a recessive disorder characterized by neurological and developmental abnormalities, growth and mental retardation, microcephaly, premature ageing and abnormal skin photosensitivity that does not lead to pigmentation changes or skin cancer (Kleijer et al. 2008).

Four subtypes of CS have been discriminated according to the type of onset and rate of progression. Molecularly, CS patients are diagnosed by a reduction of recovery of RNA and DNA synthesis in absence of UV sensitivity in fibroblasts following UV irradiation (He et al. 2017).

There are rare CS variants that combine features of CS and XP and receive the nomenclature of XPCSCD. These cases present severe UV sensitivity and cancer predisposition typical from XP and developmental abnormalities which are common in CS patients (Kashiyama et al. 2013; Natale and Raquer 2017).

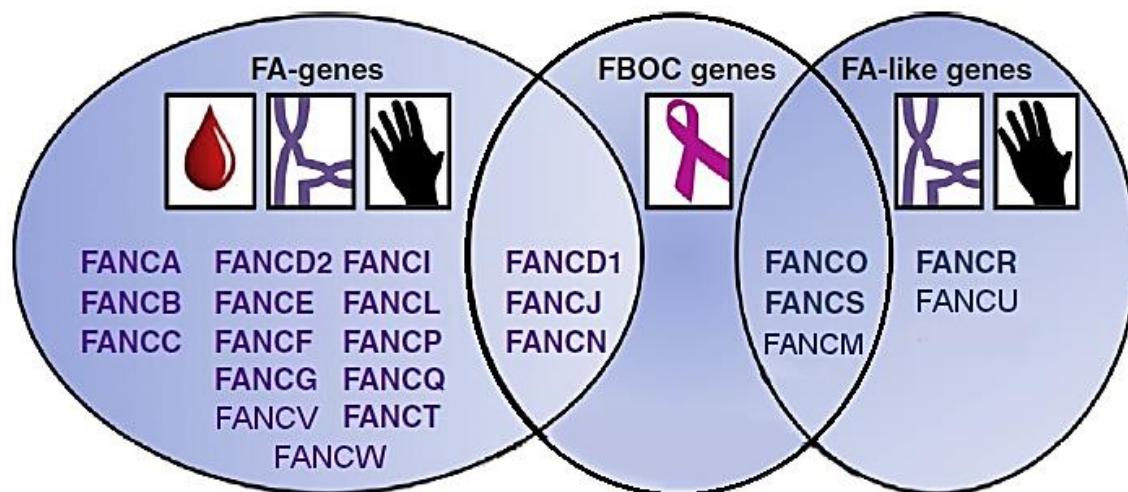
### **I.3.2 INTERSTRAND CROSSLINKS REPAIR AND FANCONI ANEMIA**

One of the most dangerous DNA lesions are the ICLs, since they covalently link the two strands of the DNA. These ICL block the DNA molecular machinery by inhibiting the strand separation which leads to the block of replication, transcription and segregation of DNA. About 40 unrepaired ICLs can kill a mammalian cell (Dronkert and Kanaar 2001). ICLs provoke a challenge for the DNA repair mechanisms because of the involvement of the two strands (McHugh et al. 2001), and, aside from leading to the cell death, they can induce mutations and DNA rearrangements which could drive to uncontrolled cell growth and tumor formation.

ICLs can be produced by the products of the cellular metabolism or by external agents like mitomycin C (MMC), diepoxybutane (DEB), cisplatin, nitrogen mustard and psoralens (Muniandy et al. 2010).

The detection and repair of these lesions require strictly organized multiple DNA repair pathways such as Fanconi anemia (FA) pathway, HR, translesion synthesis (TLS) and NER.

FA is a genetically and phenotypically heterogeneous disorder caused by germ-line mutations in genes that cooperate in the repair of DNA ICL. It is a rare disease with an incidence of 1-9 in 1,000,000 live births and an estimated carrier frequency of 1 in 250 in most populations (Mathew 2006). It was first reported by Guido Fanconi (1927) and its clinical features (all of incomplete penetrance) include bone marrow failure, pancytopenia, hyperpigmentation, skeletal malformations, small stature and urogenital abnormalities and cancer predisposition. FA cellular phenotype is characterized by extreme sensitivity to DNA cross-linking agents and chromosomal fragility (Bogliolo and Surralles 2015) and it is caused by mutations in any of the, until the date hereof, 22 identified complementation groups: *FANCA*, *FANCB*, *FANCC*, *FANCD1* (*BRCA2*), *FANCD2*, *FANCE*, *FANCF*, *FANCG* (*XRCC9*), *FANCI*, *FANCJ* (*BRIP1*), *FANCL* (*PHF9*), *FANCM*, *FANCN* (*PALB2*), *FANCO* (*RAD51C*), *FANCP* (*SLX4*), *FANCQ* (*XPF*), *FANCR* (*RAD51*), *FANCS* (*BRCA1*), *FANCT* (*UBE2T*), *FANCU* (*XRCC2*), *FANCV* (*REV7*) and *FANCW* (*RFWD3*) (Gueiderikh et al. 2017; Knies et al. 2017) from which twenty-one shows autosomal inheritance while *FANCB* maps to the X chromosome. It is expected that more associated genes will arise, although there is a controversy about whether its designation as FA genes comes along with the initial requirement of its association with a clinical case, bone marrow failure, chromosome fragility and malformations to be called FA genes. FA genes which mutations do not lead to bone marrow failure would be confined into FA-like genes while genes which biallelic mutations carriers are not linked to FA disease phenotype would be designated as FA-associated genes (Bogliolo and Surralles 2015) (Figure 6).



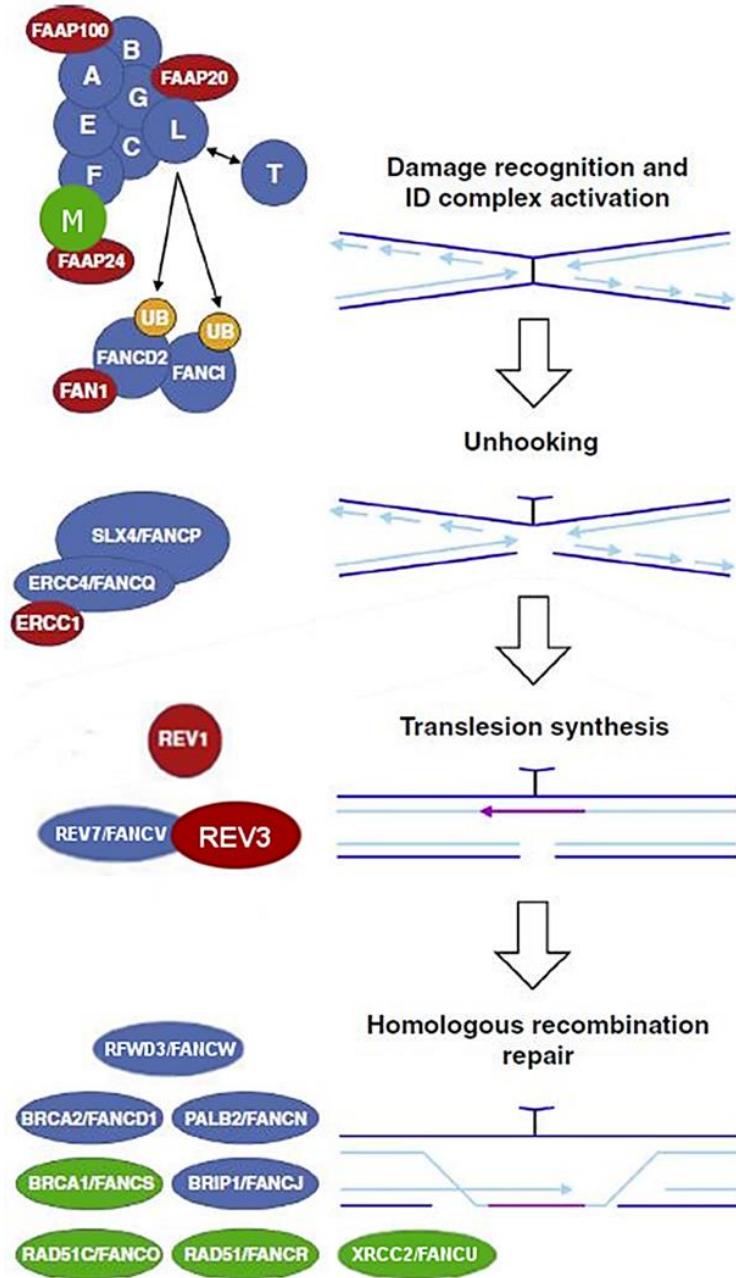
**Figure 6: Diagram of the relationship of FA-genes (biallelic mutations in these genes produce bone marrow failure, chromosome fragility and malformations), familiar breast and ovarian cancer (FBOC) genes and FA-like genes (biallelic mutations in these genes do not cause bone marrow failure) (Bogliolo et al. 2017; Catucci et al. 2017) (adapted from (Bogliolo and Surralles 2015)).**

### I.3.2.1 GROUP I: FA CORE COMPLEX

FA proteins from the twenty-two complementation groups reported have been divided in three functional groups.

Seven FA proteins (FANCA, FANCB, FANCC, FANCE, FANCF, FANCG, and FANCL) and six FA-associated proteins (FANCM/FAAP250, FAAP100, FAAP24, FAAP20, FAAP16 and FAAP10) constitute the nuclear FANCore complex. The core complex forms a high molecular weight E3 ubiquitin ligase complex, based on the ubiquitin ligase domain of FANCL. This complex function is to activate through monoubiquitination a second heterodimer formed by FANCD2 and FANCI (ID complex). The key event in initiating ICLs repair lies on the monoubiquitination of FANCD2, which is achieved through the functions of the gene product of *FANCT*, an E2 enzyme that connects the ubiquitin to the substrate, the gene product of *FANCL*, an E3 ubiquitin ligase, and the E1 ubiquitin-activating enzyme that activates the ubiquitin (van Twent et al. 2017). A mutation in any of the components of the FANCore or *FANCT* drives to an impairment in the monoubiquitination of the ID complex,

with the exception of mutations in FANCM, whose cells still show a residual FANCD2 monoubiquitination (Singh et al. 2009) (Figure 7).



**Figure 7: The FA pathway of ICLR.** Blue: proteins encoded by FA genes; green: proteins encoded by FA-like genes; red: FA-associated proteins (adapted from (Bogliolo and Surralles 2015)).

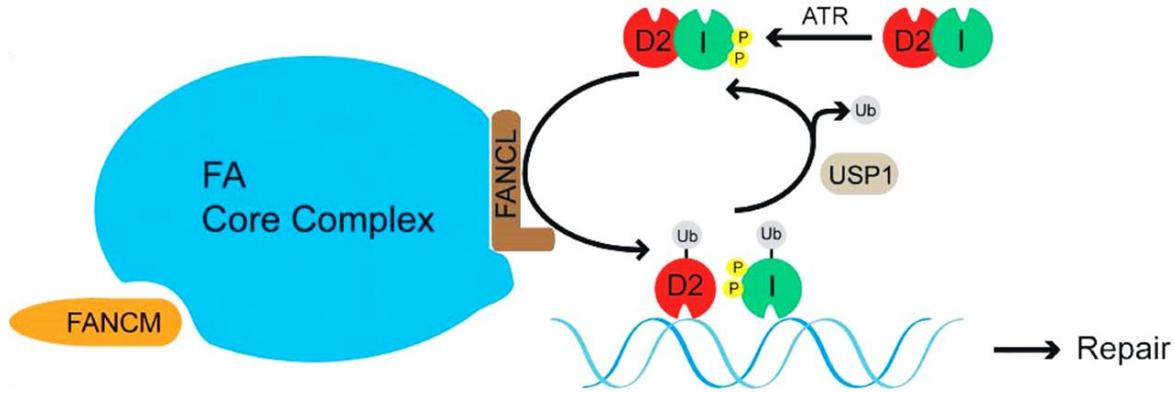
### I.3.2.2 GROUP II: ID COMPLEX

As already mentioned, group II is formed by FANCD2 and FANCI proteins associated in the so called ID complex, activated upon DNA damage by interdependent monoubiquitination. A recently discovered Tower domain of FANCD2 is crucial for a direct interaction with the DNA from the arrested replication fork, being this interaction the one that triggers the subsequently monoubiquitination (C. C. Liang et al. 2016).

Multiple constituents of the FA pathway (FANC-A, -G, -M, -D1 and the two components of ID complex) are phosphorylated by both ataxia-telangiectasia (ATM) and ATM and Rad3-related (ATR) kinases. While ATM coordinates the cellular response to DNA DSBs by phosphorylating substrates to halt cell cycle progression and proceed to damage repair, ATR kinase is activated in response to replicative stress. The activation of the FA/BRCA pathway starts with disassociation of the ID2 heterodimer, which is enhanced by ATM/ATR-mediated phosphorylation of FANCI (Boisvert and Howlett 2014).

The monoubiquitinated form of FANCD2 relocates in nuclear foci that colocalize with γH2AX in response to damage: ATR phosphorylates H2AX at stalled replication forks to recruit FANCD2 to DNA damage (Bogliolo et al. 2007).

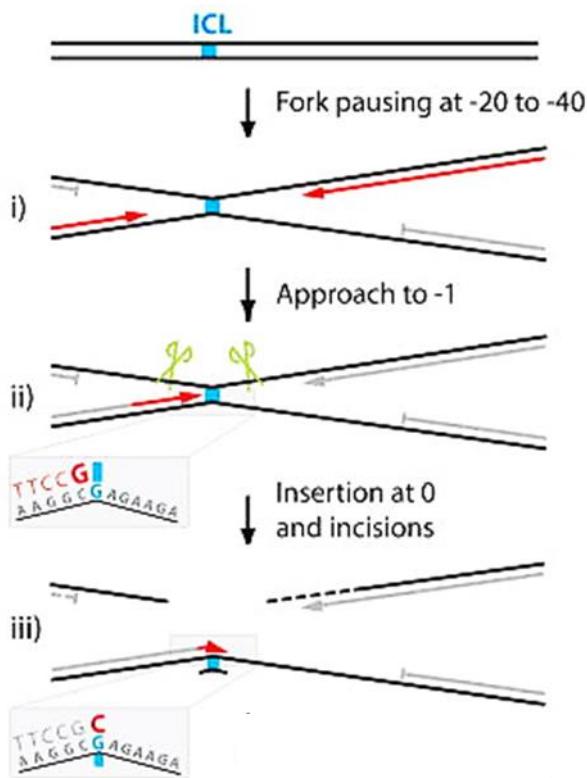
The USP1:UAF1 protein dimer is required then for the ID deubiquitination, a crucial step to complete the DNA repair process and the rescue of the stalled replication forks (Gueiderikh et al. 2017; Q. Liang et al. 2014). UAF1 protein binds and activates the ubiquitin-specific protease USP1 (Cohn et al. 2009). When the DNA damage repair is completed, the ID complex must be deubiquitinated in a critical step for the ICLR executed by USP1 deubiquitinase (Figure 8). USP1 levels are regulated at transcription level and they are highest when cells exit S phase or when the cell cycle is restored after DNA damage (Nijman et al. 2005). The lack of de-ubiquitination of the ID complex also produces ICL sensitivity, therefore suggesting that both monoubiquitination and deubiquitination of the complex are crucial steps for the pathway (J. M. Kim et al. 2009).



**Figure 8:** After the DNA damage, FANCI is phosphorylated and the ID complex is monoubiquitinated by FANCL. The ID complex is then able to be recruited to the chromatin to promote DNA repair. After its action, ubiquitination is removed by USP1 to recycle the ID complex (adapted from (Crossan and Patel 2012)).

### I.3.2.3 ICL INCISION

The principal ICLR pathway takes place in S phase of the cell cycle. Current models propose that the mechanism is initiated when two DNA replication fork collide with the linked strands and generate a new structure in which the leading strands stall about 20-40 nucleotides from the crosslink (Raschle et al. 2008) (Figure 9). One of the forks then advances up to a one nucleotide from the crosslinked base and stalls again (Knipscheer et al. 2009). The process is followed by the action of endonucleases involved in dual incisions on either side of the crosslink and “unhooks” it.

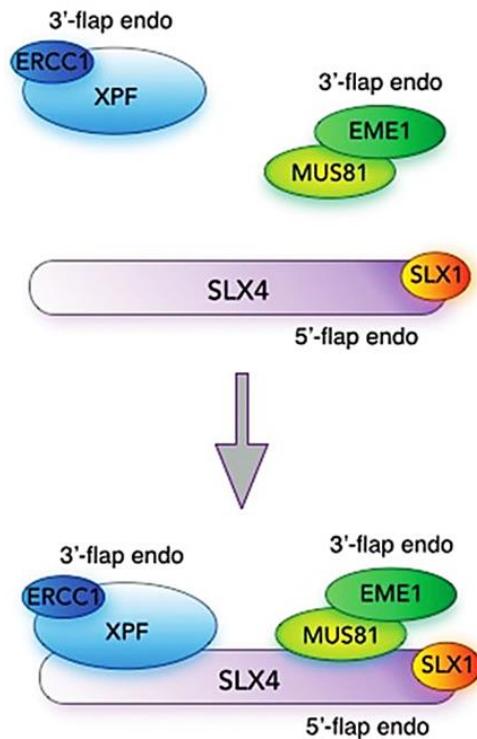


**Figure 9: Representation of fork collision and incisions in ICL.** The replication fork collides with the linked strands and gets initially arrested at -20 to -40 from the crosslink. Then approaches to it while one strand is incised at both sides of the crosslink. Insertion at 0 allows the extension of the nascent strand (adapted from (Knipscheer et al. 2009)).

The Unhooking step performed by proteins involved in dual incisions on either side is a fundamental step in ICLR of FA/BRCA pathway. Following the monoubiquitination of the ID complex six different endonucleases are involved in ICLR: SLX1-SLX4, XPF-ERCC1, MUS81-EME1, FAN1, SNM1A and SNM1B. These nucleases have different preferences for the different X-shape incisions substrates that the crosslink causes when the replication forks are stalled.

### I.3.2.3.1 SLX PROTEINS: SLX1-SLX4

SLX4 is a large protein of about 200 KDa described as a platform where FANCQ/XPF-ERCC1, MUS81-EME1 and SLX1 nucleases can dock (Figure 10) to perform their roles in repairing DNA damage induced by ICLs or TOP1 inhibitors, in HJ resolution and telomere maintenance (Sarkar et al. 2015).



**Figure 10: SLX4 is a platform for the formation of the SLX1-SLX4-MUS81-EME1-XPF-ERCC1 complex** (adapted from (West et al. 2015)).

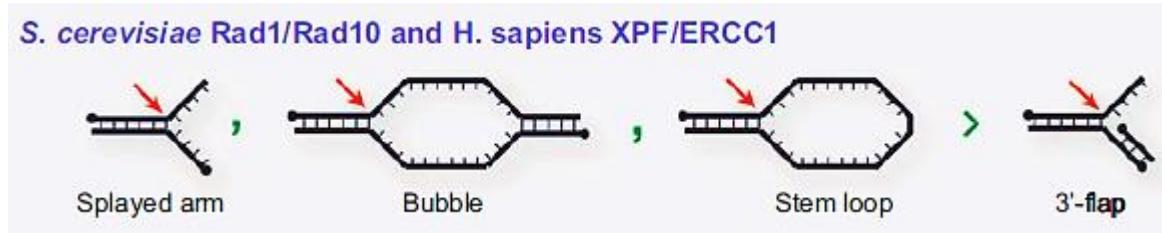
HJ are defined as four branched DNA structures that arise as intermediates of HR resolution. These intermediates must be correctly repaired to allow proper segregation during mitotic division (West et al. 2015). SLX4 contains two tandem UBZ4-type domains: UBZ-1 and UBZ-2. UBZ-1 is the required domain to recognize a poly-ubiquitylated protein at DNA damage sites and the proper recruitment of the protein to these sites, while UBZ-2 is needed for the resolution of HJ (Lachaud et al. 2014). It is the protein in charge of orchestrating the cooperation of the involved nucleases (SLX1, MUS81 and XPF) and this structure specific selective endonuclease is called SMX (Wyatt et al. 2017).

SLX1 is the catalytic subunit of the SLX4-SLX1 structure-specific endonuclease. Studies were SLX1 was depleted showed a very mild sensitivity to cross-linking agents, therefore showing that the SLX1-SLX4 heterodimer is not the crucial endonuclease for the unhooking but acts as a platform recruiting other nucleases. Despite being a promiscuous endonuclease able to cut a wide variety of DNA substrates like splayed arms, 5' flaps, 3' flaps and HJ, it has a preference for 5' flaps generated on the 5' side of the crosslink (Saito et al. 2012), and it incises them at the ssDNA-dsDNA junction when the leading strand has been extended to the -1 position.

#### I.3.2.3.2 XPF-ERCC1

Cells lacking FANCQ/XPF-ERCC1 showed a strong ICLR impairment that was restored after genetic complementation with wild type cDNA (Bogliolo et al. 2013). The physical interaction between the N-terminus of SLX4 and XPF is critical for the unhooking (Crossan et al. 2011; Hodskinson et al. 2014) and almost the entire SF2 helicase-like domain of XPF is required for the interaction (K. Hashimoto et al. 2015). Loading of XPF-ERCC1 to the crosslink was reduced in the absence of SLX4, but this did not occur in the other way around, indicating that SLX4 leads XPF-ERCC1 to the stalled replication forks (Klein Douwel et al. 2014). Some mutations found in XPF disrupt the function of this selective endonuclease in the repair of ICL without compromising its role in NER, suggesting that different mutations in the same nuclease can lead to malfunction in the different DNA repair mechanisms (Bogliolo et al. 2013).

The most efficient substrate incision structure from XPF-ERCC1 is the “splayed arm” (Figure 11). XPF cuts the DNA duplex at the base of the 3' arm. It also cuts other structures like 3' flap but its efficiency is considerably reduced especially when the leading strand is present in the junction.



**Figure 11: Substrate specificity of XPF/ERCC1.** The preferential structures of the endonuclease are splayed arms, bubbles and stem loops, following by 3'-flap structures. Red arrows indicate the sites of cleavage (modified from (Ciccia et al. 2008)).

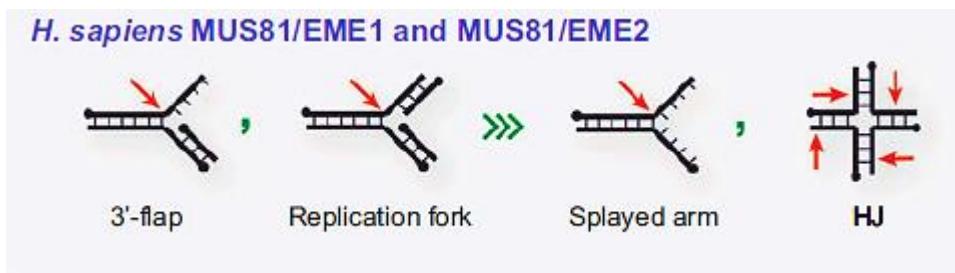
Apparently, incisions on both sides of the crosslink would be necessary for its correct removal, therefore another nuclease needs to cut at the 3' side, unless XPF-ERCC1 is able to cut at both sides. The presence of a nascent leading strand, or a lagging strand or both strands inhibits the action of the XPF-ERCC1 but it is now unrevealed how RPA protein selectively stimulates XPF-ERCC1 to overcome its inhibition in the presence of the nascent leading strand. Notwithstanding, RPA does not stimulate XPF-ERCC1 cleavage on fork substrates containing a nascent lagging strand; on the contrary, RPA occupation of the 3'-flap region of the fork while there is a nascent lagging strand at the 5'-flap region prevents XPF-ERCC1 to recognize the fork junction (Abdullah et al. 2017).

XPF-ERCC1 is a member of the XPF/MUS81 protein family and regarding their structure and function similarities there is probably a competition between XPF and MUS81 endonucleases to bind to SLX4 (I. M. Munoz et al. 2009). It has been argued about the importance of the individual contributions of the endonucleases and although this issue is still not completely solved, XPF-ERCC1 seems to be the main endonuclease for the unhooking (Klein Douwel et al. 2017). It has recently been discovered that the ubiquitin-like protein UHFR1 is involved in the recruitment of MUS81-EME1 and XPF-ERCC1 for the unhooking although the complete picture is still unclear (S. Hashimoto et al. 2016).

### I.3.2.3.3 MUS81-EME1

MUS81 is the catalytic subunit of the MUS81-EME1 heterodimer that belongs to the XPF/MUS81 nuclease family. It has a preference for the 3' flap structures that contain a 5'

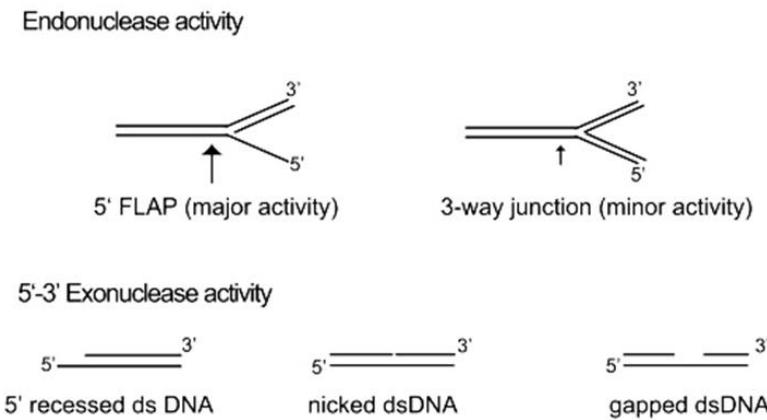
end within 4 nucleotides of the flap junction; if the 5' end is located more than 5 nucleotides away from the ICL, MUS81 cannot cleave this DNA structure (Figure 12) (Ciccia et al. 2008). Mutations in MUS81-EME1 were less sensitive to ICLs than those in XPF-ERCC1 heterodimer and this sets aside the action of MUS81 to a particular subset of ICL DNA intermediates and demonstrates that this heterodimer endonuclease plays a secondary role in crosslinks repair (J. Zhang and Walter 2014).



**Figure 12: Substrate specificity of MUS81/EME1.** Its preferential substrates are 3'-flap structures and replication forks followed by far by splayed arms and HJ structures. Red arrows indicate the sites of cleavage (modified from (Ciccia et al. 2008)).

#### I.3.2.3.4 FAN1

Fanconi Associated Nuclease 1 (FAN1) is another nuclease of the family XPF/MUS81 that is recruited to ICLs by its ubiquitin-binding (UBZ) domain to the monoubiquitinated FANCD2. Despite mutations in FAN1 do not cause severe ICL sensitivity, they still produce chromosome abnormalities due to the block of replication forks even without the presence of crosslinks (Lachaud et al. 2016; Segui et al. 2015; Trujillo et al. 2012). FAN1 preferred substrates are 5' flap DNA (Figure 13), like SLX1 does, but it cleaves 2-4 nucleotides 3' to the branch point instead of cutting at the ssDNA-dsDNA junction and it complements the cleavage activity of MUS81 by subsequently incising the lagging strand, for which MUS81 has no affinity (O'Donnell and Durocher 2010).



**Figure 13: FAN1 nuclease activities and possible specific substrates.** Its endonuclease activity has mainly affinity for 5'-flap structures and a weaker affinity for 3-way junction structures. The arrows indicate the double-stranded site of cleavage. FAN1 5'-3' exonuclease activity is performed in 5' recessed dsDNA, nicked dsDNA and gapped dsDNA structures (adapted from (O'Donnell and Durocher 2010)).

Studies performed by Knipscheer group in *X. laevis* eggs in order to discriminate which endonucleases are responsible for the unhooking of ICL showed that immunodepletion of MUS81 or FAN1 did not affect the repair pathway while removing of XPF-ERCC1 impaired the repairing efficiency, therefore pushing XPF-ERCC1 heterodimer to be the most crucial endonuclease recruited to SLX4 for the crosslinks unhooking (Klein Douwel et al. 2014).

### I.3.2.3.5 SNM1 NUCLEASES: SNM1A AND SNM1B

SNM1 nucleases are processing enzymes involved in ICLs repair. SNM1A works as an exonuclease and it loads in a nascent leading strand stalled fork substrate to digest in 5'→3' direction past the ICL, continues several nucleotides beyond the junction and completes the unhooking. SNM1A exonucleolytic action might replace the 3' incisions when the 3' side of the ICL is not ideal for cleavage by other nucleases like XPF or MUS81. Therefore, when a 5' incision takes place, SNM1A can complete the unhooking without the need for the 3' endonucleolytic cleavage. Furthermore, SNM1A could also eliminate some of the nucleotides between the incision points to a residual mono-adduct that can be detected as a substrate for the downstream TLS bypass of ICL (J. Zhang and Walter 2014).

On the other side, SNM1B is a 5' to 3' dsDNA nuclease that participates in the nucleolytic processing of ICLR intermediates; it does not seem to be important for the unhooking step but it is sought to be required for the proper localization of other repair factors such as FANCD2 (SNM1B is crucial for efficient assembly of FANCD2 into subnuclear foci), BRCA1 and RAD51. Its exonuclease activity generates single-strand regions at collapsed replication forks after XPF-ERCC1 unhooks the ICL lesion (Mason and Sekiguchi 2011).

New approaches will be necessary to completely understand the combined action of XPF-ERCC1, FAN1, MUS81, RPA and SNM1A, but it seems demonstrated that there are different ways to fix ICLs depending on the temporal and substrate context in which they are detected (A. T. Wang et al. 2011).

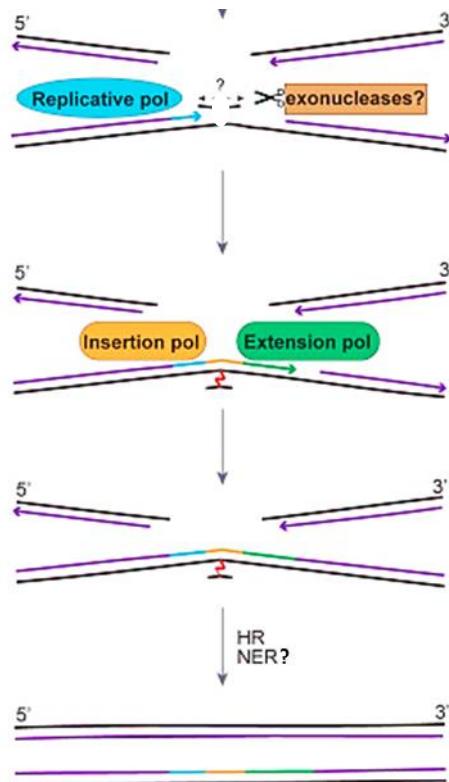
#### I.3.2.4 GROUP III: TRANSLESION SYNTHESIS AND RECOMBINATION PROTEINS

This group consists of a group of proteins whose alteration do not alter FANCD2 monoubiquitination levels in response of DNA damage, as they play their role downstream ID complex and following to the incisions performed by the nucleases.

After the unhooking processes, the 3' end will be extended from the nascent leading strand previously arrested at the -20 to -40 position from the ICL by the TLS DNA polymerases REV1 and Pol- $\zeta$ , therefore the nascent leading strand becomes a template for the repair of the opposite strand (Abdullah et al. 2017). Pol- $\zeta$  consists in polymerases REV3 and REV7 (which has been recently identified as *FANCV* gene of FA pathway) (Bluteau et al. 2017) and the accessory subunits POLD2 and POLD3 (Tomida et al. 2015) that are recruited to the stalled replication forks through their interaction with mono-ubiquitinated PCNA and the extended strand turns into a template for the repair of the opposite strand. REV1 is a cytidine-transferase which incorporates single cytosines opposite to the crosslinks, that usually occurs between guanines (Crossan and Patel 2012).

The duplication of the parental strand that has been incised from the ICL by the endonucleases produces a DSB in the daughter duplex. Meanwhile the opposite parental strand still keeps the crosslink attached and requires the polymerases from TLS to bypass it in its daughter duplex. After this step, the daughter strand gets completely filled by ligation

and a model suggests that unhooked DNA mono-adduct might be removed by NER. The repaired daughter strand becomes then the template for repair of DSB by HR (Figure 14) (Roy and Scharer 2016).



**Figure 14: Representation of lesion bypass in ICL.** After the unhooking the crosslink remains attached to the other parental strand and the duplication of it requires the TLS polymerases to bypass it. Finally the ligases will complete the daughter strand and the crosslink will be removed. This daughter duplex will be the template for the repair of the DSB (adapted from (Roy and Scharer 2016)).

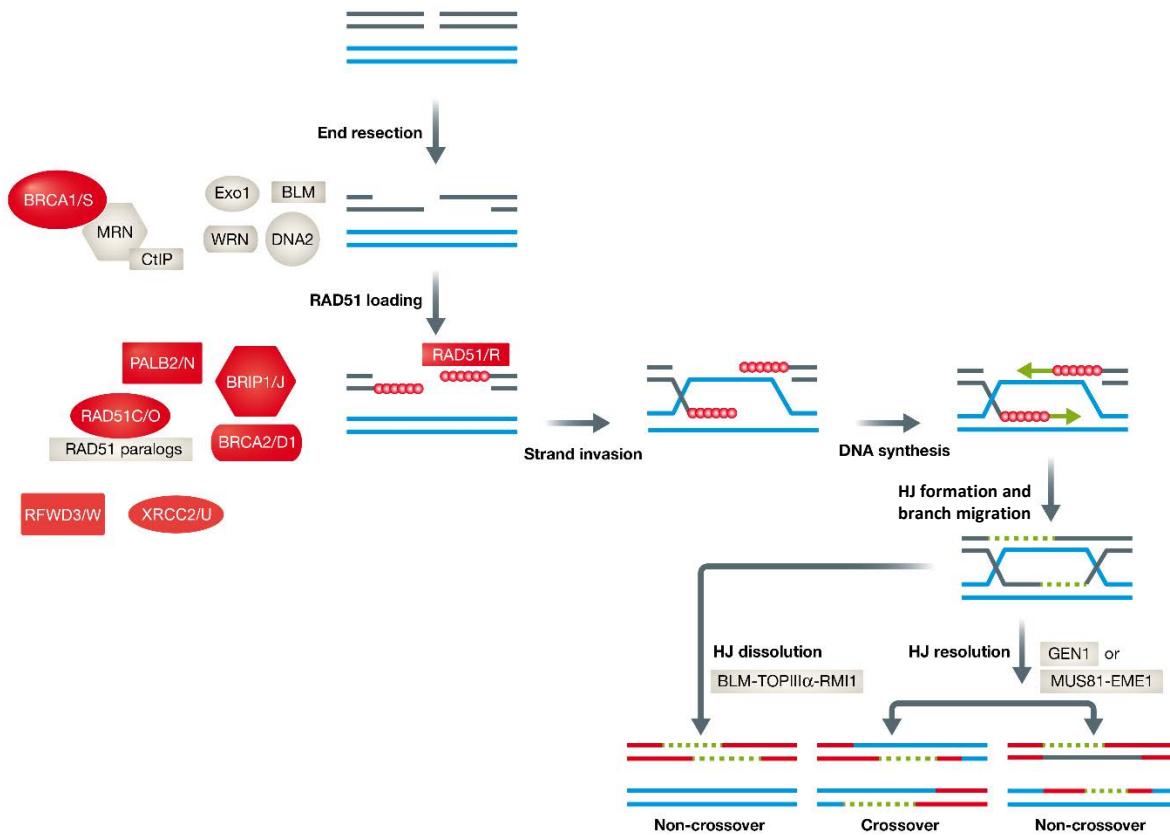
HR and NHEJ are the two basic mechanisms by which a DSB can be fixed. NHEJ links two DNA end ligation without any homology requirement neither end resection and it is active during the whole cell cycle. It is also known as canonical NHEJ (C-NHEJ) and it requires the action of the DNA-dependent protein kinase (DNA-PK) (which comprises the Ku70/80 heterodimer and the catalytic subunit DNA-PKcs), XRCC4 and ligase IV proteins. It can restore DNA sequence but often causes insertions or deletions. In contrast to NHEJ

(Bhargava et al. 2016), the HR requires the presence of a sister chromatid as an homologous intact template to faithfully copy the genetic information, and therefore is confined to S/G2 phases of the cell cycle (Katsuki and Takata 2016).

Certain genes of HR and involved in the FA pathway have been revealed, furthermore, to be involved in FBOC susceptibility. Homozygous mutations of *FANCD1* (*BRCA2*), *FANCN* (*PALB2*), *FANCO* (*RAD51C*), *FANCJ* (*BRIP1*), *FANCS* (*BRCA1*), cause FA, while heterozygous or monoallelic mutations predispose to breast and ovarian cancer. While *FANCS/BRCA1* and *FANCO/RAD51C* are considered FA-like genes involved in FBOC, *FANCR/RAD51* and *FANCU/XRCC2* are identified as FA-like genes without connection to breast and ovarian cancer predisposition (Bogliolo and Surralles 2015). Additionally, *FANCM* gene, which is involved in HR as part of the complex with BLM/RM1/Topo3A (dissolvosome complex) also contributes to FBOC (Kiiski et al. 2014; Peterlongo et al. 2015).

During HR, the DNA duplex generated by TLS is used as a template to repair the DSB originated by the unhooking. *FANCS/BRCA1* forms a complex with Abraxas and RAP80 proteins to detect damage through recognition of ubiquitinated proteins (Sawyer et al. 2015; B. Wang et al. 2007). 5' sites of the DSB must be firstly processed to generate 3' ssDNA substrates for the binding of the repair proteins.

MRN complex reaches the lesion site; it is formed by MRE11, RAD50 and Nijmegen Breakage Syndrome 1 (NBS1). This endonuclease, together with CtIP endonuclease that works associated to *FANCS/BRCA1*, cleaves at 5' to generate a ssDNA overhang. This process is called DNA end resection and allows the following search of homology on the sister chromatid (Figure 15) (Katsuki and Takata 2016).



**Figure 15: The HR pathway of DSB repair (adapted from (Michl et al. 2016)).**

Resection is necessary for HR but not for NHEJ and CtIP acts as a regulator promoting HR instead of NHEJ (Huertas and Jackson 2009). The 3' ssDNA generated during this resection step is then covered and protected by RPA. The last HR protein identified to be associated to FA is FANCW/RFWD3 (Knies et al. 2017). RFWD3 ubiquitinates RPA so that it can leave the ssDNA and go towards the proteasome (Feeney et al. 2017; Inano et al. 2017). The recruiting of RPA to ssDNA attracts ATR and its partner ATR interacting protein (ATRIP) complex to turn on the S-phase checkpoint that leads to cell cycle arrest. The ATR-ATRIP activation is mediated by TopBP1 protein and it phosphorylates FANCJ/BRIP1 helicase for its proper G2 checkpoint function (Andreassen and Ren 2009). Therefore, the formation of ssDNA-RPA requires cyclin-dependent kinase (CDK) protein activity. This CDK

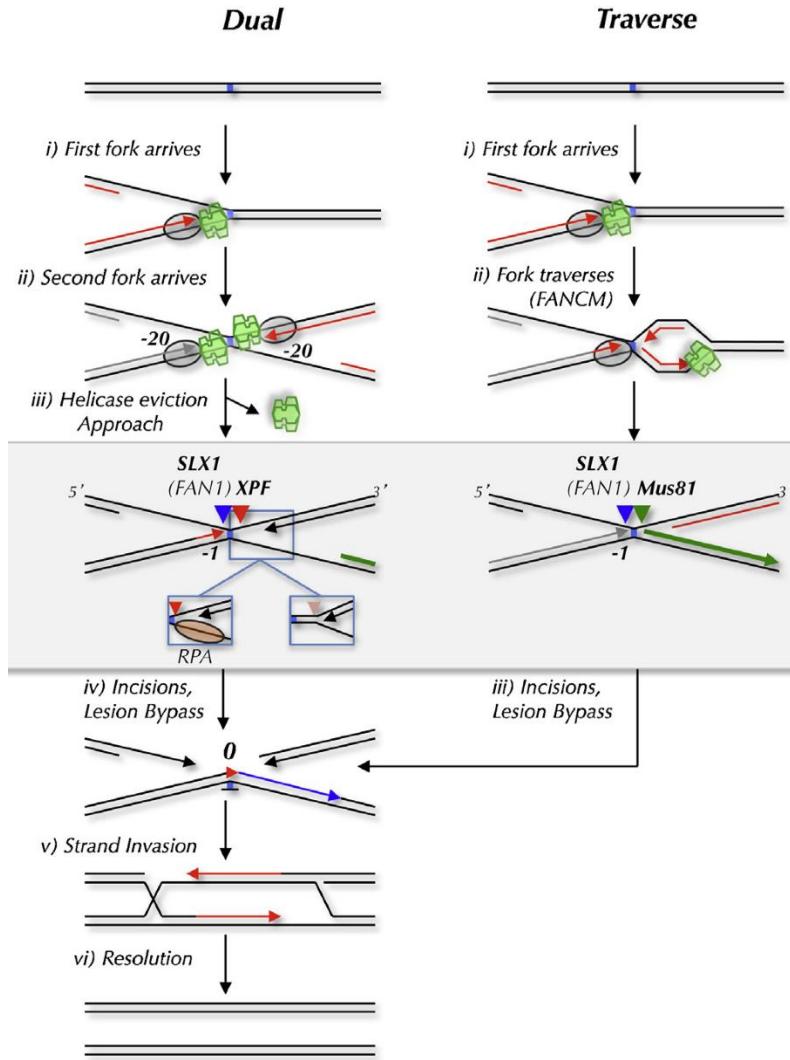
forms a complex with a cyclin that directs its phosphorylative activity and restricts it to S and G2 phases to monitor the timing of the repair mechanism (Zhao et al. 2017).

RPA is then replaced by FANCR/RAD51 thanks to a new complex formed by FANCS/BRCA1, FANCN/PALB2 and FANCD1/BRCA2. FANCD1/BRCA2 was identified as the first core HR gene involved in FA (Howlett et al. 2002) and the loading of FANCR/RAD51 stimulated by this protein is required for strand invasion. FANCN/PALB2 association to FANCS/BRCA1 is enhanced by ATR to promote HR. FANCN/PALB2 stabilizes FANCD1/BRCA2 and links together FANCD1/BRCA2 with FANCS/BRCA1 (Orthwein et al. 2015).

Five FANCR/RAD51 paralogs (RAD51B, FANCO/RAD51C (Somyajit et al. 2012), RAD51D, FANCU/XRCC2 (Park et al. 2016), and XRCC3) organized in two different complexes have been identified as FANCR/RAD51 mediators (Katsuki and Takata 2016); they are positive regulators at different steps of HR through facilitating the invasion of the sister chromatid, although the molecular mechanism of this stimulation is still unknown (Taylor et al. 2015)

The last step of HR involves extension of the filament by polymerase eta (Pol $\eta$ ), resolution of the HJ or synthesis dependent strand annealing (SDSA) mechanism. In this last mechanism a D-loop migrates while DNA is synthesized at 3' of the invading ssDNA tail ensuring that a copy of the homologous DNA template is replaced without chromosome crossover (Miura et al. 2012). HR finishes with the ligation of the DNA strand.

During last years the idea of the two replication forks stalling at the crosslink point as a requirement to initiate ICLR has been put to spotlight and a traverse model has won attention. This other replication coupled model would require the translocase activity of FANCM, and it would recruit RPA to an ICL duplex DNA, which could enhance XPF-ERCC1 to make the 5' incision (Figure 16). In summary, both ICL traverse and fork convergence events would ultimately lead to an X-shaped structure that would be post-replicatively repaired by similar unhooking and TLS mechanisms (Roy and Scharer 2016; J. Zhang and Walter 2014).



**Figure 16: Possible mechanisms of replication-coupled ICLR (adapted from (J. Zhang and Walter 2014)).**

### I.3.3 REPLICATION-INDEPENDENT ICL REPAIR

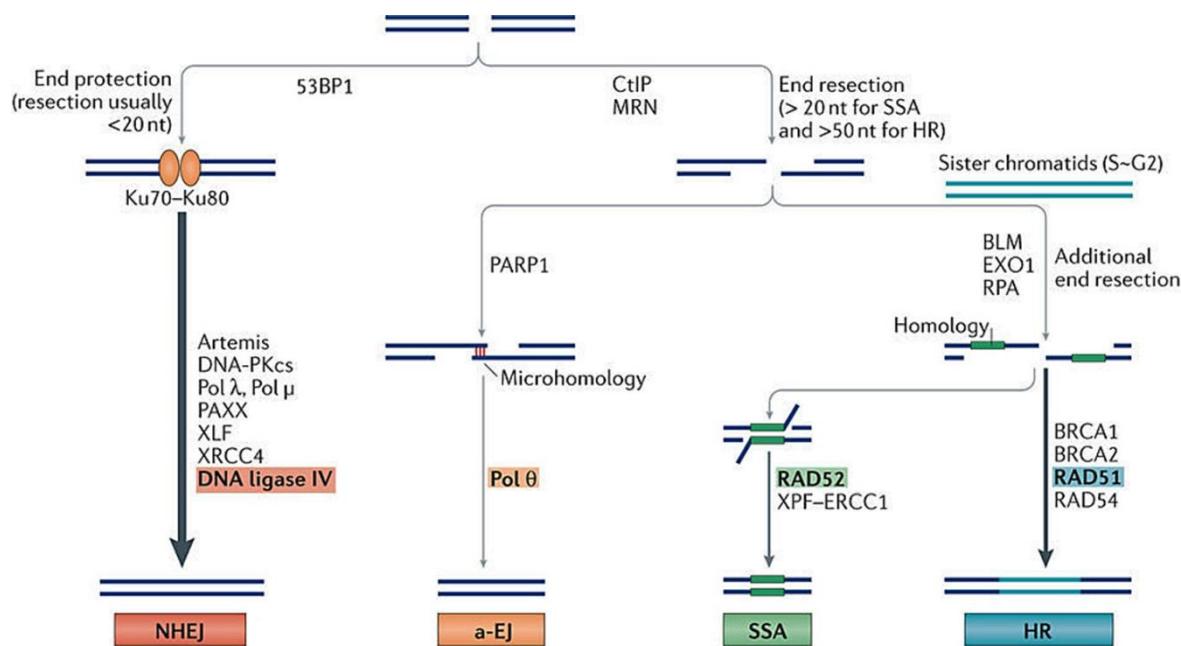
DNA crosslinks are especially damaging during replication in S-phase, however the replication-independent repair (RIR) during G0/G1 plays an important role in post-mitotic cells. Once mitosis is ended, endogenous crosslinks could block transcription of crucial genes. Moreover, ICLR in G1 reduces considerably the damage before the cells enter S-

phase (Roy and Scharer 2016). This replication and recombination-independent ICLR involves NER proteins to recognize and incise the crosslinks, but as cells with mutations in NER factors are only mildly sensitive to crosslinker agents, this pathway could be secondary in the ICLR context, and be activated only when the transcription of an essential gene is compromised (Eniou et al. 2012). Unlike replication-dependent ICL, which starts when a replication fork is stalled and needs the orchestrated action of the FA pathway proteins, RIR does not rely on FA pathway. The starting point could be a collision between a RNA polymerase and the crosslink, from where NER or mismatch repair (MMR) proteins could recognize ICLs depending on the structural nature of the ICL (Williams et al. 2013).

Actually, most of the endonucleases involved in the replication-dependent ICLR unhooking would take part in this pathway (XPF-ERCC1 with RPA, SNM1A and FAN1) and especially SNM1A is thought to be recruited to CSB during TC-NER complexes to process these transcription-dependent ICLR (Iyama et al. 2015). DSBs are not generated in RIR but they are generated in replication-dependent ICL in response to crosslinker agents, and they last much longer in cells deficient for XPF-ERCC1, which would suggest that XPF-ERCC1 is involved in something else apart from the incision, possibly in the resolution of the DSB through recombinational repair (Williams et al. 2013) which could explain the possible contribution of the heterodimer in RIR but the exact knowledge of how the ICLR pathway works independent from replication is still unclear.

#### **I.3.4 OTHER DOUBLE STRAND BREAK SUBPATHWAYS**

Any DSB that takes place in S phase may be resected; in the absence of a sister chromatid it will be processed by MMEJ or by SSA (Figure 17).



**Figure 17: Diagram of the different DSB repair pathways** (adapted from (Chang et al. 2017)).

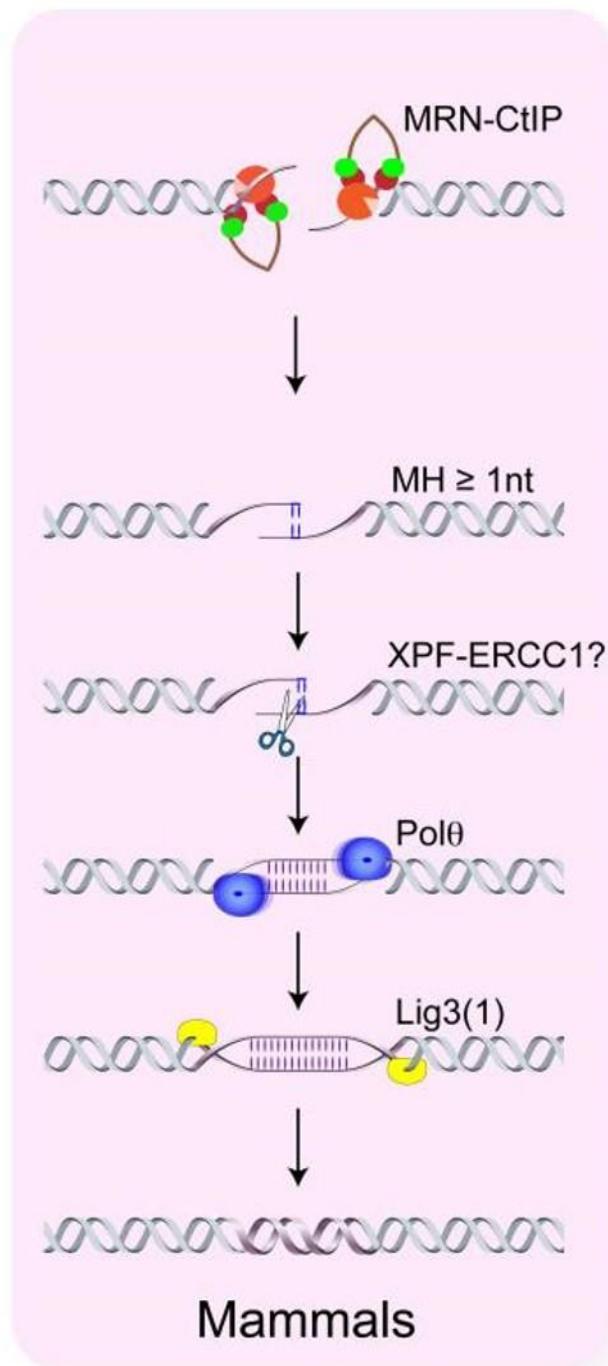
MMEJ, which is an a-EJ pathway that takes place when any of the two canonical DSB pathways cannot work or simply at 10-20% cells levels when the other two pathways are available, probably because MMEJ produces a deletion, and therefore it is a more error-prone and mutagenic pathway. MMEJ consists of the alignment of microhomologous sequences near the DSB before joining and is inherently associated with deletions flanking the breaks (Sfeir and Symington 2015).

MMEJ starts with 5' end resection through MRN complex stimulated by phosphorylated CtIP action. This complex generates 15-100 nucleotides 3' overhangs. In mammalian cells, the extension of short complementary base pairing called microhomologies (MH) is required for the MMEJ and their stretch can be as few as 1 nucleotide. MMEJ operates differently depending on the size of MH and the types of DNA breaks. Poly (ADP-ribose) polymerase 1 (PARP1) is involved in joining the linear DNA fragments through their complementary sequences after the resection. When the ends are incompatible, insertions of nucleotides at break sites copied from other chromosomes or added *de novo* generate MH that are used

to pair the DNA ends and promote diversity and increase mutagenicity (Sfeir and Symington 2015).

ERCC1-XPF is not part of the core NHEJ machinery and there are evidences that HR occurs in the absence of XPF-ERCC1. Rad10-rad1, its yeast ortholog, is required to remove the nonhomologous sequence from the 3' ends to allow the synthesis and/or ligation. In mammals, the mechanism is not totally clear but some studies demonstrate that in the absence of XPF-ERCC1, joining of homologous 3' overhangs is impaired and drives to large deletions. Furthermore, this endonuclease cleaves unprotected telomeric ends, promotes telomeric fusion through end joining and it is important for end joining in c-NHEJ deficient cells (Bhargava et al. 2016). These observations support the hypothesis that XPF-ERCC1 plays a role in MMEJ analogous to yeast Rad10-rad1 (Ahmad et al. 2008; Manandhar et al. 2015). If the homology does not naturally occurs flanking the broken end, XPF/ERCC1 removes the 3' flap and forms DNA ends which are available for the DNA synthesis and ligation, and as a consequence provokes the deletion of inter-MH sequence (Sinha et al. 2016).

The TLS polymerase theta (Pol θ) is shown to be necessary in mammals for the synthesis of nucleotides during joining. It might be involved in the displacing of RPA from the ssDNA and it interacts with RAD51 and downregulates its role as an intermediate of HR, thereby enhancing MMEJ instead (Ceccaldi et al. 2015). Pol θ is able to stabilize annealing of two 3' ssDNA overhangs with as little as 2 bp homology, simply using the annealing partner as a template. The polymerase activity contributes to prevent long resection of ends which would shunt to other DSB repair mechanisms (Chang et al. 2017). The pathway concludes with the sealing of the ends; Ligase III (Lig3) is the main protein involved in the ligation step and Ligase I (Lig1) acts as a back-up enzyme (Sfeir and Symington 2015) (Figure 18).



**Figure 18: Mechanistic model for MMEJ in mammals** (adapted from (Sfeir and Symington 2015)).

MMEJ is an extremely versatile mechanism, when MH are shorter than 5-6 bp, it resembles NHEJ, but when the homologues are larger than 15 bp, it operates more similarly to SSA (Sinha et al. 2016). Although the role of PARP1 is not precisely clarified in MMEJ, its activity, together with Pol θ role, seem to be the main directors of this pathway (Bhargava et al. 2016).

SSA is a DSB pathway that requires longer homologues than MMEJ to initiate the repair of the break. It shares the initiation step with other DSB pathways such as MMEJ and HR because it requires MRN and CtIP to generate the ssDNA tails, but the resection is more extensive and needs additional nucleases like EXO1 or Bloom syndrome RecQ-like helicase (BLM) or DNA replication helicase/nuclease 2 (DNA2). End resection is a crucial step of SSA and factors that inhibit end resection suppress this pathway (Bhargava et al. 2016).

After end resection, RPA quickly covers the single strand to prevent the formation of secondary structures and the unannealed non-homologous portions of the 3' ssDNA are cleaved by XPF-ERCC1 endonuclease and the MMR complex MSH2-MSH3, whose role is not totally understood yet. Finally, the annealing of the homologues is mediated by RAD52 (Chang et al. 2017) a DNA binding protein that specifically works in SSA mediating annealing of the 3' tails and enhancing the endonucleolytic activity of XPF-ERCC1 (McNeil and Melton 2012).

Following to the annealing, some DNA polymerases will fill the gaps and some ligases will produce the junction, but the nature of which specific proteins are involved in this step is still unclear.

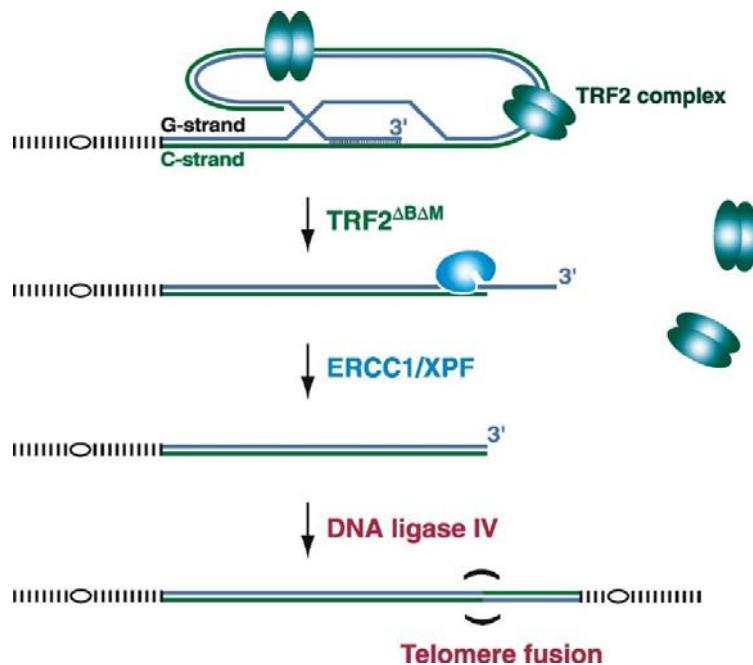
When sequence repeats are close to a DSB, SSA can be activated and produce a deletion of sequences between the repeats, which constitutes an important source of mutagenesis. Although SSA is an error-prone mechanism, it is needed for the survival of cells with DNA damage that could not be repaired in any other way (Stark et al. 2004). Consequently, SSA and MMEJ are deletion-inducers error-prone mechanisms unlike HR, which requires a sister chromatid as a template to generate a loyal copy of the strand. However, some deletions can occur while HR repair pathway when DNA tandem repeats provoke the resolution by crossing over. These deletions would be similar to the ones

produced by SSA but the key distinction element in this case would be RAD51, which is essential in HR for the strand-invasion but does not participate in SSA.

SSA could also participate in gene targeting, a process also called homology-directed gene editing that is defined by a site-specific modification of genomic DNA using an exogenous donor template that is homologous to the target site. This uncommon event might be promoted by a DSB at a target site (Bhargava et al. 2016).

### I.3.5 TELOMERE STABILITY

Telomeres are protein-DNA complexes at the ends of linear chromosomes that are important for genome stability. They are composed by TTAGGG DNA repeats that end in a single-stranded 3'-overhang of the G-strand which are protected by TRF2, a dimeric protein that binds along the duplex telomeric repeat tract. Inhibition of TRF2 activates a DNA damage response that leads to induction of apoptosis or senescence. These unprotected telomeres are processed by NHEJ, which produces mutations through telomeres fusion and multicentric chromosomes formation. TRF2 can promote telomeric loop (T-loop) formation. T loops are large duplex loops formed through the strand invasion of the single-stranded telomeric overhang. If the 3' overhang is confined into the duplex part of the telomere it remains protected from degradation and prevents fusion by NHEJ. These overhangs disappear gradually as a consequence of DNA replication, but even when cells do not progress to S phase overhang loss is observed in TRF2 inhibited cells, which suggests the activity of a nuclease cleaving these overhangs. XPF-ERCC1 is the endonuclease involved in 3' overhang telomeric cleavage (Zhu et al. 2003) (Figure 19).



**Figure 19: Model for telomere processing** (adapted from (Zhu et al. 2003)).

XPF shows interaction with TRF2 via SLX4 and cleaves 3' overhangs from uncapped telomeres, which results in telomeres shortening and ageing (Manandhar et al. 2015). SLX4 associates with telomeres in late S phase or under genotoxic stress, interacts to TRF2 by its TMB domain. The main protective role of TRF2 is to prevent overhang loss, but the mechanism by which it blocks XPF-ERCC1 from removing the 3' overhang is not yet unrevealed.

# **Objectives**



## II. OBJECTIVES

Concerning the wide range of diseases associated with changes in XPF protein and the amount of DNA repair pathways in which it plays a crucial role, the objective of this thesis was to better understand the role of *ERCC4/XPF* in DNA repair and human diseases.

The specific objectives of this work were:

- To analyze the genotype-phenotype correlation of *ERCC4* pathogenic variants.
- To uncover new XPF partners and study their role in DNA repair.



## **Materials and Methods**



## III. MATERIALS AND METHODS

### **III.1- CELL LINES AND CULTURE**

Human cell lines HEK 293T (Human Embryonic Kidney 293T cells, ATCC CRL-11268) and human osteosarcoma U2OS were used in this study.

HEK 293T *ERCC4* knocked out (HEK XPF-KO) created by TALENs (Transcription Activator-Like Effector Nuclease) technology were generated and used as the knocked *ERCC4* background in which all the mutant versions of XPF were generated.

HEK 293T *USP11* knocked out (HEK USP11-KO) and double knocked out HEK *ERCC4-USP11* (HEK XPF/USP11) were created by Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) technology.

Human osteosarcoma cell line U2OS DR-GFP was kindly provided by Dr Maria Jasin (New York). U2OS SA-GFP was previously developed by members of our laboratory using SA-GFP plasmid (Addgene Nº 41594).

All the cell lines were cultured in DMEM (Dulbecco's Modified Eagle Medium, Biowest) complemented with 10% FBS (Fetal Bovine Serum, Biowest) and 0.1 µg/mL Plasmocin (IBIAN Technologies). Cells were cultured at a controlled temperature of 37° C in 5% CO<sub>2</sub> atmosphere. All the cell lines were kept in liquid N<sub>2</sub> in a Biobank from our group.

### **III.2- ESTABLISHING OF THE HEK 293T TALEN ERCC4-KNOCKED OUT AS THE XPF-KO BACKGROUND CELL LINE**

#### **III.2.1 GENERATION OF THE HEK XPF-KNOCKED OUT CELL LINE**

HEK 293T *ERCC4*-KO was generated by TALEN. *ERCC4* exon 2 (Figure 20) was targeted by a pair of designed plasmids fused to a FokI nuclease domain that excised the sequence producing an early truncated protein completely unfunctional.

5' TCGCCGTGTAACAAATGAAATCACAAGCAACAGTCGCTATGAAGTTACACA 3'

##### **TAL Nuclease recognition sequences**

Blue= TAL Binding sites

Red= Cut region

**Figure 20: *ERCC4* TALEN recognition sequence.** The fragment corresponds to exon 2. Blue region represents TALEN binding sites and red region represents FokI endonuclease cutting sites.

Additionally, another plasmid was used: a reporter-based plasmid containing the same *ERCC4* recognition sequence, a red fluorescent protein sequence and an out of frame green fluorescent protein (GFP) that could be restored when the DSB produced by FokI was repaired by NHEJ. After 48 h post transfection the fluorescence was visible.

Double fluorescent (red and green) cells were selected by flow cytometry using the FACSAria II (BD Bioscience). Single cell cloning of these cells was performed by limit dilution (3 cells/mL), plating 3 plates of 96 wells with 100 µL of complete medium to ensure just one clone was plated per well. After 2-3 weeks three individual clones grown and were picked to check the lack of XPF protein by Western Blot (WB).

### **III.2.2 WESTERN BLOTTING VALIDATION OF THE KNOCKED OUT CLONES**

50 µL of RIPA 1x lysis buffer (Millipore 20-188) with Benzonase nuclease (10 U/mL final) (VWR International 7074-6-3) was added for each million of cells. Samples were incubated at 37°C during 10 min and then centrifuged at 13,000 rpm. The total protein concentration of the supernatant was determined by Bio-Rad Protein Assay (Biorad) according to manufacturer's instructions and 50 µg of total proteins with Laemmli 1x (Sigma S3401-10VL) were denaturalized for 10 min at 96°C and loaded in an SDS-PAGE 8% acrylamide. Proteins were transferred from the gel to the nitrocellulose membrane in the iBLOT2 (Invitrogen, ThermoFisher Scientific), blocking of the membrane was achieved by incubation with blocking solution (5% of milk in TBS+ Tween) during 1h at RT, immunodetection was performed by incubating the membranes with diluted primary antibodies in blocking solution during 16h at 4°C. Primary antibodies used were: anti-XPF (mouse monoclonal, Ab-1 219 Thermo Scientific) 1:200 and anti-Actin (rabbit polyclonal, ab1801, Abcam) 1:1,000. The following day, membranes were incubated in secondary antibodies conjugated to peroxidase during 1h at RT and revealed with Pierce ECL Western Blotting Substrate (Pierce) and digital images of the membranes were captured with the GeneGnome apparatus (Syngene Bio imaging).

### **III.2.3 CHARACTERIZATION AND SELECTION OF THE XPF-KNOCKED OUT CLONE**

Genomic DNA was extracted from  $5 \times 10^6$  clonal HEK XPF-KO cells using the DNeasy Blood & Tissue Kit (QIAGEN) according to manufacturer's instructions and DNA concentration was measured by spectrophotometry using NanoDrop ND-1000 (NanoDrop Technologies).

PCR to identify mutations inserted by TALEN was performed using the following primers (Table I):

<b>ERCC4-Ex2-F</b>	TGTAGACTGGTTGGCTGAAGT
<b>ERCC4-Ex2-R</b>	CGCCTATGTGCTTCCCAAGA

**Table I:** Primers sequences used to identify the mutations inserted by TALEN.

PCR conditions are schematized in (Table II):

Segment	Cycles	Temperature	Time
1	1	94° C	1 min
2	35	94° C	30 sec
		58° C	30 sec
		72° C	4 min
3	1	72° C	4 min

**Table II:** PCR conditions to amplify XPF modified region by TALEN.

The product size was checked by loading the sample into a 1.5% agarose gel, the DNA band was purified and subcloned by TOPO TA cloning (Thermo Fisher) according to manufacturer's, transformed in One Shot TOP10 Chemically Competent *E.coli* cells (Life Technologies) following manufacturer's instructions, amplified by performing Minipreps with NucleoSpin Plasmid (Macherey-Nagel) and sent to Sanger sequencing of Macrogen with the ERCC4\_Ex2-F primer to check the sequence mutations in the individual alleles. Sequences results were analyzed by Sequencher (Gene Codes Corporation) bioinformatics software.

### **III.3- GENERATION OF XPF CELL LINES**

#### **III.3.1 GENERATION OF THE XPF-Wt LENTIVIRAL VECTOR**

The retroviral pBABE vector (pBABE puro IRES-EGFP, Addgene 14430) with the cDNA of *ERCC4* insert was already built in our lab. *ERCC4* cDNA was modified by cloning the HA-tag epitope in C-terminal, and with two N-terminal epitopes: streptavidin binding protein (SBP) and calmodulin binding protein (CBP) (TAP tag). (From now onwards referred as pBABE XPF). Modified *ERCC4* cDNA was subcloned into a 3<sup>rd</sup> generation lentiviral vector pUltra (Addgene, 24129) downstream of the EGFP-P2A site, to produce a bi-cistronic expression of EGFP and ERCC4. P2A site, similar to IRES, is a peptide with a motif associated with cleavage-like activity through a ribosomal skip mechanism that impairs normal peptide bond formation between ERCC4 and EGFP without affecting the translation of EGFP, ensuring both proteins can be translated from a single sequence to two individual proteins. To do that, two primers were designed in order to amplify and modify the insert TAPtag-XPF-HA by including a restriction site compatible with the restriction enzymes used to cut the pUltra vector (Table III).

<b>TAPTAG-XPF-HA_S</b>	CCCTCTAGAATGAAGCGACGATGGAAAAAG
<b>TAPTAG-XPF-HA_AS</b>	ATGTGATCATCAAGCGTAATCTGGAACATCG

**Table III:** Primers sequences used to amplify and modify the insert from pBABE plasmid making compatible with the pULTRA lentiviral plasmid.

The PCR was carried out with the following conditions (Table IV):

Segment	Cycles	Temperature	Time
1	1	94° C	1 min
2	35	94° C	30 sec
		57° C	30 sec
		72° C	4 min
3	1	72° C	2 min

**Table IV: PCR conditions to amplify the insert from pBABE plasmid.**

The PCR product was then digested first with XbaI (Roche) restriction enzyme during 2h at 37°C and second with BclI (Roche) restriction enzyme during 2h at 55°C.

The pUltra vector was digested first with XbaI (Roche) as the insert and secondly with BamHI (Roche) restriction enzyme during 2h at 37°C. The vector was loaded in a 0.7% agarose gel, purified, dephosphorylated with FastAP (Thermo Fisher Scientific) and ligated to the digested PCR product of the insert TAPtag-XPF-HA with the Quick Ligation Kit (New England Biolabs) overnight on ice. The ligation product was then transformed using One Shot Stbl3 Chemically competent *E.coli* (Thermo Fisher Scientific), amplified by standard Miniprep and Maxiprep procedures and sent to Sanger sequencing of Macrogen for a primer walking (Table V).

<b>FOR1-XPF-pULTRA</b>	TGGAAAAAGAATTTCATAGCCG
<b>FOR2-XPF-pULTRA</b>	TTCTCCAGCTGCAGTGCC
<b>FOR3-XPF-pULTRA</b>	TGAGAAATCTTTGTGAGGAAA
<b>FOR4-XPF-pULTRA</b>	AAGCTTTGGTCAGAATTCAAGT
<b>FOR5-XPF-pULTRA</b>	TGGATGAAATTAGGAAGGAAGA
<b>FOR6-XPF-pULTRA</b>	GAGCTAACCTTGTCGGCA
<b>FOR7-XPF-pULTRA</b>	TCACTCCAGAAATGTGCGTG
<b>FOR8-XPF-pULTRA</b>	CTTGTAAAAATGCCAGGGG
<b>REV1-XPF-pULTRA</b>	TACCGCTACCGCTTCCAAGT

**Table V:** Primers sequences used to perform a primer walking of the pULTRA plasmid ligated to the PCR amplified insert.

### III.3.2 GENERATION OF THE XPF-LENTIVIRAL PARTICLES AND GENETIC COMPLEMENTATION OF HEK XPF-KO CELLS

Production of Lentiviral particle was achieved with the CalPhos Tranfection kit (Clonetech).  $5 \times 10^6$  packaging cells HEK293T (passages between 3 and 8) were seeded in 10 cm plates with complete medium. Three plates were needed for the production of each type of lentiviral particles. 24h hours later the medium was changed for new one and 3  $\mu\text{L}$  of chloroquine 100 mM were added per plate and the transfection solution (Table VI) was prepared per each plate.

Volume ( $\mu\text{L}$ )	Component
<b>10</b>	pUltra- TAP-XPF-HA (1 $\mu\text{g}/\mu\text{L}$ )
<b>6.5</b>	Packaging plasmid (PAX) (1 $\mu\text{g}/\mu\text{L}$ )
<b>3.5</b>	Lenti VSVG plasmid (ENV) (1 $\mu\text{g}/\mu\text{L}$ )
<b>87</b>	Calcium Solution ( $\text{CaCl}_2$ ) (Clontech)
<b>593</b>	Sterile MQ water

**Table VI:** Components of the transfection solution used to produce the lentiviral particles.

One volume of Solution B (2x, HBS Calphos, Clontech) was then added while gently vortexing the transfection solution. After 15 min at RT, this solution was added dropwise to the three dishes which remain at 37°C and 5% of CO<sub>2</sub> overnight. The third day medium was replaced with fresh complete medium. The day after the medium containing the lentiviral particles was collected and stored at 4°C and new medium was added to the plates. The fifth day the supernatant was collected again and filtered through a 0.45  $\mu\text{m}$  filter together with the supernatant volume collected the previous day. The supernatants were then centrifuged with Beckman JS-24.38 rotor at 19,500 rpm during 1.5h at 4°C or using Amicon-Ultra15 centrifugal filter units (Millipore) during 1h at 3,220 RCF. After the centrifugation the medium was carefully discarded and the lentiviral pellet was resuspended with PBS 1x up to 150  $\mu\text{L}$ , by warming them at 32°C during 2h shaking at 1,000 rpm.

180,000 HEK XPF-KO cells were seeded in a 12-wells plate, 24h later cells were infected with 40  $\mu\text{L}$  of pULTRA-TAP-XPF-HA lentiviral particles and 1.5  $\mu\text{L}$  Polybrene (Sigma-Aldrich). Three days after the infection, green fluorescence was detectable and cells that had integrated the virus with the XPF cDNA were selected by flow cytometry using the FACSaria II (BD Bioscience). Single cell cloning of these cells was performed as previously detailed and a single clone confirmed by WB was determined as HEK XPF-KO + XPF-Wt cell line.

### III.3.3 GENERATION OF THE XPF MUTANT VARIANTS

The following table summarizes the XPF-single nucleotide variants generated (Table VII):

Aminoacidic change	Nucleotidic change
p.R153P	c.458G>C
p.I225M	c.675A>G
p.L230P	c.689T>C
p.C236R	c.706T>C
p.R589W	c.1765C>T
p.R689S	c.2065C>A
p.R799W	c.2395C>T

**Table VII: Aminoacidic changes and single nucleotide changes of the XPF variants generated.**

The generation of the XPF mutant variants was achieved by two different methodologies: by subcloning of the XPF inserts from the retroviral vector pBABE to the lentiviral vector pUltra or by site directed mutagenesis from the pUltra +TAP-XPF-HA.

#### III.3.3.1 GENERATION OF XPF VARIANTS BY SUBCLONING

Two XPF variants, C236R and R589W, were moved from pBABE to pUltra following the same protocol described above for the generation of the wild type TAP-XPF-HA .The pUltra C236R and pUltra R589W vectors were sent to Sanger sequencing (Macrogen) to check the introduced variants. The primers used are detailed in Table IV.

### III.3.3.2 GENERATION OF XPF VARIANTS BY SITE DIRECTED MUTAGENESIS

The XPF variants: R153P, I225M, L230P, R689S and R799W were generated by site directed mutagenesis from the lentiviral vector carrying the wild type version of XPF (pUltra + TAP-XPF-HA) by designing with the software from Agilent Technologies the mutagenic primers that contained the desired single nucleotide variant (Table VIII).

Mutation	Primers
p.Arg153Pro	CATCTTGCAGCCTCTTCCCCAGAAAAACAAACGTG CACGTTGTTTCTGGGGAAAGAGGCGCAAGATG
p.Ile225Met	CCTACCATGCTTGCTATGCAGACTGCTATACTGG CCAGTATAGCAGTCTGCATAGCAAGCATGGTAGG
p.Leu230Pro	TAGACATGCATTAAAATGTCCGGTATAGCAGTCTGTATAGCAAG CTTGCTATAACAGACTGCTATACCGGACATTTAAATGCATGTCTA
p.Arg689Ser	GAAGCTCACTTCGAAATTCACTCATATCCACAACTATGCTTG CAAAGCATAGTTGTGGATATGAGTGAATTCTGAAGTGAGCTTC
p.Arg799Trp	CTTCACCTCCCCAGACTATGGATTCTCTGGTGC GCACCCAGAGAATCCATAGTCTGGGGAAAGTGAAG

**Table VIII:** Primers sequences used to introduce the exact single nucleotide variant of *XPF* by site directed mutagenesis.

The QuickChange II XL Site-Directed Mutagenesis Kit (Agilent Technologies) was used to introduce the variants in the wt cDNA of *XPF* by a PCR amplification (Table IX).

<b>5 µL of 10x reaction buffer</b>
<b>X µL (10 ng) of dsDNA template</b>
<b>X µL (125 ng) of oligonucleotide primer #1</b>
<b>X µL (125 ng) of oligonucleotide primer #2</b>
<b>1 µL of dNTP mix</b>
<b>3 µL of QuickSolution</b>
<b>ddH<sub>2</sub>O to a final volume of 50 µL</b>

**Table IX:** Parameters used for the amplification and introduction of the nucleotide variants.

And ideal cycling parameters appear in table X:

Segment	Cycles	Temperature	Time
1	1	95° C	1 min
2	18	95° C	50 sec
		60° C	50 sec
		68° C	12 min
3	1	68° C	7 min

**Table X:** Cycling parameters used for the PCR of the site directed mutagenesis.

After amplification DpnI restriction enzyme was added during 1h at 37°C to digest the dsDNA template, as this enzyme is able to cleave at methylated sites, therefore cutting the template plasmid, methylated by the bacteria, though leaving the PCR product intact.

The PCR product was transformed by XL10-Gold Ultracompetent Cells (Agilent Technologies), plated in LB Agar with Ampicillin (100 µg/mL), DNA from single colony minipreps was controlled by Sanger sequencing (Macrogen) with primers from Table XI to check if the variants were correctly introduced.

Variant	Primer name	Sequence
R153P	c.458G>C(p.R153P)	GGAGTTGAACACCTCCCTCG CTGCTACATGGAACCTTGGC
I225M	c.675A>G(p.I225M)	GGCCAAGGTTCCATGTAGCA CCGTTGCTCTCAGAGATTCCA
L230P	c.689T>C(p.L230P)	CTGTGGCCAAGGTTCCATGT TCCGTTGCTCTCAGAGATTCC
C236R	c.689T>C(p.L230P)	CTGTGGCCAAGGTTCCATGT TCCGTTGCTCTCAGAGATTCC
R589W	c.1765C>T (p.R589W)	ATCCATCCGCTTCTGGGTTG TGCAGATGCTGTGCCCTTA
R689S	c.2065C>A(p.R689S)	GCAAGCATGGTTGTCCCTGA AGGCTTGCTAGGGTCAAACTC
R799W	R799W-S786F	AAAACTCATAAGGGAAAAAGCAAGC CTCTGACTCGGGAAAGGGTTT

**Table XI:** Primers sequences used to check the single nucleotide variants introduced by site directed mutagenesis.

The CalPhos-Clontech transfection Kit (Clontech) was used to generate the lentiviral particles with all the XPF variants following the same steps as for the XPF-wt and HEK XPF-KO cells were transduced with each individual variant as described in section III.3.2.

### **III.4- GENERATION OF USP11 KNOCKED OUT AND ERCC4/USP11 DOUBLE KNOCKED OUT BY CRISPR/Cas9 TECHNOLOGY**

#### **III.4.1 TARGET SEQUENCE SELECTION**

The selected plasmid to generate the USP11 knocked out was PX458 (pSpCas9(BB)-2A-GFP (PX458), Addgene 48138) (Ran et al. 2013). This is a mammalian expression vector

containing an U6 promoter, a sgRNA scaffold site next to the oligo cloning site, the coding sequence of the humanized *S. pyogenes* Cas9 endonuclease and a fused 2A-GFP to allow screening of the transfected cells.

This plasmid needed to be ligated to a specific sequence of 20 nucleotides to guide the Cas9 endonuclease to the complementary sequence of USP11 genomic DNA. The selected sgRNA sequence was obtained from bioinformatics analyses carried out by Cong (Cong et al. 2013) that were optimized to minimize the off-target modifications. The selected sgRNA sequence for USP11 was 5'GCAGTGGGAGGCATACGTGC3' (Sigma-Aldrich) and it targets the exon 2 of *USP11*. sgRNA sequences showed in the Cong table have a PAM (Protospacer Adjacent Motif) motif (a 5'-NGG -3'motif) that was deleted from the selected sequence to design the pair of oligos. This PAM motif is present in the target gene next to the DNA sequence corresponding to the sgRNA sequence and allows Cas9 to bind and cut properly. Its removal from the designed oligos avoids the nuclease to cut the sgRNA sequence of the plasmid.

The two sgRNA (sense and antisense) needed to be 5' phosphorylated to insert in the PX458 linearized with *Bbs*I (New England Biolabs) and dephosphorylated (see below). The final sequences (5' to 3') are in Table XII.

<b>CRISPR USP11_S</b>	[Phos] <b>CACCGCAGTGGGAGGCATACGTGC</b>
<b>CRISPR USP11_AS</b>	[Phos] <b>AAACGCACGTATGCCTCCCACTGC</b>

**Table XII: sgRNA sequences for *USP11*.**

### III.4.2 PRIMERS ANNEALING AND VECTOR CONSTRUCTION

10 µL of both selected primers mixed at a final concentration of 100 µM were annealed by three sequential incubations of 30 min at 37°C, 5 min at 95°C and 10 min at RT. 10 µg of PX458 plasmid were digested with 10 units of *Bbs*I restriction enzyme (New England Biolabs) during 3.5 h at 37°C in buffer 2.1 1x (New England Biolabs), followed by its loading in a 0.8% agarose gel run at 100 V.

The corresponding band was cut and purified using NucleoSpin Gel and PCR Clean-up kit (Macherey-Nagel) and dephosphorylated by FastAP Thermosensitive Alkaline Phosphatase (1U/ $\mu$ L) (ThermoFisher Scientific).

50 ng of the digested PX458 were ligated with 1  $\mu$ L of diluted 1:200 annealed sgRNA by 1  $\mu$ L of Quick Ligase (New England Biolabs) and Quick Ligation Reaction Buffer 1x (Quick Ligation Kit, New England Biolabs) on ice overnight.

From this ligation product, 5  $\mu$ L was used to transform One Shot TOP10 Chemically Competent *E.coli* cells (Life Technologies) following manufacturer's instructions and plating them overnight in LB agar with Ampicillin.

Minipreps from these transformed colonies were carried out using NucleoSpin Plasmid purification kit (Macherey-Nagel) according to manufacturer's instructions, the final product was quantified by NanoDrop ND- 1,000 (NanoDrop Technologies) and sent to Sanger sequencing (Macrogen) using the primer 5' – TTTCTTGGGTAGTTGCAG – 3'.

After confirming the sequences, Maxiprep was carried out using NucleoBond PC 500 Kit (Macheneray- Nagel) according to manufacturer's instructions, the DNA concentration was quantified by NanoDrop ND- 1,000 and sent to Sanger sequencing service of Macrogen using the same primer as the one used for the Minipreps sequencing.

### **III.4.3 CELL TRANSFECTION AND CLONE SELECTION**

500,000 HEK 293T clonal cells and 500,000 HEK XPF-KO cells were seeded in a well of a 6-wells plate with 2 mL of complete medium. Six hours later cells transfections were carried out using 7.5  $\mu$ L of Lipofectamine 2,000 (Life Technologies) in 250  $\mu$ L of Opti-MEM (Gibco, Life Technologies) and mixing these with 2.5  $\mu$ g of the CRISPR vector in 250  $\mu$ L of Optimem. Transfection medium (2.5 mL) was changed for complete medium after incubation overnight. 48 h after transfections cells were detached and analyzed by flow cytometry using the FACS Aria II (BD Bioscience) to select those that have included the plasmid based on the fluorescence emission conferred by the GFP protein encoded by the PX458 vector. Single sorted cells were seeded in three 96-wells plates with 200  $\mu$ L of complete medium.

After 2-3 weeks individual grown clones were checked by WB to confirm the absence of USP11 protein in HEK and in HEK XPF-KO cell lines

### III.4.4 GENOTYPING

Genomic DNA was extracted from  $5 \times 10^6$  cells of each cell line following the lack of USP11 confirmation by WB using the DNeasy Blood & Tissue Kit (QIAGEN) according to manufacturer's instructions and DNA concentration was measured by spectrophotometry using NanoDrop ND-1,000 (NanoDrop Technologies).

PCR to identify mutations in the target region of CRISPR clones was performed. Primers to amplify a *USP11* region in the genomic DNA were designed (Table XIII) and ordered to Sigma-Aldrich.

<b>USP11_KO_Clones_S</b>	TTGGGCATGGGAAGTTGTAT
<b>USP11_KO_Clones_AS</b>	GCACACAGAATGGGCTTATG

**Table XIII:** Primers sequences used for the PCR to amplify a *USP11* region in the genomic DNA.

Ideal parameters for the reaction are summarized in table XIV.

<b>1x Reaction Buffer (Bioron GmbH)</b>
<b>50 ng of genomic DNA</b>
<b>10 µM of primer USP11_KO_Clones_S</b>
<b>10 µM of primer USP11_KO_Clones_AS</b>
<b>10 mM of dNTP mix</b>
<b>25 mM of MgCl<sub>2</sub></b>
<b>0,5 Units of Taq DNA Polymerase (Bioron GmbH)</b>
<b>ddH<sub>2</sub>O to a final volume of 25 µL</b>

**Table XIV:** PCR reaction components.

PCR cycling conditions used are shown here (Table XV).

<b>Segment</b>	<b>Cycles</b>	<b>Temperature</b>	<b>Time</b>
<b>1</b>	1	94° C	1 min 30 sec
<b>2</b>	35	94° C	45 sec
		54° C	45 sec
		72° C	45 sec
<b>3</b>	1	72° C	7 min

**Table XV: PCR cycling conditions to amplify a *USP11* region of the genomic DNA.**

PCR amplified product was loaded in a 1.5% agarose and purified using ExoSAP-IT for PCR Product Cleanup (Affymetrix) as indicated by manufacturer's instructions. The product was subcloned by TOPO TA cloning (Thermo Fisher) following protocol instructions and sent to Sanger sequencing to check the sequence mutations in the individual alleles.

### **III.5- FUNCTIONAL STUDIES**

#### **III.5.1 UVC SENSITIVITY SURVIVAL ASSAY**

In order to determine the sensitivity of the cell lines to the nucleotide dimers lesions produced by UVC irradiation, HEK 293T cells of each transduced line were seeded in 2 mL of complete medium per duplicate in a 6-well plate. 24h afterwards, medium was removed, cells were washed with PBS and UVC irradiated (254 nm; 15 W UVC Lamp G15-T18 Philips) at the doses of 0, 2, 5, 10 and 15 J/m<sup>2</sup>. Complete medium was then added and survival cells were counted after 72 h by the Beckman-Coulter Cell Counter. At least three independent assays were performed and results were expressed as a percentage of irradiated viable cells versus (vs) viability of the untreated controls.

### III.5.2 UNSCHEDULED DNA SYNTHESIS ASSAY

In order to evaluate the repair activity of the GGR subpathway of NER (GG-NER), Unscheduled DNA Synthesis (UDS) assays were performed. A mix of poly-L and poly-D Lysine (Sigma-Aldrich) was used to coat 96-well plates to improve cell attachment. Afterwards,  $8 \times 10^4$  cells of each transduced cell line were seeded in 100  $\mu\text{L}$  of complete medium, seeding 10 replicate wells per cell line, from which half of them are UVC irradiated (254 nm) at 20  $\text{J/m}^2$  after 16 h while the other half remains as non-treated controls. After UV irradiation cells were incubated in DMEM without FBS with 5-ethynyl-2'-deoxyuridine (EdU) (Invitrogen) conjugated with fluorescent-azide. After 2h of incubation at 37°C, cells are fixed in 100  $\mu\text{L}$  of Fixation Buffer (Table XVI) and incubated for 20 min on ice. EdU is detected by adding 41  $\mu\text{L}/\text{well}$  of EdU Detection Solution (Table XVI) that exposes it to Alexa Fluor 488-azide coupling solution (Invitrogen) and nuclei were counterstained with DAPI (Dojindo) during 1h. Then, cells were washed in PBS 0.05% Tween-20 during 40 min, fixed with 100  $\mu\text{L}$  Formalin/PBS (1:10) during 20 min and image acquisition and data processing achieved using a high content screening (HCS) system, the ARRAY<sup>SCAN</sup>VTI (Thermo Scientific) Plates were scanned with a CCD camera-equipped fluorescence microscope and the images were processed with the software Cellomics Scan (Cellomics). At least three independent UDS assays were performed and results are represented as fluorescence intensity of treated and non-treated cells.

Fixation Buffer		Detection Buffer	
Final Conc.	Component	Final Conc.	Component
<b>300 mM</b>	Sucrose	<b>50 mM</b>	Tris-HCl pH 7.3
<b>2%</b>	Formalin	<b>4 mM</b>	CuSO <sub>4</sub>
<b>0.5%</b>	Triton X-100	<b>10 mM</b>	Ascorbate (Na)
	PBS	<b>10 µM</b>	Alexa 488 azide
			MQ H <sub>2</sub> O
		<b>20 ng/mL</b>	DAPI

**Table XVI: Fixation and Detection Buffers detailed components used in the UDS assay.**

### III.5.3 RECOVERY OF RNA SYNTHESIS ASSAY

To evaluate the repair activity of the TCR subpathway of NER (TC-NER), Recovery of RNA synthesis (RRS) assays were performed with all the transduced cell lines. Coating and seeding of the cells were performed as for the UDS assay. After 16h, half of the cells were UVC irradiated (20 J/m<sup>2</sup>) and incubated during 8h at 37°C in DMEM with 1% FBS serum (to allow RRS) before incubation during 2h at 37°C in 60 µL/well of serum-free DMEM supplemented with 5-ethynyluridine (EU) (100 µM). Afterwards, cells were fixed, EU was detected and image acquisition was done as in the UDS assay. At least three independent RRS assays were performed and results are represented as fluorescence intensity of treated and non-treated cells. Detailed UDS and RRS methodologies were provided by Dr Ogi group (Jia et al. 2015).

### III.5.4 DEB SENSITIVITY SURVIVAL ASSAY

To evaluate the sensitivity of the cell lines to ICL, 2 x 10<sup>5</sup> HEK 293T cells of each transduced line were seeded in 2 mL of complete medium per duplicate in a 6-well plate. 24h afterwards, DEB was added at a final concentration of 0, 0.025, 0.05, 0.1 and 0.2 µg/mL.

Cells were grown during the time needed by the untreated to perform at least three population doublings. Afterwards cells were harvested by trypsinization and counted with a Beckman-Coulter Cell Counter. At least three independent assays were performed and results are expressed as a percentage of treated viable cells vs the untreated controls.

### **III.5.5 DEB-INDUCED G2/M CELL CYCLE ARREST**

$10^6$  cells of each cell line were seeded in 3 x 25 cm<sup>2</sup> flasks (F25) in 5 mL of complete medium. 24 h afterwards DEB was added at doses of 0, 0.005, 0.01, 0.025 and 0.05 µg/mL. 48 h after the treatment, cells were trypsinized, PBS washed and resuspended in 200 µL of PBS. 2 mL of ice-cold ethanol 70% was added and cells were placed for 30 min on ice. Ethanol was removed by centrifugation and substituted with 2 mL of Staining solution (40 µg/mL Propidium Iodide, PI, Invitrogen; Pure Link RNasa A 0.1 mg/mL, Invitrogen; PBS 1x). Cell cycle population distribution depending on DNA amount was analyzed by flow cytometry with FACSCalibur (BD Biosciences) as in (Bogliolo et al. 2013). 15,000 events were registered per sample and data was analyzed by FlowJo VX software.

### **III.5.6 MICRONUCLEI- FRAGILITY TEST**

Micronuclei (MN) are small nuclei close to the main nucleus. The origin of MN is due to mal-segregation of whole chromosomes or acentric chromosome fragments which makes them unable to be properly distributed by the mitotic spindle during cell division.

Thus, MN are a sign of genotoxic events and chromosomal instability generated by chromosomal breaks produced either by errors during replication or by ICL agents exposure.

Traditional chromosome aberrations analyses are highly time-consuming and require more technical expertise. Notwithstanding, MN scoring system is one of the most widely used methods to measure genome damage in humans (Fenech et al. 2011). MN assay associated to flow cytometry represents a fast and robust system to obtain cytogenetic damage information (Avlasevich et al. 2006).

Around  $3 \times 10^5$  cells from each cell line were seeded in 6-well plates. Twenty four hours later cell cultures were untreated or treated with 0.01 µg/mL of DEB and they were kept in culture enough time for at least one population doubling. Cells were then sequentially stained in order to distinguish MN from apoptotic debris and necrotic chromatin. First of all a staining with Ethidium Monoazide Bromide (EMA) (0.025 mg/mL) and secondly a staining with Sytox green (0.2 µM). EMA is a dye which covalently binds to chromatin of dying and dead cells after a photo-activation step achieved by keeping cells under a 60 W light bulb (about 30 cm distance) for 20 min. EMA allows to detect dying cells because it just can reach the cells whose membranes are compromised. After the photo-activation period, a wash with cold PBS with 2% FBS was added to remove EMA that was not covalently bound to DNA. After that, a lysis step with 250 µL of lysis solution 1 (0.584 mg/mL NaCl, 1 mg/mL sodium citrate, 0.3 µg/mL IGEPAL, 1 mg/mL RNase A and 0.2 µM Sytox green in deionized water) during 1h at RT was done. Later, a second lysis step was done adding 250 µL of solution lysis 2 (85.6 mg/mL sucrose, 15 mg/mL citric acid and 0.2 µM Sytox green in deionized water) during 30 min at RT. Lysis produces degradation of cytoplasmic membrane that allow penetrance of Sytox green dye that stains nuclei and MN. In this way, this process allowed a differentiation of particles labeled with Sytox green (nuclei and MN of alive cells) or Sytox green and EMA (apoptotic cells). After lysis, samples were stored at 4 °C until being processed by flow cytometry (up to two days).

Data acquisition was performed by flow cytometry with FACSCalibur; Sytox-associated fluorescence was detected by FL1 channel while EMA-associated fluorescence was detected by FL3 channel. Collected data was analyzed by Flow Jo VX software.

The data of MN presented in this work represents results from five independent experiments each one in duplicate.

### **III.6- PROTEIN PURIFICATION**

#### **III.6.1 TANDEM AFFINITY PURIFICATION**

HEK XPF-KO cell line expressing the wt version of XPF with the tandem N-terminal tags SBP and CBP and the HEK XPF-KO cell line expressing an empty vector (EV) tagged were grown until reaching  $1 \times 10^8$  cells of each cell line. Cells expressing TAP-XPF were either non treated or subjected to MMC (1  $\mu$ M) for 16h or to UVC irradiation (150 J/m<sup>2</sup>) and detached after 1h. Cell pellets were processed and purified using InterPlay Mammalian TAP System (Agilent Technologies) according to manufacturer's guidelines except that streptavidin resin was substituted with Streptavidin Magnetic Sepharose beads (GE Healthcare Life Sciences) (40  $\mu$ l/sample). Calmodulin resin from the last purification step was washed three times in Tris 1M pH 8.5 and 2/3 of the resin volume was sent to mass spectrometry service (PRBB Proteomics- MS facilities) and 1/3 was boiled with Laemmli buffer and loaded in a standard WB gel.

#### **III.6.2 HA-PURIFICATION**

HEK XPF-KO cells infected with the XPF-HA were used for XPF protein purification through its C-terminal HA-tag. HEK XPF-KO was used as a control. Two 20-cm culture dishes of HEK XPF-KO were seeded and six 20-cm dishes for HEK XPF-KO +XPF-HA. Out of these six, two dishes were treated with MMC (1.5  $\mu$ M) during 16h and two with 50 J/m<sup>2</sup> of UVC. UVC treated cells were incubated during 1h at 37°C before being processed. Cells were trypsinized and lysed with 2 mL of Lysis Buffer (Table XVII) per sample during 15 min at 4°C. Later, 8 mL of Dilution Buffer (Table XVII) were added before sample sonication (2 x 1 min) and centrifuged at 13,000g during 20 min at 4°C.

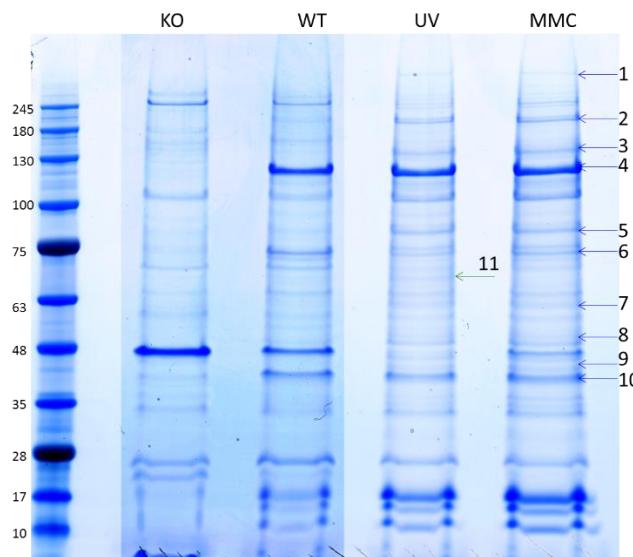
Lysis Buffer		Dilution Buffer	
Final Conc.	Component	Final Conc.	Component
<b>50 mM</b>	Tris HCl, pH 7.5	<b>50 mM</b>	Tris HCl, pH 7.5
<b>150 mM</b>	KCl	<b>150 mM</b>	KCl
<b>1 mM</b>	EDTA	<b>1 mM</b>	EDTA
<b>1%</b>	NP-40	<b>0.1%</b>	Triton X-100
<b>10%</b>	Glycerol	<b>10%</b>	Glycerol
<b>1x</b>	Proteases inhibitor cocktail	<b>1x</b>	Proteases inhibitor cocktail
	H <sub>2</sub> O		H <sub>2</sub> O

**Table XVII: Lysis and Dilution Buffers detailed components used for the HA-purification.**

Supernatants were filtered with 0.45 µm filters (Millex Syringe Filters) and total protein quantified by Bio-Rad Protein Assay (Biorad). Samples concentrations were adjusted to 10 mg of total protein per tube using 1:5 Lysis:Dilution Buffer. 100 µL of these samples were kept frozen to be used as controls (whole cell extracts (WCE)). Protein purification was performed adding 40 µL of Pierce anti-HA magnetic beads (Thermo Scientific) previously washed in 1:5 Lysis:Dilution buffer. Beads were added to each sample tube and kept rotating overnight at 4°C. The following day tubes were centrifuged twice at 2,500 g during 5 min at 4°C, supernatants were discarded and anti-HA magnetic beads were washed ten times in 1:5 buffer before 200 µL of HA Synthetic Peptide (Thermo Fisher) was added. Samples were incubated shaking at 37°C for 10 min to elute the proteins from the magnetic beads. 10 µL of eluted samples were loaded in a 4-12% agarose gradient WB gel (Mini-PROTEAN Precast gels, Bio-Rad) to check the protein controls. Antibodies used for this WB were: XPF (Mouse Monoclonal Ab-1 (219) Thermo Scientific 1:200) and Tubulin (Mouse, ab7291, Abcam 1:10,000).

### III.6.2.1 PROTEIN BAND EXTRACTION FROM SDS GEL

The remaining samples were concentrated by boiling until reducing the volume to around 50 µL and loaded in a 4-12% SDS gel, run at 0.15 Amp for 1:10h. The gel was stained with Coomassie Blue Staining (Thermo Scientific) for around 5h and destained in H<sub>2</sub>O overnight. The following day individual protein bands were selected to be cut out from the gel with a scalpel (Figure 21).



**Figure 21:** Blue Coomassie SDS gel of XPF-HA purification.

Each individual band was cut in four fragments and dissolved first in 500 µL of Destaining buffer 1 (Table XVIII) during 30 min shaking at 40°C and secondly in 500 µL of Destaining buffer 2 (Table XVIII).

Destaining buffer 1	
Final Conc	Component
25 mM	Ammonium Bicarbonate
50%	Metanol
	H <sub>2</sub> O

Destaining buffer 2	
Final Conc	Component
25 mM	Ammonium Bicarbonate
50%	Acetonitrile
	H <sub>2</sub> O

**Table XVIII:** Distaining buffers 1 and 2 detailed components used for the protein band extraction of SDS gel.

Then, fragments were washed twice in 500 µL of 25 mM Ammonium Bicarbonate during 2 min shaking, the solution was aspirated and 200 µL of Acetonitrile were added during 15 min to remove the water from the gel. Acetonitrile was then removed by a speed vacuum during 10 min. 100 µL of 1:10 Dithiothreitol (DTT) prepared in Ammonium Bicarbonate were added to the samples during 1 h shaking at 56°C. DTT was then removed and samples were washed in 200 µL 25 mM Ammonium Bicarbonate shaking at RT during 10 min. Iodoacetamide (IAA) diluted in 25 mM Ammonium Bicarbonate was added during 45 min in darkness shaking at 25°C. After a brief spin, the supernatant was removed and the samples were washed twice in 500 µL for 2 min, then in 200 µL of Acetonitrile for 15 min at RT and dried with the speed vacuum at RT for 15 min. Samples were then digested with 30 µL of Trypsine prepared in 25 mM Ammonium Bicarbonate during 10 min. Afterwards, 40 µL of 25 mM Ammonium Bicarbonate were added and kept during 16h at 37°C. Later on, supernatants were transferred to new eppendorfs, 40 µL of 50% Acetonitrile were added and samples were kept in a water bath sonicator for 10 min. The supernatant was removed again and exchanged by 40 µL of 1% Formic Acid during 10 min on the water bath sonicator. These two last steps were repeated and the supernatants were collected. A final water bath sonicator step of 10 min with 70% Acetonitrile was carried and the total supernatant volume was filtered with Centrifugal Filter Units (Millipore) during 1 min at 13,000 g and evaporated until 20 µL of final volume per sample. This volume was finally sent to mass spectrometry service, the ions from the sample were separated according to their mass-to-charge ratio and results were displayed as spectra of the relative abundance of detected ions as a function of the mass-to-charge ratio by specialized softwares.

### **III.7- STABLE ISOTOPE LABELING WITH AMINO ACIDS IN CELL CULTURE**

Stable isotope labeling with aa in cell culture (SILAC) is a technique for *in vivo* incorporation of a label into proteins for mass spectrometry (MS)-based quantitative proteomics (Ong et al. 2002). HEK XPF-KO transduced with XFP-Wt were left untreated or treated with UVC (40J/m<sup>2</sup>, 1h recovery time) or MMC (0.5 µg/mL for 21h) treatments in identical culture media except for a ‘light’ (HEK XPF-KO cells untreated), ‘middle’ (HEK XPF-

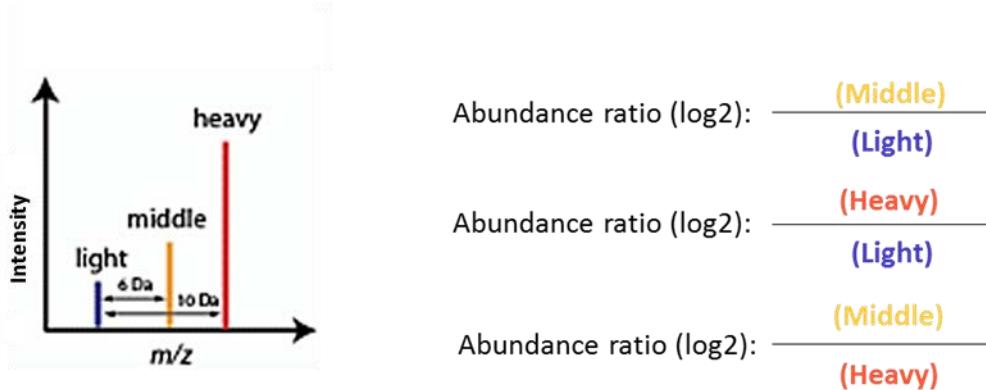
KO + XPF-Wt treated) or ‘heavy’ (HEK XPF-KO + XPF-Wt untreated) forms of lysine and arginine.

Cells were cultured for 14 days in lysine- and arginine-free DMEM (Thermo Fisher Scientific) containing 10% dialyzed FBS with 200 mg/L L-proline (Sigma). Light media was supplemented with 100 mg/L L-lysine (Thermo Fisher Scientific) and 100 mg/L L-arginine (Thermo Fisher Scientific), middle media was supplemented with 100 mg/L  $^{15}\text{N}_4$ -L-lysine (Thermo Fisher Scientific) and 100 mg/L  $^{13}\text{C}_6$ -L-arginine (Wako Chemicals, Japan), heavy media was supplemented with 100 mg/L  $^{13}\text{C}_6,^{15}\text{N}_2$ -L-lysine (Wako Chemicals, Japan) and 100 mg/L  $^{13}\text{C}_6,^{15}\text{N}_4$ -L-arginine (Wako Chemicals, Japan).

Cells were lysed in EBC buffer supplemented with protease inhibitor cocktails (Nacalai Tesque, Japan) with brief sonication, and HA-XPF and its interacting proteins were enriched using anti-HA magnetic beads (Thermo Fisher Scientific). Proteins were resolved by SDS-PAGE and in-gel digested with trypsin by standard protocol. Peptide fractions were analyzed on an Easy-nLC1000 coupled to a Q-Exactive instrument (Thermo Fisher Scientific) equipped with a nanoelectrospray source. Peptides were separated on a 50 cm analytical column (75  $\mu\text{m}$  inner diameter) with 2  $\mu\text{m}$  C18 beads (Thermo Fisher Scientific). The column temperature was maintained at 50°C using an integrated column oven (EASY-Spray, Thermo Fisher Scientific). Each peptide fraction was separated using a 65 min gradient ranging from 0% buffer B (100% ACN and 0.1% formic acid) to 35% buffer B in 60 min and ramped to 95% buffer B in 5 min at a flow rate of 300 nL/min. The washout followed at 95% buffer B for 10 min. Spray voltage was set to 2 kV, s-lens RF level at 50, and heated capillary temperature 320°C. All experiments were performed in the data-dependent acquisition mode to automatically isolate and fragment top10 multiply-charged precursors (+2, +3 and +4) according to their intensities. Former target ions were dynamically for 15 seconds excluded and all experiments were acquired using positive polarity mode. Full scan resolution was set to 70,000 at m/z 200 and the mass range was set to m/z 380-1500. Full scan ion target value was 3E6 allowing a maximum fill time of 60 ms. Higher-energy collisional dissociation (HCD) fragment scans was acquired with optimal setting for parallel acquisition using 1.6 m/z isolation width and normalized collision energy of 27.

The data were analyzed using SEQUEST in Proteome Discoverer 2.1 (Thermo Fisher Scientific) and searched in the complete human proteome database (Swiss-Prot). The mass tolerances for the precursor and fragment were 10 ppm and 0.02 Da, respectively. Cysteine carbamidomethylation was included as a fixed modification, and N-terminal protein acetylation and methionine oxidation were included as variable modifications. Peptide identification was filtered at a false discovery rate (FDR) < 1%.

This technique provided quantitative information about the interactions established by each cell condition based on the mass difference of the different stable-isotope composition. The ratio of peak intensities reflects the relative protein abundance (Figure 22). Abundance ratios were considered reliable over 1.



**Figure 22: SILAC experiment with three cell populations labeled. Ratios between samples are calculated comparing the differences in the intensities of the peaks (adapted from (Geiger et al. 2011)).**

### **III.8- COIMMUNOPRECIPITATIONS**

In order to confirm protein complexes interactions of XPF and USP11 proteins, endogenous coimmunoprecipitations (CoIP) were performed. 2 x 20 cm Petri dishes were seeded of each cell line. 48h afterwards, cells were trypsinized and lysed using the same buffers as detailed in HA-purification section. After protein quantification and adjustment of the samples to 10 mg in 10 mL, they were incubated overnight with primary antibodies in a final concentration of 1  $\mu$ g/mL. Antibodies used for the endogenous IP were: XPF (Mouse

Monoclonal Ab-1 (219) Thermo Scientific) and USP11 (Rabbit Monoclonal ab109232, Abcam). The following day, 50 µL of Protein G Magnetic Sepharose beads (GE Healthcare, Life Sciences) were incubated during 3 h rotating at 4°C with the samples containing the antibodies. After this time antibodies were attached to the beads and these were washed 10 times in 1:5 Dilution buffer as detailed in HA-purification section. Protein complexes were then detached from magnetic beads by boiling them in 40 µL of Laemmli buffer and the eluted proteins were loaded in a MiniProtean Precast gels (Bio-Rad) and standard WB were performed to check CoIP of both proteins.

Besides increasing the amount of XPF-HA tagged in the sample, HA-purifications were also performed to confirm CoIP of USP11 protein following the same protocol.

### **III.9- IN VIVO DOUBLE STRAND BREAKS REPAIR ASSAYS**

#### **III.9.1 HOMOLOGOUS RECOMBINATION REPAIR ASSAY**

To test the contributive role of XPF and USP11 in HR repair pathway, U2OS carrying the chromosome-integrated reporter DR-GFP (provided by Dr Jasin) were used. This reporter is designed to express the GFP when the HR pathway is activated by the co-expression of the I-SceI endonuclease and GFP fluorescence can be detected and quantified by flow cytometry (Figure 23).

#### **DR-GFP**



**Figure 23: Scheme of the inducible Sce-I reporter system for HR pathway.** The reporter contains a GFP gene interrupted by a I-SceI restriction site and a fragment of 5' and 3' truncated of the GFP gene (adapted from (Bennardo et al. 2008)).

siRNA oligos were designed (Sigma) to downregulate XPF and USP11. Negative control was the siRNA of Luciferase and positive control was the siRNA of BRCA2 (Table XIX).

$5 \times 10^5$  U2OS-DR cells were seeded in 6-well plates with 2 mL of DMEM medium. siRNA transfections were performed using Lipofectamine RNAiMAX (Invitrogen) (Table XIX) Transfection Reagent and Opti-MEM (Gibco, Life Technologies) following manufacturer's instructions during two days in a row per duplicate.

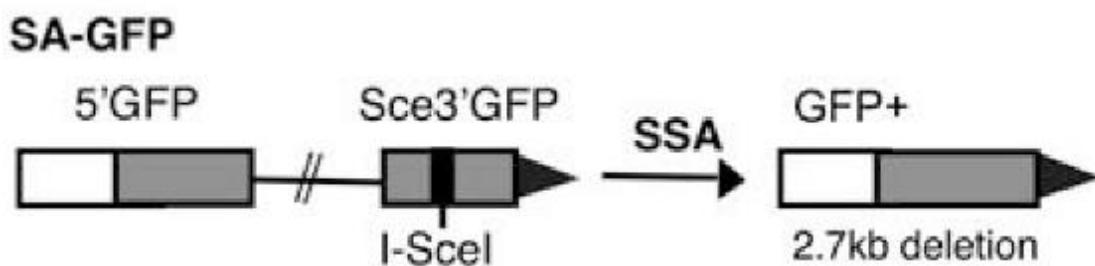
siRNA	Sequence	siRNA concentration (nM)	Volume RNAiMAX ( $\mu$ L/well)
Luciferase	5'-CGUACGCGGAAUACUUCGATT-3'	30	7.5
XPF	5'-GCACAAACCUGAAGGUUGUA-3'	30	7.5
USP11	5'-CAGAGAUGAAGAAGCGUUA-3'	10	2.5
BRCA2	5'-GGAUUAUACAUUUUCGCA-3'	10	2.5

**Table XIX:** siRNA oligo sequences of each gene to target protein downregulations, siRNA concentrations and Lipofectamine RNAiMAX volumes used for the U2OS-DR transfections.

Three different vectors were used to obtain the frequency of transfected cells and the repair efficiency: an EV (CAGGS) as negative control, a GFP expression vector (NZE-GFP) as a positive control and I-Sce endonuclease expression vector (CBAS, kindly provided by Dr Maria Jasin, New York). Cells downregulated with siRNA were transduced with 2.5  $\mu$ g of one of the three vectors, Polyethylenimine (PEI) (Sigma) and Opti-MEM at a ratio 1:4 DNA/PEI. Two days later cells were analyzed by cell cytometry using the FLH1 channel for the green fluorescence. Data was finally analyzed with FlowJo VX software.

### III.9.2 SINGLE STRAND ANNEALING REPAIR ASSAY

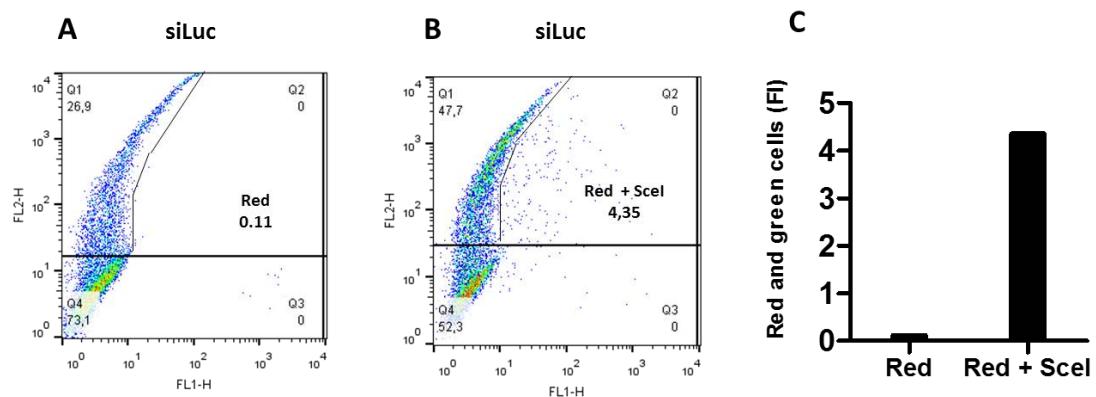
U2OS cells used in this assay were carrying the chromosome-integrated reporter SA-GFP to control the SSA induced by the I-SceI endonuclease (Addgene N° 41594). The reporter was designed to be restored when the SSA repair pathway works (Figure 24); the GFP gene recovers its structure and fluorescence can be detected and quantified by flow cytometry.



**Figure 24: Scheme of the inducible Sce-I reporter system for SSA pathway.** The reporter contains a 5' fragment of GFP gene and a 3' fragment of GFP gene with the I-SceI site (adapted from (Bennardo et al. 2008)).

Same siRNA sequences as the ones used in the previous *in vivo* described assay were used to evaluate the role of these proteins in SSA.

All the cells were transduced with 2.5 µg of a vector named pLKO5.sgRNA.EFS.tRFP (Addgene 57823) which produces a red fluorescent protein, and one replicate of the U2OS-SA cells transfected with all the siRNA downregulated genes was, besides, transduced with 2.5 µg of the vector containing the sequence for the I-SceI endonuclease (CBAS). The transfection was performed with PEI (Sigma) and Opti-MEM at a ratio 1:4 DNA/PEI. Two days later cells were analyzed by cell cytometry using the FLH2 channel for the red fluorescence and FLH1 for the green fluorescence. Data was analyzed with FlowJo VX software; double red+green fluorescence values were analyzed for each siRNA transfection transduced either with red or red+SceI vectors (Figure 25); fluorescence values from the red transduced vector were then subtracted from the double transduced red+SceI values and data were represented relative to the negative control (siLuc).



**Figure 25: Double fluorescence intensity values in the control (siLuc) sample.** For each siRNA transfection, the number of red+green fluorescent cell was analyzed when transducing the cells either with (A) red vector or (B) red+Scel vectors. (C) Fluorescence intensity (FI) in the double transduced sample was almost 40 times higher than in the red vector transduced sample.

## **Results and Discussion**



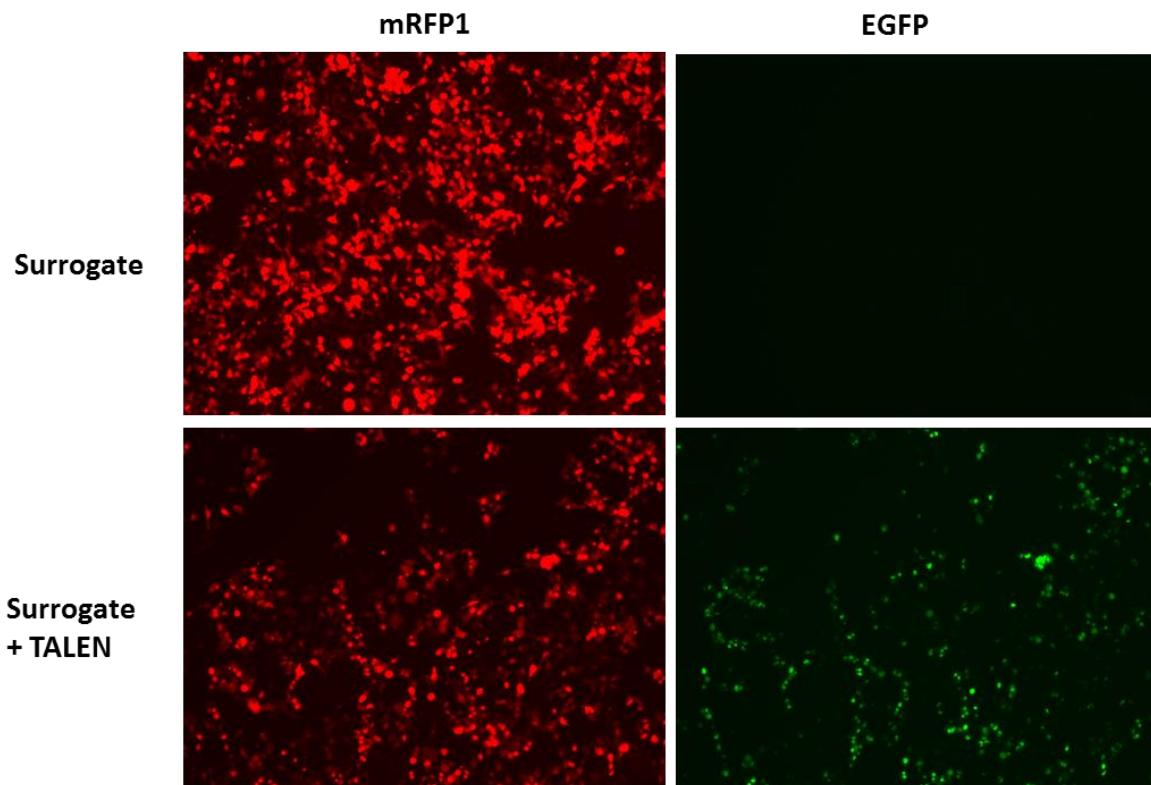
## IV. RESULTS AND DISCUSSION

### **PROJECT 1**

#### **IV.1- ESTABLISHING OF THE HEK 293T TALEN *ERCC4*-KNOCKED OUT AS THE XPF-KO BACKGROUND CELL LINE**

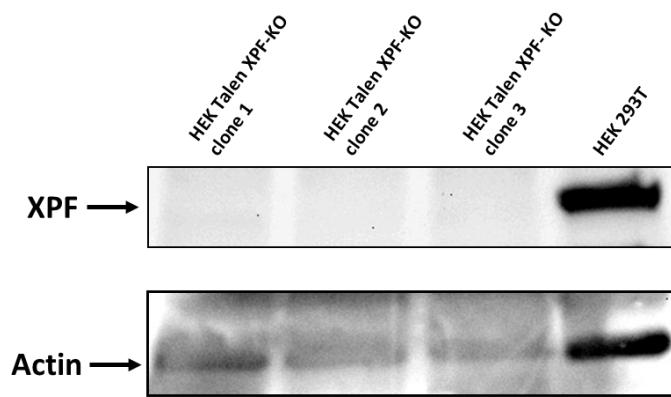
The purpose of this project was to analyze the genotype-phenotype correlation of *ERCC4* pathogenic variants. As it is well known, genetic background influences phenotypes and the creation of isogenic cell lines has been a key point of this study. TALEN technology was used to edit the genome of HEK 293T cells. TALEN nucleases were engineered to bind to an *ERCC4* specific sequence to disrupt the gene sequence and generate a human cell line KO for this gene.

Our system consisted of different vectors, a pair of TALEN plasmids (forward and reverse) with the target sequence fused to a FokI nuclease domain and a reporter-based plasmid containing the same *ERCC4* recognition sequence, the red fluorescent protein sequence and an out of frame GFP that could be restored when the DSB was repaired by NHEJ. The fluorescence was visible 48 h post transfection with TALEN and surrogate plasmids. Cells where TALEN proteins were cutting the DNA target site showed red and green fluorescence as showed in figure 26.



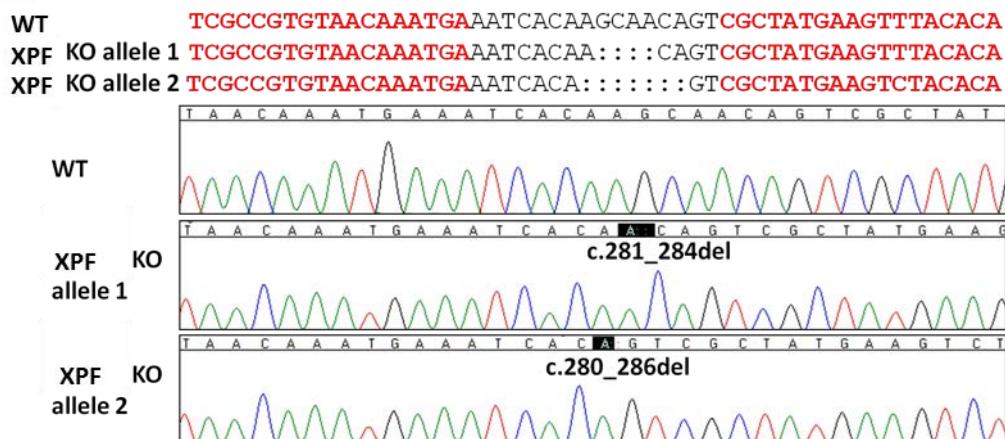
**Figure 26: Analysis of TALEN endonuclease activity by fluorescence microscopy.** Cell cultures images 48h after transfection with either Surrogate vector or Surrogate + TALEN vectors are shown. Red fluorescence is detected in cells transfected with Surrogate (RFP+) vector; red and green fluorescence is detected in cells transfected with Surrogate + TALEN (RFP+GFP+) vector. No presence of double positive cells was found in cell cultures transfected just with the Surrogate vector.

After cell sorting to select the double fluorescent cells, single cell cloning was performed and, 2-3 weeks later, clones were picked and checked by WB to identify any *ERCC4* KO ones. Three clones showed no XPF band (Figure 27) and clone number 1 was selected to continue the KO validation.



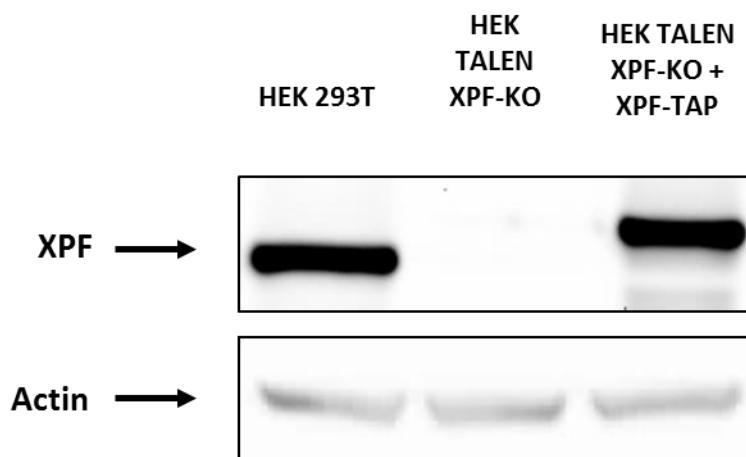
**Figure 27: WB analysis of XPF TALEN clones.** None of the three clones showed an XPF band meaning that genome edition in order to create a KO was successful. Actin band is shown as a loading control.

Sequencing analysis of genomic DNA from clone 1 revealed that the mutations introduced by the TALEN were two different deletions of 4 and 7 bp respectively: c.281\_284del and c.280\_286del (Figure 28). Primer walking sequencing of the whole *ERCC4* gene confirmed that the rest of the sequence remained unaltered.



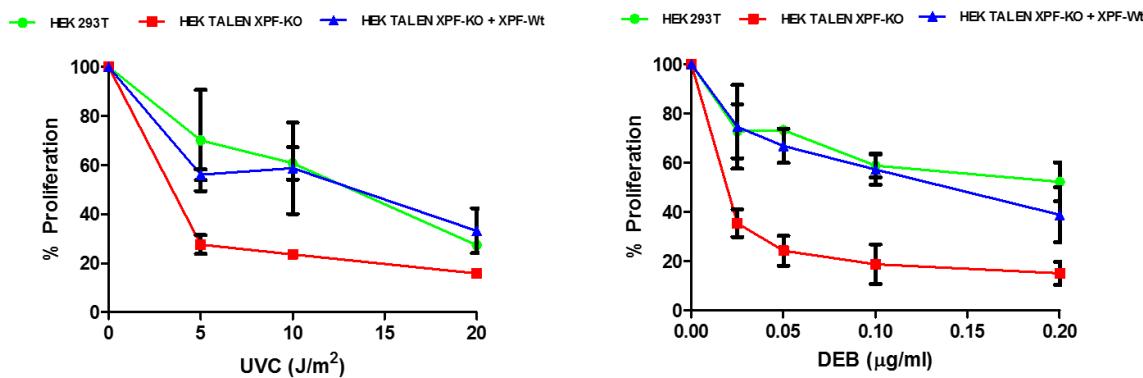
**Figure 28: Sequencing reaction chromatograms of heterozygous mutant clones.** The KO clone was a compound heterozygote for a 4 bp deletion (c.281\_284del) and a 7 bp deletion (c.289\_286del) both in exon 2.

The wt cDNA of *ERCC4* modified with SBP and CBP epitopes in N-terminal and HA epitope in C-terminal sites cloned in the pULTRA vector (see materials and methods) was transduced in the HEK XPF-KO cell line (Figure 29). Subsequent, GFP sorting by flow cytometry allowed selection of the infected cells.



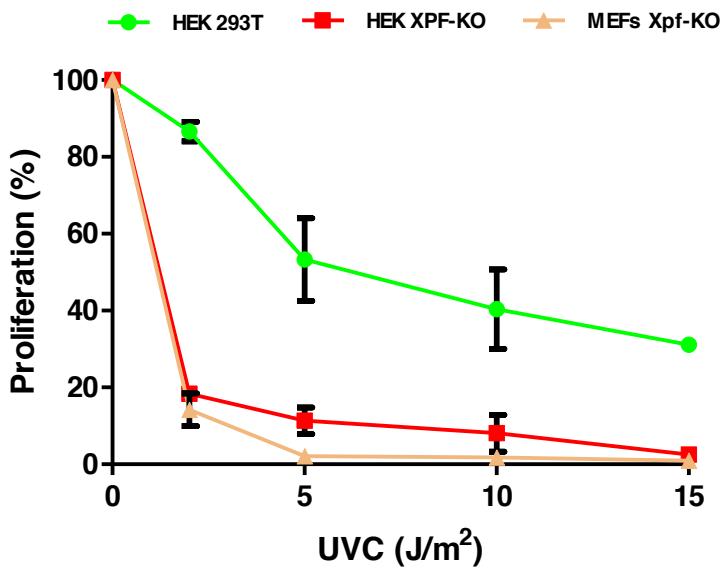
**Figure 29: WB validation of the HEK XPF-KO cell line transduced with XPF-tagged viral vectors.**  
XPF band is clearly detectable in HEK 293T and in HEK TALEN XPF-KO + XPF-TAP. The slight difference in the molecular weight from the endogenous XPF of HEK 293T to the exogenous one was due to the TAP and HA epitopes. Actin band is shown as a loading control.

To test whether the exogenous XPF was complementing the cellular phenotype, UVC and DEB sensitivity survival assays of these cell lines were performed. According to expectations, the HEK XPF-KO cell line was hypersensitive to UVC or ICL treatments. HEK XPF-KO was more than 4 times more sensitive to UVC irradiation and more than 10 times to DEB treatment than the control line HEK 293T, however XPF-KO complemented with the XPF-Wt restored the cellular phenotype after both treatments (Figure 30). This genetic complementation evidenced TAP and HA epitopes were not affecting XPF function.



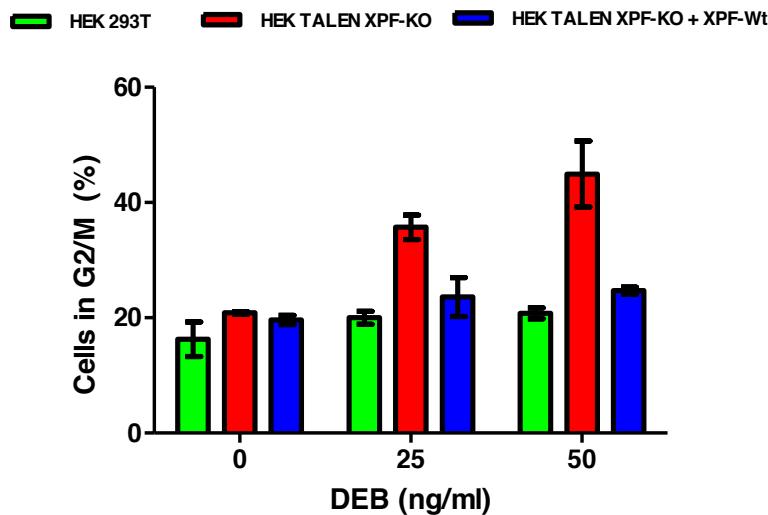
**Figure 30: Functional studies of HEK XPF-KO transduced with XPF-Wt compared to positive and negative controls.** The proliferation percentage of HEK XPF + XPF-Wt cells was similar to the positive control under UVC irradiation or DEB treatment as exogenous XPF restores the cell phenotype. Graphs represent proliferation percentage media of two independent experiments of two replicates with standard deviation (SD).

Although the phenotype of cells lacking *ERCC4* was previously studied in MEFs (Bogliolo et al. 2013), this was the first successful attempt to edit a human cell line to achieve an *ERCC4* KO. Previous studies comparing prevalence of *ERCC4* mutation frequency observed in patients with the mutation frequency expected to obtain according to Hardy-Weinberg principle revealed under representations of biallelic combinations of *ERCC4*, indicating that most of biallelic mutations in humans are embryo lethal (Osorio et al. 2013). Actually, all the patients presenting biallelic mutations in XPF retain at least a hypomorphic allele (Bogliolo et al. 2013; Kashiyama et al. 2013). Regarding the similar embryo lethality described in MEFs Xpf-KO, an additional survival including this MEF Xpf-KO cell line was performed to compare the sensitivity to UVC irradiation of the two XPF-KO lines among the different cell models. Consistently, HEK XPF-KO were as sensitive as MEFs Xpf-KO (Figure 31).



**Figure 31: UVC cell survival to compare XPF-KO in different cell lines.** The TALEN edited HEK XPF-KO showed levels of UVC sensitivity comparable to MEFs Xpf-KO, previously reported as sensitive. Graph represents proliferation percentage media and SD of two independent experiments of two replicates.

If left unrepaired, chromosome alterations caused by ICL produce a block of the cell cycle in G2/M phase. The HEK XPF-KO phenotype was tested for the percentage of cells stalled in G2/M after being treated with two different doses of DEB. As shown in figure 32, KO cells showed higher amounts of G2/M arrested cells due to the inability of repairing the accumulation of alterations in the chromosomes. The same phenotype was observed in MEFs Xpf-KO (Osorio et al. 2013). Furthermore, analyses of the cell cycle also confirmed that accumulation of cells in G2/M phase of the XPF-KO was restored to the normality after introduction of the XPF-Wt.

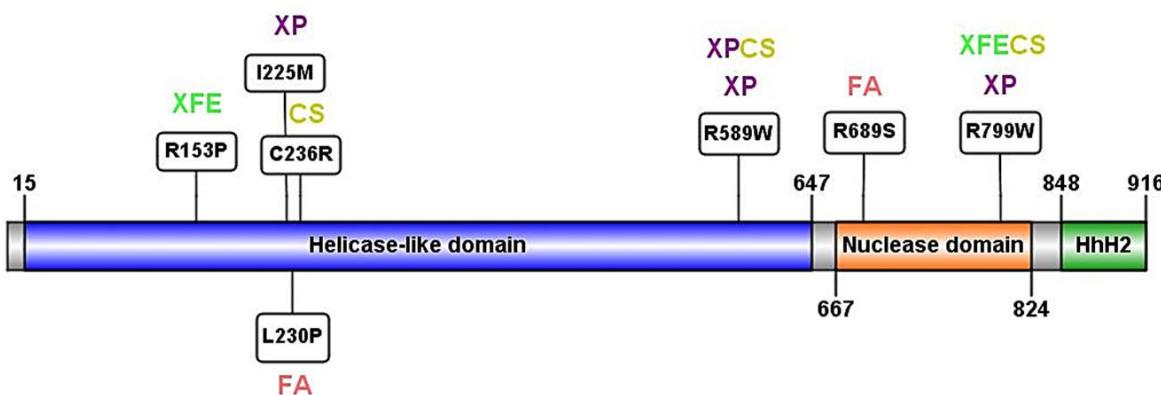


**Figure 32: Cell cycle study of HEK XPF-KO cells transduced with exogenous XPF-Wt protein.**  
XPF-KO cells transduced with XPF-Wt reduced their percentage of cells stalled in G2/M phase to similar levels to the HEK 293T. Graph represents the media of two independent experiments with SD.

#### **IV.2- ANALYSES OF DIFFERENT XPF VARIANTS IN AN XPF-KO BACKGROUND**

##### **IV.2.1 SELECTION OF THE XPF SINGLE NUCLEOTIDE VARIANTS TO STUDY**

Among the several mutations described in patients we selected seven different single nucleotide changes of *ERCC4* that had been associated with a characteristic clinical phenotype (Figure 33).



**Figure 33: Schematic view of the XPF domains with the selected mutations.** Seven single XPF disease-associated variants were selected for the study. Five of them correspond to the helicase-like domain and two of them are located in the nuclease domain. In green, XFE; in purple: XP; in pink, FA; in mustard: CS. The patient XPCSCD had an allele with C236R and the other with R589W. R799W mutation found in homozygosis in XP patients was also found in a cohort of patients with XFE sharing CS features. This figure was created with a protein designing software from (Liu et al. 2015).

The aminoacidic non-conservative substitution of a highly conserved arginine for a proline, R153P, was caused by a nucleotide change (c.458G>C) found in homozygosity in an XFE patient. This aminoacidic change affects the helicase domain, a leucine-rich region involved in the interaction with SLX4 and DNA binding (Klein Douwel et al. 2017). The patient's main feature was the accelerated ageing phenotype and the disease was named as XFE progeroid syndrome (Niedernhofer et al. 2006). Catalytic activity of XPF<sup>R153P</sup> was almost intact but the mutant was unable to properly locate into the nucleus and remained mislocalized in the cytoplasm due to protein misfolding (Ahmad et al. 2010).

The aminoacidic change I225M caused by a nucleotide substitution (c.653A>G) was found in heterozygosity with another missense mutation (c.1515G>A; G513R) in a patient showing mild XP features (Matsumura et al. 1998). Although I225M is located in the same domain as the R153P, XPF is stable and its interacting ability with SLX4 is not affected (K. Hashimoto et al. 2015). The other missense mutation found in the patient, G513R, is located

C-terminal further in the same domain but it does not lead to complete dysfunction in NER, which explains the mild XP phenotype of XPF<sup>I225M</sup> (Matsumura et al. 1998).

The aminoacidic change of a highly conserved residue L230P caused by the nucleotide change c.689T>C was found in the paternal allele of an FA patient carrying a 28 bp duplication in exon 11 of the maternal allele (c.2371\_2398dup28 [p.Ile800Thrfs\*24]). The duplication truncates XPF by missing the HhH<sub>2</sub> domain (Bogliolo et al. 2013). The HhH<sub>2</sub> domain truncated by the duplication is essential for ERCC1 binding while the analyzed mutation affects SLX4 binding (K. Hashimoto et al. 2015).

The aminoacidic change C236R (c.706T>C) was found in heterozygosity in two different patients: a patient with CS disease carried this mutation located in the N-terminal helicase domain and an exon 8 frameshift insertion which generated a premature stop codon (c.1730\_1731insA [p.Tyr577\*]) in the other allele. A second patient carried c.706T>C mutation in heterozygosity with another missense mutation (c.1765C>T, p.R589W), previously found in XP individuals. This second patient showed a combined syndrome of CS and XP named CSXPCD (Kashiyama et al. 2013). Interestingly, XPF<sup>C236R</sup> does not affect SLX4 binding despite being in N-terminal region very close to other involved mutations, while XPF<sup>R589W</sup> severely affects its structure and SLX4 binding ability (K. Hashimoto et al. 2015). R589W change was previously associated with different XP clinical phenotypes: combined with a deletion in exon 3 of the second allele caused severe XP while in heterozygosity with the missenses mutations causing R799W or P379S aminoacidic changes caused mild XP (Ahmad et al. 2010; Kashiyama et al. 2013).

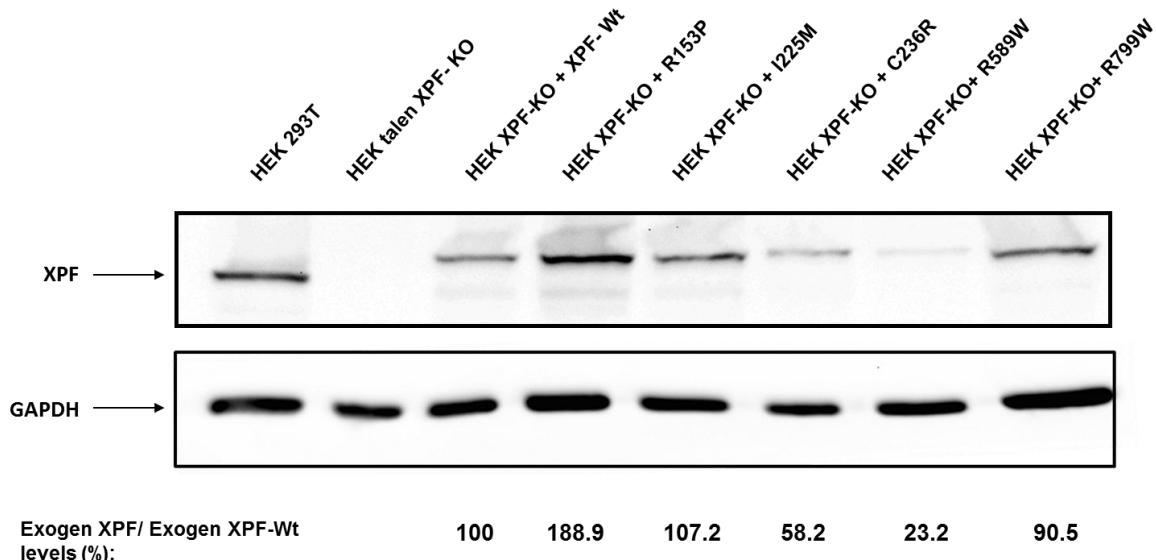
In another FA patient a new *ERCC4* missense mutation was found in exon 11 of one allele (c.2065C>A [p.Arg689Ser]) while the other allele had a 5 bp deletion in exon 8 (c.1484\_1488delCTCAA) that leads to a frameshift and a premature stop codon (p.Thr495Asnfs\*6) (Bogliolo et al. 2013). This mutation is in the nuclease domain and it affects its excision activity.

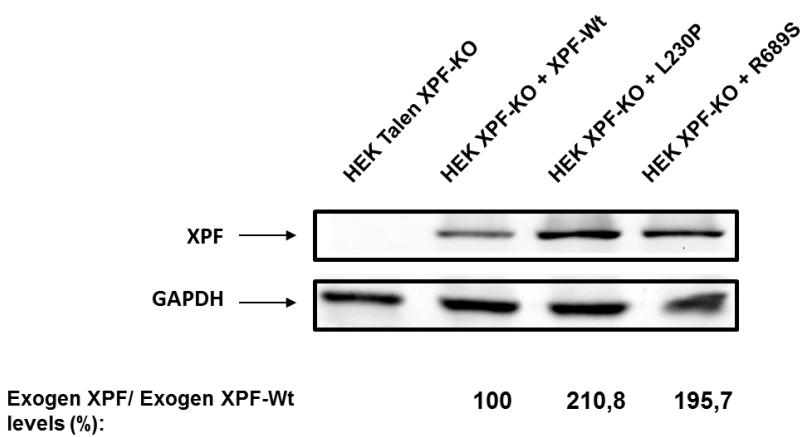
The c.2395C>T (R799W) was found in homozygosis in a patient (Sijbers et al. 1998) diagnosed with XP disease but recently was found in heterozygosity with a truncated allele (c.388+1164\_792+795del (p.Gly130Aspfs\*18)) in a patient from a cohort of patients with

progeria syndromes. The patient showed low RRS levels typical of CS phenotype (Mori et al. 2018). Thus is possible that, regarding also the XP phenotype found in the heterozygote R799W/R589W, a dosage effect of XPF<sup>R799W</sup> can influence the phenotypical outcomes of the patients.

#### **IV.2.2 EXPRESSION OF THE XPF MUTANT VARIANTS IN AN XPF-KO BACKGROUND**

The expression levels of all the mutant XPF proteins (XPF<sup>R153P</sup>, XPF<sup>I225M</sup>, XPF<sup>L230P</sup>, XPF<sup>C236R</sup>, XPF<sup>R589W</sup>, XPF<sup>R689S</sup> and XPF<sup>R799W</sup>) were assessed by WB (Figure 34).





**Figure 34: WB levels of each single variant XPF mutant transduced cell line.** Levels of XPF proteins are expressed as a percentage relative to the exogenous XPF-Wt. GAPDH is used as a loading control.

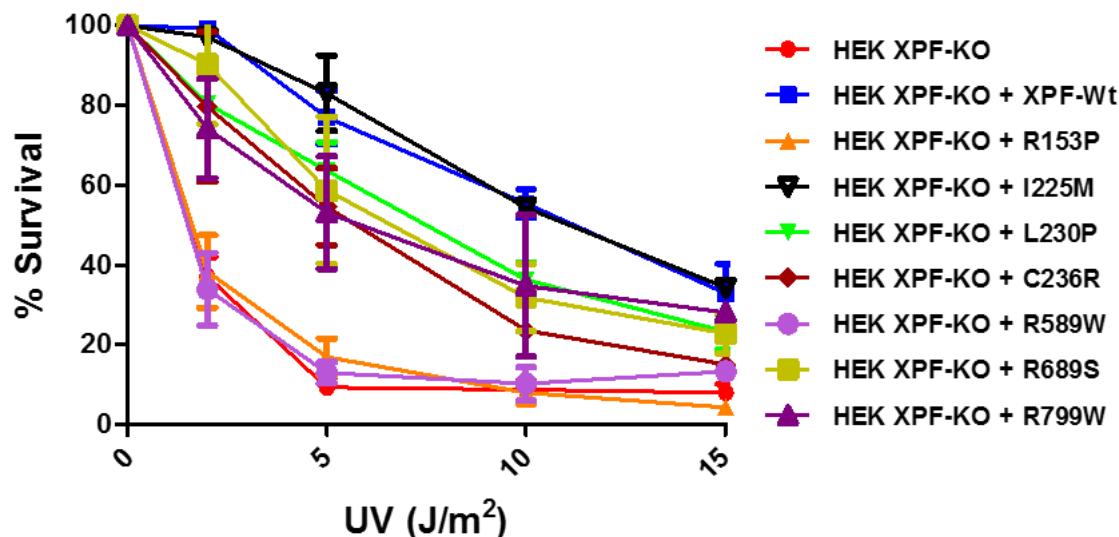
All XPF variants were detectable by WB and some of them had levels of expression similar to XPF-Wt ( $\text{XPF}^{\text{L225M}}$  and  $\text{XPF}^{\text{R799W}}$ ).  $\text{XPF}^{\text{R153P}}$ ,  $\text{XPF}^{\text{L230P}}$  and  $\text{XPF}^{\text{R689S}}$  showed increased levels of protein production compared to the XPF-Wt. This was in contrast to the reduced levels observed in patients (Bogliolo et al. 2013; Mori et al. 2018; Niedernhofer et al. 2006; Sijbers et al. 1998). The increased levels detected in vitro ensure that the cellular phenotype-genotype correlation is not due to a matter of protein quantity, but depends on the single mutation that alters the essential protein in a specific way.  $\text{XPF}^{\text{C236R}}$  and  $\text{XPF}^{\text{R589W}}$ , however, showed decreased levels relative to the Wt-XPF even in vitro, which could further explain the cellular phenotypes of these mutations.

### IV.2.3 FUNCTIONAL ANALYSES OF XPF VARIANTS

#### IV.2.3.1 NER PATHWAY

Several assays were performed to test and compare the behavior of the pathogenic XPF variants after UVC irradiation in an isogenic background: UV sensitivity, UDS and RRS.

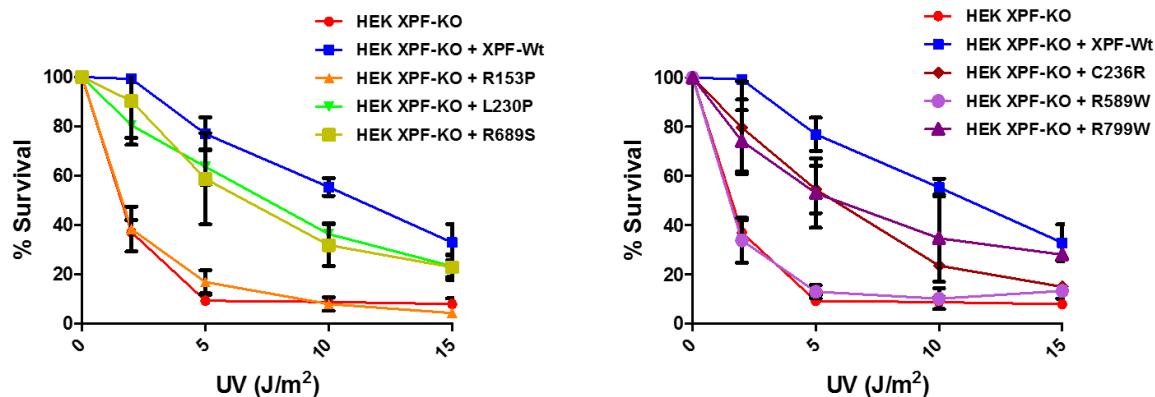
First of all we tested the sensitivity of our cell lines to UVC irradiation: figure 35 shows the percentage of surviving cells of each cell line after different doses of UVC radiation.



**Figure 35: Percentage of surviving cells after increasing doses of UV irradiation.** Graph represents the media of at least two independent experiments with two replicates and the standard error of the mean (SEM).

All the variants showed increased levels of sensitivity in comparison with the XPF-Wt with the exception of XPF<sup>I225M</sup>, that was in contrast with previous studies (Matsumura et al. 1998). Matsumura group found mRNA expression of this variant was similar to Wt fibroblasts but they were unable to detect the mutant protein by WB, suggesting a rapid degradation of the mutant variant. In contrast, our XPF<sup>I225M</sup> levels were comparable to the Wt as shown in figure 35, which could justify this unexpected resistance. Considering that the clinical phenotype associated to this mutant could be a matter of protein level production that was not corresponding to our XPF<sup>I225M</sup> protein level, this variant was eliminated from all the successive analyses. For a better understanding, the data has been split into two different graphs, the first one including controls and XPF variants R153P (XFE), L230P (FA) and

R689S (FA); and a second graph with controls and XPF variants C236R (CS), R589W (CS/XP) and R799W (XP and XFE/CS) (Figure 36).



**Figure 36: Percentage of surviving cells after increasing doses of UV irradiation.** The previous data has been split in two different graphs for a better understanding.

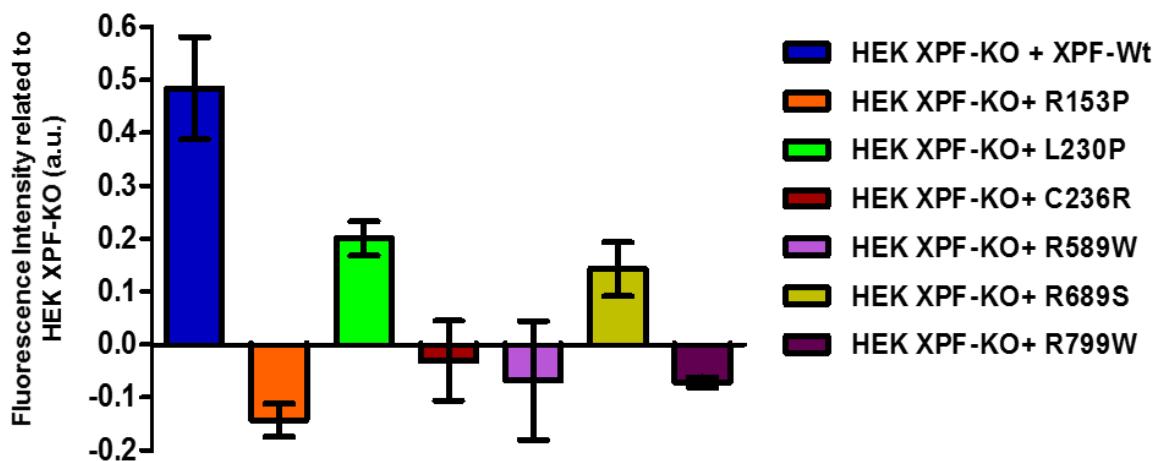
Cells producing XPF<sup>R153P</sup> and XPF<sup>R589W</sup> are very sensitive to UV, at similar levels to the XPF-KO. These results are in concordance with the phenotype of the patient in which these mutations were identified, as XPF<sup>R153P</sup> was found in homozygosity in an XFE progeroid patient that showed skin photosensitivity (Niedernhofer et al. 2006), and XPF<sup>R589W</sup> was found in XP patients (Gregg et al. 2011) and in an XPCSCD patient providing the mutant allele that confers the XP phenotype (Kashiyama et al. 2013). XPF<sup>R153P</sup> results for UV sensitivity were essentially identical to the previously reported studies that identified it as 10x more sensitive to UV (Niedernhofer et al. 2006). XP patients main feature is the extreme sensitivity to UV irradiation, although there are different levels of sensitivity from the proteins involved in this disease (Matsumura et al. 1998). Unexpectedly, XPF<sup>R589W</sup> sensitivity was higher than the previously reported for the XP condition that presented 2-3x more sensitive than XPF-Wt (Gregg et al. 2011), pointing the importance of the residual activity of the second mutant allele Pro379Ser (P379S) in NER. XPF<sup>R799W</sup> was initially found in homozygosity in an XP patient (Sijbers et al. 1998) but it was recently reclassified as a CS mutant with XFE features when the second allele had an early truncating mutation (Mori et al. 2018). In line with these last results, XPF<sup>R799W</sup> just showed a mild sensitivity to UVC very similar to the sensitivity of

the Cockayne associated XPF<sup>C236R</sup> variant. This result also provides an explanation to the mild XP phenotype of the double mutant R589W-R799W (Kashiyama et al. 2013) and indicates that XPF<sup>R799W</sup> retains the majority of its functions in UVC induced DNA repair. Another hypothesis that could explain the behavior of R799W variant could be a gene dosage effect: in homozygosity it would be associated to mild XP (Sijbers et al. 1998) while in heterozygosity with a null allele it would be associated to atypical XFE disease sharing features of CS (Mori et al. 2018). In heterozygosity with R589W, it would be responsible for the patient's CS features while the R589W would be the cause of the XP phenotype also present in the patient (Fassihi et al. 2016). This last hypothesis highlights the importance of the genetic background of the patients in the modulation of the phenotype.

The two XPF variants found in FA patients, XPF<sup>L230P</sup> and XPF<sup>R689S</sup> showed a marked resistance to UVC compared to the defective XPF. These results confirmed previous studies which proved that these two mutations do not seem to completely impair XPF to participate in NER pathway (Bogliolo et al. 2013).

Regarding XPF<sup>C236R</sup>, previously associated with CS, showed mild UVC sensitivity, in accordance with the low UVC sensitivities of CS patients derived cells (Kashiyama et al. 2013).

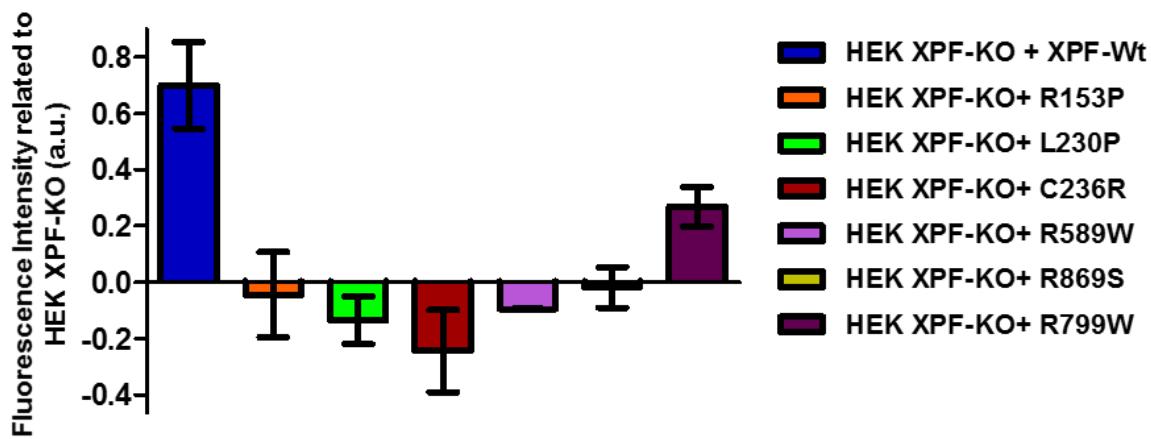
To further discern among the XPF variants whose mutation impairs NER pathway, functional analysis to individually evaluate GG-NER and the TC-NER were performed with the whole set of variants. Figure 37 shows UDS assay data representing the ability of each XPF variant cell line to synthesize DNA to repair a damage induced by UVC in the G1 phase of the cell cycle.



**Figure 37: UDS assay of the of XPF mutants set.** Fluorescence intensity is represented relative to fluorescence intensity of HEK XPF-KO. Graph represents the media of at least three independent experiments with SD.

These assay results allowed a clear distinction from the XPF mutations that conferred UV sensitivity. Despite all the variants showed a certain grade of impairment in UDS, the two FA associated mutations,  $XPF^{L230P}$  and  $XPF^{R689S}$  retained around 30-40% of their UDS abilities when compared to the wt-corrected cells.  $XPF^{R153P}$  showed the lowest UDS value and the CS and the XP associated mutations,  $XPF^{C236R}$ ,  $XPF^{R589W}$  and  $XPF^{R799W}$ , showed UDS levels similar to the XPF-KO. These observations are in line with the UDS levels of the patients-derived cells bearing these mutations (Ahmad et al. 2010; Gregg et al. 2011; Kashiyama et al. 2013; Mori et al. 2018). These results demonstrate that XFE, XP or CS associated mutations impair GG-NER much more then FA related mutations.

The TC-NER subpathway activity of the set of XPF mutants was evaluated by a RRS assay after UV induced damage. The graph from figure 38 represents the ability of each cell line to synthesize RNA after the damage in comparison with the XPF-KO cells.



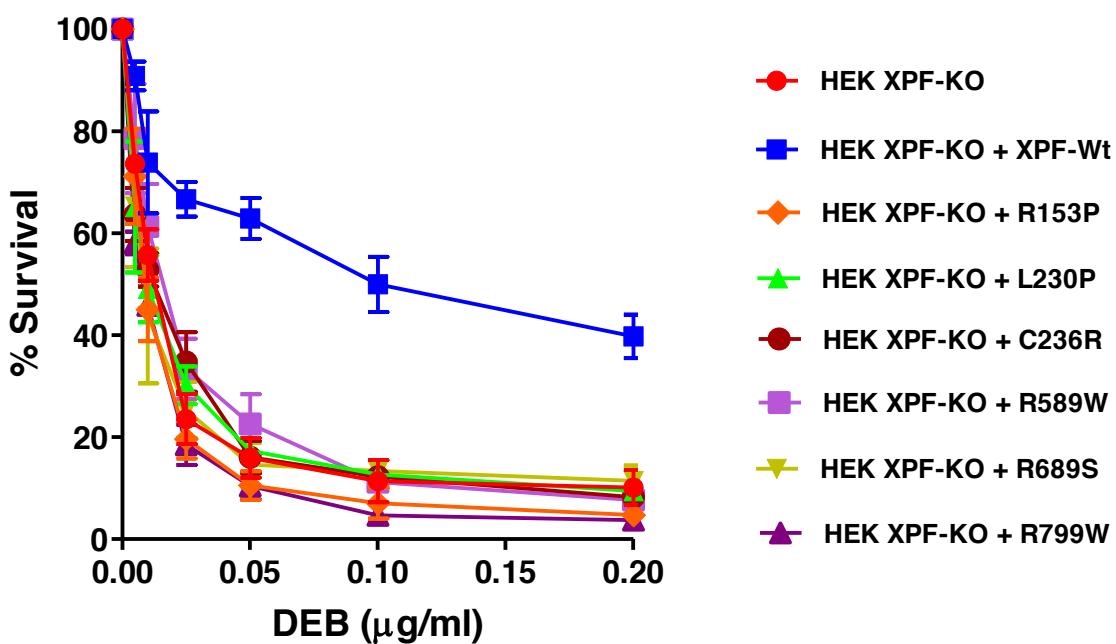
**Figure 38: RRS assay of XPF mutants set.** Fluorescence intensity is represented relative to fluorescence intensity of HEK XPF-KO. Graph represents the media of at least three independent experiments with SD.

The mutation  $\text{XPF}^{\text{R}153\text{P}}$  had low levels of RRS as expected by its inability to reach the nucleus and the mutation associated to CS,  $\text{XPF}^{\text{C}236\text{R}}$ , showed the strongest impairment in concordance with literature (Kashiyama et al. 2013); notwithstanding  $\text{XPF}^{\text{L}230\text{P}}$  and  $\text{XPF}^{\text{R}689\text{S}}$  involved in FA, have a high impact in the RRS after UV-induced damage. The XP associated mutation  $\text{XPF}^{\text{R}799\text{W}}$  is the only one from the studied variants with a residual ability to synthesize RNA after the damage, which drives to reconsider this assay as a useful tool to discern mutations associated with typical CS pathology rather than XP. Actually, it has been recently put up for debate if RRS levels should be used as a determinant feature for CS classification regarding the phenotype variability of the patients which has driven to the enlargement of diagnosis criteria focusing in their clinical features instead (Wilson et al. 2016). Consequently, with our results, RRS assay would not be able neither to provide conclusive data for pathological XPF variants associated with FA. Interestingly, some of the FA proteins such as BRCA1, FANCD1, FANCD2, FANCA and FANCM have been reported to be involved in resolving RNA-DNA ICLs, which are known as R-loops (Garcia-Rubio et al. 2015; Lafuente-Barquero et al. 2017). These loops are produced when the stalled replication fork forms a hybrid with the transcriptional nascent mRNA. According to the low RRS levels showed by  $\text{XPF}^{\text{L}230\text{P}}$  and  $\text{XPF}^{\text{R}689\text{S}}$ , one hypothesis could be that XPF was also

one of the FA proteins involved in the removal of these R-loops, therefore XPF mutations could affect its ability to participate in RNA-DNA ICLs repair, resulting in TCR impairment.

#### IV.2.3.2 ICLR PATHWAY

The repair of the two covalently linked strands of DNA caused by products of the metabolism or external agents is crucial for the survival of cells. The role of XPF as a 5' endonuclease involved in the excision of this damage is largely identified. To test the phenotypical behavior of the XPF variants in the repair of DNA crosslinks, a DEB survival test with increasing doses was performed (Figure 39).



**Figure 39: Percentage of surviving cells under increasing doses of DEB.** Graph represents the media of at least two independent experiments with two replicates and the SEM.

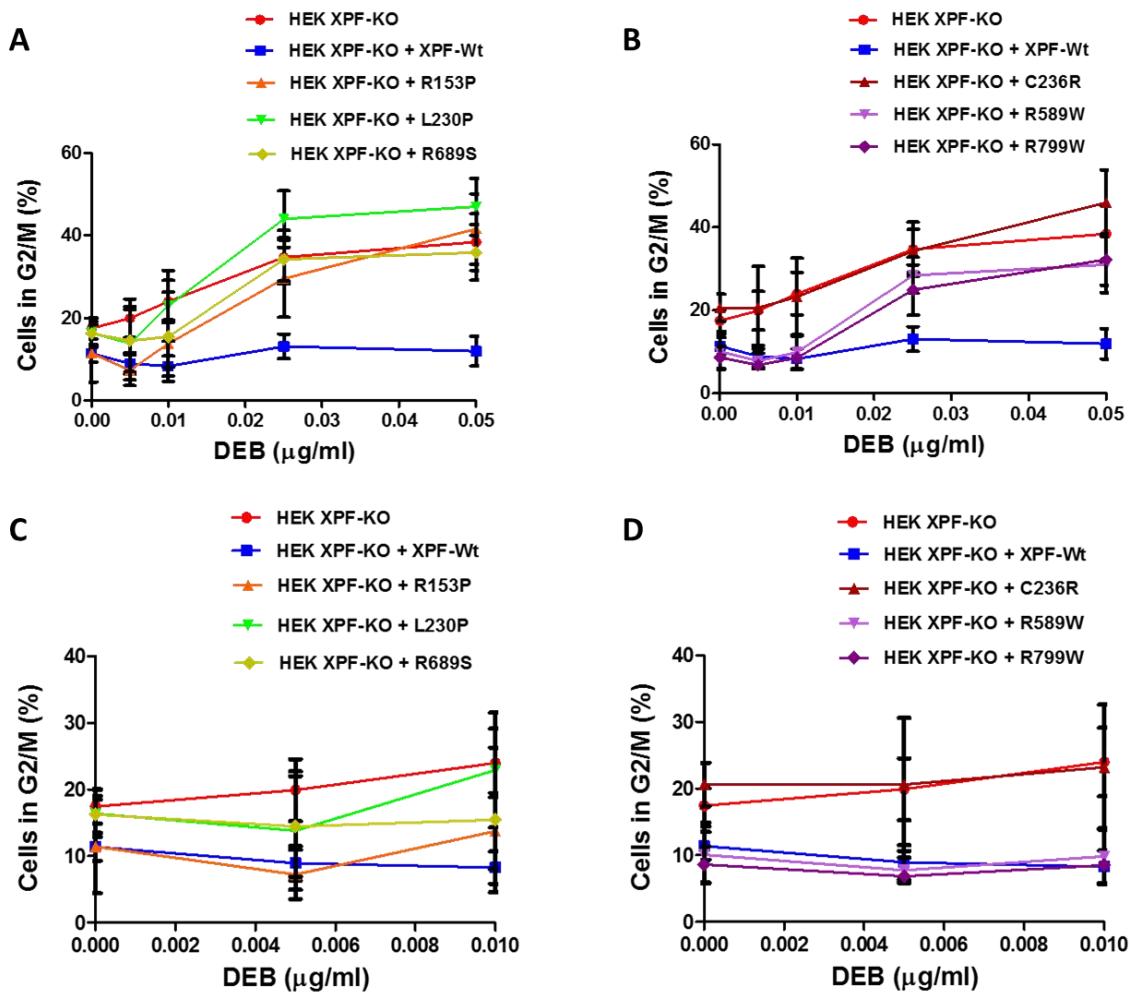
All the XPF variants cells showed a marked ICL sensitivity, resembling cells lacking XPF. XFE Progeria associated variant XPF<sup>R153P</sup> was already reported to impair ICL

(Niedernhofer et al. 2006) due to its improper folding and its inability to reach the nucleus. XPF<sup>R799W</sup> showed a strong sensitivity to DEB, more significant than the around 60% sensitivity to MMC that was considered to classify it as an XFE mutation (Mori et al. 2018). The two FA associated mutants XPF<sup>L230P</sup> and XPF<sup>R689S</sup> which were previously reported to impair ICL (Bogliolo et al. 2013) were also very sensitive as the CS associated variant XPF<sup>C236R</sup>, which was additionally reported to impair ICL (Klein Douwel et al. 2017).

The only XPF variant whose ICLR could be expected to be less affected was XPF<sup>R589W</sup>, as it has been classified as an XP or XPCSCD variant, but the DEB sensitivity assay did not point towards this hypothesis. One hypothesis to explain this marked DEB sensitivity could be that crosslinks inducers are also responsible for generating DNA mono-adducts, which are repaired by NER (Grant et al. 1998) and therefore XPF mutations affecting NER would be also impairing ICLR. Additionally, it is thought that during TLS in FA pathway, the bypassed mono-adduct is repaired by NER, thereby mutations affecting NER would be also impairing ICLR.

#### **IV.2.3.2.1 ICL- INDUCED G2/M ARREST**

The accumulation of chromosome alterations caused by ICL agents exposure is associated with a delay of the cell cycle to come into mitosis phase. A DEB-induced G2/M cell cycle arrest assay was performed to test if the different XPF variants could prevent the cells from being arrested in G2/M phase of the cell cycle due to the DEB-induced chromosome alterations. For a better understanding of the results, data have been plotted in two different graphs sharing the same control cell lines and the same cell lines have been then plotted in two different graphs showing a more accurate view of low DEB doses results (Figure 40).



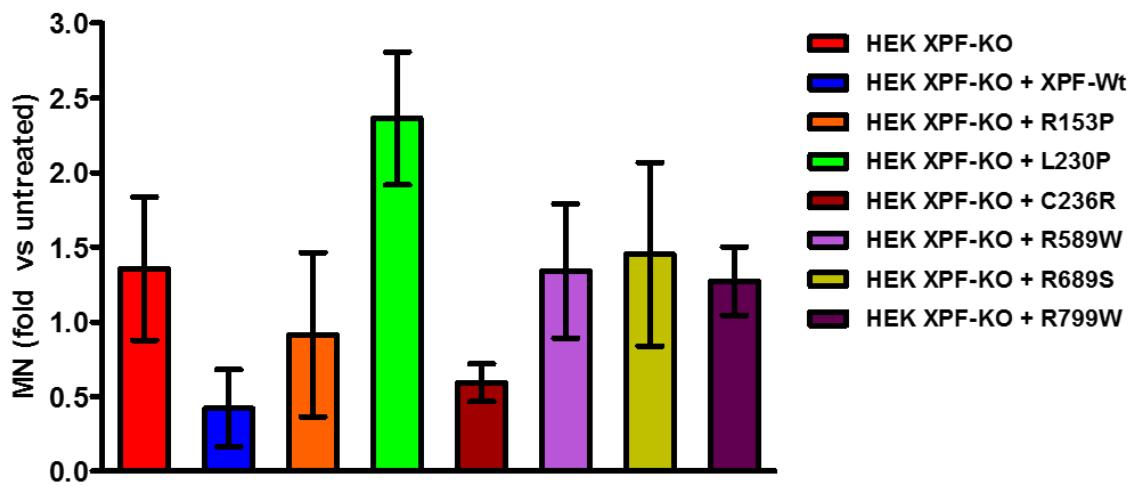
**Figure 40: Percentage of cells stalled in G2/M phase after DEB exposition.** A and B: Half of the XPF variants are represented in each graph with the positive and negative controls. Each graph represents the media of at least two independent experiments with the SEM. C and D: Plots of low DEB doses are represented here for a better understanding. Each graph represents the media of at least two independent experiments with the SEM.

All the XPF variants showed a higher percentage of G2/M arrested cells than the XPF-Wt. The two mutations associated with FA disease, XPF<sup>L230P</sup> and XPF<sup>R689S</sup> followed a similar pattern and, in concordance with previous studies (Osorio et al. 2013) and the DEB survival assay data, display about the same percentage of arrested cells than the negative control. Additionally, progeria associated mutation XPF<sup>R153P</sup> and CS associated mutation XPF<sup>C236R</sup>

levels of arrestment were as well very high. This could be easily understood for the XFE mutation but it was less clear for the CS mutation, although the criteria to identify a CS variant covers now a wider range of phenotypes (Wilson et al. 2016) including ICL sensitivity. Our original data show that the G2/M block after ICL treatment could be another useful parameter to be considered in the diagnosis of CS. Regarding the remaining variants, XPFR<sup>589W</sup> and XPFR<sup>799W</sup>, showed a very similar phenotype, less severe than the rest of the mutants but still showing large number of arrested cells. Indeed, at low doses of DEB these two cell lines were behaving as the XPF-Wt (Figure 40), which could be explained because R589W is reported to be an XP associated disease, and by hence its main affected DNA repair pathway could be NER instead of ICLR; and R799W, associated to XP and to XFE with CS features, could have a minor impact on ICLR pathway and the cell sensitivity to DEB could be more related (and similar to XPFR<sup>589W</sup>) to the repair of DEB mono-adducts than to the ICLs.

#### **IV.2.3.2.2 ANALYSIS OF ICL INDUCED CHROMOSOME FRAGILITY**

Crosslinker agents produce in ICLR deficient cells chromosome breaks that can be left behind in anaphase and produce MN in the daughter cells. The MN-Fragility test (Avlasevich et al. 2006) provided useful information about the levels of chromosome breaks measured by counting the number of MN. MN formed after treating the set of our XPF variants with DEB is shown in the figure 41.



**Figure 41: MN test after DEB (0.01 µg/mL) exposure of all XPF variants.** Data is represented in fold changed vs untreated cells. Graph represents the media of at least three independent experiments with SD.

XPF<sup>L230P</sup> (FA) showed the highest number of MN after treatment, in concordance with the chromosome fragility phenotype of the FA patient from which it was identified (Bogliolo et al. 2013). The other FA associated mutation, XPF<sup>R689S</sup> showed similar levels of MN to the XPF-KO cell line. XPF<sup>R589W</sup> (XP), and XPF<sup>R799W</sup> (XP, XFE/CS) variants also showed fragility levels resembling the ones of the XPF-KO cell line. Interestingly, R589W and R799W mutations showed high levels of MN even if the percentages of G2/M arrested cells were moderate, indicating that most of the cells that achieved to reach the mitosis phase accumulated many chromosomal breaks. XPF<sup>C236R</sup>, CS associated variant, does not show chromosome fragility under DEB exposure, meaning that DEB causes less chromosome alterations probably because more DNA damage is successfully resolved during the prolonged G2/M block (see figure 39) and the percentage of cells reaching telophase do not present so many chromosome breaks. In contrast, the high number of MN found in XPF<sup>L230P</sup> (and in the other cell lines) suggests that a higher percentage of cells are unable to repair the DNA damage during the G2/M block and that the cells that escape the block carry a heavy burden of breaks. The SD of XPF<sup>R153P</sup> sample is too large to reach any definitive conclusion.

In the case of R799W and R589W at these doses of DEB the G2/M block is similar to the wt indicating that the G2/M check point in these cells could not be activated. This would explain the increased levels of DNA damage that is not repaired that is found in the daughter cells.

#### **IV.3- OVERVIEW OF THE ANALYZED XPF VARIANTS CELLULAR PHENOTYPES**

This has been the first time that the function of several XPF variants associated to distinct clinical phenotypes has been compared in isogenic human cell lines *in vivo*. This enables to analyze if the variability of the phenotypes is correlated with the specific nucleotide changes or it is influenced by other factors such as the genetic background, the ability of the protein to fold properly and reach the nucleus or the possible interactions with other proteins.

At first glance, it is remarkable how the position of the aa substitution along XPF sequence does not define the disease of the patient as it was illustrated in figure 33.

As previously explained, mutation I225M did not show a pathological cellular phenotype and has not been considered in these analyses.

Four variations were confined in the helicase-like domain and two in the nuclease domain but the patients' phenotypes were not determined by these positions. Notwithstanding, none of the mutations were affecting the C-terminal HhH2 domain from which XPF binds to ERCC1 to conform the stable endonuclease heterodimer and mutations in that domain have not been identified in patients to date (McNeil and Melton 2012), since this interaction is essential for XPF functions.

The interaction between XPF and SLX4 is vital for the recruitment of XPF to ICL sites and the activation of the incision. The XPF region involved in SLX4 binding comprises most of the helicase-like domain, from aa 12 to 650. Four of the selected mutations were confined in this section: R153P, L230P, C236R and R589W. Hashimoto and colleagues performed a yeast two-hybrid assay to unravel whether these mutations were preventing SLX4 binding

and found out that R153P, L230P and R589W were abolishing XPF binding ability to SLX4 while C236R was not. Consistently, they also examined if these mutants were disrupting the ERCC1 binding even if their location was not affecting ERCC1 binding site and found that the same three XPF mutations were affecting this interaction, pointing to the magnitude of the aa changes for the viability of XPF (K. Hashimoto et al. 2015). Despite this interpretation relies too heavily in the amount of target proteins obtained by the yeast two-hybrid assay, which is usually poor and could overestimate the significance of the variants, this mapping provides some hints for the phenotype interpretation diminishing the importance of the location of the aa change along the sequence and moreover highlights the importance of further similar studies performed in human cell lines.

In parallel, recent studies have also focused in the impact of generating an XPF-KO human cell line via CRISPR edition and have brought to light the importance of the XPF-ERCC1 heterodimer transference from the cytoplasm to the nucleus (J. Lehmann et al. 2017). Cells lacking XPF retained ERCC1 in the cytoplasm. These studies endorse the previous observations of some XPF mutations phenotypes that were unable to reach the nucleus due to an improper protein folding, such as XPF<sup>R153P</sup>, XPF<sup>R799W</sup> (Ahmad et al. 2010) and XPF<sup>C236R</sup> (K. Hashimoto et al. 2015). In this sense, a more comprehensive study of the heterodimer localization should include all the XPF mutations selected in this project.

Table XX summarizes the cellular phenotype of our variants:

Nucleotide change	XPF mutation	Patient disease	UV sensitivity	UDS	RRS	DEB sensitivity	G2/M	MN
c.458G>C	R153P	XFE	Red	Red	Light Red	Red	Red	Not conclusive
c.2395C>T	R799W	XP/XFE?/CS?	White	Red	White	Red	Light Red	Red
c.1765C>T	R589W	XP/ XPCSCD	Red	Red	Red	Red	Light Red	Red
c.706T>C	C236R	CS	Light Red	Light Red	Red	Red	Red	White
c.689T>C	L230P	FA	White	White	Red	Red	Red	Red
c.2065C>A	R689S	FA	White	White	Light Red	Red	Red	Red

1	2	3	4	5
White	Light Red	Light Red	Red	Red

**Table XX: Summary table of the whole set of XPF variants and their cellular phenotypes.** For each *ERCC4* nucleotide change, the XPF mutation and the pathology associated found in the patient where it was identified are described. The results of the functional assays performed with cells lacking XPF containing the variants are shown in a red scale from 1 (less affected) to 5 (more affected).

Regarding the analysis of XPF activity in the NER pathway, the UV sensitivity survival provided reliable information to discern mutations more associated to ICLR impairment such as L230P and R689S, which showed marked resistance to UV and considerable levels of UDS. In contrast, XFE associated R153P and XP mutation R589W were clearly disrupting XPF role in NER and ICLR. An interesting finding was the atypical behavior of XPF<sup>R799W</sup>, who did not show sensitivity to UV. This variant was found in homozygosis in XP patients (Sijbers et al. 1998) and thus a defined phenotype of NER disruption was expected. This variant was also identified in heterozygosis in a patient presenting XFE with CS features thus enforcing the expectations of high levels of UV sensitivity such as the R153P causing XFE. Our results support the hypothesis that the phenotypes associated with XPF mutations are related to other elements of the cellular context like genetic background and/or interacting proteins. The third assay executed to evaluate the role of XPF in NER, the RRS assay, did not contribute significantly to discern among the XPF phenotypes but indicated that the CS associated mutation XPF<sup>C236R</sup> was the most abruptly affected as expected by its

inability to participate in TCR of NER (Kashiyama et al. 2013). Besides, the unexpected RRS low levels of FA associated XPF variants enable the possibility for the first time of considering XPF as another FA protein involved in the resolving of ICLs in RNA-DNA hybrids (R-loops). R-loops are present in telomeres and contribute to telomere maintenance (Garcia-Rubio et al. 2015; Lafuente-Barquero et al. 2017). XPF plays a role in telomere maintenance; it is known to be involved in the excision of T-loops (Zhu et al. 2003) and it is responsible for telomeres loss when TRF2 is overexpressed (P. Munoz et al. 2005) thus contributing to ageing and genome instability. Although additional studies should be performed to assess the role of XPF in R-loops resolving, RRS assay results points towards its involvement.

A limitation for the NER assays is the cell line used in our studies; HEK 293T are a good human cell line for genome editing and thus were chosen for this study but, on the other hand, are tricky to work with for NER evaluating assays since they weakly adhere to the substrate.

Concerning ICLR analysis, there is a more marked sensitivity in FA associated variants  $XPF^{L230P}$  and  $XPF^{R689S}$  as reported (Bogliolo et al. 2013) and in the XFE related variants  $XPF^{R153P}$ ,  $XPF^{R799W}$  (Mori et al. 2018; Niedernhofer et al. 2006). Slightly less sensitive are the variants associated with CS features as  $XPF^{C236R}$  and  $XPF^{R589W}$  (XP).

A very recent study performed by Popp and colleagues classifies for the first time R589W as an FA mutation (Popp et al. 2018). The patient from which it was identified was presented as an FA clinical phenotype, however, bone marrow failure, one of the crucial features to be classified as FA, was absent. Despite the atypical FA clinical phenotype, it was the first FA patient to develop skin photosensitivity. The second allele of *ERCC4* presented a novel splice site mutation (c.793-2A>G) which produced a premature termination of translation (p.Thr265Valfs\*13), hence originating a null allele as happened with the previously reported FA XPF variants (Bogliolo et al. 2013). Contrary to Hashimoto group, who found out R589W mutation was abruptly affecting XPF structure and its SLX4 binding (K. Hashimoto et al. 2015) Popp and colleagues detected a residual proportion of  $XPF^{R589W}$  escaping from protein misfolding, able to reach the chromatin (Popp et al. 2018)

as happened with the reported FA XPF variants. Interestingly and in concordance to our results this mutant had reduced UVC irradiation resistance, UDS and RRS levels, although contrary to our results, XPF<sup>R589W</sup> showed higher UVC irradiation resistance levels than XPF<sup>L230P</sup> (Popp et al. 2018). This combination of findings provides some support for the conceptual premise that there is a link between ICL and NER pathways, reinforce our findings of XP associated variants presenting ICL impairment and endorse the theory that mutations found in specific locations of the sequence should not be associated with discrete DNA repair pathways impairment.

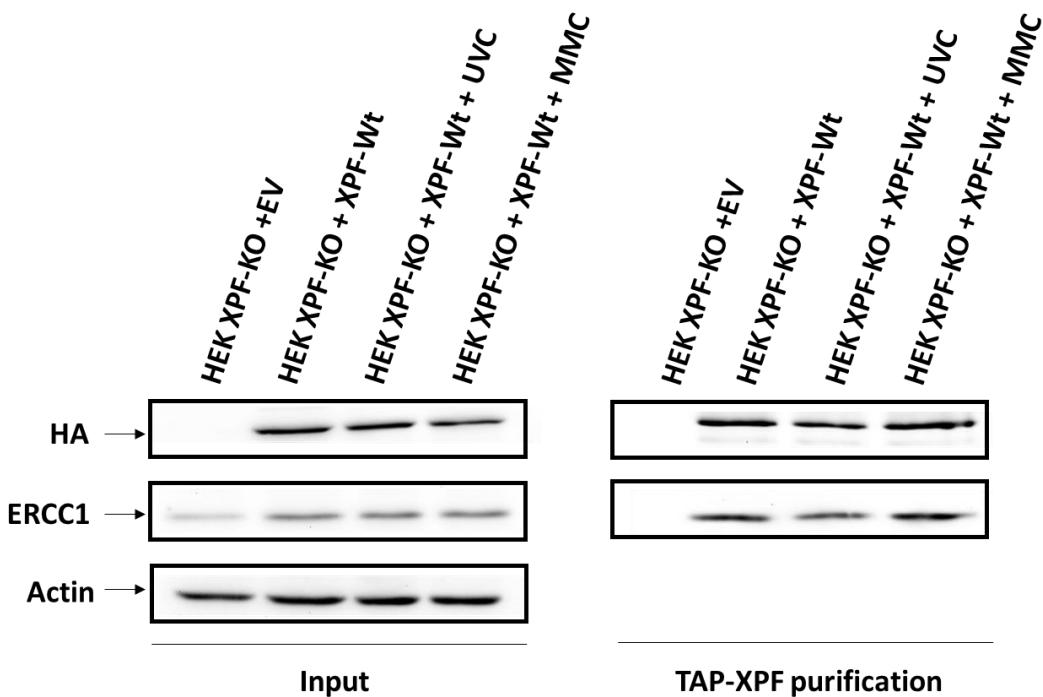
While the G2/M arrest assay evaluates the inability of the cells to reach mitosis due to the chromosome aberrations, the MN-Fragility test focuses in the breaks that chromosomes experience during anaphase and are unable to progress to the daughter cells. For instance, FA associated variants XPF<sup>L230P</sup> and XPF<sup>R689S</sup> presented high percentage of G2/M arrested cells and high number of MN, indicating the marked ICLR impairment of these variants, while XPF<sup>R589W</sup> and XPF<sup>R799W</sup> showed a mild arrest but a high number of MN, what suggests that in the cells expressing XP associated variants the G2/M checkpoint is not activated by the DNA damage and the cells can relatively easily reach mitosis with a damaged DNA.

The message that stands out from our studies after analyzing a set of XPF variants in a homogenous cellular and genetic background is that despite the cellular functional analyses sometimes provide promising suggestions to understand the genotype, a defined cellular phenotype cannot be correlated to each XPF mutation; functional analysis might help to correlate the genotypes to the cellular phenotypes but definitive statements about the contribution of XPF variants to the phenotype must take in account other factors such as the level of XPF, cell localization, possible interactors that can affect its role in DNA damage repair and the different genetic background of the different patients that could modulate the genotype-phenotype correlations.

## PROJECT 2

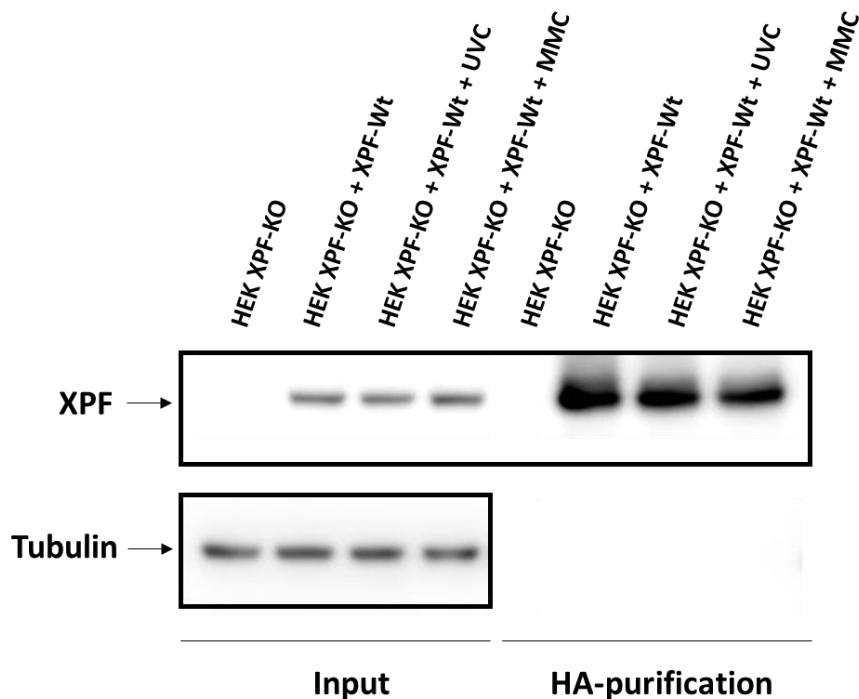
### **IV.4- PROTEOMIC STUDY OF XPF INTERACTOME**

TAP were performed in order to identify potential XPF interactors that could have a relevant role in DNA repair. The Streptavidin Calmodulin tag placed in the N-terminal site of XPF was used for a double step of XPF purification. HEK XPF-KO cells infected with the tagged XPF or with an EV were grown untreated, treated with MMC or UVC irradiation to obtain protein candidates able to interact with XPF when it is actively participating in ICL or NER pathways. A WB confirming the affinity purification is shown in figure 42.



**Figure 42: WB validation of the XPF purification through the N-terminal tags.** One third of the Streptavidin-Calmodulin XPF purification was tested by WB to confirm XPF presence through HA tag and ERCC1 interaction in transduced HEK XPF-KO under different treatments: UVC (150J/m<sup>2</sup>, detached after 1h) or MMC (1 μM for 16h). Actin was used as a loading control detectable in the input but not in the purification fraction.

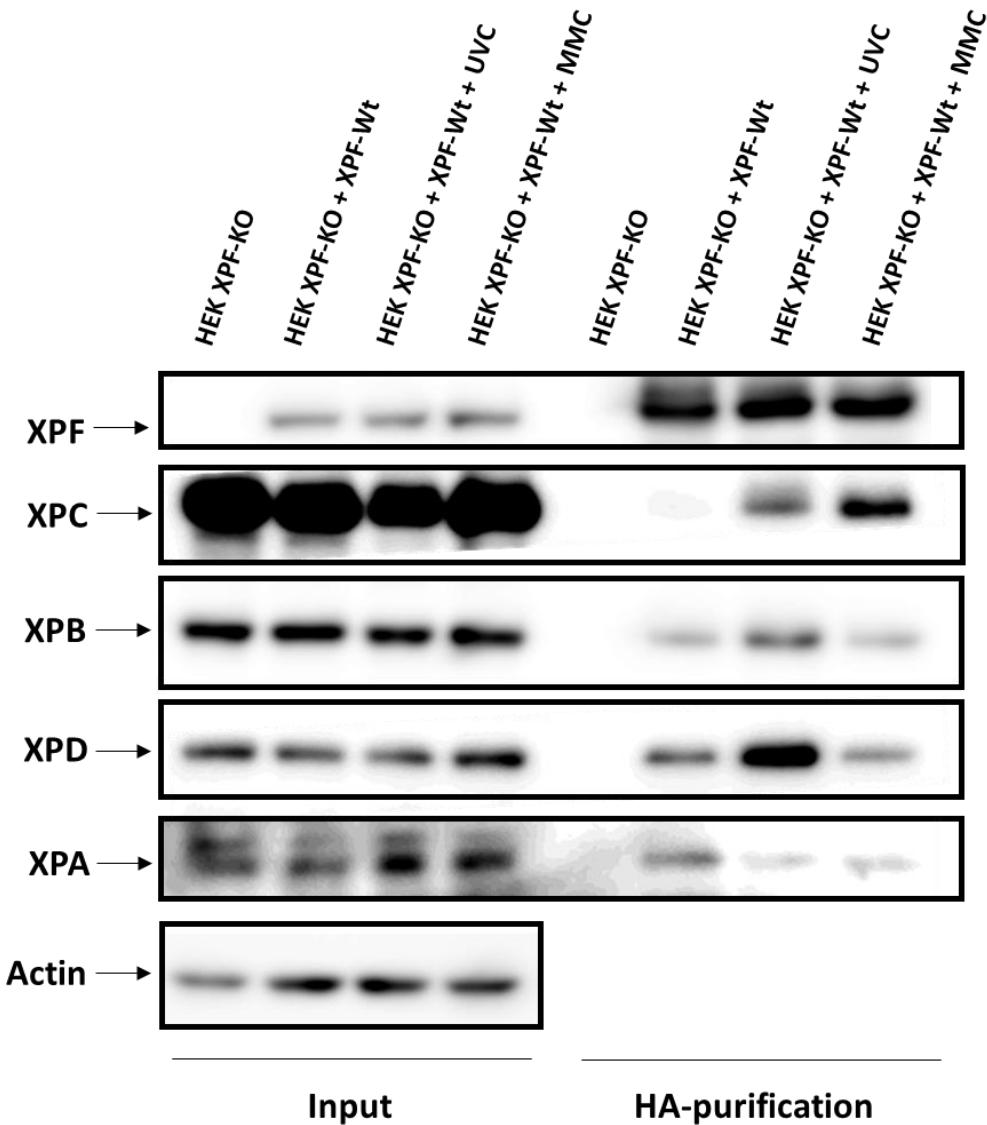
Proteins purified through the N-terminal tag under different conditions that were identified by mass spectrometry are shown in the annex (Annex Table I). Some proteins already known to interact with XPF in DNA repair pathways in which XPF takes part appeared, such as ERCC1, SLX4, XPG and XPA but it was not possible to further validate by WB the XPF interaction with new candidates (hnRNPU, ZC3H11A, CCDC81A, PPM1A) found by mass spectrometry, thereby a different proteomic approach based in XPF purification through the C-terminal HA-tag was taken on. This approach consisted in the immunoprecipitation of the HA-tag achieved by a magnetic bead surface coated with anti-HA antibody and allowed to pool down XPF. The WB confirmation of the XPF-HA purification is shown in figure 43.



**Figure 43: WB validation of the XPF purification through the C-terminal tag.** 5% total of purification volume was used to confirm by WB the XPF purification through the HA-tag in transduced HEK XPF-KO under different treatments: UVC ( $50\text{J/m}^2$ , detached after 1h) or MMC ( $1.5\ \mu\text{M}$  for 16h). Tubulin was used as a loading control detectable in the input but not in the purification fraction.

XPF-HA purifications ran in gels and stained with Coomassie Blue Staining to identify and cut single bands from each sample lane were processed by mass spectrometry. A list of protein candidates is shown in annex (Annex Table II).

CoIP to check proteins found by mass spectrometry involved in DNA repair were performed as shown in figure 44.



**Figure 44: CoIP of XPF and some of the interactors found by mass spectrometry.** The detection of XPC, XPB, XPD and XPA confirmed the mass spectrometry results from the HA-XPF purification in the HEK

XPF-KO transduced cell line under different treatments (see figure 43). XPF was used as a precipitation control. XPC interaction seems to be increased under both treatments. XPB and XPD interactions seems to increase under UVC treatment. Actin was used as a loading control detectable in the input but not in the purification fraction.

XPA, XPB, XPC and XPD are proteins involved in NER (Marteijn et al. 2014) and therefore unsurprising to appear as XPF candidate interactors although those interactions could be indirect. XPA is known to interact with XPF-ERCC1 heterodimer through ERCC1 (Su et al. 2012) and XPB, XPC and XPD appear most likely because the indirect cooperation they establish in NER cascade.

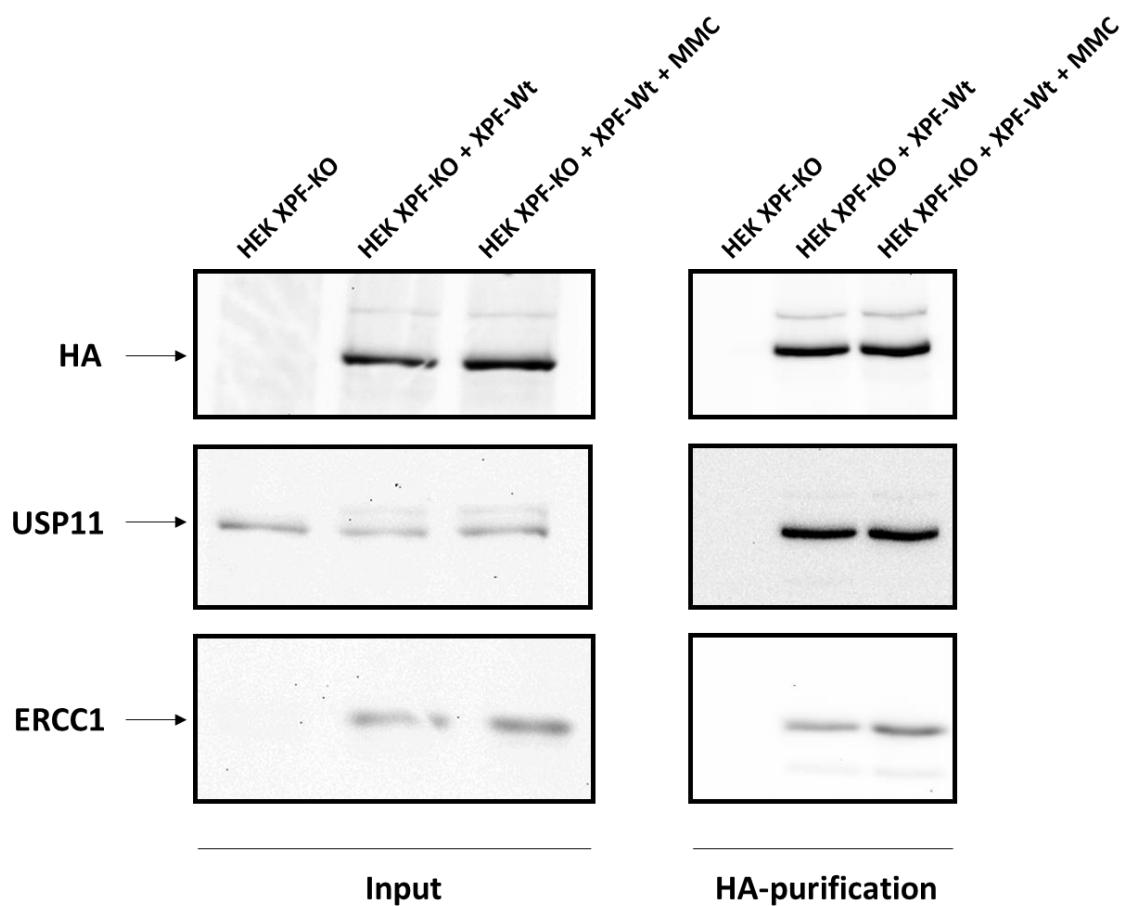
One unanticipated finding was that the mass spectrometry analysis of the two methods of XPF affinity purification (through TAP or HA XPF-tags) did not unravel the same protein candidates. A possible explanation for this apparent inconsistency might be that the different tags by themselves are interacting with specific proteins or covering up possible XPF interactions. Additionally, the UV irradiation dose was not the same for the two purifications; for the TAP-XPF purification the UV dose was 150 J/m<sup>2</sup>, which was then reduced to 50 J/m<sup>2</sup> in the HA-XPF purification assay in case the irradiation was too high and some interactions were impaired. Furthermore, mass spectrometry is a highly delicate technique, able to detect many false positive contamination proteins which could also mask real XPF interactors. The fact that the two mass spectrometry batches were performed in different spectrometer services might contribute as well to the variability, although it may strengthen the validity of the proteins found by both mass spectrometry assays.

Unfortunately, the CoIP performed did not confirm any new interaction of XPF with the candidates (XRCC5/Ku80 and XRCC6/Ku70) found by mass spectrometry of single bands gel digestion. Consequently, another proteomic approach was implemented based on stable isotopes labeling, SILAC (Baple et al. 2014), to get a quantitative and more accurate output of the protein candidates to interact with XPF. HEK XPF-KO transduced with XFP-Wt were cultured under UVC (40J/m<sup>2</sup>, detached after 1h) or MMC (0.5 µg/mL for 21h) treatments with different isotopes of lysine and arginine in order to obtain quantitative data of the possible

XPF interactors under different conditions. A list of the mass spectrometry identified proteins is shown in annex (Annex Table III) (Annex Table IV).

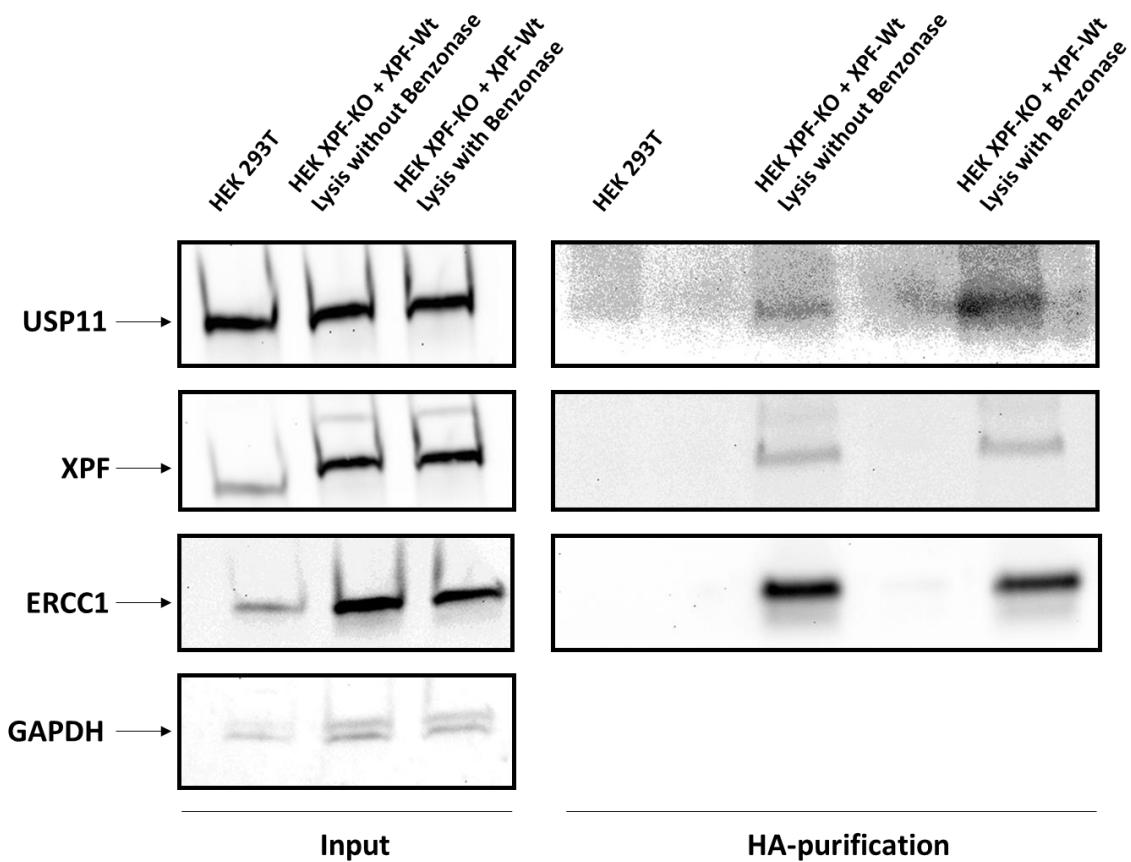
Taking into account XPF involvement in DNA repair, we focused on proteins from the obtained data which could have a role in DNA repair and identified USP11 protein as an XPF candidate interactor. *USP11* is located in chromosome X p11.3 and turns into USP11, a deubiquitinase that removes conjugated ubiquitin from target protein and polyubiquitin chains and inhibits proteasome degradation of target proteins (Ideguchi et al. 2002). It participates in DNA repair regulation after DSB (Wiltshire et al. 2010); it is thought to deubiquitilate BRCA2 (Schoenfeld et al. 2004) although this interaction *in vivo* is not totally understood. Besides, USP11 deubiquitilates and stabilizes p53 (Ke et al. 2014) and interacts with other deubiquitylases (DUBs) like USP7, with the transcriptional elongation factors TCEAL1 and TCEAL4 and with the NRF2 regulatory protein KEAP1 (Schoenfeld et al. 2004). All these data support the idea USP11 participates in DNA repair. Recently, it has been reported to interact with XPC (Shah et al. 2017), being this the first time USP11 is seen to take part in NER. Notwithstanding, USP11 is a relatively non studied protein which until now had never been associated with XPF and regarding its reported roles in DSB and NER pathways, it sparks interest in characterizing this potentially relevant XPF interactor in the context of DNA repair.

USP11 showed up to interact constitutively with XPF with a slight tendency to reinforce this interaction under MMC treatment. SILAC assay was able to detect two or three unique peptides of USP11 independently of the conditions, however the parameter which assesses the probability, the Sum PEP Score, was not so high in two out of the three experiments, which might denotes XPF-USP11 interaction is weak or indirect. Concerning these signs, USP11 was considered as a potential candidate whose interaction with XPF should be validated by WB (Figure 45).



**Figure 45: CoIP of USP11 to confirm SILAC results.** XPF-HA immunoprecipitation showed a clear USP11 band in HEK XPF-KO + XPF-Wt with and without MMC treatment (1.5  $\mu$ M for 16h). HA and ERCC1 were used as positive controls.

In order to unravel if USP11-XPF interaction is mediated by the DNA or in contrast, it is a protein-protein interaction, a new CoIP was set adding a DNase to degrade this potential intermediary. As shown in figure 46, the addition of benzonase to the lysis buffer of the purification did not change USP11 detection, proving in this way that USP11-XPF interaction is not mediated by DNA.

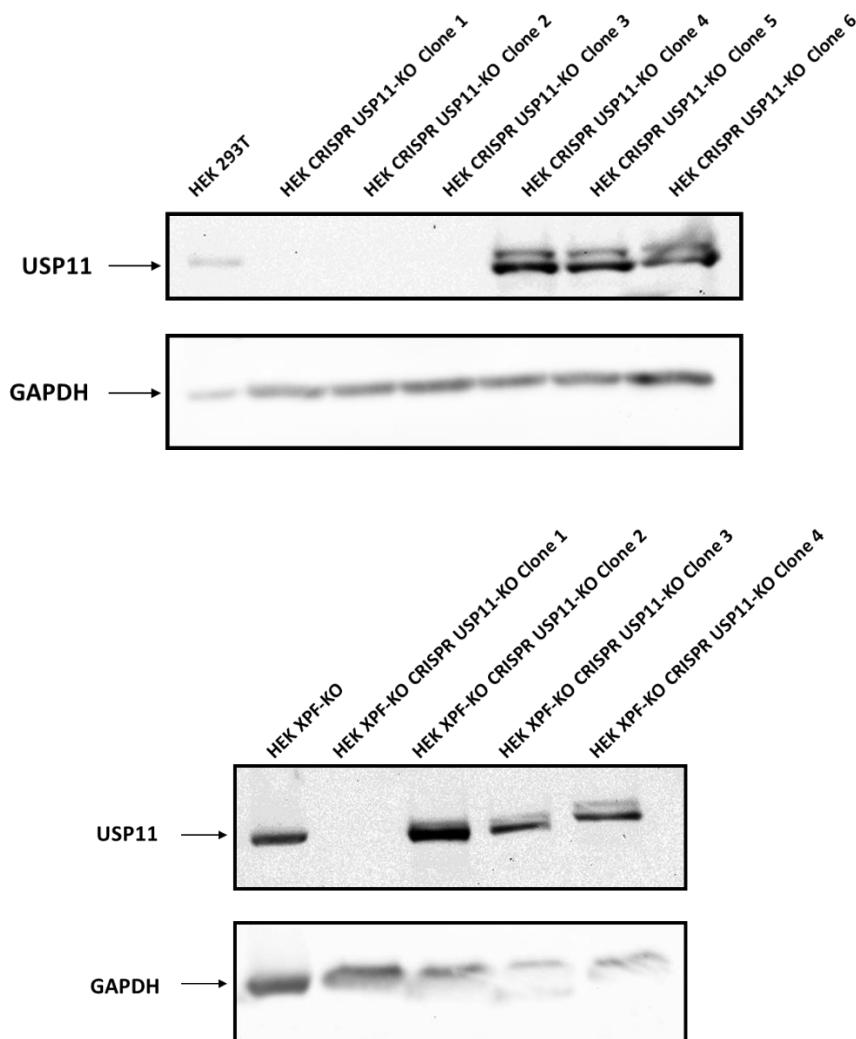


**Figure 46: CoIP of USP11 to test the nature of the XPF interaction.** HEK XPF-KO+XPF-Wt were treated with a DNase like benzonase to check if USP11-XPF interaction was mediated by DNA or it was direct. USP11 band is shown independently of benzonase, proving in this way the interaction is DNA independent.

#### **IV.5- GENERATION OF THE HEK USP11-KO AND THE DOUBLE KO HEK XPF/USP11 BY CRISPR TECHNOLOGY**

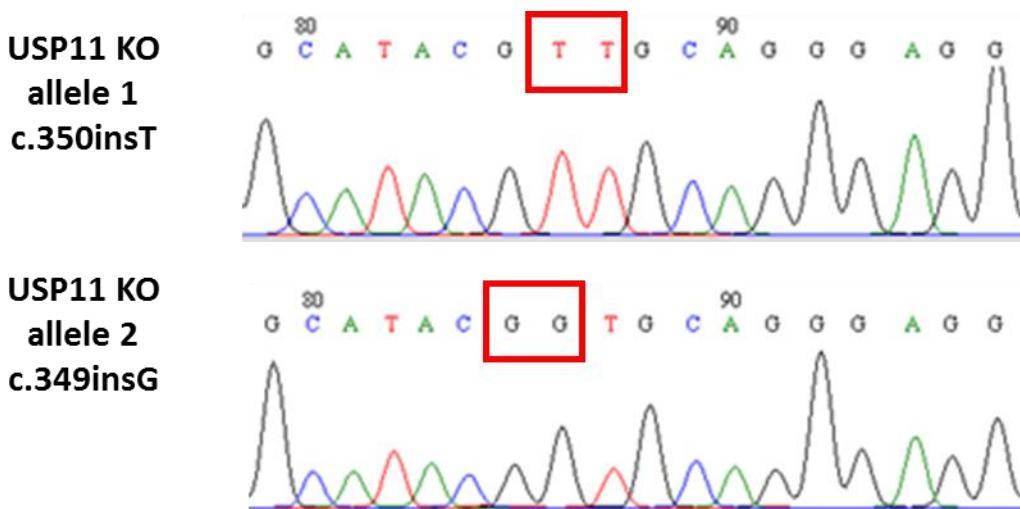
Gene editing CRISPR technology has emerged and improved constantly for the last years. It allows the easily generation of KO, introduction of point mutations, deletions and insertions in specific sites of the genome. It consists of a designed gRNA which leads the nuclease Cas9 to cut in a specific site of the genome that is subsequently repaired in an error prone way that ends up disrupting or modifying the protein reading frame.

CRISPR/Cas9 technique was implemented to obtain a KO of *USP11* in HEK 293T and a double KO HEK XPF/*USP11*. PX458 mammalian expression vector designed to cut exon 2 of *USP11* was used. Infected cells were sorted and divided in 96-well plates to achieve single KO clones of HEK USP11-KO and HEK XPF-KO / USP11-KO that were tested by WB (Figure 47).



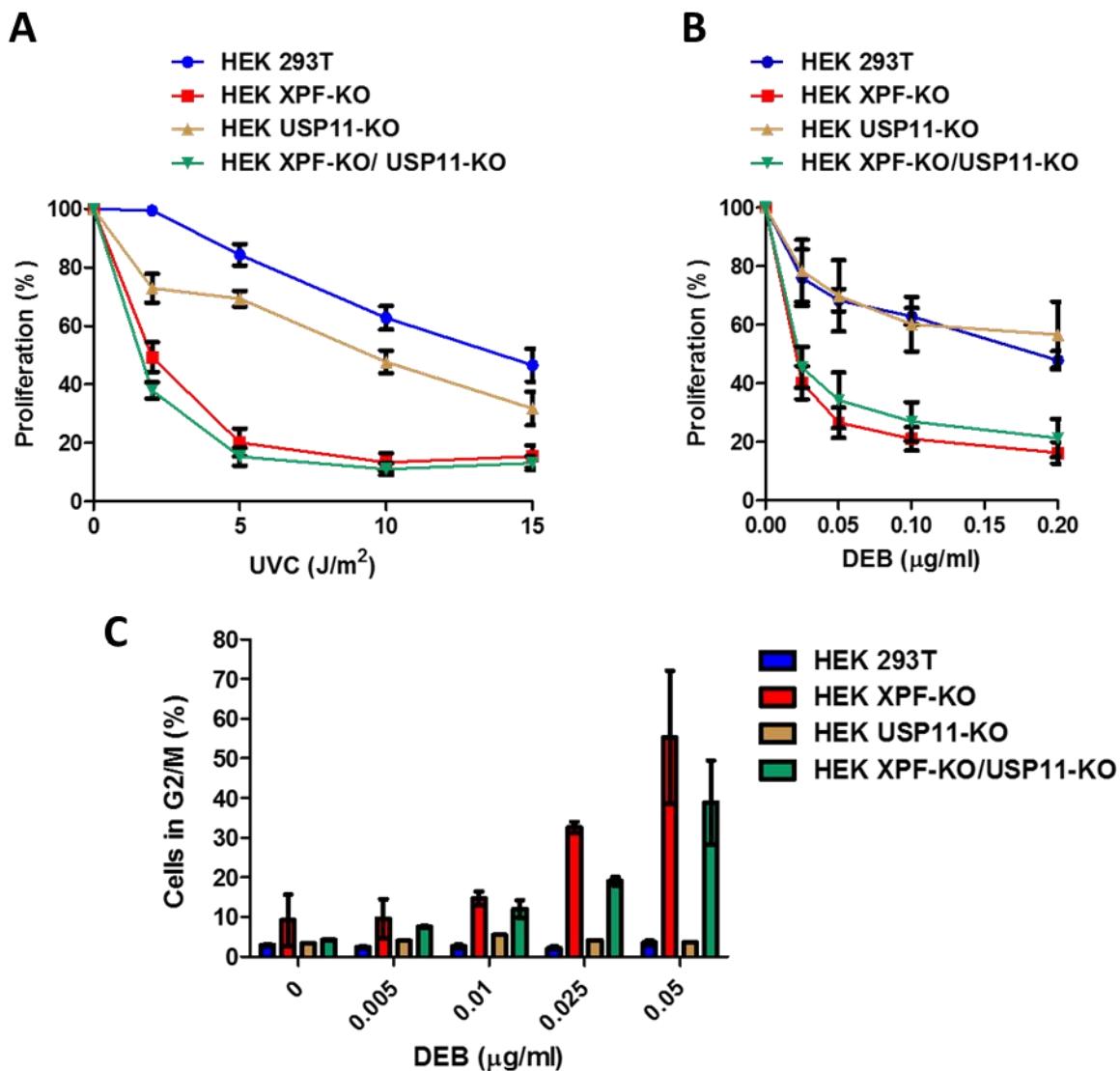
**Figure 47: WB analysis of USP11 CRISPR clones in HEK 293T cells (above) and in HEK XPF-KO cells (below).** Clones 1 from both WB which did not show USP11 band were selected. GAPDH band is shown as a loading control.

The sequencing analyses of the genomic DNA extracted and amplified by PCR from clone 1 of each cell line revealed that the mutations introduced by the CRISPR were two different single nucleotide insertions, respectively: c.350instT (c.350dup) and c.349insG (c.349dup) (Figure 48).



**Figure 48: Sequencing reaction chromatograms of heterozygous mutant clones.** The USP11-KO clone showed a 1 bp insertion (c.350dup) in one allele and a different 1 bp insertion (c.349dup) in the second allele, both in exon 2.

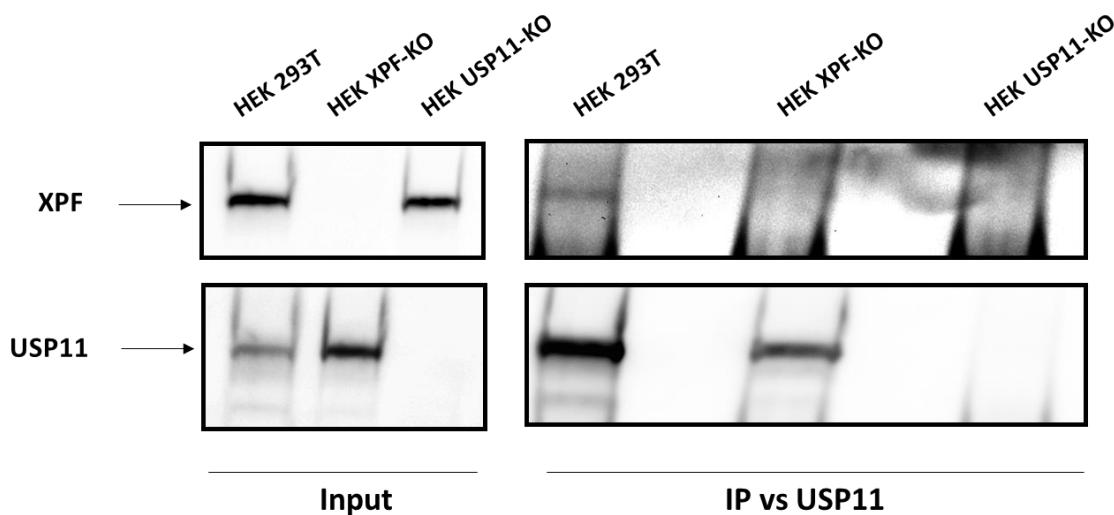
Functional studies of single and double USP11-KO cells were then performed to test their phenotype under treatments that impair DNA repair pathways where XPF participates. It can be seen from data in figure 49 that HEK USP11-KO cells have a mild reduction of survival after UVC irradiation, however, they were as resistant as the Wt to DEB treatment in terms of percentage of G2/M arrested cells and DEB sensitivity. Double KO XPF/USP11 mostly behaves as the HEK XPF-KO, suggesting that its sensitivity to UVC and DEB treatments, or its inability to reach mitosis is mainly due to the lack of XPF.



**Figure 49: Functional studies of USP11 role in NER and ICLR.** UVC cell survival (Figure A) and DEB cell survival (Figure B). The double KO HEK XPF-KO/USP11-KO sensitivity is similar to the HEK XPF-KO levels. USP11 cells showed a slight but consistent sensitivity to UVC. Graphs show proliferation percentage media and SEM of at least three independent experiments of two replicates. Cell cycle study (Figure C) shows the percentage of cells stalled in G2/M phase after DEB exposition. HEK USP11-KO behaves almost as the wild

type HEK 293T and HEK XPF-KO/USP11-KO arrested cells levels are more likely HEK XPF-KO. This graph represents the media of two independent experiments with the SD.

The new generated HEK USP11-KO cell line was used as a control to confirm by an endogenous USP11 immunoprecipitation that XPF-USP11 interaction was not an artifact of HA-tag of XPF (Figure 50). This outcome, together with the rejection of the assumption that USP11 was interacting with XPF throughout the DNA as shown in figure 46, strengthens the confirmation of USP11 as a novel XPF interactor.



**Figure 50: Immunoprecipitation vs USP11.** XPF band is shown in the IP of HEK 293T but is absent in the two negative controls. USP11 band is shown in HEK 293T and HEK XPF-KO samples. This endogenous IP vs USP11 confirms the interaction takes place *in vivo* and is not mediated by HA-tag of XPF.

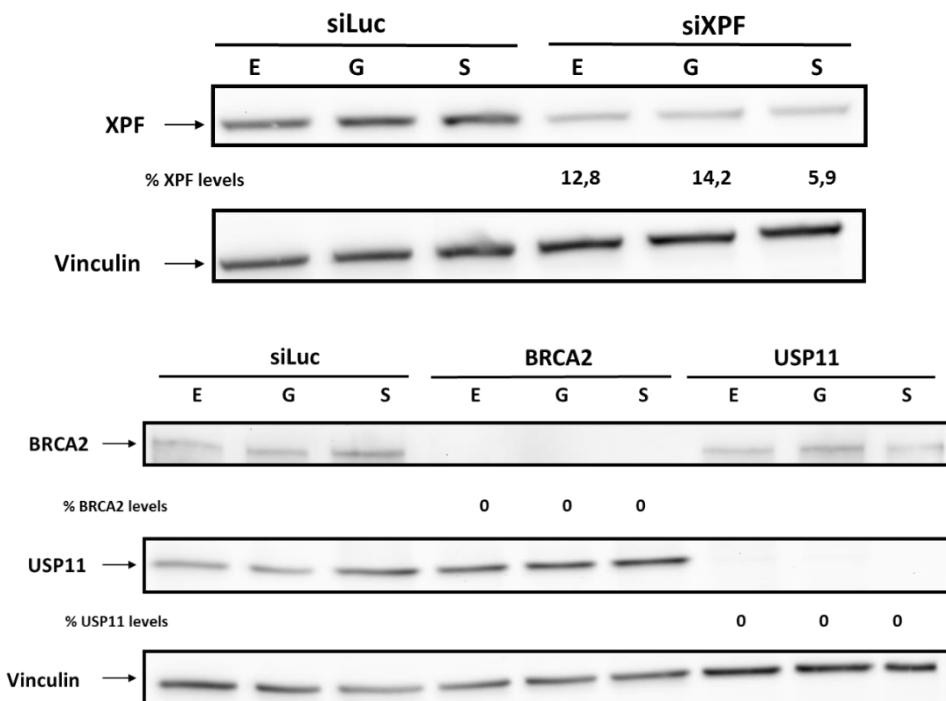
#### **IV.6- ANALYSIS OF XPF AND USP11 ROLES IN *IN VIVO* DOUBLE STRAND BREAKS REPAIR**

XPF is known to participate in some DSB repair pathways besides its proved role in NER and ICLR. Considering this new identified XPF-USP11 interaction, we wanted to unravel the role of these two proteins in HR and SSA.

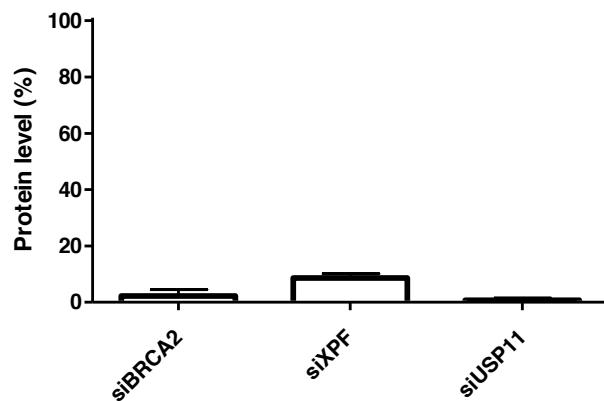
#### IV.6.1 STUDY OF HOMOLOGOUS RECOMBINATION REPAIR

In order to characterize the role of USP11 in DNA repair by HR, a study using U2OS DR-GFP cell line was performed. This cell line carries a chromosome-integrated reporter to restore a GFP expression cassette by HR when a unique DSB is induced by I-SceI endonuclease expression (Bennardo et al. 2008).

The indicated genes were knockdown by siRNA and depletion efficiency was analyzed by WB (Figure 51). The following figures correspond to one individual assay out of the five HR experiments. *XPF* was downregulated with around 90% of efficiency; *USP11* and *BRCA2* were downregulated with around 100% of efficiency. Figure 52 represents the mean of siRNA transfection efficiency of three WB.

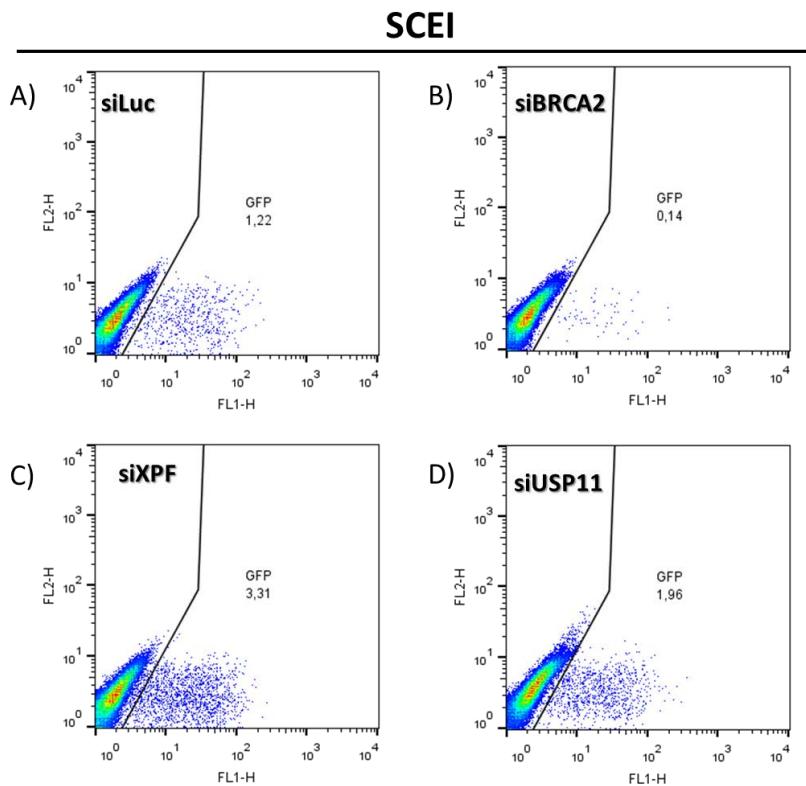


**Figure 51: siRNA efficiency in DR-GFP experiments analyzed by WB.** Representative WB obtained from a single DR-GFP assay. Percentage of protein level is represented for each downregulated gene respect to siLuc control (considered as a mean of 100%). E= EV; G= GFP vector; S= Sce-I vector. Vinculin was used as a loading control.



**Figure 52: siRNA efficiency in three DR-GFP experiments analyzed by WB.** Bars represent the percentage of protein level for each downregulated gene corresponding to the mean of three individual WB data with SEM.

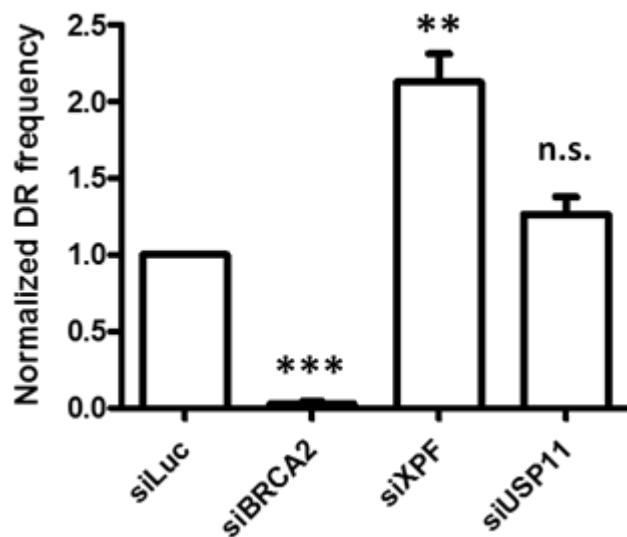
GFP fluorescence was then analyzed by flow cytometry (Figure 53).



**Figure 53: In vivo HR pathway in U2OS DR-GFP stable cell line measured by cell cytometry.** Following to siRNA and vector transfection, fluorescence of cells was analyzed by cell cytometry. Graphs show the amount

of GFP detected after endonuclease vector (I-SceI) transfection. Samples with EV and GFP vector were used as controls (used to normalize data obtained with I-SceI vector). A) siRNA luciferase (Luc), B) siBRCA2 control, C) siXPF, D) siUSP11.

As can be seen in figure 54, the depletion of XPF or USP11 does not decrease the HR frequency compared to the control siLuc, meaning these two proteins do not play an essential role in HR. In contrast, reduction of BRCA2, our positive control, is clearly impairing HR as expected (Howlett et al. 2002; Michl et al. 2016).



**Figure 54: *In vivo* HR pathway in U2OS DR-GFP stable cell line.** The flow cytometry analysis of GFP fluorescence is represented for each gene downregulated U2OS DR-GFP cell line. Luc is used as a control and data is normalized respect to it. The mean of five independent experiments is represented with the SEM. Paired t-test is represented, statistical differences were considered when p-value < 0,005 (n.s.: no significant).

The unambiguous outcome of siXPF HR frequency proves XPF is not essential for HR as it had been previously reported (Ahmad et al. 2008) and HR can take place in absence of XPF (Figure 54). Actually, XPF downregulation increased HR frequency, which turns it as a possible HR negative regulator. DSB repair is crucial for genome stability maintenance and XPF has been reported to participate in MMEJ and SSA, so downregulation of this gene would reduce the backup DSB repair mechanisms, hence increasing the HR of DSB. One possible explanation of XPF as a negative regulator of HR could be that, due to XPF

predisposition to cut bubble structures (Ciccia et al. 2008), it could cut DNA structures formed during HR. These late HR structures excised by XPF would proceed to mitosis containing gaps or breaks which could be then processed by MUS81-EME1, or even by ERCC1, fixing the possible formation of chromosome bridges during anaphase segregation (Naim et al. 2013).

USP11 knockdown does not reduce the HR neither increases it compared to the control (Figure 54), which sets USP11 aside from this DSB repair subpathway.

USP11 has been reported to interact with BRCA2 and PALB2, indeed, it seems to be essential for the formation of the complex BRCA1-PALB2-BRCA2 which is recruited to the DSB site in HR (Orthwein et al. 2015). It deubiquitilates PALB2 so that it can be bound to BRCA1. Orthwein and co-workers saw a reduction of HR when USP11 was downregulated and the subsequent increase of HR when those siUSP11 cells were transfected with a USP11 vector resistant to the siRNA. This is in contrast with our findings: one explanation could be in the levels of USP11 inhibition; while we show complete USP11 inhibition (Figure 52) Orthwein and co-workers have still detectable levels of the band. Despite levels of USP11 inhibition are not shown for that assay they performed an immunoprecipitation vs PALB2 where they also tried to inhibit USP11 but still showed detectable levels of the protein. To unravel if the different inhibition levels are the clue, a complete inhibition should be performed, knocking out *USP11* in this U2OS-DR cells.

Another interesting approach could come from cell cycle. Orthwein and co-workers stated USP11 is cell cycle regulated by proving its loss in G1 after DNA damage was more marked than in S or G2 phases (Orthwein et al. 2015). In this sense, it would be interesting to compare our outcomes with cells synchronized in S phase.

#### **IV.6.2 STUDY OF SINGLE STRAND ANNEALING REPAIR**

One of the less characterized repair abilities of XPF-ERCC1 is its role in SSA. Their orthologues in *S. cerevisiae* Rad1-Rad10 have been well understood to participate in MMEJ and SSA. It has been reported that the mammalian heterodimer also participates in DSB

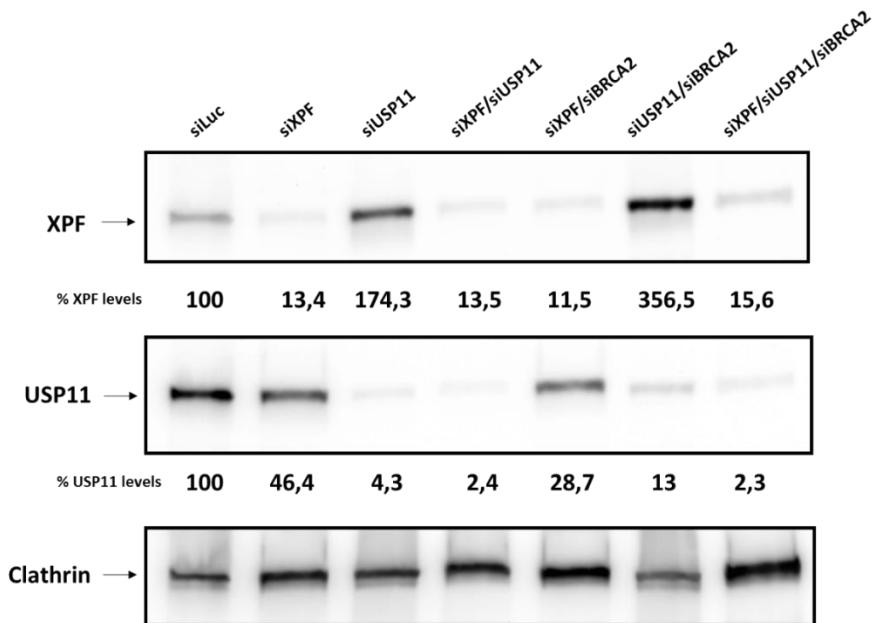
repair by SSA (Ahmad et al. 2008; Chang et al. 2017) but further studies will be needed to elucidate XPF-ERCC1 role in this DNA repair subpathway. Under the assumption that XPF interacts with USP11 and considering there is no report about the role of USP11 in SSA we decided to unravel the role, if any, of USP11 in SSA.

For this purpose, U2OS cells carrying the chromosome-integrated reporter SA-GFP to control the SSA induced by the I-SceI endonuclease were used (Bennardo et al. 2008). Similarly to the DR-GFP system, the GFP sequence is restored by SSA in response to the I-SceI-induced DSB. Additionally in this assay, cells are infected either just with a red fluorescent protein vector (as a control of the efficiency of the transfection), or with a red fluorescent protein vector and I-SceI vector, and both red and green fluorescence are analyzed by cell cytometry.

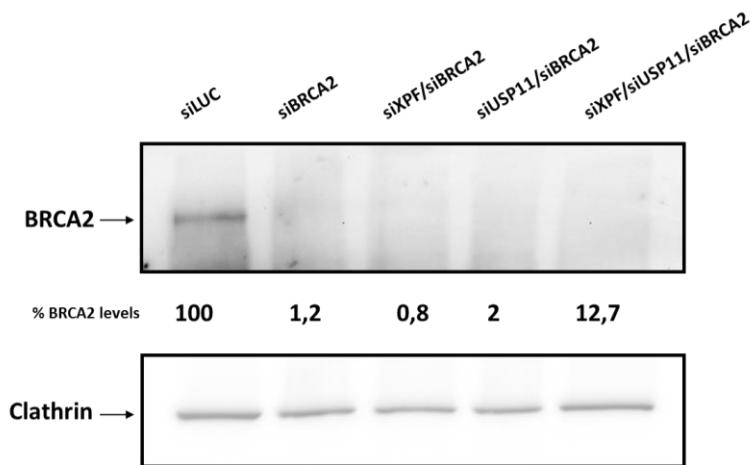
*BRCA2* is known to suppress SSA (Stark et al. 2004), therefore it was used as a control, as its reduction is expected to increase SSA and by consequence GFP fluorescence. Besides individual *XPF*, *USP11* and *BRCA2*, the combinations of two or three genes were also studied.

Genes were knocked-down by siRNA and transfection efficiency was checked by WB (Figure 55) (Figure 56). siXPF downregulated *XPF* more than 80%; *USP11* was downregulated more than 90% and *BRCA2* was downregulated more than 95% in single or combined siRNA downregulations (Figure 57). There was a rather surprising outcome derived from the WB levels of *XPF*: *USP11* downregulation substantially increased *XPF* levels. This was observed for the individual inhibition of *USP11* and the double inhibition *USP11/BRCA2*. The opposite effect, an increase of *USP11* levels when inhibiting *XPF*, however, was not observed. One hypothesis to explain this situation could be that, *USP11* would be directly involved the regulation of XPF-ERCC1 heterodimer levels. Supporting this conjecture, the importance of the ubiquitination of the XPF binding domain of ERCC1, (HhH)<sub>2</sub>, for the stability of both ERCC1 and XPF proteins has been recently reported (Yang et al. 2017). ERCC1 seems more likely to be altered by ubiquitination than XPF, although the stability of XPF would be also affected by this modification. Additional experiments to

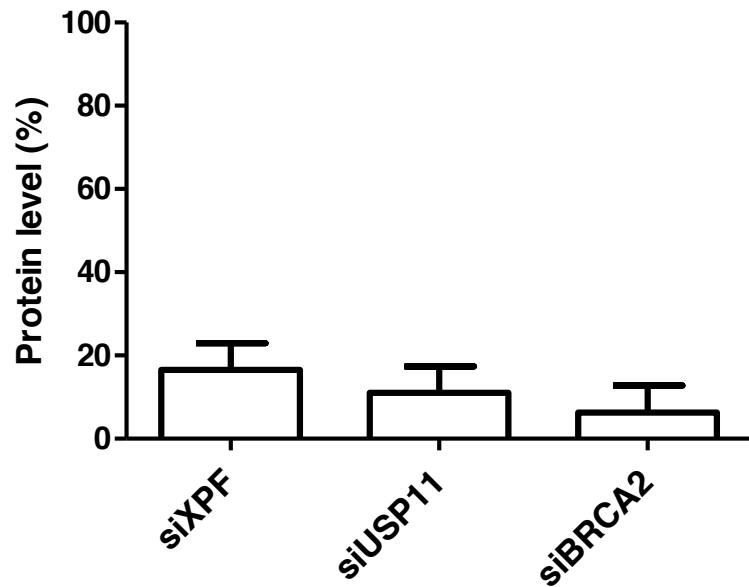
asses ERCC1 ubiquitination in SSA context could uncover the nature of XPF/USP11 epistasis.



**Figure 55: siRNA efficiency in SA-GFP experiments analyzed by WB.** Representative WB obtained from a single SA-GFP assay. Percentage of protein level is represented for each downregulated gene, *XPF* or *USP11*, respect to siLuc control. Clathrin was used as a loading control.

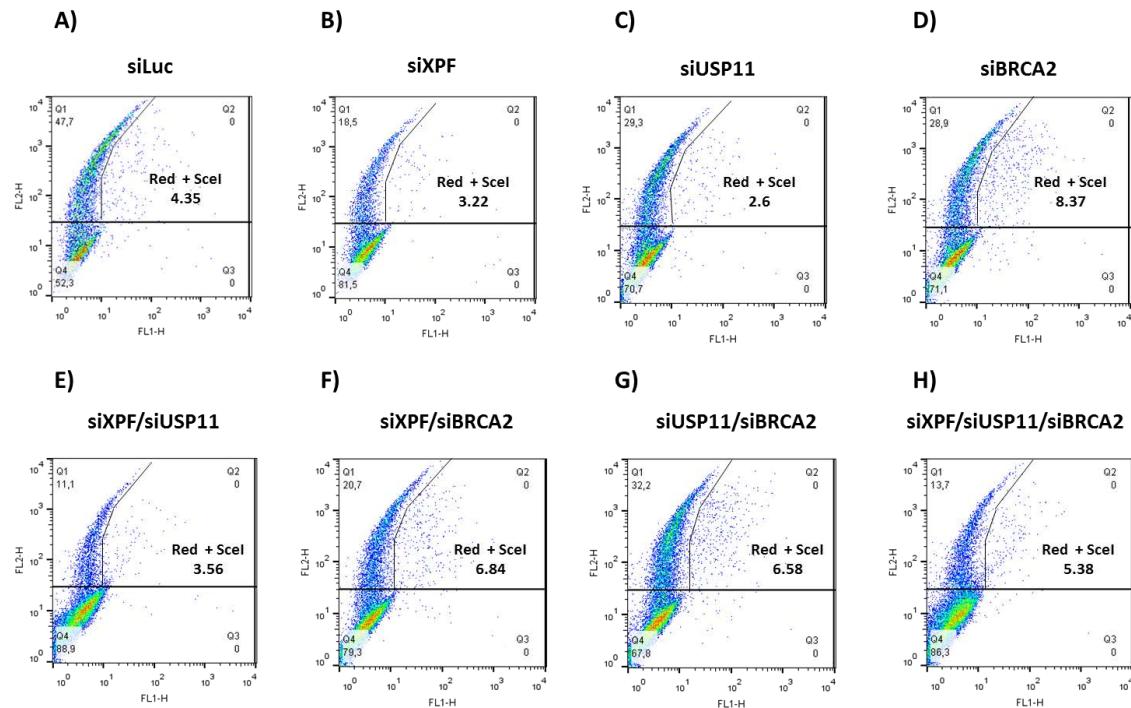


**Figure 56: siRNA efficiency in SA-GFP experiments analyzed by WB.** Representative WB obtained from a single SA-GFP assay. Percentage of protein level is represented for *BRCA2* downregulated gene, respect to siLuc control. Clathrin was used as a loading control.



**Figure 57: siRNA efficiency in four SA-GFP experiments analyzed by WB.** Bars represent the percentage of protein level for each downregulated gene corresponding to the mean of four individual WB data with SD.

The combination of red fluorescence and GFP fluorescence was analyzed by flow cytometry (Figure 58).

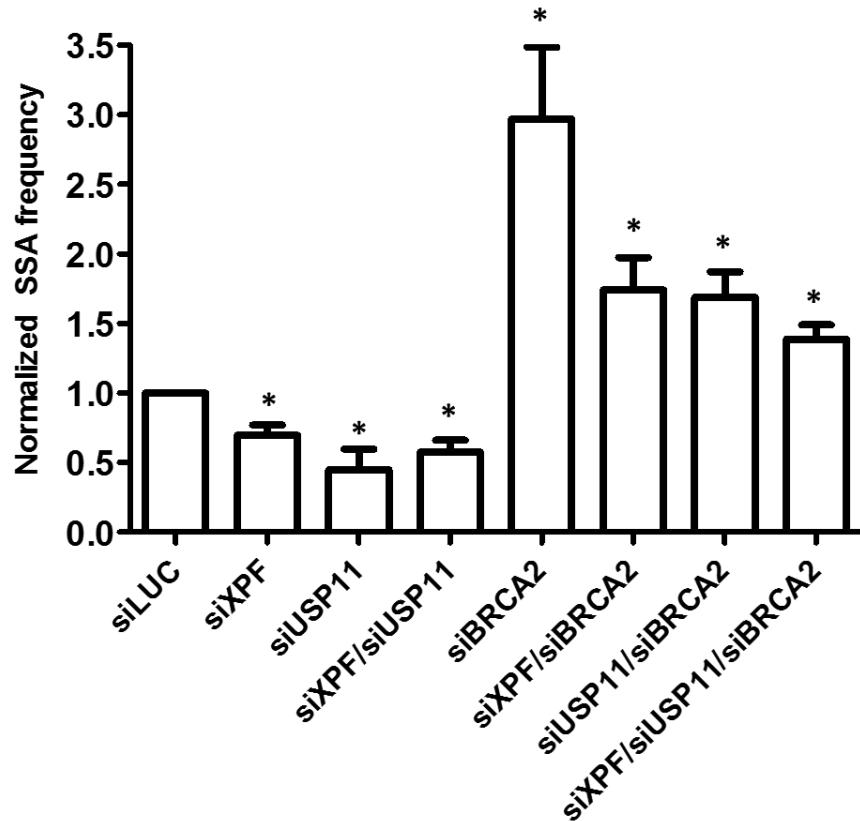
**SCEI**

**Figure 58: *In vivo* SSA pathway in U2OS SA-GFP stable cell line measured by cell cytometry.**

Following to siRNA and vector transfection, fluorescence of cells was analyzed by cell cytometry. Graphs show the amount of red fluorescence and GFP detected after red fluorescence vector and endonuclease vector (I-SceI) transfection. All the samples transfected with the same siRNA were infected with red fluorescence protein and half of them were additionally infected with the I-SceI. Red fluorescence was therefore used as a control to normalize data. A) siLuc B) siXPF; C) siUSP11; D) siBRCA2; E) siXPF/siUSP11; F) siXPF/siBRCA2; G) siUSP11/siBRCA2; H) siXPF/siUSP11/siBRCA2.

The results of the SSA fluorescence analysis are set out in figure 59. What stands out in the graph is the remarkable high frequency of SSA when *BRCA2* is downregulated, which confirms its role as a negative regulator of SSA. Comparing the individual knockdown of *XPF* to the siLuc, it results in a drop of SSA frequency about 30%, which confirms a role for *XPF* in this DSB pathway (Bhargava et al. 2016). Interestingly, *USP11* knockdown drove to similar outcomes, showing a fall of more than 50% respect to the siLuc; which proves for the

first time this protein is involved in SSA. A point of interest arises in the understanding of the downregulation combination of *XPF* and *USP11* at the same time.



**Figure 59: *In vivo* SSA pathway in U2OS SA-GFP stable cell line.** The flow cytometry analysis of GFP fluorescence is represented for each gene downregulated U2OS SA-GFP cell line. The mean of four independent experiments is represented with the SEM. Paired t-test is represented, statistical differences were considered when p-value < 0,005.

As it can be deduced from the figure X, the double knockdown of *XPF* and *USP11* experienced a fall in SSA frequency with values between siXPF and siUSP11 frequency values, pointing out that both proteins might be epistatic in positively regulating SSA. This is the most striking result to emerge from the data, as allows to classify the new XPF interactor USP11 as a protein involved in SSA.

Similar reduction levels are observed with the combination of *BRCA2* downregulation together with *XPF* or *USP11* downregulations, again proving *XPF* and *USP11* have similar roles in SSA. No significant reduction is found in the triple downregulated *BRCA2*, *XPF* and *USP11* compared to the double *BRCA2* knockdowns, pointing again to the epistatic effect of *XPF* and *USP11*.

#### **IV.7- UNDERSTANDING THE BIOLOGICAL IMPACT OF XPF-USP11 INTERACTION**

Considering *USP11* is a deubiquitylase involved in DNA damage response (DDR) (Hendriks et al. 2015) is not inconceivable it could play a role in DNA repair together with *XPF* both in NER and SSA.

Recent research has suggested *USP11* participates in NER regulation by deubiquitinating *XPC*. *XPC* acts as a probe checking DNA damage; first, it is polyubiquitinated after UV exposure to enhance its binding to the damage site, secondly it is sumoylated and finally it experiences a second ubiquitination event to be released from the damage site. In order to recover *XPC* protein and avoid proteasome degradation, *USP11* deubiquitylates it, thereby contributing to the NER regulation positively (Shah et al. 2017).

A similar scenario could be envisaged for *XPF*. Actually, another DUB, *USP45* deubiquitylase, has been proved to regulate *XPF-ERCC1* activity by deubiquitylating *ERCC1*. Cells lacking *USP45* were hypersensitive to UV irradiation and DNA ICLs and the recruitment of *XPF-ERCC1* to the damaged DNA seemed to be regulated by its proper deubiquitination, which would be set out by *USP45* (Perez-Oliva et al. 2015). In this case, *ERCC1* ubiquitination would control its activity instead of regulating *XPF-ERCC1* stability as previously discussed (Yang et al. 2017). In this sense, *USP11* outcomes from this research differ from the ones from Perez-Oliva group, as cells lacking *USP11* were not hypersensitive to UVC or ICL exposure, but the results stand out for another interesting possibility of participating together with *XPF* in SSA, which certainly highlights the view that proteins involved in DSB repair are often influenced by a web of inhibitions/activations or by post-

translational modifications rather than individually taking part in the pathways (Surralles et al. 2004).

A study of potential USP11 de-ubiquitinating sites should be performed, although not just focusing in XPF-ERCC1 heterodimer; assuming its epistasis, USP11 could deubiquitylate other proteins associated to SSA repair, regulating in that way their participation in DSB repair pathways. This would not be against the idea of the physical interaction between XPF-ERCC1 and USP11, which has been widely proved in this study, but gives a hint of the possibilities of USP11 to be deubiquitinating other surrounding proteins maybe by taking advantage of the interaction with XPF heterodimer and travelling recruited to it or, in contrast, making USP11 unable to proper deubiquitylate target proteins since the union to XPF-ERCC1 could avoid USP11 deubiquitinating ability.

Actually, some target proteins of USP11 involved in DSB repair have already been reported, such as PALB2 (Orthwein et al. 2015) and BRCA2 (Schoenfeld et al. 2004), whose reduction increases the frequency of SSA (Stark et al. 2004). Additionally, these genes are considered FBOC genes (Bogliolo and Surralles 2015), which could spur the study of other target proteins of USP11 which could be involved in cancer.

This study has provided a deeper insight into the molecular biology of XPF in DSB repair. The identification of USP11 as an XPF interactor lays the groundwork for future research into characterizing this interaction and points towards the study of post-translational modifications in SSA DNA repair context.

# Conclusions



## V. CONCLUSIONS

1. Functional analyses reveal that the position of the amino acid substitution along XPF sequence by itself does not clearly define which DNA repair pathway is impaired and therefore neither determines the patient's clinical phenotype. Such outcomes imply that other factors must be taken in account including the levels of XPF, cell localization and possible interactors that can affect its role in DNA damage repair.
2. XPF variants identified in patients with FA are able to retain mild NER activity, while XPF variants identified in patients with other diseases present both ICL and UV sensitivities. This could be due to intrastrand lesions that also require XPF to be repaired .
3. Cells with FA associated variants hardly reach mitosis and, the ones that do, experience chromosome breaks; on the contrary, cells from XP and CS associated variants can relatively easily reach mitosis with damaged DNA.
4. USP11 is a deubiquitinase involved in DNA repair that emerges as a novel XPF interactor. XPF-USP11 interaction is DNA damage-independent and is not mediated by DNA.

5. *ERCC4* and *USP11* are epistatic genes involved in SSA. *USP11* does not participate in other DNA repair pathways such as ICL and HR but could have a minor role in NER that requires further investigation.
6. XPF plays a role in both HR and SSA pathways by positively regulating SSA and negatively regulating HR.

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# **Annex**



Accession	Description	Control				WT			
		MMJS _01	MMJS _05	MMJS _09	MMJ S_02	MMJ S_06	MMJ S_10	Con trol	WT
P07992	DNA excision repair protein ERCC1 OS=Homo sapiens GN=ERCC1 PE=1 SV=1 - [ERCC1_HUMAN]	Σ# PSMs 44	A2: Area	C2: Area	D2: Area	E2: Area	F2: Area	COU NTS	COU NTS
Q92889	DNA repair endonuclease XPF OS=Homo sapiens GN=ERCC4 PE=1 SV=3 - [XPF_HUMAN]	117			1,729E 8	1,064E 8	3,324E 8	0	3
P22090	40S ribosomal protein S4, Y isoform 1 OS=Homo sapiens GN=RPS4Y1 PE=1 SV=2 - [RS4Y1_HUMAN]	7			2,537E 8	1,571E 8	4,117E 8	0	3
P46782	40S ribosomal protein S5 OS=Homo sapiens GN=RPS5 PE=1 SV=4 - [RS5_HUMAN]	4			3,893E 6	2,673E 6	2,380E 7	0	3
P62263	40S ribosomal protein S14 OS=Homo sapiens GN=RPS14 PE=1 SV=3 - [RS14_HUMAN]	2			1,585E 6	9,882E 6	6	0	2
P84098	60S ribosomal protein L19 OS=Homo sapiens GN=RPL19 PE=1 SV=1 - [RL19_HUMAN]	2			2,089E 6	2,379E 6	6	0	2
P62280	40S ribosomal protein S11 OS=Homo sapiens GN=RPS11 PE=1 SV=3 - [RS11_HUMAN]	2			1,281E 6	3,855E 6	4,551E 6	0	2
P61247	40S ribosomal protein S3a OS=Homo sapiens GN=RPS3A PE=1 SV=2 - [RS3A_HUMAN]	2			1,315E 7	2,058E 7	0	0	2
P11142	Heat shock cognate 71 kDa protein OS=Homo sapiens GN=HSPA8 PE=1 SV=1 - [HSP7C_HUMAN]	4			4,199E 6	4,368E 6	2,928E 6	0	2
Q75152	Zinc finger CCCH domain-containing protein 11A OS=Homo sapiens GN=ZC3H11A PE=1 SV=3 - [ZC11A_HUMAN]	2			2,281E 5	3,475E 5	0	0	2
P36578	60S ribosomal protein L4 OS=Homo sapiens GN=RPL4 PE=1 SV=5 - [RL4_HUMAN]	2			3,671E 6	3,522E 6	0	0	2
Q00839	Heterogeneous nuclear ribonucleoprotein U OS=Homo sapiens GN=HNRNPU PE=1 SV=6 - [HNRPU_HUMAN]	2			5,227E 6	3,908E 6	0	0	3
Q81Y92	Structure-specific endonuclease subunit SLX4 OS=Homo sapiens GN=SLX4 PE=1 SV=3 - [SLX4_HUMAN]	3			2,479E 6	6,354E 6	0	0	2
Q9H1R3	Myosin light chain kinase 2, skeletal/cardiac muscle OS=Homo sapiens GN=MYLK2 PE=1 SV=3 - [MYLK2_HUMAN]	2			1,466E 7	2,643E 7	0	0	2
P21333	Filamin-A OS=Homo sapiens GN=FLNA PE=1 SV=4 - [FLNA_HUMAN]	2			1,002E 8	1,951E 5	0	0	2
P98198	Probable phospholipid-transferring ATPase ID OS=Homo sapiens GN=ATP8B2 PE=2 SV=2 - [AT8B2_HUMAN]	2			1,866E 7	5,789E 7	0	0	2

Accession	Description	Control				WT + UV				Cont rol	WT - UV	
		Σ# PSMS	A2: Area	B2: Area	C2: Area	MMJ S_01	MMJ S_05	MMJ S_09	MMJ S_07	MMJ S_11		
Q92889	DNA repair endonuclease XPF OS=Homo sapiens GN=RCC4 PE=1 SV=3 - [XPFI_HUMAN]	147				4,315E	5,509E	3,951E	8	8	0	3
P07992	DNA excision repair protein ERCC1 OS=Homo sapiens GN=ERCC1 PE=1 SV=1 - [ERCC1_HUMAN]	58				3,315E	4,984E	3,328E	8	8	0	3
P28715	DNA repair protein complementing XP-G cells OS=Homo sapiens GN=ERCC5 PE=1 SV=3 - [ERCC5_HUMAN]	8				1,965E	1,135E	6	7	7	0	2
P62701	40S ribosomal protein S4, X isoform OS=Homo sapiens GN=RPS4X PE=1 SV=2 - [RS4X_HUMAN]	14				1,070E	1,177E	1,289E	7	7	0	3
Q00839	Heterogeneous nuclear ribonucleoprotein U OS=Homo sapiens GN=HNRNPU PE=1 SV=6 - [HNRNPU_HUMAN]	7				1,934E	1,928E	5,071E	6	6	0	3
P0CW22	40S ribosomal protein S17-like OS=Homo sapiens GN=RPS17L PE=2 SV=1 - [RS17L_HUMAN]	9				4,070E	4,136E	3,558E	7	7	0	3
P61247	40S ribosomal protein S3a OS=Homo sapiens GN=RPS3A PE=1 SV=2 - [RS3A_HUMAN]	5				2,499E	1,582E	1,171E	7	7	0	3
P46783	40S ribosomal protein S10 OS=Homo sapiens GN=RPS10 PE=1 SV=1 - [RS10_HUMAN]	7				1,009E	1,009E	1,009E	7	7	0	2
P15880	40S ribosomal protein S2 OS=Homo sapiens GN=RPS2 PE=1 SV=2 - [RS2_HUMAN]	5				1,249E	1,885E	1,513E	7	7	0	2
Q8IY92	Structure-specific endonuclease subunit SLX4 OS=Homo sapiens GN=SLX4 PE=1 SV=3 - [SLX4_HUMAN]	5				1,951E	6,336E	4,696E	6	6	0	3
P27635	60S ribosomal protein L10 OS=Homo sapiens GN=RPL10 PE=1 SV=4 - [RL10_HUMAN]	3				7,522E	1,312E	6	7	7	0	3
P36578	60S ribosomal protein L4 OS=Homo sapiens GN=RPL4 PE=1 SV=5 - [RL4_HUMAN]	4				3,284E	4,739E	3,700E	6	6	0	3
P11142	Heat shock cognate 71 kDa protein OS=Homo sapiens GN=HSPA8 PE=1 SV=1 - [HSP7C_HUMAN]	7				9,205E	9,211E	2,947E	6	6	0	3
Q92878	60S ribosomal protein L6 OS=Homo sapiens GN=RPL6 PE=1 SV=3 - [RL6_HUMAN]	3				1,803E	2,788E	6	6	6	0	3
P62851	40S ribosomal protein S25 OS=Homo sapiens GN=RPS25 PE=1 SV=1 - [RS25_HUMAN]	4				5,834E	2,770E	7,636E	5	5	0	3
P84098	60S ribosomal protein L19 OS=Homo sapiens GN=RPL19 PE=1 SV=1 - [RL19_HUMAN]	3				5,632E	5,045E	6	6	6	0	2
P49411	Elongation factor Tu, mitochondrial OS=Homo sapiens GN=TUFM PE=1 SV=2 - [EFTU_HUMAN]	5				9,372E	3,568E	3,114E	5	5	0	3
P46782	40S ribosomal protein S5 OS=Homo sapiens GN=RPSS5 PE=1 SV=4 - [RSS5_HUMAN]	3				4,975E	1,162E	1,162E	6	6	0	2

Q08J23	tRNA (cytosine(34)-C(5))-methyltransferase OS=Homo sapiens GN=NSUN2 PE=1 SV=2 - [NSUN2_HUMAN]	4			1,021E-6	1,217E-6	0	2
P62249	40S ribosomal protein S16 OS=Homo sapiens GN=RPS16 PE=1 SV=2 - [RPS16_HUMAN]	2			2,393E-7	1,328E-7	0	2
P61254	60S ribosomal protein L26 OS=Homo sapiens GN=RPL26 PE=1 SV=1 - [RPL26_HUMAN]	3			6,980E-6	3,566E-6	0	2
P09651	Heterogeneous nuclear ribonucleoprotein A1 OS=Homo sapiens GN=HNRNPA1 PE=1 SV=5 - [ROA1_HUMAN]	2			5,022E-5	3,483E-5	0	2
P83881	60S ribosomal protein L36a OS=Homo sapiens GN=RPL36A PE=1 SV=2 - [RPL36A_HUMAN]	2			2,109E-6	1,542E-6	0	2
P12004	Proliferating cell nuclear antigen OS=Homo sapiens GN=PCNA PE=1 SV=1 - [PCNA_HUMAN]	2			2,012E-6	2,881E-6	0	2
Q04637	Eukaryotic translation initiation factor 4 gamma 1 OS=Homo sapiens GN=EIF4G1 PE=1 SV=4 - [EIF4G1_HUMAN]	2			1,115E-6	5,542E-5	0	2
P62750	60S ribosomal protein L23a OS=Homo sapiens GN=RPL23A PE=1 SV=1 - [RPL23A_HUMAN]	2			1,485E-6	2,941E-6	0	2
P50402	Emerin OS=Homo sapiens GN=EMD PE=1 SV=1 - [EMD_HUMAN]	3			8,070E-5	4,876E-5	2,784E-5	0
P30153	Serine/threonine-protein phosphatase 2A 65 kDa regulatory subunit A alpha isoform OS=Homo sapiens GN=PPP2R1A PE=1 SV=4 - [2AAA_HUMAN]	2			1,003E-6	6,167E-5	0	2
P62263	40S ribosomal protein S14 OS=Homo sapiens GN=RPS14 PE=1 SV=3 - [RPS14_HUMAN]	3			8,509E-6	8,947E-6	3,640E-6	0
P67809	Nuclease-sensitive element-binding protein 1 OS=Homo sapiens GN=YBX1 PE=1 SV=3 - [YBOX1_HUMAN]	2			7,346E-5	9,559E-5	0	2
P42677	40S ribosomal protein S27 OS=Homo sapiens GN=RPS27 PE=1 SV=3 - [RPS27_HUMAN]	2			4,567E-6	6,884E-6	0	2
P62910	60S ribosomal protein L32 OS=Homo sapiens GN=RPL32 PE=1 SV=2 - [RPL32_HUMAN]	3			1,752E-6	3,238E-6	2,216E-6	0
Q9UNF1	Melanoma-associated antigen D2 OS=Homo sapiens GN=MAGED2 PE=1 SV=2 - [MAGED2_HUMAN]	3			7,165E-5	7,848E-5	2,134E-5	0
Q9Y5J1	U3 small nucleolar RNA-associated protein 18 homolog OS=Homo sapiens GN=UTP18 PE=1 SV=3 - [UTP18_HUMAN]	2			2,718E-6	4,786E-6	0	3
O75152	Zinc finger CCCH domain-containing protein 11A OS=Homo sapiens GN=ZC3H11A PE=1 SV=3 - [ZC3H11A_HUMAN]	3			6,005E-5	5,476E-5	3,293E-5	0
P23025	DNA repair protein complementing XP-A cells OS=Homo sapiens GN=XPA PE=1 SV=1 - [XPA_HUMAN]	3			5,317E-6	4,451E-6	0	2
Q8VWWK9	Cytoskeleton-associated protein 2 OS=Homo sapiens GN=CKAP2 PE=1 SV=1 - [CKAP2_HUMAN]	2			3,597E-5	3,692E-5	0	2

Accession	Description	Control			MMC			WT_UV
		MMJS _01	MMJS _05	MMJS _09	MMJS _04	MMJS _08	MMJS _12	
Q92889	DNA repair endonuclease XPF OS=Homo sapiens GN=ERCC4 PE=1 SV=3 - [XPF_HUMAN]	Σ# PSMs	A2: Area	B2: Area	C2: Area	E2: Area	F2: Area	COUN TS
P07992	DNA excision repair protein ERCC-1 OS=Homo sapiens GN=ERCC1 PE=1 SV=1 - [ERCC1_HUMAN]	104			1,994E8	7,553E7	4,130E8	0
Q6UWP8	Suprabasigin OS=Homo sapiens GN=SBSN PE=2 SV=2 - [SBSN_HUMAN]	38			1,630E8	5,050E7	2,996E8	0
P36578	60S ribosomal protein L4 OS=Homo sapiens GN=RPL4 PE=1 SV=5 - [RL4_HUMAN]	2			2,634E5		2,536E5	0
P35813	Protein phosphatase 1A OS=Homo sapiens GN=PPM1A PE=1 SV=1 - [PPM1A_HUMAN]	3			1,228E6	1,808E6	3,566E6	0
Q00839	Heterogeneous nuclear ribonucleoprotein U OS=Homo sapiens GN=HNRNPU PE=1 SV=6 - [HNRPU_HUMAN]	3			2,941E7	6,363E7	4,977E7	0
Q9H1R3	Myosin light chain kinase 2, skeletal/cardiac muscle OS=Homo sapiens GN=MLK2 PE=1 SV=3 - [MLK2_HUMAN]	2			2,618E6	2,267E6	2,208E6	0
Q6ZN84	Coiled-coil domain-containing protein 81 OS=Homo sapiens GN=CCDC81 PE=2 SV=2 - [CCDC81_HUMAN]	2			1,869E7		2,015E7	0
					1,991E8	2,485E8		0
								2

**Annex Table 1: Mass spectrometry data from XPF-TAP purification.** Interactors of XPF purified samples without treatment, with UVC irradiation (150J/m<sup>2</sup> 1h of recovery) or MMC treatment (1.5 µM, 16h) per triplicate are shown here in respect to the control (XPF-KO). PSMs: total number of identified peptide spectra matched for the protein.

	Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMs	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pI	Score Sequest HT	# Peptides Sequest HT
	High	P49454	Centromere protein F [OS=Homo sapiens]	0	3.04047	1.3084112	3	4	3	1	3210	367,537	5,07	3,65088141	3
<b>B1</b>	Medium	O9UQ35	serine/arginine repetitive matrix protein 2 [OS=Homo sapiens]	0.01613	1.232225	0.4722837	1	1	1	1	2752	299,438	12,06	1,79620183	1
	Medium	Q14676	Mediator of DNA damage checkpoint protein 1 [OS=Homo sapiens]	0.01613	2.56698	1,4836636	3	3	3	1	2089	226,529	5,47	0	3
	Medium	Q8TD10	Chromodomain-helicase-DNA-binding protein 5 [OS=Homo sapiens]	0.01613	0.97757	0.6655019	1	1	1	1	1954	222,909	6,15	0	1

	Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMs	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pI	Score Sequest HT	# Peptides Sequest HT
	High	Q14676	Mediator of DNA damage checkpoint protein 1 [OS=Homo sapiens]	0	52,3107	14,600287	18	43	18	1	2089	226,529	5,47	67,8228009	18
	High	A6NH9-1	Structural maintenance of chromosomes flexible hinge domain-containing protein RRP5 homolog [OS=Homo sapiens]	0	60,46448	15,112219	26	55	26	1	2005	226,231	7,3	69,6142339	26
	Medium	Q14690	PH-interacting protein [OS=Homo sapiens]	0.02924	1.43321	0.5344735	1	3	1	1	1871	208,57	8,87	0	1
	High	Q8WW20	Structure-specific endonuclease subunit SLX4 [OS=Homo sapiens]	0	2,60642	0.7138935	1	2	1	1	1821	206,56	8,85	0	1
	High	Q8Y92	Chromodomain-helicase-DNA-binding protein 1 [OS=Homo sapiens]	0	8,67743	3,217012	5	11	5	1	1834	199,889	6,06	11,9492532	5
	High	O14646	Chromodomain-helicase-DNA-binding protein 1 [OS=Homo sapiens]	0	20,0259	5,497076	9	18	9	1	1710	196,567	7,14	20,7677811	9
	High	P51532-1	Transcription activator BRG1 [OS=Homo sapiens]	0	12,5927	4,1894353	6	11	6	1	1647	184,53	7,88	5,92011726	6
	High	Q02880	DNA topoisomerase 2-beta [OS=Homo sapiens]	0	270,988	46,125461	65	357	51	1	1626	183,152	8	703,432699	65
	High	Q8N127-1	THO complex subunit 2 [OS=Homo sapiens]	0	41,5965	13,370998	19	49	19	1	1593	182,659	8,44	56,8713735	19
	High	P11388-4	Isoform 4 of DNA topoisomerase 2-alpha [OS=Homo sapiens]	0	33,40182	46,401985	78	535	64	1	1612	182,567	8,63	1038,95821	78
	High	Q9NRL2	Bromodomain adjacent to zinc finger domain protein 1A [OS=Homo sapiens]	0	63,7098	22,622108	29	70	29	1	1556	178,592	6,6	8,9467206	29
	High	O63036	Intron-binding protein aquarius [OS=Homo sapiens]	0	11,8733	4,3779744	7	9	7	1	1485	171,186	6,37	1,61474156	7
	High	Q9UIGO	Tyrosine-protein kinase BAZ1B [OS=Homo sapiens]	0	14,0301	5,9339177	8	12	8	1	1483	170,796	8,48	3,98667264	8
	High	P38880-3	Isoform 3 of Homeobox protein cut-like 1 [OS=Homo sapiens]	0	53,3713	2,176781	3	3	3	1	1516	165,587	5,86	1,81655965	3
	High	Q9UIJ0	ATPase family AAA domain-containing protein 2B [OS=Homo sapiens]	0	7,48184	2,7434842	4	7	1	1	1458	164,811	6,8	8,00571334	4
	High	Q9NT15	Sister chromatid cohesion protein PDSS5 homolog B [OS=Homo sapiens]	0	12,9433	6,219765	7	11	7	1	1447	164,563	8,47	8,11630535	7
	High	Q96T23	remodeling and spacing factor 1 [OS=Homo sapiens]	0	2,5626	11,936155	12	21	12	1	1441	163,72	5,01	12,5643468	12
	High	Q6PL18-1	ATPase family AAA domain-containing protein 2 [OS=Homo sapiens]	0	12,9099	4,672529	6	10	3	1	1390	158,456	6,32	13,860622	6
	High	Q7ZK3-1	Pogo transposable element with ZNF domain [OS=Homo sapiens]	0	10,6252	4,822695	6	7	6	1	1410	155,245	7,4	5,38452554	6
	High	Q9P2E9-1	Ribosome-binding protein 1 [OS=Homo sapiens]	0	41,4923	15,533915	18	35	18	1	1410	152,381	8,6	25,3741769	18
	High	Q9UKV3-1	Apoptotic chromatin condensation inducer in the nucleus [OS=Homo sapiens]	0	273,671	41,983594	45	359	2	1	1341	151,771	6,43	808,990564	45
	High	Q9UKV3-5	Isoform 4 of Apoptotic chromatin condensation inducer in the nucleus [OS=Homo sapiens]	0	264,112	41,415663	45	358	2	1	1328	150,467	6,43	780,37653	45
	High	Q9BQGQ-2	Isoform 2 of Myb-binding protein 1A [OS=Homo sapiens]	0	2,49976	0.7507508	1	2	1	1	1332	149,274	9,28	0	1
	High	P28715-1	DNA repair protein complementing XP-G cells [OS=Homo sapiens]	0	9,02981	4,0472175	4	7	4	1	1186	133,027	5,22	3,68164706	4

	Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMs	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pI	Score Sequest HT	# Peptides Sequest HT
	High	Q9UKV3-1	Apoptotic chromatin condensation inducer in the nucleus [OS=Homo sapiens]	0	11,2331	4,1014169	6	9	6	1	1341	151,771	6,43	6,36114132	6
	High	O75533-1	Splicing factor 3B subunit 1 [OS=Homo sapiens]	0	6,02079	2,3006135	3	5	3	1	1304	145,738	7,09	3,58734798	3
	High	Q13045-1	protein flightless-1 homolog [OS=Homo sapiens]	0	7,95636	2,3640662	3	5	3	1	1269	144,659	6,05	5,3245815	3
	High	Q08211	Atp-dependent rna helicase a [OS=Homo sapiens]	0	209,245	41,968504	50	237	50	1	1270	140,869	6,84	421,519841	50
	Medium	P54098	DNA polymerase subunit gamma-1 [OS=Homo sapiens]	0,02985	1,20377	0.8878128	1	2	1	1	1239	139,473	6,9	0	1
	High	Q14562	ATP-dependent RNA helicase ddx8 [OS=Homo sapiens]	0	19,8116	8,278885	8	11	8	1	1220	139,227	8,32	13,1515679	8
	High	Q7L2E3-2	Isoform 2 of Putative ATP-dependent RNA helicase DHX30 [OS=Homo sapiens]	0	17,9869	9,2471358	10	15	10	1	1222	136,03	8,6	7,8582495	10
	High	Q9Y2K7	Lysine-specific demethylase 2A [OS=Homo sapiens]	0	35,1578	14,457831	15	32	15	1	1162	132,708	7,58	31,1919024	15
	Medium	Q9UNV4-1	transcription termination factor 2 [OS=Homo sapiens]	0,02985	1,12819	1,032022	1	1	1	1	1162	129,508	8,37	0	1
	High	P35251-1	Replication factor C subunit 1 [OS=Homo sapiens]	0	5,5051	3,222965	3	4	3	1	1148	128,175	9,36	0	3

High	Q697Q0-1	Cytospin-A [OS=Homo sapiens]	0	19,2347	10,02858	10	18	10	1	11,17	124,525	5,72	13,4054443	10
High	Q9H2P0	Activity-dependent neuroprotector homeobox protein [OS=Homo sapiens]	0	4,9846	1,9056261	2	3	2	1	1102	123,485	7,34	1,74325895	2
High	P23370-1	Probable global transcription activator SNF2L1 [OS=Homo sapiens]	0	29,1232	14,516129	16	26	9	1	1054	122,527	8,09	23,9319128	16

Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMs	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pl	Score	Peptides Sequest HT
B4	Q16531	DNA damage-binding protein 1 [OS=Homo sapiens]	0	1,656	0,6140351	1	1	1	1	1140	126,887	5,26		0
	Q9Y5B9	FACT complex subunit SPT16 [OS=Homo sapiens]	0	236,278	56,3148	53	224	53	1	1047	119,838	5,66	421,827204	53
	P42285	Suppressor of viralicidic activity 2-like 2 [OS=Homo sapiens]	0	13,0166	7,0057582	6	9	6	1	1042	117,729	6,52	5,31437433	6
	Q9H0A0	N-acetyltransferase 10 [OS=Homo sapiens]	0	41,6117	20,487805	18	43	18	1	1025	115,657	8,27	35,1097795	18
	Q14527	Helicase-like transcription factor [OS=Homo sapiens]	0	17,9357	7,4333021	7	15	7	1	1009	113,857	8,6	11,3782876	7
	P03874	Poly [ADP-ribose] polymerase 1 [OS=Homo sapiens]	0	918,825	79,092702	101	1894	101	1	1014	113,012	8,88	5095,15786	101
	P49916-1	DNA ligase 3 [OS=Homo sapiens]	0	6,574	4,2616452	4	4	4	1	1009	112,835	9,01	1,73698843	4
	Q8N5C6	S1 RNA-binding domain-containing protein 1 [OS=Homo sapiens]	0	5,33347	2,9145729	3	3	3	1	995	111,705	8,72	4,23382056	3
	Q75400	pre-mRNA-processing factor 40 homolog A [OS=Homo sapiens]	0	1,90355	0,940389	1	1	1	1	957	108,737	7,56		0
	Q9Y2W1	Thyroid hormone receptor-associated protein 3 [OS=Homo sapiens]	0	2,09887	1,2565445	1	1	1	1	955	108,601	10,15		0
Medium	O96137-1	Putative RNA-binding protein 15 [OS=Homo sapiens]	0,01875	1,31885	1,9447288	1	1	1	1	977	107,124	10,08		0
High	Q9NYF8-1	Bcl-2-associated transcription factor 1 [OS=Homo sapiens]	0	9,44791	3,9130435	4	13	4	1	920	106,059	9,98	5,59469521	4
High	Q01831	DNA repair protein complementing XP-C cells [OS=Homo sapiens]	0	1,91222	0,7446809	1	1	1	1	940	105,887	8,9	1,74018943	1
High	Q9Y5B6	PAX3- and PAX7-binding protein 1 [OS=Homo sapiens]	0	1,52871	1,1995638	1	2	1	1	917	104,739	5,68		0
High	Q9Z889	DNA repair endonuclease XPF [OS=Homo sapiens]	0	561,256	69,432314	68	1005	68	1	916	104,42	6,93	2501,60384	68

Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMs	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pl	Score	Peptides Sequest HT
B5	Q94559	Cell division cycle 5-like protein [OS=Homo sapiens]	0	2,79891	2,1197007	2	2	2	1	802	92,194	8,18		0
	P13387	DNA topoisomerase 1 [OS=Homo sapiens]	0	62,2559	30,980392	22	74	22	1	765	90,669	9,31	74,3361579	22
	P19447	TFIIB basal transcription factor complex helicase XPB subunit [OS=Homo sapiens]	0	2,40935	2,8132992	2	2	2	1	782	89,221	7,23		0
	P18074-1	Tfiib basal transcription factor complex helicase xpd subunit [OS=Homo sapiens]	0	77,3474	45,789474	26	73	26	1	760	86,854	7,15	97,8966215	26
	P13010	X-ray repair cross-complementing protein 5 [OS=Homo sapiens]	0	222,706	65,437158	41	231	41	1	732	82,652	5,81	504,7268853	41
	Q92499	ATP-dependent RNA helicase DDX1 [OS=Homo sapiens]	0	9,10576	5	6	5	1	740	82,38	7,23	1,96563959	5	
	P49959-3	Isoform 3 of Double-strand break repair protein MRE11A [OS=Homo sapiens]	0	4,69188	2,6722925	2	2	2	1	711	81,02	6,15		0
	Q13330-1	Metastasis-associated protein MTA1 [OS=Homo sapiens]	0	16,36339	9,3700294	6	11	6	1	715	80,75	9,26	13,4378451	6
	Q92841	Probable ATP-dependent RNA helicase DDX17 [OS=Homo sapiens]	0	77,1622	37,722908	23	82	22	1	729	80,222	8,27	110,649328	23
	P00571	ATP-dependent RNA helicase DDX3X [OS=Homo sapiens]	0	3,41329	3,4743202	2	3	1	1	662	73,198	7,18	1,6029582	2
High	Q06787	Fragile X mental retardation protein 1 [OS=Homo sapiens]	0	3,11035	3,3227848	2	4	1	1	632	71,131	7,42	1,849424	2
Medium	P12956	X-ray repair cross-complementing protein 6 [OS=Homo sapiens]	0,02849	1,25088	1,6420361	1	2	1	1	609	69,759	6,64		0
High	P51114-1	Fragile X mental retardation syndrome-related protein 1 [OS=Homo sapiens]	0	3,38758	4,1867955	2	4	1	1	621	69,678	6,15	1,849424	2
High	P18887	DNA repair protein XRCC1 [OS=Homo sapiens]	0	120,491	40,442338	22	152	22	1	633	69,434	6,39	318,499228	22

Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMs	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pl	Score	Peptides Sequest HT
Medium	O94776	Metastasis-associated protein MTA2 [OS=Homo sapiens]	0,02457	1,68173	1,497006	1	1	1	1	668	74,976	9,66		0
High	P12956	X-ray repair cross-complementing protein 6 [OS=Homo sapiens]	0	293,384	66,502463	43	290	43	1	609	69,759	6,64	604,855505	43
High	P51114-1	Fragile X mental retardation syndrome-related protein 1 [OS=Homo sapiens]	0	23,0774	13,204509	8	18	7	1	621	69,678	6,15	14,782655	8

	High	P18887	DNA repair protein XRCC1 [OS=Homo sapiens]	0	104819	6,6350711	5	9	5	1	633	69,434	6,39	4,35622644	5
	High	P17844	probable ATP-dependent RNA helicase DDX5 [OS=Homo sapiens]	0	54,984	26,384365	15	59	8	1	614	69,105	8,92	98,3988343	15
<b>B6</b>	High	Q06787_9	Isoform 9 of Fragile X mental retardation protein 1 [OS=Homo sapiens]	0	56,2177	25,855247	13	42	1	1	611	68,924	7,42	60,3885313	13
High	Q9NUW8	tyrosyl-DNA phosphodiesterase 1 [OS=Homo sapiens]	0	13,9941	12,5	6	9	6	1	608	68,377	7,65	11,6622963	6	
High	P27694	Replication protein A 70 kDa DNA-binding subunit [OS=Homo sapiens]	0	7,25115	5,68,18182	3	3	3	1	616	68,095	7,21	4,03435028	3	
High	Q06787_2	Isoform 1 of Fragile X mental retardation protein 1 [OS=Homo sapiens]	0	57,1791	27,605428	13	44	1	1	594	66,93	7,23	58,2799848	13	
Medium	Q96IB3_1	hyppermethylated in cancer 2 protein [OS=Homo sapiens]	0,02338	1,99913	1,6260163	1	2	1	1	615	66,114	6,38	4,9058175	1	
High	Q86UE4	protein LYRIC [OS=Homo sapiens]	0	35,0493	17,697595	8	18	8	1	582	63,799	9,32	20,924956	8	

Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMS	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pI	Score	Peptides Sequest HT
High	Q96T60	Bifunctional polynucleotide phosphatase/kinase [OS=Homo sapiens]	0	29,0737	19,765674	9	25	9	1	521	57,04	8,46	26,465871	9
High	Q9UMS4	Pre-mRNA-processing factor 19 [OS=Homo sapiens]	0	57,0677	44,6442857	15	76	15	1	504	55,146	6,61	10,2,408488	15
High	Q53EL6_1	Programmed cell death protein 4 [OS=Homo sapiens]	0	4,36609	5,9701493	2	2	2	1	469	51,703	5,21	1,77377677	2
<b>B7</b>	P54127	UV excision repair protein RAD23 homolog B [OS=Homo sapiens]	0	3,73334	4,400978	2	3	2	1	409	43,145	4,84	0	2
High	P35659_1	Protein DEK [OS=Homo sapiens]	0	4,35982	5,0666667	2	7	2	1	375	42,648	8,56	5,44552088	2

Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMS	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pI	Score	Peptides Sequest HT
High	P18754_2	Isoform 2 of Regulator of chromosome condensation [OS=Homo sapiens]	0	100,901	44,026549	14	72	14	1	452	48,115	8,16	136,571179	14
High	O5VVV7	protein SX41P [OS=Homo sapiens]	0	5,71772	7,1078431	2	3	2	1	408	45,524	9,42	2,58791447	2
High	P31689_1	DnaJ homolog subfamily A member 1 [OS=Homo sapiens]	0	4,27253	9,5717884	2	2	2	1	397	44,839	7,08	0	2
High	P36559_1	Protein DEK1 [OS=Homo sapiens]	0,00521	1,263376	2,6666667	1	1	1	1	375	42,648	8,56	0	1
High	Q8IW50_1	PHD finger protein 6 [OS=Homo sapiens]	0	12,9506	24,9351507	7	14	7	1	365	41,264	8,68	11,8915551	7
High	P07992_3	Isoform 3 of DNA excision repair protein ERCC1 [OS=Homo sapiens]	0	13,5667	17,027864	5	17	5	1	323	35,541	8,82	22,9325138	5

Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMS	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pI	Score	Peptides Sequest HT
High	Q13888_1	General transcription factor IIH subunit 2 [OS=Homo sapiens]	0	13,0229	12,655228	5	12	5	1	395	44,39	6,76	15,955723	5
High	Q7ZZE3_1	Aprrataxin [OS=Homo sapiens]	0	18,5507	23,876404	6	22	6	1	356	40,714	9,17	33,1994686	6
High	P40938	replication factor C subunit 3 [OS=Homo sapiens]	0	26,1118	32,303371	8	17	8	1	356	40,53	8,34	12,86112	8
High	P35249	replication factor C subunit 4 [OS=Homo sapiens]	0	17,1125	22,865014	7	19	7	1	363	39,657	8,02	24,973071	7
<b>B9</b>	O9NWY4	UPF0609 protein C4orf27 [OS=Homo sapiens]	0	2,95832	7,2254335	2	4	2	1	346	39,411	6,8	0	2
High	P35250_1	Replication factor C subunit 2 [OS=Homo sapiens]	0	33,0394	38,135593	11	31	11	1	354	39,132	6,44	48,434146	11
High	P40937_1	Replication factor C subunit 5 [OS=Homo sapiens]	0	5,65723	9,7058824	3	5	3	1	340	38,472	7,2	3,63496196	3
High	P06746	DNA polymerase beta [OS=Homo sapiens]	0	56,6169	53,433836	18	58	18	1	335	38,154	8,95	96,6008033	18
High	P07992_3	Isoform 3 of DNA excision repair protein ERCC1 [OS=Homo sapiens]	0	46,1709	37,613	10	55	10	1	323	35,541	8,82	107,632045	10

Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMS	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pI	Score	Peptides Sequest HT
High	P40938	replication factor C subunit 3 [OS=Homo sapiens]	0	6,52059	9,8314607	3	4	3	1	356	40,53	8,34	1,71456885	3
High	P35249	replication factor C subunit 4 [OS=Homo sapiens]	0	46,314	47,107438	14	37	14	1	363	39,657	8,02	52,1136779	14

<b>B10</b>	High	P35250-1	Replication factor C subunit 2 [OS=Homo sapiens]	0	2,54741	8,4745763	2	3	2	1	354	39,132	6,444	0	2
	High	P40937-1	Replication factor C subunit 5 [OS=Homo sapiens]	0	28,3904	41,764706	12	25	12	1	340	38,472	7,2	24,0994389	12
	High	P06746	DNA polymerase beta [OS=Homo sapiens]	0	14,5683	26,268657	7	15	7	1	335	38,154	8,95	5,78062606	7
	High	P07992-1	DNA excision repair protein ERCC-1 [OS=Homo sapiens]	0	196,393	83,164983	26	282	26	1	297	32,542	6,25	644,410699	26
	High	P23025	DNA repair protein complementing XP-A cells [OS=Homo sapiens]	0	32,036	26,375626	5	18	5	1	273	31,348	6,73	38,937887	5

Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMs	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pI	Score Sequest HT	# Peptides Sequest HT
High	P18887	DNA repair protein XRCC1 [OS=Homo sapiens]	0,00559	1,11323	1,5797788	1	2	1	1	633	69,434	6,39	1,778366733	1
High	P27694	Replication protein A 70 kDa DNA-binding subunit [OS=Homo sapiens]	0	5,44427	7,7922078	4	5	4	1	616	68,095	7,21	6,93858244	4
High	Q9UGN5-1	Poly [ADP-ribose] polymerase 2 [OS=Homo sapiens]	0	5,76665	5,8319039	3	4	3	1	583	66,164	8,88	1,97141039	3
High	Q96IB3-1	Hypermethylated in cancer 2 protein [OS=Homo sapiens]	0	1,7073	1,6260163	1	2	1	1	615	66,114	6,38	2,33989549	1
High	P32780	General transcription factor IIH subunit 1 [OS=Homo sapiens]	0	44,0655	40,875912	16	43	16	1	548	61,993	8,66	77,8674933	16
High	Q13112	Chromatin assembly factor 1 subunit B [OS=Homo sapiens]	0	3,13674	3,9355993	2	2	2	1	559	61,454	7,5	0	2
High	Q96NV9	crossover junction endonuclease MUS81 [OS=Homo sapiens]	0,00559	0,81192	1,0886292	1	1	1	1	551	61,135	9,73	0	1

Protein FDR Confidence	Accession	Description	Exp. q-value	Sum PEP Score	Coverage	# Peptides	# PSMs	# Unique Peptides	# Protein Groups	# AAs	MW [kDa]	calc. pI	Score Sequest HT	# Peptides Sequest HT
<b>B12</b>	P23375-1	Lysine-specific demethylase 5A [OS=Homo sapiens]	0	0,59172	1	1	1	1	1	1690	191,97	6,49	0	1
	Q92623	Tetratricopeptide repeat protein 9A [OS=Homo sapiens]	0	4,05405	5	1	1	1	1	222	24,364	8,98	0	1
	Q129121	Lymphoid-restricted membrane protein [OS=Homo sapiens]	0	1,44144	1	4	1	1	1	555	62,083	5,85	0	1
	P19237	Tropomodulin I, slow skeletal muscle [OS=Homo sapiens]	0	4,27807	1	3	1	1	1	187	21,679	9,58	6,1745	1
	Q9NVE4	Coiled-coil domain-containing protein 87 [OS=Homo sapiens]	0	0,8245	1	1	1	1	1	849	96,342	8,59	0	1

**Annex Table II: Mass spectrometry data from XPF-HA purification from individual bands of SDS gel.** Interactors of XPF purified samples with UV irradiation (50J/m<sup>2</sup>; 1h of recovery) (Band 11; B11) or under MMC treatment (1.5 μM, 16h) (the rest of bands except B12) are shown here. (B12 came from a gel fragment without protein band, negative control). Proteins identified in each single band are ordered according to the MW expected from the SDS gel band size. Q-value: minimal false discovery rate at which the identification is considered correct. Posterior Error Probability (PEP) score: probability that the observed peptide spectrum matches (PSM) is incorrect. Coverage: percentage of the protein sequence covered by identified peptides (confident coverage threshold >1). Peptides: total number of distinct peptide sequences identified in the protein group. Unique peptides: The number of peptide sequences that are unique to a protein group. Protein Groups: number of proteins displayed in the Protein Group Members view. Sequest-HT: Algorithm to identify collections of tandem mass spectra to peptide sequences that have been generated from databases of protein sequences.

Protein FDR Confidence	Accession	Description	Sum PEP Score	Coverage	# Peptides	# PSMs	# Unique Peptides	# AAs	MW [kDa]	Gene ID	Abundance Ratio (log2): [Medium]/[Light]	Abundance Ratio (log2): [Heavy]/[Light]	Abundance Ratio (log2): [Medium]/[Heavy]	# Razor Peptides
High	O56816	BAG family molecular chaperone regulator 2 [OS=Homo sapiens]	6.852504856	11.37440706	2	2	211	23.757	BAG2	10.82	11.37	-0.55	0	0
High	O56281	Beta-actin-like protein 2 [OS=Homo sapiens]	133.465686	17.8191189	8	55	1	376	41.976	ACTB12	11.17	11.17	0	0
High	P26295	DNA repair protein complementing XP-A cells [OS=Homo sapiens]	5.55021313	7.69230769	1	1	1	273	31.348	XPA	8.08	9.72	-1.64	0
Low	O92552	Isotform 2 of 28S ribosomal protein S27, mitochondrial [OS=Homo sapiens]	1.95506845	2.57009346	1	1	1	428	49.119	MRPS27	8.72	8.87	-0.15	0
Medium	O75165	DnaJ homolog subfamily C member 13 [OS=Homo sapiens]	2.98399909	0.49041462	1	2	1	2243	254.252	DNAJC13	8.22	8.24	-0.02	0
High	P07992	DNA excision repair protein ERCC-1 [OS=Homo sapiens]	369.8239473	62.6262626	25	140	25	297	32.542	ERCC1	4.67	4.79	-0.12	0
High	Q70790	acyl-CoA dehydrogenase family member 11 [OS=Homo sapiens]	94.1642326	26.4102564	18	29	18	780	87.228	ACAD11	4.18	4.54	-0.37	0
High	P49759	Isotform 3 of Dual specificity protein kinase CLK1 [OS=Homo sapiens]	109.692773	37.0722433	17	34	15	526	61.718	CLK1	1.47	4.31	-5.78	2
High	Q8V192	Structure-specific endonuclease subunit SLX4 [OS=Homo sapiens]	643.363168	53.655217	91	232	90	1834	199.989	SLX4	4.13	4.29	-0.17	2
High	Q28899	DNA repair endonuclease XRF-1 [OS=Homo sapiens]	1100.2576	75.3275109	84	842	84	916	104.42	ERCC4	4.24	4.27	-0.02	0
High	Q96C7	Isocitrate/isozyme domain-containing protein 1 [OS=Homo sapiens]	19.9897381	22.4832115	4	5	4	298	32.216	ISOC1	3.2	4.14	-0.94	0
Medium	Q7272W4	Zinc finger CCH-type antiviral protein 1 [OS=Homo sapiens]	4.5168.1823	2.2172949	2	2	2	902	101.367	ZCHAV1	0.85	4.03	-3.17	0
High	Q9H1R3	Mysin light chain kinase 2, skeletal/cardiac muscle [OS=Homo sapiens]	22.7916367	1.84563758	2	21	2	596	64.644	MLYK2	3.56	3.78	-0.22	0
Low	Q9BPX3	Condensin complex subunit 3 [OS=Homo sapiens]	0.97183558	0.38521267	1	1	1	1015	114.262	NAPG	-1.41	3.66	-5.07	0
High	P19338	Nucleolin [OS=Homo sapiens]	59.6385982	21.2676056	14	17	14	710	76.568	NCL	1.72	3.01	-1.29	0
Low	Q14956	Transmembrane glycoprotein NMB [OS=Homo sapiens]	1.09658441	3.14685315	1	1	1	572	63.882	GNMB	2.19	2.79	-0.6	0
High	Q96HS1	Serine/threonine-protein phosphatase Pgam5, mitochondrial [OS=Homo sapiens]	59.1695502	14	26	14	289	31.985	PGAM5	2.84	2.44	0.41	0	
High	Q43390	Isotform 2 of Heterogeneous nuclear ribonucleoprotein R [OS=Homo sapiens]	11.596465	6.91836399	4	6	4	636	71.17	HNRNPR	-0.24	2.36	-2.6	0
Medium	Q8R1L5	Isotform 2 of Cytochrome c oxidase protein 20 homolog [OS=Homo sapiens]	4.29174911	10.7692308	1	1	1	130	146.659	COK2D0	2.33	2.36	-0.03	0
High	Q6P5K6	60S ribosomal protein L21-like 1 [OS=Homo sapiens]	85.4034051	64.7540984	8	47	7	122	145.98	RPL2L1	2.41	2.31	0.11	1
High	O60934	Nibrin [OS=Homo sapiens]	5.75092142	3.0509788	3	3	3	754	84.906	NBN	1.41	2.06	-0.65	0
High	O15460	Prolyl 4-hydroxylase subunit alpha 2 [OS=Homo sapiens]	6.50951015	3.73831776	2	2	2	535	60.864	RPHA2	2.53	2.04	-0.51	0
High	P18077	60S ribosomal protein L35a [OS=Homo sapiens]	15.3926509	40	6	9	6	110	12.53	RPL35A	1.18	1.92	-0.74	0
High	P05387	60S acidic ribosomal protein P2 [OS=Homo sapiens]	23.3201849	33.9130395	3	7	3	115	11.658	RPLP2	1.54	1.91	-0.37	0
High	Q16480	Protein RPLP5 homolog [OS=Homo sapiens]	4.76666781	1.65686799	3	3	3	181	104.026	RPLD1	0.96	1.91	-0.95	0
High	Q22878	60S ribosomal protein L6 [OS=Homo sapiens]	40.204729	48.6111111	15	29	15	288	32.708	RPL16	1.02	1.82	-0.8	0
High	Q96EV4	translational machinery-associated protein 16 [OS=Homo sapiens]	70.7767715	46.3054187	11	22	11	203	23.849	TMA16	0.4	1.79	-1.39	0
High	P49327	Fatty acid synthase [OS=Homo sapiens]	22.111601	26.3639884	55	76	54	2511	27.254	FASN	0.14	1.78	-1.64	2
High	Q43844	Putative GFP-binding protein [OS=Homo sapiens]	22.03315609	12.0155039	6	7	6	516	56.948	RPLP6	1.72	1.75	-0.03	0
High	Q06787	fragile X mental retardation protein 1 [OS=Homo sapiens]	28.3418144	9.49367089	5	9	1	632	71.131	FMR1	1.3	1.74	-0.43	4
High	P26578	60S ribosomal protein L4 [OS=Homo sapiens]	41.3524971	21.54566774	9	20	9	427	47.667	RPL4	0.93	1.72	-0.78	0
High	P67021	ribosomal L1 domain-containing protein 1 [OS=Homo sapiens]	7.85722872	6.32653061	2	2	2	490	54.939	RSL1D1	0.83	1.68	-0.84	0
High	P49599	Isotform 3 of Double-strand break repair protein MRCE1A [OS=Homo sapiens]	9.679331699	7.87632066	4	4	4	711	81.02	MRCE1A	0.67	1.66	-1	0
Low	Q9H1X3	dnal homolog subfamily C member 25 [OS=Homo sapiens]	1.053646	2.22222222	1	1	1	360	42.377	DNAJC25	1.52	1.66	-0.14	0
Low	Q30099	Ubiquitin carbony-terminal hydrolase 7 [OS=Homo sapiens]	22.0315609	12.0155039	7	6	6	516	56.948	RPLP6	1.72	1.75	-0.03	0
High	Q06477	Isotform 9 of Eukaryotic translation initiation factor 4 gamma 1 [OS=Homo sapiens]	28.3418144	9.49367089	3	3	3	632	71.131	FMR1	1.3	1.74	-0.43	4
High	P26578	60S ribosomal protein L4 [OS=Homo sapiens]	41.3524971	21.54566774	9	20	9	427	47.667	RPL4	0.93	1.72	-0.78	0
High	P67021	ribosomal L1 domain-containing protein 1 [OS=Homo sapiens]	7.85722872	6.32653061	2	2	2	490	54.939	RSL1D1	0.83	1.68	-0.84	0
High	P49599	Isotform 3 of Double-strand break repair protein MRCE1A [OS=Homo sapiens]	9.679331699	7.87632066	4	4	4	711	81.02	MRCE1A	0.67	1.66	-1	0
Medium	P04075	Isotform 2 of Fructose-bisphosphate aldolase A [OS=Homo sapiens]	12.4608942	10.5263158	4	4	4	418	45.232	ALDOA	-0.33	1.52	1.66	-0.14
Medium	P05455	Lipid La protein [OS=Homo sapiens]	3.50473236	3.43323755	1	1	1	408	46.928	SLSB	0.37	1.56	-1.19	0
High	Q9BRG2	SH3 domain-containing protein 3A [OS=Homo sapiens]	7.08464757	5.38194444	4	4	4	576	63.054	SH2D3A	1.6	1.54	0.07	0
Medium	P52294	Importin subunit alpha 5 [OS=Homo sapiens]	4.39328385	3.15985513	2	2	2	538	60.184	KNAA1	2.28	1.63	-5.96	0
High	P61313	protein AF-9 [OS=Homo sapiens]	30.3092459	22.5490196	9	4	4	204	24.131	RPL15	0.93	1.61	-0.77	0
High	P40475	putative helicase MOY-10 [OS=Homo sapiens]	3.97676778	2.293312064	2	2	2	1003	11.3599	MILT11	0.29	1.5	-1.92	0
Medium	P60832	H/ACA ribonucleoprotein complex subunit 4 [OS=Homo sapiens]	3.14736602	3.69649805	2	2	2	514	57.638	DKC1	0.82	1.49	-0.67	0
High	Q81ZL8	Proline-, glutamic acid- and leucine-rich protein 1 [OS=Homo sapiens]	5.0323952	5.08140708	2	3	2	1130	119.624	PELP1	-0.58	1.47	-2.06	0
Medium	P5386	acidic ribosomal protein P1 [OS=Homo sapiens]	4.9097426	14.0538087	1	2	1	114	115.075	RPLP1	0.84	1.47	-0.62	0
High	Q9H2U1	ATP-dependent RNA helicase DHX36 [OS=Homo sapiens]	3.79692681	1.98412989	2	2	2	1008	114.688	DHX36	0.25	1.44	-1.18	0
High	P72277	Protein disulfide-isomerase [OS=Homo sapiens]	23.875236	11.023622	5	7	5	508	57.081	P4HB	.29	1.43	0.35	0
High	P12277	Creatine kinase B-type [OS=Homo sapiens]	28.9101846	21.7847769	6	8	6	381	42.617	CKB	-1.9	1.41	-3.31	0
High	P83396	ATP-citrate synthase [OS=Homo sapiens]	83.1565296	22.3432343	19	28	19	1101	120.762	ACLY	-0.7	1.39	-2.09	0
High	P50914	60S ribosomal protein 13 [OS=Homo sapiens]	24.378671	15.8139353	3	7	3	215	23.417	RPL14	0.9	1.38	-0.47	0
High	A11010	Acetolactate synthase-like protein [OS=Homo sapiens]	6.16887451	4.27225159	2	2	2	632	67.825	IILVBL	1.37	1.38	0	0
High	P05388	60S ribosomal protein P0 [OS=Homo sapiens]	33.0680237	24.6056782	7	11	7	317	34.252	RPLP0	0.58	1.27	-0.69	0
Medium	P40429	60S ribosomal protein 13a [OS=Homo sapiens]	3.08338731	7.881734	2	2	2	203	23.562	RPL3A	0.7	1.27	-0.57	0
Low	O75600	Isotform 2 of 2-amino-3-ketobutyrate coenzyme A ligase, mitochondrial [OS=Homo sapiens]	1.08620327	2.24719101	1	1	1	445	47.944	GCAT	1.08	1.27	-0.2	0
Low	P24241	Transcription elongation factor II polypeptide 3 [OS=Homo sapiens]	2.21245868	0.87719298	1	1	1	798	89.853	TCEB3	0.63	1.26	-0.63	0
Medium	P51784	Ubiquitin carbonyl-terminal hydrolase 11 [OS=Homo sapiens]	4.61891396	2.01068536	2	2	2	103.147	USP11	0.96	1.25	-0.28	0	
Medium	Q9h583	HEAT repeat-containing protein 1 [OS=Homo sapiens]	3.6471774	1.07276119	2	2	2	2144	242.215	HEATR1	-0.04	1.25	-1.29	0
High	Q00839	Heterogeneous nuclear ribonucleoprotein U [OS=Homo sapiens]	108.99708	21.33333333	19	34	18	825	90.528	HNRPNU	0.11	1.23	-1.12	1
High	P25205	Isotform 2 of DNA replication licensing factor MCM3 [OS=Homo sapiens]	26.8692348	7.97186401	6	10	6	853	95.848	MCM3	0.2	1.22	-1.02	0

Medium	Q95985	DNA topoisomerase 3-beta-1 [OS=Homo sapiens]	3.98000686	1.85614849	2	2	862	96.599	TOP3B	.102	1.21	-0.19	0	
High	P42424	60S ribosomal protein L7a [OS=Homo sapiens]	78.4262425	41.7293233	11	11	266	29.977	RPL7A	.038	1.19	-0.81	0	
High	Q92978_2	Isoform 2 of DNA repair protein RAD50 [OS=Homo sapiens]	53.189161	16.09391047	21	21	1318	154.191	0.56	1.19	-0.53	0		
High	P18124	60S ribosomal protein L7 [OS=Homo sapiens]	46.602609	39.516129	8	17	8	248	29.207	RPL7	0.49	1.19	-0.7	0
High	H07020	60S ribosomal protein L18 [OS=Homo sapiens]	48.7779873	36.1702128	6	11	6	188	21.621	RPL18	0.48	1.17	-0.69	0
High	Q75616_1	GTPase Era, mitochondrial [OS=Homo sapiens]	10.7406698	6.63615561	2	4	2	437	48.319	ERAL1	.58	1.15	0.43	0
High	Q8Y6N4	Cytochrome c oxidase assembly protein COX11, mitochondrial [OS=Homo sapiens]	10.3175539	12.3188406	2	3	2	276	31.41	COX11	.28	1.15	0.12	0
High	P02786	Transferrin receptor protein 1 [OS=Homo sapiens]	47.6267254	14.2105263	9	16	9	760	84.818	TFRC	.101	1.12	-0.11	0
High	P06737_1	Glycogen phosphorylase, liver form [OS=Homo sapiens]	50.5342359	15.7024793	12	16	12	847	97.087	PYGL	-.97	1.11	-2.07	0
Medium	Q99832	T-complex protein 1 subunit 2 [OS=Homo sapiens]	4.34288829	2.946533	2	2	543	59.329	CC7	-.38	1.11	-1.49	0	
High	Q08211	ATP-dependent RNA helicase 1 [OS=Homo sapiens]	82.6228654	14.094488	15	26	15	1270	140.869	DHH9	-.049	1.1	-1.59	0
Low	Q13907_2	Isoform 2 of isopentenyl-diphosphate Delta-isomerase 1 [OS=Homo sapiens]	2.09057406	3.165901408	1	1	1	284	32.465	ID1	0.79	1.1	-0.31	0
High	P99438	Probable ATP-dependent RNA helicase DDX28 [OS=Homo sapiens]	89.9438566	42.4074074	19	32	19	540	59.545	DDX28	-.28	1.09	0.19	0
High	H07233_2	Isoform 2 of Putative ATP-dependent RNA helicase DHX30 [OS=Homo sapiens]	11.093543	3.737315876	3	5	3	1222	136.03	DHX30	0.47	1.09	-0.62	0
High	Q9NR30_1	Nucleolar RNA helicase 2 [OS=Homo sapiens]	63.0303361	19.412516	14	24	11	783	87.29	DX21	0.47	1.08	-0.62	0
High	P05253	60S ribosomal protein L18a [OS=Homo sapiens]	11.2915939	26.1363636	5	5	5	176	20.749	RPL18A	.063	1.07	-0.44	0
Medium	P08175_1	RNA-binding protein 10 [OS=Homo sapiens]	2.55783391	0.75268817	1	1	1	930	103.469	RB10	-.008	1.07	-1.14	0
Low	Q81D6N6	Ribosome biogenesis protein BRXL homolog [OS=Homo sapiens]	2.22076437	1.88300283	1	1	1	353	41.375	BRLX1	0.11	1.05	-0.94	0
Low	A0D1E9_1	GTP-binding protein 10 [OS=Homo sapiens]	1.76776648	2.3253584	1	1	1	387	42.906	GTPBP10	.089	1.05	-0.16	0
High	P98YF8_1	Bcl-2-associated transcription factor 1 [OS=Homo sapiens]	133.332621	33.030435	28	51	26	920	106.059	BCLAF1	.078	1.04	-0.26	2
High	P35680	AP-2 complex-subunit 1 sigma [OS=Homo sapiens]	6.54311266	10.5633803	2	3	2	142	17.007	AP2S1	.033	1.04	-0.21	0
Low	Q5L717_2	Isoform 2 of ELAV-like protein 1 [OS=Homo sapiens]	1.58703573	2.54957507	1	1	1	353	38.972	EFLV1	0	1.03	-1.03	0
High	P22102_1	Trifunctional purine biosynthetic protein adenosine-3' [OS=Homo sapiens]	13.6824871	6.63960296	6	7	6	1010	107.699	GART	-.036	1.02	-1.88	0
High	P08170	Serinearginine-rich splicing factor 4 [OS=Homo sapiens]	6.90898988	6.66801619	3	4	3	494	56.645	SRSF4	0.18	1.02	-0.84	0
High	H053H1_2	Borealin [OS=Homo sapiens]	5.5438905	6.78571429	2	2	2	280	31.304	CDC48	-.47	1.02	-1.49	0
High	Q9H0D5_1	5'-3' exonuclelease 2 [OS=Homo sapiens]	5.58731186	2.52631579	2	2	2	950	108.513	XRN2	0.47	1	-0.53	0
High	P29023	60S ribosomal protein L3 [OS=Homo sapiens]	87.4042255	39.7022333	15	26	15	403	46.08	RPL3	.053	0.97	-0.44	0
High	P55795	heterogeneous nuclear ribonucleoprotein H2 [OS=Homo sapiens]	30.3362148	23.362148	8	10	2	49	49.232	HNRPH2	0.04	0.97	-0.53	0
High	P06744_2	Isoform 2 of Glucosidase phosphatase isomerase 1 [OS=Homo sapiens]	24.9978941	11.0720562	6	8	6	569	64.284	GPI	-.204	0.97	-3.01	0
High	P22087	tRNA 2'-O-methyltransferase fibrillarin [OS=Homo sapiens]	12.8098375	14.3302181	3	3	3	321	33.763	FBL	0.04	0.97	-0.32	0
High	Q8WIC1	MAP7 domain-containing protein 3 [OS=Homo sapiens]	57.7526684	16.7808219	13	20	13	876	98.368	MAP7D3	.03	0.95	0.08	0
High	H12659	Elavl-binding factor 2 [OS=Homo sapiens]	73.1288712	20.39762021	15	24	14	858	95.277	EF2	.031	0.94	-0.53	0
High	Q92900	Regulator of nonsense transcripts 1 [OS=Homo sapiens]	54.720061	18.5119575	19	21	19	1129	124.267	UPF1	0.02	0.94	-0.92	0
High	P56270_2	Isoform 2 of Myc-associated zinc finger protein [OS=Homo sapiens]	27.5303771	13.3874239	6	9	6	493	51.04	MALZ	0.79	0.94	-0.15	0
High	P07676	Uncharacterized protein C2orf47, mitochondrial [OS=Homo sapiens]	10.8446663	4.81096656	1	2	1	291	35.524	C2orf47	0.98	0.94	0.04	0
High	P12532_2	Isoform 2 of Creatine kinase U-type, mitochondrial [OS=Homo sapiens]	5.99960545	7.8125	2	2	2	448	50.389	[MNT1A; CRMT1]	-.162	0.94	-2.56	0
High	Q96AY2_2	Isoform 2 of Crossover junction endonuclease EME1 [OS=Homo sapiens]	18.9419538	9.7701544	6	9	6	583	64.738	EME1	1	0.92	0.08	0
High	P08170	Regulator of nonsense transcripts 1 [OS=Homo sapiens]	27.5303771	13.3874239	12	17	12	465	52.677	UPF1	0.02	0.94	-0.92	0
High	Q13085_4	Isoform 4 of Acetyl-CoA-carboxylase 1 [OS=Homo sapiens]	5.20645921	1.5107008	3	3	3	283	47.685	AHCY	-.838	0.91	-0.15	0
High	H040402	Emerin [OS=Homo sapiens]	49.0795040	8.21591685	8	13	8	254	82.333	ACACA	0.31	0.9	-0.59	0
High	H07043	Mitochondrial ribonuclease protein 1 [OS=Homo sapiens]	30.5471088	24.8123058	8	13	8	403	47.217	TRMT1OC	.005	0.88	0.17	0
High	P46777	60S ribosomal protein L5 [OS=Homo sapiens]	38.9390321	29.6296296	8	13	8	297	34.341	RPL5	-.025	0.87	-1.12	0
High	P38159_1	RNA-binding motif protein X chromosome [OS=Homo sapiens]	37.0054949	16.3582864	6	13	6	391	42.306	RBMX	0.21	0.87	-0.65	0
High	P33992	DNA replication licensing factor ncm5 [OS=Homo sapiens]	29.7339159	12.8065395	8	13	8	734	82.233	MCMV5	0.56	0.87	0.33	0
High	P76768	Gen-associated protein 4 [OS=Homo sapiens]	6.36507932	1.79584121	2	3	2	1058	119.96	GEMIN4	0.54	0.87	-0.33	0
High	P0DMV8	heat shock 0 kDa protein 1A [OS=Homo sapiens]	371.935961	53.1981279	37	13	31	641	70.009	SP18; HSPA1	.086	0.86	0.01	0
High	P11940_1	Polypeptidate-binding protein 1 [OS=Homo sapiens]	120.75939	32.3839971	18	34	18	636	70.626	PABPC1	0.3	0.85	-0.57	0
High	P0U4F8	Inslulin receptor substrate 4 [OS=Homo sapiens]	212.672866	31.9809696	34	103	34	1257	133.685	IRSp4	0.98	0.85	0.13	0
Low	O14979_1	Heterogeneous nuclear ribonucleoprotein D-like [OS=Homo sapiens]	1.84042281	1.9047619	1	2	1	420	46.409	IRNPDL; HNR	-.025	0.85	-1.1	0
High	P62906	60S ribosomal protein L10A [OS=Homo sapiens]	26.4217161	27.1889401	6	11	6	217	24.816	RPL10A	0.46	0.84	-0.38	0
High	Q9BX55_2	Isoform 2 of Ap-1 complex subunit mu-1 [OS=Homo sapiens]	10.2764377	6.88655172	4	5	4	435	49.809	AP1M1	0.51	0.84	-0.32	0
High	P82272	Heterogeneous nuclear ribonucleoprotein M [OS=Homo sapiens]	136.702608	39.3150585	27	60	27	730	77.464	HNRPNM	0.36	0.82	-0.46	0
High	Q92947_1	Glutaryl-CoA dehydrogenase, mitochondrial [OS=Homo sapiens]	7.20032097	4.33789954	2	2	2	438	48.096	GCDH	0.57	0.82	-0.25	0
High	P46940	Ras GTPase-activating-like protein IQGAP1 [OS=Homo sapiens]	14.0226222	3.433995172	5	7	5	1657	189.134	IQGAP1	-.7	0.81	-1.51	0
Medium	Q92979_1	Symplekin [OS=Homo sapiens]	10.2764377	6.88655172	2	2	2	1274	141.059	SyMPK	-.058	0.8	-1.39	0
High	Q86N99	crossover junction endonuclease MUS31 [OS=Homo sapiens]	41.6275074	18.5117967	9	13	9	551	61.135	MUS31	0.71	0.79	-0.38	0
High	P84090	Enhancer of rudimentary homolog [OS=Homo sapiens]	32.8473623	36.5384615	4	12	4	104	12.251	ERH	0.51	0.79	-0.29	0
High	Q45656	DNA replication licensing factor MCm6 [OS=Homo sapiens]	31.9595037	11.6393072	10	12	10	821	92.831	MCM6	0.03	0.79	-0.76	0
High	Q92948_2	Isoform 2 of Mitochondrial import inner membrane translocase subunit Tim50	30.9981278	10.9649123	5	9	5	456	50.433	TIMM50	0.28	0.78	-0.5	0
Low	P50570_1	Dynamamin-2 [OS=Homo sapiens]	1.90235967	2.18390805	1	1	1	870	98.003	DNM2	-0.29	0.78	-1.07	0
High	P21333	Filamin-A [OS=Homo sapiens]	7.49881062	2.15338119	4	4	4	2647	280.564	F11A	-0.49	0.77	-1.26	0
Low	P62714	serine/threonine-protein phosphatase 2A catalytic subunit beta isoform [OS=Hc]	1.9722428	2.58898676	1	1	1	309	35.552	PP2CB	-.005	0.75	-0.8	0

Medium	Q13459-1	Unconventional myosin-Xb [OS=Homo sapiens]	4,430640423	1,854427245	3	3	3	2157	243,249	M109B	0.61	0.74	-0.12	0
Medium	P075419-3	Isotform 3 of Cell division control protein 45 homolog [OS=Homo sapiens]	3,9665794624	1,8353946488	1	1	1	598	68,724	CDC45	0.39	0.74	-0.35	0
High	Q96EV1-1	DnaJ homolog subfamily A member 3, mitochondrial [OS=Homo sapiens]	59,00738841	25,83333333	11	16	11	480	52,456	DNAJA3	0.98	0.72	0.26	0
High	Q96MU7-1	YTH domain-containing protein 1 [OS=Homo sapiens]	31,2702196	13,6176066	10	14	10	727	84,649	YTHDC1	-0.08	0.72	-0.81	0
High	P50395-1	Rab GDP dissociation inhibitor beta [OS=Homo sapiens]	8,65867616	8,533923284	3	3	3	445	50,631	GDI2	-1.61	0.71	-2.32	0
High	Q14961	Small subunit processive component 20, mitochondrial [OS=Homo sapiens]	8,355240881	1,04129264	2	4	2	2785	318,182	UTP20	0.9	0.71	0.19	0
High	P128288	Cytochrome b-c1 complex subunit 8 [OS=Homo sapiens]	29,063243	35,3658337	4	8	4	82	9,9	UQCRC	0.9	0.7	0.2	0
High	P001648	ATP-binding cassette sub-family D member 3 [OS=Homo sapiens]	7,09154458	2,8831563	2	4	2	659	75,428	AEDD3	0.62	0.7	-0.08	0
High	P06748	ATP-dependent RNA helicase DDX39A [OS=Homo sapiens]	29,49448252	13,5831582	6	12	6	427	49,098	DIX39A	0.58	0.69	-0.11	0
High	Q9UNM00-1	Nucleophosmin [OS=Homo sapiens]	16,296628	19,7278912	5	7	5	294	32,555	NPM1	-0.3	0.69	-0.99	0
High	Q9P016	39S ribosomal protein S36, mitochondrial [OS=Homo sapiens]	5,36073646	13,592233	2	4	2	103	11,777	MRPL36	0.08	0.69	-0.61	0
Low	Q14157-5	Isotform 5 of Ubiquitin-assortilated protein 2-like [OS=Homo sapiens]	1,95108901	0,81521739	1	1	1	1104	116,369	UBAP2L	-1.23	0.69	-1.91	0
High	P17844	Probable ATP-dependent RNA helicase DDX5 [OS=Homo sapiens]	69,2435524	23,7785016	15	24	10	614	69,105	DDX5	0.19	0.68	-0.49	0
High	P000567	Nucleolar protein 56 [OS=Homo sapiens]	11,4611035	6,22895623	3	4	3	594	66,009	NDP56	0.7	0.68	0.02	0
Low	Q9UNM00-2	Transmembrane and coiled-coil domain-containing protein 1 [OS=Homo sapien	1,4,4819149	4,25531915	1	1	1	188	21,161	TMC01	0.48	0.68	-0.2	0
High	P202020	Plasma membrane calcium-translocating ATPase 1 [OS=Homo sapiens]	13,0283900	4,45151033	4	5	4	1258	138,668	ATPBP1	-0.25	0.67	-0.92	0
High	Q9L233	Non-POU domain-containing octamer-binding protein [OS=Homo sapiens]	6,93734064	6,7940552	3	3	2	471	54,197	NONO	-0.29	0.67	-0.96	1
Medium	Q13620	Cullin-4B [OS=Homo sapiens]	4,23202126	2,08105148	2	3	2	913	103,916	CUL4B	-0.85	0.67	-1.53	0
Medium	P78371-1	T-complex protein 1 subunit beta [OS=Homo sapiens]	3,08181234	2,80373832	1	1	1	535	57,452	CCT7	-0.54	0.66	-1.19	0
Low	Q6KCM7-3	Isotform 3 of Calcium-binding mitochondrial carrier protein SCaMC-2 [OS=Homo sapien	1,05223239	1,55339806	1	1	1	515	56,846	SLC25A25	0.18	0.66	-0.49	0
High	P25398	40S ribosomal protein S12 [OS=Homo sapiens]	14,765532	12,878979	2	6	2	132	14,605	RP512	-0.2	0.65	-0.85	0
High	Q09161	Nuclear cap-binding protein subunit 1 [OS=Homo sapiens]	18,3601143	6,83544304	5	6	5	790	91,781	NCBP1	-0.19	0.64	-0.82	0
High	P088431	Histone H3.1 [OS=Homo sapiens]	16,4936742	47,7941176	5	11	5	136	15,394	T1H3H; HIST	-0.03	0.64	-0.67	0
Medium	Q9Y312	Protein AA2 [OS=Homo sapiens]	2,32239395	1,82291667	1	1	1	384	43,444	AAR2	-0.24	0.64	-0.89	0
High	P07900-2	Isotform 2 of Heat shock protein HSP 90 alpha [OS=Homo sapiens]	235,821952	42,0374707	34	68	19	854	98,039	HSP90AA1	-0.87	0.63	-1.49	16
Low	Q9Y483	Metal-response element-binding transcription factor 2 [OS=Homo sapiens]	2,24078557	1,68634064	1	1	1	593	67,063	MTF2	0.56	0.63	-0.07	0
High	P41252	Isoleucine-tRNA ligase, cytoplasmic [OS=Homo sapiens]	47,4870102	8,003316957	10	23	10	1262	144,406	LRPS	-0.24	0.62	-0.85	0
High	P67809	Nuclease-sensitive element-binding protein 1 [OS=Homo sapiens]	10,0952	14,1975309	4	4	4	324	32,593	YBX1	0.12	0.62	-0.51	0
High	Q8NVDV7	Trinucleotide repeat-containing gene 6A protein [OS=Homo sapiens]	14,2993901	4,688909276	7	8	7	1962	210,169	TNR6A	0.75	0.61	0.15	0
High	P49593	Protein phosphatase 1F [OS=Homo sapiens]	31,9034703	21,5859031	7	9	7	454	49,8	PPM1F	0.7	0.59	0.11	0
Medium	Q9YNM3	50S ribosomal protein L11, mitochondrial [OS=Homo sapiens]	4,42950899	5,10948905	1	2	1	137	15,373	MRPL41	0.65	0.59	0.06	0
High	P2050-1	60S ribosomal protein L12 [OS=Homo sapiens]	16,7215657	42,4242924	5	6	5	165	17,808	RP112	-0.18	0.58	-0.4	0
High	P04843	Dolichyl-diphosphooligosaccharide-protein glycosyltransferase subunit 1 [OS=Homo sapiens]	1,798014377	2,65450462	2	3	2	607	68,527	RPN1	-0.36	0.58	-0.94	0
High	Q9E751-1	DNA-directed RNA polymerase I subunit RPA49 [OS=Homo sapiens]	6,6670128	6,02910603	3	3	3	481	53,928	POLR1E	-1.47	0.58	0.89	0
Medium	Q9Y379	Nucleolar complex protein 1, homolog [OS=Homo sapiens]	3,88243468	1,201670214	1	2	1	749	84,866	NOC2L	0	0.58	-0.58	0
High	P07834	Bifunctional glutamate/proline-tRNA ligase [OS=Homo sapiens]	72,7000529	14,9470899	20	33	20	1512	170,483	EPRLS	-0.35	0.57	-0.91	0
High	P09651-1	Heterogeneous nuclear ribonucleoprotein A1 [OS=Homo sapiens]	21,6208456	15,0537634	4	5	4	372	35,723	HNRNPA1	-1.19	0.57	-2.47	0
High	Q9ET42-1	ATP-dependent zinc metalloprotease YME1L1 [OS=Homo sapiens]	16,7215657	42,4242924	5	6	5	165	17,808	RP112	-0.18	0.58	-0.4	0
High	Q9Y4833	Isotform 1 of Importin 2 [OS=Homo sapiens]	138,395671	8,73185312	37	54	37	4684	531,466	YME1L1	0.42	0.57	-0.16	0
High	Q9UNX3	60S ribosomal protein L26-like 1 [OS=Homo sapiens]	67,5424633	51,7241379	11	34	3	145	17,246	RP126L1	0.62	0.56	-0.33	0
Medium	Q53366-1	Ubiquitin-protein ligase E3C [OS=Homo sapiens]	4,61078263	2,216065648	3	3	3	1083	123,344	UBC3C	0.49	0.56	-0.07	0
Medium	Q9Y26-2	Isotform 2 of Importin 11 [OS=Homo sapiens]	4,28029914	0,38522167	1	2	1	1015	116,932	IPQ11	-0.32	0.56	-0.89	0
High	P03210-2	Isotform 2 of AR2 complex subunit beta [OS=Homo sapiens]	54,8470912	20,1892244	19	26	19	951	105,625	AP2B1	0.2	0.55	0.35	0
High	P533007	Tricarboxylate transport protein, mitochondrial [OS=Homo sapiens]	108,070017	34,7266881	10	41	10	311	33,991	SLC25A1	0.69	0.54	0.16	0
High	Q9BYV4	Nucleolar complex protein 1 [OS=Homo sapiens]	15,7158570	10,6589147	4	6	4	516	58,431	NOCAL	0.88	0.54	0.34	0
High	P04181-1	Ornithine aminotransferase, mitochondrial [OS=Homo sapiens]	32,12520503	17,676338	6	6	6	439	48,504	OAT	0.91	0.53	0.38	0
High	P05141	ADP/ATP translocase 2 [OS=Homo sapiens]	485,7120126	61,7449664	26	245	9	298	32,831	SLC25A5	0.63	0.52	0.1	37
High	Q14204	Cytoplasmic dynein 1 heavy chain 1 [OS=Homo sapiens]	411,506234	27,707706	106	156	106	4646	532,072	DYNC1H1	-0.25	0.52	-0.77	0
High	P02805	histone H4 [OS=Homo sapiens]	134,7215646	58,2524722	10	50	10	103	11,36	ST1H4H; HIST	-0.07	0.52	-0.59	0
High	P27704	Leucine-rich PR motif-containing protein, mitochondrial [OS=Homo sapiens]	92,0336161	18,361549	25	37	25	1394	157,805	LPRPRC	0.03	0.52	-0.48	0
High	Q96P5-1	E3 ubiquitin-protein ligase ZFP91 [OS=Homo sapiens]	65,32837312	24,5614035	14	25	14	570	63,406	ZFP91	0.62	0.52	0.11	0
High	Q129067	Isotform 7 of Interleukin enhancer-binding factor 3 [OS=Homo sapiens]	64,022626	17,5946548	14	21	14	898	95,748	ILF3	-0.36	0.52	-1.08	0
High	Q9Y743-3	Isotform 3 of Serine/threonine-protein phosphatase 6 catalytic subunit [OS=Homo sapiens]	14,8199864	6,14035088	2	4	2	342	38,921	PPP6C	0.51	0.52	-0.01	0
High	Q8VNM0	Dimethyladenosine transferase 1, mitochondrial [OS=Homo sapiens]	125,967786	42,7745665	13	33	33	346	39,518	TBIM	0.7	0.51	0.19	0
High	Q9P215	Leucine-tRNA ligase, cytoplasmic [OS=Homo sapiens]	9,73536639	2,12585034	3	4	3	1176	134,379	LARS	-0.88	0.51	-1.39	0
High	Q9Y2W1	Thyroid hormone receptor-associated protein 3 [OS=Homo sapiens]	119,821558	30,8900524	23	42	21	955	108,601	TRAP3	0.3	0.5	-0.2	0
Medium	P09N86	Ras GTP-activating protein-binding protein 2 [OS=Homo sapiens]	4,60907301	2,69709544	1	2	1	482	54,088	G3BP2	0.07	0.5	-0.42	0
High	P04350	Tubulin beta-4A chain 1 [OS=Homo sapiens]	710,74432	82,432424	27	351	4	444	49,554	TUBB4A	0.5	0.48	0.01	2
High	P11142-1	Heat shock cognate 11 kDa protein [OS=Homo sapiens]	509,349529	60,907121	39	167	31	646	70,854	HSP40	0.22	0.48	-0.26	13
High	P06891	ribose-phosphate pyrophosphokinase 1 [OS=Homo sapiens]	75,3384937	53,345012	9	21	9	318	34,812	PRPS1	0.37	0.48	-0.11	0
High	P26592	Isotform 2 of Threonyl-tRNA ligase, cytoplasmic [OS=Homo sapiens]	9,6334493	4,4975545	3	4	3	756	86,806	TSR1	-1.18	0.48	-1.96	0
High	P233991	DNA replication licensing factor MCW44 [OS=Homo sapiens]	91,6360134	23,986095	16	25	16	863	96,498	MCW44	0.01	0.47	-0.46	0
High	P05347-1	structural maintenance of chromosomes protein 2 [OS=Homo sapiens]	56,9692243	16,2071346	16	22	16	1197	135,572	SMC2	0.15	0.47	-0.32	0
High	P61978-2	Isotform 2 of Heterogeneous nuclear ribonucleoprotein K [OS=Homo sapiens]	31,5770267	23,93224138	9	10	9	464	50,996	HNRPK	-0.97	0.47	-1.43	0

High	P17812	CTP synthase 1 [OS=Homo sapiens]	65.8080667	20.4737733	11	19	11	591	66.648	CTPS1	0.27	0.46	-0.19	0
High	Receptor expression-enhancing protein 4 [OS=Homo sapiens]	22.6655918	16.7315175	3	7	3	257	29.376	REEP4	0.72	0.46	0.26	0	
High	Isotform 2 of Eukaryotic translation initiation factor 3 subunit B [OS=Homo sapiens]	10.0921844	6.66552119	2	3	3	873	98.967		-1.12	0.46	-1.58	0	
High	Isotform 4 of Pyruvate dehydrogenase E1 component subunit alpha, somatic for	67.4920218	6.54205607	3	3	3	428	47.549	PDHA1	0.07	0.46	-0.39	0	
High	Tubulin alpha-1C chain [OS=Homo sapiens]	581.614935	68.8195991	28	247	2	449	49.863	TUBA1C	0.41	0.45	-0.04	3	
High	Q9UJK0	Ribosome biogenesis protein Ts13 homolog [OS=Homo sapiens]	87.7325813	13.8333333	11	25	11	312	33.575	TSR3	0.85	0.45	0.4	0
High	Isotform 3 of L-lactated dehydrogenase A chain [OS=Homo sapiens]	44.6651869	29.362809	9	13	8	361	39.812	LDH1A	-1.49	0.45	-1.95	1	
High	Q96P70	Impritin-9 [OS=Homo sapiens]	27.5111096	8.3573487	9	12	9	1041	115.889	IP09	-0.13	0.45	-0.58	0
Low	Isotform 3 of E3 ubiquitin-protein ligase lissterin [OS=Homo sapiens]	1.0345631	0.71743929	1	1	1	1812	205.046	LTN1	0.23	0.45	-0.23	0	
High	Q9Z841	Probable ATP-dependent RNA helicase DDX17 [OS=Homo sapiens]	84.8348018	25.3772291	16	33	11	729	80.222	DDX17	0.06	0.44	-0.39	6
High	Martin-3 [OS=Homo sapiens]	52.4504746	23.7308146	13	18	13	847	94.565	MATR3	-0.14	0.44	-0.58	0	
High	Pre-mRNA-processing factor 6 [OS=Homo sapiens]	35.11497802	13.4962806	11	13	11	941	106.558	PRPF6	-0.78	0.44	-1.22	0	
High	Isotform 3 of Rapamycin-insensitive companion of mTOR [OS=Homo sapiens]	9.722753939	1.67464649	2	3	2	1732	194.888	RICTOR	2	0.44	0.57	0	
High	P55998	26S protease regulatory subunit 7 [OS=Homo sapiens]	4.74193098	5.08083141	2	2	2	433	48.603	PRMC2	-0.4	0.44	-0.84	0
High	Q9NQ29	putative RNA-binding protein Luc7-like 1 [OS=Homo sapiens]	422.486496	47.1698113	26	198	18	371	43.701	LUC7L	0.54	0.43	0.11	0
High	Histone H2B type 1-j [OS=Homo sapiens]	137.994765	57.9365079	9	58	2	126	13.896	HIST1H2Bj	0.06	0.43	-0.36	0	
High	Cellular tumor antigen p53 [OS=Homo sapiens]	56.8627829	45.9872777	10	14	10	393	43.625	TP53	-0.23	0.43	-0.66	0	
High	Surfeit locus protein 6 [OS=Homo sapiens]	15.8096652	12.1883657	5	6	5	361	41.426	SURF6	-0.83	0.43	-1.26	0	
High	Q9NZ01-1	Very long-chain enoyl-CoA reductase [OS=Homo sapiens]	12.3669352	7.14285714	2	4	2	308	36.011	TECR	0.62	0.43	0.2	0
Low	Q13045-1	protein flightless-1 homolog [OS=Homo sapiens]	1.112397	0.5161545	1	1	1	1269	144.659	FLL	-0.51	0.43	-0.94	0
High	P11566	C-1-tetrahydrofolate synthase; cytoplasmic [OS=Homo sapiens]	98.9235701	29.5718766	23	34	23	935	101.495	MTHFD1	-0.77	0.42	-1.19	0
High	P05782	AP-2 complex subunit alpha-1 [OS=Homo sapiens]	54.6050431	14.7389869	14	19	9	977	107.478	AP2A1	0.46	0.42	0.04	5
High	Q96CW1	AP-2 complex subunit mu [OS=Homo sapiens]	14.5115814	9.1954023	5	7	5	435	49.623	AP2M1	0.39	0.42	-0.93	0
High	Q9E425	Insulin-like growth factor 2 mRNA-binding protein 2 [mRNA-binding protein 7] [OS=Homo sapiens]	12.4439133	6.7357513	4	5	1	579	63.666	IGFBP3	-0.56	0.41	-0.97	0
High	Q13595-1	Transformer-2 protein homolog alpha [OS=Homo sapiens]	12.28664123	8.15602837	2	3	2	282	32.669	TRAF2A	-0.51	0.41	-0.92	0
High	P51553-1	isocitrate dehydrogenase (NAD) subunit gamma, mitochondrial [OS=Homo sapiens]	8.82556394	5.8524173	2	3	2	393	42.767	IDH3G	0.15	0.41	-0.26	0
Low	Q9Y247-2	Isotform 2 of Nck-associated protein 1 [OS=Homo sapiens]	1.21781413	1.2345679	1	1	1	1134	129.433	NCKAP1	-1.48	0.41	-1.9	0
High	P05782	AP-2 complex subunit alpha-1 [OS=Homo sapiens]	54.6050431	14.7389869	14	19	9	977	107.478	AP2A1	0.46	0.42	0.04	5
High	Q96CW1	AP-2 complex subunit mu [OS=Homo sapiens]	14.5115814	9.1954023	5	7	5	435	49.623	AP2M1	0.39	0.42	-0.93	0
High	Q9E425	Insulin-like growth factor 2 mRNA-binding protein 2 [mRNA-binding protein 7] [OS=Homo sapiens]	12.4439133	6.7357513	4	5	1	579	63.666	IGFBP3	-0.56	0.41	-0.97	0
High	P11566	Transformer-2 protein homolog alpha [OS=Homo sapiens]	12.28664123	8.15602837	2	3	2	282	32.669	TRAF2A	-0.51	0.41	-0.92	0
High	Q13595-1	isocitrate dehydrogenase (NAD) subunit gamma, mitochondrial [OS=Homo sapiens]	8.82556394	5.8524173	2	3	2	393	42.767	IDH3G	0.15	0.41	-0.26	0
Low	Q9Y247-2	Isotform 2 of Nck-associated protein 1 [OS=Homo sapiens]	1.21781413	1.2345679	1	1	1	1134	129.433	NCKAP1	-1.48	0.41	-1.9	0
High	P149458	Signal peptide	26.632907	43.203258	4	8	4	86	101.105	SRP9 SRP19p1	0.28	0.4	-0.12	0
High	P060884	DnaJ homolog subfamily A member 2 [OS=Homo sapiens]	19.1206483	8.37786408	3	7	3	412	45.717	DNAJ1	0.9	0.4	0.49	0
High	Q9EUJ2-1	Heterogeneous nuclear ribonucleoprotein U-like protein 1 [OS=Homo sapiens]	16.6011478	6.42533564	5	6	4	856	95.679	HNRNPL1	-0.09	0.4	-0.49	0
High	P14697-2	Transformer-2 protein homolog alpha [OS=Homo sapiens]	52.36783306	16.4596723	14	20	14	966	109.369	GNA1B	-0.75	0.39	-1.14	0
High	P105399	Trifunctional enzyme phosphodiesterase esterases [OS=Homo sapiens]	37.148877	12.97508383	9	11	9	763	83.947	HADHA	-0.39	0.38	-0.58	0
High	P62937	peptidyl prolyl cis-trans isomerase A [OS=Homo sapiens]	23.1816634	32.7272272	4	8	4	165	18.001	PPIA	-2.13	0.39	-2.52	0
High	P131943	Heterogeneous nuclear ribonucleoprotein H [OS=Homo sapiens]	65.2501612	34.298441	11	18	4	449	49.198	HNRNPH1	-0.36	0.38	-0.74	8
High	P018455	Histon H2A.Z [OS=Homo sapiens]	27.7682082	12.15253008	3	17	2	128	13.545	H2AFZ	0.1	0.38	-0.28	0
High	P57088	Transmembrane protein 33 [OS=Homo sapiens]	11.62228385	8.50202429	2	3	2	247	27.96	TMEM33	-0.12	0.38	-0.05	0
High	Q96D09	G-protein coupled receptor-associated sorting protein 2 [OS=Homo sapiens]	6.86761294	5.019193317	3	4	3	838	93.715	2; AMCK5-G	0.05	0.38	-0.33	0
High	QKWHB-1	1-phosphatidylinositol 4,5-bisphosphate phosphodiesterase esterases [OS=Homo sapiens]	3.93465876	4.78440638	7	9	7	1693	189.104	PICHL	0.86	0.37	0.49	0
Medium	Q9EG43	protein L1V1 homolog [OS=Homo sapiens]	1.21781413	1.2345679	4	2	2	475	54.821	L1V1	0.4	0.37	0.03	0
High	P62935	transformer-2 protein homolog beta [OS=Homo sapiens]	17.8804254	18.75	5	7	5	288	33.646	TRA2B	0.1	0.36	-0.26	0
High	P26593-3	Isotform 3 of Polyprymidine tract-binding protein 1 [OS=Homo sapiens]	6.609083917	5.02692998	2	3	2	557	59.596	PTRBP1	-0.64	0.36	-1	0
High	P0610-1	Clathrin heavy chain 1 [OS=Homo sapiens]	141.746152	25.0149524	35	48	35	1675	191.493	CLTC	-1.8	0.35	-2.16	0
High	Q7L34	Zinc finger protein 771 [OS=Homo sapiens]	120.732428	54.7641325	16	37	16	317	35.670	WIF171	0.35	0.35	0.24	0
High	Q9ETCO	Death-inducer obligator 1 [OS=Homo sapiens]	68.2259411	10.9821429	20	30	20	2240	243.723	DIDO1	-0.3	0.35	-0.65	0
High	P62836	GTP-binding nuclear protein RAN [OS=Homo sapiens]	31.2253856	19.9074074	4	10	4	216	24.408	RAN	-0.51	0.35	-0.86	0
High	Q8TEX9-2	Isotform 2 of importin-4 [OS=Homo sapiens]	16.7647043	6.00184672	5	8	5	1083	118.826	KIF11	-0.02	0.35	-0.37	0
Medium	Q76666	KH domain-containing RNA-binding protein	4.70311552	3.16027088	1	1	1	443	48.197	KHDRBS1	-0.16	0.35	-0.51	0
Medium	Q15003	Condensin complex subunit 2 [OS=Homo sapiens]	2.804931	1.75438396	1	1	1	741	82.511	NCPAP1	0.39	0.35	0.04	0
Low	P27694	Replication protein A 70 kDa DNA-binding subunit [OS=Homo sapiens]	2.08102657	2.5974026	1	1	1	616	68.095	RPA1	2.11	0.35	1.76	0
High	Q9NT13-1	Structural maintenance of chromosomes protein 4 [OS=Homo sapiens]	58.9267891	15.6832298	18	24	18	1288	147.091	SMC3	0.15	0.33	-0.19	0
High	Q8N27-1	T-HO complex subunit 2 [OS=Homo sapiens]	19.2861286	4.45699937	8	11	8	1593	182.659	THOC2	-0.36	0.34	-0.7	0
High	Q15070-1	Mitochondrial inner membrane protein OXA11 [OS=Homo sapiens]	9.650533729	2.75862069	1	3	1	435	48.516	OXA11L	0.48	0.34	0.13	0
High	P04411	DNA-directed RNA polymerase, mitochondrial [OS=Homo sapiens]	7.40508261	2.68292683	3	3	3	1230	138.332	POLRMT	-0.03	0.34	-0.37	0
High	Q95W79-1	Centrosomal protein of 170 kDa [OS=Homo sapiens]	6.7064532	1.51515152	2	4	2	1584	175.187	CEP170	0.22	0.34	-0.12	0
High	P40227-1	T-complex protein 1 subunit zeta [OS=Homo sapiens]	13.1626273	12.4293785	5	9	5	531	57.988	CCT6A	-0.54	0.33	-0.87	0
High	P63167	Dynein light chain 1, cytoplasmic [OS=Homo sapiens]	33.0885794	39.3258427	4	7	1	89	10.359	DYNLL1	-0.29	0.32	-0.61	0
High	Q7K2T4	Staphylococcal nucleic domain-containing protein 1 [OS=Homo sapiens]	14.6742746	8.35164835	6	8	6	910	101.934	SND1	-1.26	0.32	-1.58	0
High	P460874	Isotform 4 of Probable 28S rRNA cytosine(4447)-C(5')-methyltransferase [OS=Homo sapiens]	11.309058	5.68047337	4	4	4	845	92.803	NOP2	0.07	0.32	-0.25	0
High	Q99460	26S proteasome non-ATPase regulatory subunit 1 [OS=Homo sapiens]	9.512162638	3.38740818	3	3	3	953	105.769	P5M01	-1.44	0.32	-1.77	0
Medium	P28933	28S ribosomal protein A 70 kDa [OS=Homo sapiens]	3.78038457	4.7979798	2	2	2	396	45.806	MRPS9	0.53	0.32	0.21	0
Low	P00487	26S proteasome non-ATPase regulatory subunit 14 [OS=Homo sapiens]	1.183449356	2.25806452	1	1	1	310	34.555	PSMD14	-0.7	0.32	-1.02	0
High	P22695	Cytochrome b-c1 complex subunit 2, mitochondrial [OS=Homo sapiens]	213.9320233	43.9293598	17	60	17	453	48.413	UCRC2	0.68	0.31	0.36	0
High	P13010	X-ray repair cross-complementing protein 5 [OS=Homo sapiens]	22.1491202	11.4754098	8	9	8	732	82.652	XRC5	-0.19	0.31	-0.49	0
High	Q9EQ33-1	Structure-specific endonuclease subunit SLX1 [OS=Homo sapiens]	19.1436987	18.1818182	4	5	4	275	30.751	SLX1; SLX1A	0.31	0.31	10.37	0

High	P28715-1	DNA repair protein complementing XP-G cells [OS=Homo sapiens]	18.1256284	7.92580101	7	7	1186	133.027	ERCC5	951	0.31	9,2	0	
High	P63220	40S ribosomal protein s21 [OS=Homo sapiens]	14.4428081	24.0963855	3	5	83	9.106	RPS21	0.35	0.31	0.04	0	
High	P10744-1	Tifin basal transcription factor complex helicase subunit [OS=Homo sapiens]	8.36561353	4	4	4	760	86.8454	ERCC2	11.76	0.31	11,45	0	
High	Q5B758-1	Neutral amino acid transporter B6(O) [OS=Homo sapiens]	7.579075141	4.99075786	2	2	541	56.562	SCLC1A5	0.38	0.31	0.07	0	
High	Q8BPW8	Protein NipSnap homolog 1 [OS=Homo sapiens]	7.46599399	11.2676056	2	3	284	33.289	NIPSNAP1	11.49	0.31	11,18	0	
High	Q13155	aminoacyl tRNA synthetase complex-interacting multifunctional protein 2 [OS=Hc]	7.42540602	2.8125	1	2	320	35.326	AMPP2	-0.16	0.31	-0.46	0	
Medium	Q01844-5	Isotropin 5 of RNA-binding protein EWS [OS=Homo sapiens]	4.588664927	5.74886536	2	2	661	68.923	EWSR1	-1.26	0.31	-1.67	0	
Medium	Q9N256	Formin-2 [OS=Homo sapiens]	3.44867201	0.92915215	1	1	1722	179.993	FMN2	12.59	0.31	12,27	0	
Low	O75477	erlin-1 [OS=Homo sapiens]	1.71783122	2.02312139	1	1	346	38.901	ERLIN1	11.31	0.31	11	0	
Low	O72715-1	TRIM1-like protein [OS=Homo sapiens]	1.67015948	1.09140518	1	1	733	81.695	TRIM1	0.03	0.31	-0.28	0	
High	P21127-2	Isotropin SV1 of Cyclin-dependent kinase 11B [OS=Homo sapiens]	228.406249	45.7800512	40	85	782	91.276	CK1B	-0.72	0.3	-1.02	0	
High	P16403	Histone H1.2 [OS=Homo sapiens]	155.134886	30.9859155	12	58	213	21.352	HIST1H1C	0.27	0.3	-0.93	0	
High	P11387	DNA topoisomerase 1 [OS=Homo sapiens]	49.6176493	16.0784314	11	20	765	90.669	TOP1	-0.35	0.3	-0.65	0	
Medium	Q8N0V3-1	Putative ribosoma-binding factor A, mitochondrial [OS=Homo sapiens]	4.20953986	7.38862974	2	2	343	38.335	RBFA	0.56	0.3	0.26	0	
Low	O75494-1	Serine/arginine-rich splicing factor 1, mitochondrial [OS=Homo sapiens]	1.45754805	4.96183206	1	1	262	31.282	LOC10095	-0.18	0.3	-0.48	0	
High	P08238	Heat shock protein HSP 90-beta [OS=Homo sapiens]	213.172398	49.1712707	34	65	17	724	83.212	HSP90AB1	-1.09	0.29	-1.38	2
High	P3919	40S ribosomal protein S19 [OS=Homo sapiens]	52.5103314	5.9310345	9	19	145	16.051	RPS19	-0.04	0.28	-0.33	0	
High	P07195	L-lactate dehydrogenase B chain [OS=Homo sapiens]	60.88955389	29.6407186	8	15	7	334	36.615	LDHB	-1.58	0.27	-1.85	0
High	O75147	Obscurin-like protein 1 [OS=Homo sapiens]	53.1553534	8.33333333	13	18	13	1896	206.817	OBSL1	0.33	0.27	0.06	0
High	P00308-3	Probable ubiquitin carboxy-terminal hydrolase FAF-X [OS=Homo sapiens]	10.4449573	1.75097276	4	4	2570	292.094	USP9X	-0.31	0.27	-0.58	0	
Low	P49750-4	Isotropin 4 of YP-motif-containing protein 1 [OS=Homo sapiens]	1.3317477	0.4193849	1	1	2416	241.495	YPM1	0.27	0.27	-0.13	0	
High	P06733-1	alpha-enolase [OS=Homo sapiens]	195.468706	61.5207373	22	74	22	434	47.139	ENO1	-0.28	0.26	-0.54	0
High	P26277	40S ribosomal protein S27 [OS=Homo sapiens]	124.575195	41.6666667	6	37	3	84	9.455	RPS27	0.38	0.26	0.13	6
High	P072905	Interleukin enhancer-binding factor 2 [OS=Homo sapiens]	23.889616	13.3333333	5	7	5	390	43.035	ILF2	-0.72	0.26	-0.98	0
Medium	Q8UKF5-1	Traf2 and Nck-interacting protein kinase [OS=Homo sapiens]	2.36032579	1.02941176	2	2	1360	154.848	TNIK	0.55	0.26	0.29	0	
High	P38646	Stress-70 protein, mitochondrial [OS=Homo sapiens]	230.731237	43.2986961	33	66	32	679	73.635	HSPA9	0.21	0.25	-0.04	0
High	P61513	60S ribosomal protein L37a [OS=Homo sapiens]	101.565689	59.7826087	7	39	7	92	10.268	RPL37A	0.21	0.25	-0.03	0
High	P9HC1B6	Spondin-1 [OS=Homo sapiens]	23.8337319	11.7719595	8	10	8	807	90.194	SPON1	0.47	0.25	0.22	0
High	P62273-2	Isotropin 2 of 40S ribosomal protein S29 [OS=Homo sapiens]	13.6165669	20.8955224	2	12	2	67	8.082	RPS29	0.26	0.25	0.01	0
High	O14735	CDP-diacylglycerol-inositol 3-phosphatidyltransferase [OS=Homo sapiens]	11.3432692	6.10328338	1	3	213	23.523	CDPT	0.2	0.25	-0.06	0	
High	P233993-1	DNA replication licensing factor Mcm7 [OS=Homo sapiens]	27.531774	32	96	32	719	81.257	MCM7	0.28	0.24	0.04	0	
High	P08670	Vimentin [OS=Homo sapiens]	97.4455192	54.0564378	31	24	466	53.619	VIM	-1.04	0.24	-1.28	1	
High	P63244	Guanine nucleotide-binding protein subunit beta-2-like 1 [OS=Homo sapiens]	96.5711932	48.8958991	14	25	14	317	35.055	GNB2L1; RACK	-0.18	0.24	-0.42	0
High	O95821-1	Apoptosis-inducing factor 1, mitochondrial [OS=Homo sapiens]	69.0894727	27.0799494	14	24	14	613	66.859	AIFM1	0.11	0.24	-0.13	0
High	P07157-1	Putative RNA-binding protein 15 [OS=Homo sapiens]	31.9715381	10.07062	8	11	8	977	107.124	RBM15	-0.12	0.24	-0.36	0
High	Q9NY20	39S ribosomal protein L16, mitochondrial [OS=Homo sapiens]	19.7175095	11.1553785	2	5	2	251	28.431	MRPL16	0.01	0.24	-0.23	0
High	P39656	Dolichyl-diphosphoinosaccharide-protein glycosyltransferase 48 kDa subunit [OS=Homo sapiens]	17.7708371	6.79824561	3	6	3	456	50.769	DGOST	-0.11	0.24	-0.35	0
High	P38E7X	Puretrophin-1 [OS=Homo sapiens]	18.0057681	2.38380697	4	7	3	1191	130.72	PLK1G4	-0.47	0.23	-0.7	1
High	Q5E1G-1	Programmed cell death protein 4 [OS=Homo sapiens]	16.66339925	5.54371002	2	5	2	469	51.703	PDGD4	0.39	0.23	0.16	0
High	Q92499	ATP-dependent RNA helicase DDX1 [OS=Homo sapiens]	16.1549105	12.1621622	6	7	6	740	82.38	DDX1	-0.02	0.23	-0.25	0
High	P15880	40S ribosomal protein S21 [OS=Homo sapiens]	224.033265	56.995587	18	80	18	293	31.305	RP52	0.04	0.22	-0.18	0
High	P38919	Eukaryotic initiation factor 4E-III [OS=Homo sapiens]	34.0416254	22.1411192	8	16	6	411	46.841	EIF4A3	-0.65	0.22	-0.87	2
High	Q9Y318	60S ribosomal protein L36 [OS=Homo sapiens]	34.9423542	30.47619305	4	20	4	105	12.246	RP136	0.04	0.22	-0.22	0
High	Q9N218	Insulin-like growth factor 2 mRNA-binding protein 1 [OS=Homo sapiens]	21.5563802	10.745234	6	7	3	577	63.441	IGFBP1	-0.58	0.22	-0.8	3
High	P22636	heterogeneous nuclear ribonucleoprotein A2/B1 [OS=Homo sapiens]	19.6928298	13.8810198	5	7	5	353	37.407	HNRNPA2B1	-1.44	0.22	-1.67	0
High	P519R7-5	Isotropin 5 of Serine/threonine-protein phosphatase 6 regulatory subunit 3 [OS=	13.3962014	3.29920364	3	5	3	879	98.423	PPP6R3	0.17	0.22	-0.05	0
High	P08VP6-1	cullin-associated neddy-18-disociated protein 1 [OS=Homo sapiens]	97.73897236	23.3577236	24	35	23	1230	136.289	CAND1	-0.78	0.21	-0.99	0
High	P32969	60S ribosomal protein L9 [OS=Homo sapiens]	77.4642314	25.5208333	4	20	4	192	21.85	RP19	0.01	0.21	-0.19	0
High	P49368-1	T-complex protein 1 subunit gamma [OS=Homo sapiens]	52.6208938	20.3669725	11	19	11	545	60.495	CCT3	-0.02	0.21	-0.23	0
High	P25297	Heterogeneous nuclear ribonucleoprotein E [OS=Homo sapiens]	31.973683	28.4337349	8	14	6	415	45.643	HRNF1	-0.12	0.21	-0.33	0
High	P20876	DNA-directed RNA polymerase I subunit RP2 [OS=Homo sapiens]	16.9875402	5.36626917	6	7	6	1174	133.811	POLR2B	0.42	0.21	0.21	0
Medium	Q8N5N7	39S ribosomal protein L50, mitochondrial [OS=Homo sapiens]	4.05173378	9.49367089	1	1	1	158	18.313	MRPL50	0.97	0.21	0.76	0
Low	P08574	Cytochrome c1, heme protein, mitochondrial [OS=Homo sapiens]	1.41128022	2.15384615	1	1	1	325	35.399	CYC1	0.37	0.21	0.15	0
High	P27708	CAD protein [OS=Homo sapiens]	384.055207	31.101236	58	122	58	2225	24.829	CAD	-0.17	0.2	-0.37	0
High	P33621-2	Isotropin 2 of Coatomer subunit alpha [OS=Homo sapiens]	102.807609	18.08369692	20	31	20	1233	139.235	COPA	-0.11	0.2	-0.31	0
High	P27108	Signal recognition particle 14 kDa protein [OS=Homo sapiens]	59.1449498	36.0294118	5	17	5	366	14.561	SRP14	0.31	0.2	0.11	0
High	Q15366-2	Isotropin 2 of Pofc(r) binding protein 2 [OS=Homo sapiens]	53.7269251	27.868825	9	21	4	366	38.627	PCBP2	-0.93	0.2	-1.13	0
High	Q5S8E5	Nucleoporin NUP188 homolog [OS=Homo sapiens]	9.2223557	1.88679245	4	5	4	1749	195.917	NUP188	-0.22	0.2	-0.42	0
Medium	Q9Y6D6	Brefeldin A-inhibited glutamine nucleotide-exchange protein 1 [OS=Homo sapiens]	4.22694481	0.75716604	2	3	2	1849	208.634	ARFGEF1	0.03	0.2	-0.16	0
High	Q5B759	Complex I assembly factor TIMMDCL, mitochondrial [OS=Homo sapiens]	23.4508825	62.116041	19	52	19	285	32.158	TIMMDCL	0.31	0.2	0.11	0
High	P17987	Mk67/TFIIP domain-interacting nucleophosmin [OS=Homo sapiens]	180.796683	59.352518	25	47	25	556	60.304	TFIIP	0.04	0.19	-0.15	0
High	P06493	Cyclin-dependent kinase 1 [OS=Homo sapiens]	44.4991901	43.7710438	11	16	10	297	34.074	CDK1	0.21	0.19	0.02	0
High	P55084	Trifunctional enzyme subunit beta, mitochondrial [OS=Homo sapiens]	17.8853126	9.07172996	4	6	4	474	51.262	HDHB	-0.17	0.19	-0.36	0

High	Q7Z6Z7	[E3 Ubiquitin-protein ligase HUWE1 [OS=Homo sapiens]	5.956823398	0.858877	3	3	3	4374	481.589	HUWE1	-1.44	0.19	-1.63	0
High	Q9HDCS	junctophilin-1 [OS=Homo sapiens]	48.9181275	16.4901664	10	21	10	661	71.642	JPH1	0.61	0.18	0.44	0
High	O72325	28S ribosomal protein S12, mitochondrial [OS=Homo sapiens]	46.0383071	44.9525362	6	25	6	138	15.163	MPP512	0.29	0.18	0.11	0
High	P61927	60S ribosomal protein L37 [OS=Homo sapiens]	27.7161047	37.1134021	7	43	7	97	11.071	RPL37	0.09	0.18	-0.17	0
High	Q9BRX2	Protein-pedata homolog [OS=Homo sapiens]	27.6033797	12.2077922	4	7	4	385	43.332	FELO	0.44	0.18	0.27	0
High	Q8YK81-1	DnaJ homolog family C member 10 [OS=Homo sapiens]	11.4203451	3.65698674	3	5	3	793	91.021	DNAJC10	0.39	0.18	0.21	0
High	Q8YX11	Mitochondrial Rhod-GTPase 2 [OS=Homo sapiens]	8.29722164	2.7508906	2	3	2	618	68.075	RHOT2	-0.21	0.18	-0.4	0
Medium	Q9P010-2	Isotform 2 of NADH dehydrogenase (ubiquinone) 1 alpha subcomplex subunit 1 [OS=Homo sapiens]	8.59980836	6.30630631	1	1	1	222	24.866	TSHZ	0.28	0.18	0.1	0
High	Q6UN15-3	Isotform 3 of Pre-mRNA 3'-end-processing factor FIP1 [OS=Homo sapiens]	409.9780285	53.6538462	29	154	1	520	58.341	FIP1L1	0.33	0.17	0.16	0
High	P2396-1	40S ribosomal protein S3 [OS=Homo sapiens]	376.0782444	81.8930041	27	161	27	243	26.673	RPS3	0.15	0.17	-0.02	0
High	P62266	40S ribosomal protein S23 [OS=Homo sapiens]	308.656194	72.027972	18	442	18	143	15.798	RPS23	0.11	0.17	-0.07	0
High	P22344	Ubiquitin-like modifier-activating enzyme 1 [OS=Homo sapiens]	112.677626	25.8034026	21	35	21	1058	117.774	UBA1	-2.47	0.17	-2.64	0
High	Q9UGG63-2	Isotform 2 of ATP-binding cassette subfamily Y member 2 [OS=Homo sapiens]	93.0703672	23.9747634	15	26	15	634	72.397	ABCFL2	0.43	0.17	0.26	0
High	Q96C36	Isotform 2 of Protein-Li-isopentadecapeptide (D-aspartate)-O-methyltransferase [OS=Homo sapiens]	52.4612817	16.251	4	11	3	320	33.616	PRCR2	0.29	0.17	0.13	0
High	P22061-2	Isotform 2 of Pyroline-5-Carboxylate reductase 2 [OS=Homo sapiens]	24.5156763	10.5263158	2	4	2	228	24.664	POMT1	0.17	0.17	0	0
Medium	P47897	Glutamine-tRNA ligase [OS=Homo sapiens]	3.283034242	3.09677319	2	2	2	775	87.743	QARS	-0.7	0.17	-0.88	0
Medium	Q6ZK3	protein Fam127A [OS=Homo sapiens]	2.59192971	6.19469027	1	1	1	113	13.163	FAM127A	-0.25	0.17	-0.42	0
Low	Q16650	T-box brain protein 1 [OS=Homo sapiens]	1.25758912	1.75953079	1	1	1	682	74.007	TBR1	-0.02	0.17	-0.19	0
High	P60762	Dolichol-phosphate mannose transferase subunit 1 [OS=Homo sapiens]	42.8954581	25.7692308	5	12	5	260	29.616	DPM1	0.35	0.16	0.2	0
High	P05277-5	Isotform 5 of Protein transport protein Sec16A [OS=Homo sapiens]	24.5116763	5.81553839	9	12	9	2201	235.607	-	-0.09	0.16	-0.25	0
High	P62318	Small nuclear ribonucleoprotein snRNP B [OS=Homo sapiens]	12.866505	31.7465317	3	6	3	126	13.927	SNRNP3	-0.32	0.16	-0.47	0
High	P00846	ATP synthase subunit A [OS=Homo sapiens]	4.73960463	4.42477876	1	2	1	226	24.801	ATP6	-0.36	0.16	-0.53	0
Medium	Q96F07	Cytoplasmic FMR1-interacting protein 2 [OS=Homo sapiens]	3.18963305	0.93896714	1	1	1	1278	148.302	CIFIP2	-1.32	0.16	-1.48	0
Low	Q9Y5V0	Zinc finger protein 706 [OS=Homo sapiens]	1.18336106	14.4736842	1	1	1	76	8.492	DNF706	0.25	0.16	0.09	0
High	P62269	40S ribosomal protein S18 [OS=Homo sapiens]	317.720305	63.8157895	24	283	24	152	17.708	RPS18	0.31	0.15	0.17	0
High	P00571	ATP-dependent RNA helicase DBP3X [OS=Homo sapiens]	305.088109	54.6827795	32	101	32	662	73.198	DDX3X	0.38	0.15	0.23	0
High	P14739	Lamin-B receptor [OS=Homo sapiens]	63.3255368	17.94146568	10	21	10	615	70.658	LBR	0.2	0.15	0.05	0
High	P50991	T-complex protein 1 subunit delta [OS=Homo sapiens]	34.61203547	13.9146568	7	11	7	539	57.888	CT74	-0.09	0.15	-0.25	0
High	P13674-2	Isotform 2 of Protein 4-hydroxylase subunit alpha-1 [OS=Homo sapiens]	34.4502465	18.5393258	8	9	8	534	60.929	PABPA1	0.9	0.15	0.75	0
High	Q9hf6F5	Colled-coil domain-containing protein 36 [OS=Homo sapiens]	13.4800511	16.9444444	4	5	4	360	40.211	CCDC86	0.51	0.15	0.36	0
High	Q8WWS0-1	Proline-rich finger protein 6 [OS=Homo sapiens]	210.311968	56.3424658	19	97	6	365	41.264	PRH6	0.2	0.14	0.06	22
High	P07910-1	Heterogeneous nuclear ribonucleoprotein C/C2 [OS=Homo sapiens]	106.8430515	36.2745098	14	28	14	306	33.65	HRNRC	-0.84	0.14	-0.38	0
High	P62289	60S ribosomal protein L13 [OS=Homo sapiens]	99.3194423	57.6	10	55	10	125	14.454	RPL31	0.06	0.14	-0.09	0
Medium	Q15645	Pachytene checkpoint protein 2 homolog [OS=Homo sapiens]	4.59049056	4.62962963	2	2	2	432	48.52	TRIP13	-0.11	0.14	-0.25	0
Medium	CC4-NOT	Transcription complex subunit 1 [OS=Homo sapiens]	4.38205283	0.84175084	2	3	2	2376	266.768	CNOT1	-0.73	0.14	-0.88	0
Low	Q9H013	Magnesium transporter protein 1 [OS=Homo sapiens]	2.15658033	2.688656716	1	1	1	335	38.011	MAGI1	-0.07	0.14	-0.22	0
Low	P59834	Doublecortin domain-containing protein 1 [OS=Homo sapiens]	1.65915945	1.97740113	1	1	1	354	39.834	DCDC1	0.16	0.14	0.02	0
High	P26373-1	60S ribosomal protein S13 [OS=Homo sapiens]	200.828523	55.250237	17	92	17	211	24.247	RPL13	0.01	0.13	-0.12	0
High	P31689-1	DnaJ homolog subfamily A member 1 [OS=Homo sapiens]	73.1133989	30.1369863	12	28	10	397	44.839	DNAJA1	0.37	0.13	0.24	0
High	P10412	Histone H1A [OS=Homo sapiens]	141.051392	30.1369863	12	56	2	219	21.852	HIST1H1E	0.21	0.12	0.09	14
High	P46778	60S ribosomal protein L21 [OS=Homo sapiens]	68.99330494	50.6025	8	22	8	160	18.553	RPL21	0.12	0.12	0	0
High	P3246-1	DNA mismatch repair protein MSH2 [OS=Homo sapiens]	23.958652	6.63865163	6	10	6	934	104.677	MSH2	0.13	0.12	0.01	0
High	Q9NE4	Isoleucine-tRNA ligase, mitochondrial [OS=Homo sapiens]	15.30578678	7.50869565	6	6	6	1012	113.719	MRS2	-0.42	0.12	0.04	0
High	P04844-1	Dolichyl-diphosphooligosaccharide--protein glycosyltransferase subunit 2 [OS=Homo sapiens]	9.69973069	5.70522979	2	2	2	631	69.241	RPN2	0.13	0.12	0	0
High	Q9Y4W6	AFG3-like protein 2 [OS=Homo sapiens]	8.07888628	3.767411543	3	3	3	797	88.528	AFG3L2	0.11	0.12	-0.01	0
High	Q14864	Ribosomal RNA processing protein 1 homolog B [OS=Homo sapiens]	6.44506295	6.67282322	3	3	3	758	84.375	RPL1B	-0.34	0.12	-0.46	0
High	P62249	40S ribosomal protein S16 [OS=Homo sapiens]	242.362634	75.3424658	18	142	18	146	16.435	RPS16	0.19	0.11	0.08	0
High	P09747	Poly(Ap-ribosyl polymerase 1 [OS=Homo sapiens]	72.0148884	22.2879684	18	27	18	1014	113.012	PARP1	-0.54	0.11	-0.65	0
High	P66779	60S ribosomal protein L21 [OS=Homo sapiens]	38.7556982	12.6021004	8	13	8	857	15.738	TELO2	0.11	0.11	0	0
High	P10809	60 kDa heat shock protein, mitochondrial [OS=Homo sapiens]	36.2888628	17.9755672	9	10	9	573	61.016	HPD14	-0.33	0.11	-0.44	0
High	Q14839-2	Isotform 2 of Chromodomain-helicase-DNA-binding protein 4 [OS=Homo sapiens]	32.5065063	5.61885567	9	12	9	1940	220.709	CHD4	-0.17	0.11	-0.28	0
High	Q8Y6AA	Cilia- and flagella-associated protein 20 [OS=Homo sapiens]	23.4732414	27.9792746	5	8	5	193	22.76	STAG2	-0.63	0.11	-0.74	0
High	P16104	Isotform 4 of Hsc70 cell factor 1 [OS=Homo sapiens]	9.9993526	1.49110149	3	4	3	2079	213.272	HCC1	-1.95	0.11	-2.06	0
Medium	Q9Y4H8	telomere length regulation protein TE12 homolog [OS=Homo sapiens]	3.70257982	2.15053763	2	3	2	837	91.689	TEL02	0.14	0.11	0.03	0
Medium	Q9UHB9	Signal recognition particle subunit SRP68 [OS=Homo sapiens]	3.15247234	2.07336623	1	1	1	627	70.686	SRP68	-0.7	0.11	-0.82	0
Medium	Q9N666-1	Neuroblast differentiation-associated protein ANNAK [OS=Homo sapiens]	2.91793307	1.6808194	1	2	1	5890	628.699	ANNAK	-2.87	0.11	-0.28	0
Medium	Q8N3U4-2	Isotform 2 of Cohesin subunit SA-2 [OS=Homo sapiens]	2.50390101	1.02523659	1	1	1	1268	145.658	STAG2	-0.63	0.11	-0.74	0
High	P68371	Tubulin beta-4 chain [OS=Homo sapiens]	925.9493084	83.1460674	31	441	1	445	49.799	TUBB4B	0.09	0.1	-0.02	85
High	P62293	Ubiquitin-40S ribosomal protein S27 [OS=Homo sapiens]	284.809998	58.3333333	13	134	11	156	17.953	RPS27A	-0.04	0.1	-0.14	3
High	P62193	60S ribosomal protein L11 [OS=Homo sapiens]	146.813096	65.703371	13	67	13	20.24	RPL11	0.11	0.1	0.01	0	
High	Q5GNW6-2	Isotform 2 of Histone H2B type 2-F [OS=Homo sapiens]	148.497379	54.4776119	9	61	2	134	14.832	HIST2H2BF	-0.14	0.1	-0.23	19
High	P92616	translational activator GCN1 [OS=Homo sapiens]	90.8958214	13.1037065	28	37	28	2671	292.572	GCN11;GCN	-0.33	0.09	-0.43	0
High	P29372	DNA-3-methyadenine glycosylase [OS=Homo sapiens]	84.763927	48.6577181	11	21	11	298	32.848	MGF	0.1	0.09	0.01	0

High	Q16891	MtCOS complex subunit Mit60 [OS=Homo sapiens]	49.2335534	17.8100264	13	18	13	758	83.626	IMMIT	-0.03	0.09	-0.12	0
High	P14776-1	Transcription elongation regulator 1 [OS=Homo sapiens]	44.2653366	9.74499089	11	18	11	1098	123.823	TCEG1	-0.14	0.09	-0.23	0
High	P193673	Isoform 3 of Hoxd10 gene	26.097799	7.770901194	7	11	7	921	102.672	HKL1	-0.87	0.09	-0.95	0
High	Q9H710-3	Isoform 3 of Methyltransferase-like protein 17, mitochondrial [OS=Homo sapiens]	27.66266337	17.1548117	6	9	6	478	53.017	METTL17	0.1	0.09	0	0
High	O57200-1	Zinc finger CCH domain-containing protein 13 [OS=Homo sapiens]	9.382780788	1.49880096	2	3	2	1668	196.519	Z3H13	-0.38	0.09	-0.46	0
Medium	O5168-1	NADH dehydrogenase (ubiquinone) 1 beta subcomplex subunit 4 [OS=Homo sapiens]	3.852271783	10.0775194	1	2	1	129	15.199	NDUFB4	-0.4	0.09	-0.49	0
High	Q8N1E7-1	Nuclear pore complex protein Nup93 [OS=Homo sapiens]	155.049555	44.4444444	30	50	30	819	93.43	NUP93	0.01	0.08	-0.07	0
High	Q9Y285	Phenylalanine-tRNA ligase alpha subunit [OS=Homo sapiens]	73.4261591	23.4251969	11	25	11	508	57.528	FARS1A	0.22	0.08	0.15	0
High	P105783	40S ribosomal protein S10 [OS=Homo sapiens]	33.7745212	40	5	11	5	165	18.886	RPS10	-0.08	0.08	-0.16	0
High	P10340	Succinate dehydrogenase (ubiquinone) flavoprotein subunit, mitochondrial [OS=Homo sapiens]	15.8977593	3.76506024	2	4	2	664	72.645	SDHA	-0.91	0.08	-0.99	0
High	Q07955-1	Serine/arginine-rich splicing factor 1 [OS=Homo sapiens]	6.87424786	7.66129032	2	3	2	248	27.728	SRSF1	-0.07	0.08	-0.15	0
High	P12857	40S ribosomal protein L28 [OS=Homo sapiens]	114.4403437	57.971045	7	49	7	69	7.836	RPS28	0.22	0.07	0.15	0
High	P142911	60S ribosomal protein L18 [OS=Homo sapiens]	114.180018	63.8132296	14	41	14	257	28.007	RPL8	-0.48	0.07	-0.55	0
High	P162371	60S ribosomal protein L24 [OS=Homo sapiens]	104.750705	42.6751929	11	61	11	157	17.768	RPL24	0.04	0.07	-0.03	0
High	P14625	Endoplasmic reticulum [OS=Homo sapiens]	42.4473988	10.9589041	11	13	9	803	92.411	HS9P081	-1.27	0.07	-1.34	0
High	Q9Y678	Creatomer subunit gamma-1 [OS=Homo sapiens]	10.59290606	4.919900847	3	3	3	874	97.655	COPG1	-0.31	0.07	-0.38	0
High	P17306-1	NAH dehydrogenase (ubiquinone) 1 beta subunit, mitochondrial [OS=Homo sapiens]	6.46905912	3.45572354	2	3	2	463	52.512	NDUFV2	0.06	0.07	0	0
High	P12857	elongation factor Tu, mitochondrial [OS=Homo sapiens]	422.348415	68.3628319	27	150	27	452	49.51	TUFM	0	0.06	-0.06	0
High	P16244	40S ribosomal protein S15a [OS=Homo sapiens]	231.724544	81.5384615	15	104	15	130	14.83	RPS15A	-0.05	0.06	-0.11	0
High	P105269	serine palmitoyltransferase 1 [OS=Homo sapiens]	7.30319321	4.66257928	3	5	3	473	52.71	SPAT1C1	0.54	0.06	0.48	0
Medium	P55369	NHP2-like protein 1 [OS=Homo sapiens]	2.56365	9.375	1	1	1	128	141.65	[HPL21; SNR1]	-0.15	0.06	-0.21	0
High	Q8WUQ7-2	Isoform 2 of Cactin [OS=Homo sapiens]	382.740797	40.3208556	42	114	42	935	107.871	CPG1	0.68	0.05	0.63	0
High	Q69Y4N	protein virilizer homolog [OS=Homo sapiens]	7.46947176	3.75275938	5	5	5	1812	201.898	KIAA1429	-0.64	0.05	-0.69	0
High	Q9HAYA	exportin-5 [OS=Homo sapiens]	6.97451921	6.8362819	3	4	3	1204	136.222	XPO5	-0.55	0.05	-0.61	0
High	P18779-1	Isoform 4 of Neuropterin target esterase [OS=Homo sapiens]	4.77990803	1.45454545	1	2	1	1375	150.859	NPFLA6	0.96	0.05	0.91	0
Medium	P12766	Casein kinase I isoform alpha [OS=Homo sapiens]	2.88107425	2.96735905	1	1	1	337	38.89	CSNK1A1	0.21	0.05	0.15	0
High	P162766	60S ribosomal protein S35 [OS=Homo sapiens]	227.6407669	42.2764228	12	139	12	123	145.543	RPL35	0.12	0.04	0.08	0
High	P162850	40S ribosomal protein S15 [OS=Homo sapiens]	193.259336	56.0205253	17	73	73	158	18.419	RPS11	0.01	0.04	-0.03	0
High	Q94761	ATP-dependent DNA helicase Q4 [OS=Homo sapiens]	51.0714502	15.1490066	13	19	13	1208	132.993	RECQL	0.26	0.04	0.22	0
High	Q9NR40-5	Isoform 5 of Sphingosine kinase 2 [OS=Homo sapiens]	34.0813181	8.40998866	5	9	5	761	80.154	SHPRH2	0.18	0.04	0.14	0
High	P17367-1	Core histone macro-H2A.1 [OS=Homo sapiens]	21.4336573	16.6666667	5	7	3	372	39.592	H2AFY	-0.33	0.04	-0.37	2
High	Q8WUW0	Nucleolar pore complex protein Nup123 [OS=Homo sapiens]	5.52015456	3.11418885	3	3	3	156	128.998	NUP123	0.97	0.04	0.93	0
Medium	P27695	DNA-(apurinic or apyrimidinic site) lyase [OS=Homo sapiens]	3.0714723	2.83018868	1	2	1	318	35.532	APEX1	-1.61	0.04	-1.65	0
High	P162829	60S ribosomal protein L23 [OS=Homo sapiens]	487.879021	77.14282871	17	379	17	140	148.856	RPL23	0.15	0.03	0.13	0
High	P17CM4-2	Isoform 2 of Band 4.1-like protein 5 [OS=Homo sapiens]	47.5094086	74.0594059	32	173	6	505	57.849	EPA115	0.64	0.03	0.61	0
High	P17635	60S ribosomal protein L10 [OS=Homo sapiens]	393.7334049	63.5514019	18	242	18	214	24.588	RPL10	0.06	0.03	0.02	0
High	P162847-4	Isoform 4 of 40S ribosomal protein S24 [OS=Homo sapiens]	226.98215	16.6089965	6	78	6	289	32.41	RPS24	0.05	0.03	0.02	0
High	Q126233	Transcription intermediary factor 1-beta [OS=Homo sapiens]	59.50991919	19.8802935	15	22	15	835	88.493	TRIM28	-0.77	0.03	-0.8	0
High	P162428	splicing factor 3B [OS=Homo sapiens]	12.9133633	3.01724318	1	3	1	464	49.224	SF3A2	-0.42	0.03	-0.46	0
High	P162811	Basigin [OS=Homo sapiens]	9.96651648	6.23376623	2	3	2	385	42.174	B5G	-0.04	0.03	-0.07	0
High	P162277	40S ribosomal protein S7 [OS=Homo sapiens]	212.882488	71.6494845	18	89	18	194	22.113	RPS7	0.1	0.02	0.08	0
High	P162861	40S ribosomal protein S30 [OS=Homo sapiens]	59.83496912	33.8892051	5	52	5	59	6.644	PAU	0.04	0.02	0.02	0
High	P162842	Eukaryotic initiation factor 4A-I [OS=Homo sapiens]	33.7751252	15.270936	6	16	4	406	46.125	EIF4A1	-0.74	0.02	-0.76	0
High	Q143143	Precursor mRNA-splicing factor ATP-dependent RNA helicase DHX15 [OS=Homo sapiens]	202.790699	44.0751572	31	68	30	795	90.875	DHH15	-0.36	0.01	-0.37	0
High	P16280	putative methyltransferase C9orf114 [OS=Homo sapiens]	200.792255	192.406114	19	70	70	448	41.982	C9orf114	0.6	0.01	0.59	0
High	P16781	40S ribosomal protein S9 [OS=Homo sapiens]	191.1565858	62.8865879	21	107	21	194	22.578	RPS9	0.02	0.01	0.01	0
High	Q15393-1	Splicing factor 3B subunit 3 [OS=Homo sapiens]	146.268112	26.5406738	25	40	25	1217	135.492	SF3B3	-0.57	0.01	-0.57	0
High	Q8Y37	Probable ATP-dependent RNA helicase DHX37 [OS=Homo sapiens]	42.3648134	12.6188418	13	17	13	1157	129.464	DHX37	0.07	0.01	0.05	0
Low	P15394-4	Isoform 3 of phosphatidylinositol-4-phosphate [OS=Homo sapiens]	1.65545144	1.57894437	1	1	1	570	62.902	PGM3	-0.54	0.01	-0.54	0
High	P162427	Structural maintenance of chromosomes protein 3 [OS=Homo sapiens]	459.233344	78.8333333	28	180	28	264	29.926	RPS3A	0.02	0	0.02	0
High	P168366	Tubulin alpha-4A chain [OS=Homo sapiens]	427.842433	53.125	22	174	4	448	49.892	TUBA4A	-0.22	0	-0.22	0
High	P165060-1	Exportin-2 [OS=Homo sapiens]	85.0827423	25.7466229	21	30	21	971	110.346	CSE1L	-1.1	0	-1.1	0
Low	P17237-1	Protein MON2 homolog [OS=Homo sapiens]	1.34677849	0.58241118	1	1	1	1717	190.237	MON2	-0.34	0	-0.34	0
High	P164880	Microsomal glutathione S-transferase 3 [OS=Homo sapiens]	46.601784	32.8894768	4	13	4	152	16.506	MGST3	-0.12	0.02	-0.12	0
High	Q9UQK7	Structural maintenance of chromosomes protein 3 [OS=Homo sapiens]	35.2356268	9.77814297	11	16	11	1217	141.454	SMC3	-0.22	0	-0.22	0
High	Q16718	NADH dehydrogenase (ubiquinone) 1 alpha subcomplex subunit 5 [OS=Homo sapiens]	6.75117745	7.75862069	1	3	1	116	13.45	NUDFA5	-0.12	0	-0.12	0
Low	P14137	Ribosome biogenesis protein B0P1 [OS=Homo sapiens]	2.01597417	4.28954424	2	2	2	746	83.577	B0P1	0	0	0	0
Low	P17237-1	Protein MON2 homolog [OS=Homo sapiens]	73.373403	17.3333333	16	109	16	135	15.54	RPS17	0.03	-0.01	0.04	0
High	P163381	40S ribosomal protein L36a [OS=Homo sapiens]	170.5330436	48.1132075	13	81	4	106	12.433	RPL36A	0.07	0.01	0.08	16
High	P11498	pyruvate carboxylase, mitochondrial [OS=Homo sapiens]	9.22038567	3.65025467	3	3	3	1178	129.551	PC	-0.05	-0.01	-0.04	0
High	P162701	40S ribosomal protein S4-X isoform [OS=Homo sapiens]	489.085896	75.2851711	28	250	28	263	29.579	RPS4X	-0.05	-0.02	-0.03	0
High	P162831	40S ribosomal protein S25 [OS=Homo sapiens]	96.6725104	52	10	81	10	125	13.734	RPS25	0.02	-0.02	0.04	0

High	P20700	Laminin-B1 [OS=Homo sapiens]	62.8090822	26.7918089	14	24	11	586	66.368	LMNB1	-0.16	-0.02	-0.14	1
High	P30837	Aldehyde dehydrogenase X, mitochondrial [OS=Homo sapiens]	58.0319497	22.0502901	11	16	9	517	57.17	ALDH1B1	-0.17	-0.02	-0.15	2
High	P61006	Ras-related protein Rab-8A [OS=Homo sapiens]	13.8449926	9.17874396	2	4	2	207	23.653	RAB8A	-0.24	-0.02	-0.23	0
Low	P10745	Retinol-binding protein 3 [OS=Homo sapiens]	1.51927462	0.56134723	1	1	1	1247	135.278	RBP3	0.03	-0.02	0.05	0
High	POC058	histone H2A type 1 [OS=Homo sapiens]	145.05545	41.5384615	8	65	4	130	14.083	H2AFY2A	-0.19	-0.03	-0.15	9
High	P14232	Translational initiation factor eIF-2B subunit alpha [OS=Homo sapiens]	42.3441892	27.868825	7	13	7	305	33.691	EIF2B1	0.1	-0.03	-0.14	0
High	P25201	DNA mismatch repair protein MSH6 [OS=Homo sapiens]	17.7564335	5.66176471	7	10	7	1360	152.689	MSH6	-0.06	-0.03	-0.03	0
High	Q9P055	Very-long-chain (3R)-hydroxyacyl-CoA dehydratase 1 [OS=Homo sapiens]	14.5533332	9.94475138	3	5	3	362	43.132	PLAD1_HAC1	-0.54	-0.03	-0.51	0
High	P15151	translocon-associated protein subunit delta [OS=Homo sapiens]	11.483404	28.9017341	4	4	4	173	18.987	SP4	0.73	-0.03	0.76	0
High	Q9Y277_2	Isotform 2 of Voltage-dependent anion-selective channel protein 3 [OS=Homo sapiens]	5.61592745	6.33802817	2	2	2	284	1.77	VDAC3	-0.37	-0.03	-0.34	0
High	P78527	DNA-dependent protein kinase catalytic subunit [OS=Homo sapiens]	1063.16165	45.3246124	187	364	187	4128	468.788	PRKDC	-0.53	-0.04	-0.49	0
High	P05023	Sodium/potassium-translocating ATPase subunit alpha-1 [OS=Homo sapiens]	212.019335	34.3108504	28	55	28	1023	112.324	ATP1A1	-0.24	-0.04	-0.2	0
High	P63173	60S ribosomal protein L38 [OS=Homo sapiens]	147.312909	57.142871	8	67	8	70	8.213	RPL38	0.11	-0.04	0.14	0
High	P08662	Isotform 2 of 40S ribosomal protein S20 [OS=Homo sapiens]	99.823293	36.619783	7	43	7	142	15.995	RPS20	0.1	-0.04	0.14	0
High	P61254	60S ribosomal protein L26 [OS=Homo sapiens]	97.4425938	51.7241379	12	44	4	145	17.248	RPL26	0.01	-0.04	0.05	9
High	Q60264	SWI/SNF-related matrix-associated actin-dependent regulator of chromatin subunit 2B [OS=Homo sapiens]	32.9211240	11.3117871	11	14	11	1052	121.828	SMARCA5	-0.5	-0.04	-0.46	0
High	Q98XPS	serine/arginine repetitive matrix protein 2 [OS=Homo sapiens]	9.98014123	4.63789954	4	5	3	100.604	SRRT	-0.3	-0.04	-0.26	0	
High	Q95433	activator of 90 kDa heat shock protein ATPase homolog [OS=Homo sapiens]	8.606494933	10.9467456	4	5	4	338	38.25	AHS1	-0.36	-0.04	-0.33	0
High	Q9NRP0_2	Isotform 2 of Oligosaccharidyltransferase complex subunit OSTC [OS=Homo sapiens]	79.1917474	7.01754386	1	2	1	171	19.158	OSTC	-0.36	-0.04	-0.32	0
Low	P1532_1	Transcription activator BRG1 [OS=Homo sapiens]	1.84375381	0.85003036	1	1	1	1647	184.53	SMARCA4	-1.24	-0.04	-1.21	0
High	Q10570	Cleavage and polyadenylation specificity factor 1B [OS=Homo sapiens]	665.501345	58.1427581	71	303	71	1443	160.782	CPSF1	0.22	-0.05	0.27	0
High	Q9UQ35	serine/arginine repetitive matrix protein 1 [OS=Homo sapiens]	328.17988	27.2165698	56	141	56	2752	299.438	SRM2	-0.01	-0.05	0.04	0
High	Q9BQG0_2	Isotform 2 of Myb-binding protein 1A [OS=Homo sapiens]	28.8933827	7.95795796	11	15	11	1332	149.274	MYBBP1A	-0.53	-0.05	-0.48	0
High	Q9EA33_1	Coiled-coil domain-containing protein 47 [OS=Homo sapiens]	15.7809464	11.6024222	4	5	4	483	22.835	CCDC47	-0.02	-0.05	0.03	0
High	P1621_3	Isotform 3 of 60S ribosomal protein L17 [OS=Homo sapiens]	475.473928	53.9473684	19	351	19	228	26.356	P117C180RF	-0.02	-0.06	0.03	0
High	P46782	40S ribosomal protein S5 [OS=Homo sapiens]	278.628357	57.35294912	16	143	16	204	22.862	RPS2	0.08	-0.06	0.14	0
High	P46776	60S ribosomal protein L27a [OS=Homo sapiens]	118.8677126	42.5675676	8	55	8	148	16.551	RPL27A	0.07	-0.06	0.12	0
High	Q9WVX2	Calmodulin homeostatic endoplasmic reticulum protein [OS=Homo sapiens]	87.2589452	14.8471616	11	23	11	916	103.637	CHERP	-0.43	-0.06	-0.37	0
High	Q92621	Nuclear pore complex protein Nup205 [OS=Homo sapiens]	65.6068653	11.2823062	21	27	21	2012	227.776	NUP205	-0.2	-0.06	-0.14	0
High	Q9Y23X	Nucleolar protein 58 [OS=Homo sapiens]	17.1620414	12.2873346	5	5	5	529	55.541	NP58	0.26	-0.06	0.32	0
High	P46PKG0_1	60S ribosomal protein L1 [OS=Homo sapiens]	6.268144	5.25474453	3	3	3	1096	123.434	LARP1	-0.2	-0.06	-0.14	0
Low	Q9H3UJ	Protein unC5-homolog 4 [OS=Homo sapiens]	1.2128230	0.9533983	1	1	1	944	103.011	UNC5A	-0.1	-0.06	0.16	0
High	P56134	ATP synthase subunit f, mitochondrial [OS=Homo sapiens]	28.3098247	25.5319149	2	8	2	94	10.911	ATP5J2	0.05	-0.07	0.12	0
Medium	Q6P4A7_1	Sideroflexin-4 [OS=Homo sapiens]	4.19399256	7.712166172	2	2	2	337	37.974	SKXN4	-0.14	-0.07	-0.07	0
High	P7136	tubulin alpha-1A chain [OS=Homo sapiens]	57.685576	73.17079137	29	236	2	451	50.104	TUBA1A	-0.1	-0.08	-0.07	48
High	Q75964	ATP synthase subunit g, mitochondrial [OS=Homo sapiens]	59.9613605	46.6019417	4	11	4	103	11.421	ATP5L	-0.15	-0.08	-0.07	0
High	P49792	E3 SUMO-protein ligase RanBP1/2 [OS=Homo sapiens]	36.7097046	5.64516129	12	20	12	3224	357.974	RANBP2	-0.44	-0.08	-0.36	0
High	Q9H259	Zinc finger protein Es1 [OS=Homo sapiens]	15.7913962	5.388290598	3	4	3	585	64.065	IKZF4	-0.36	-0.08	-0.29	0
High	P43307	Translocon-associated protein subunit alpha 1 [OS=Homo sapiens]	10.1932122	6.634705664	2	4	2	286	32.215	SRP54	-0.06	-0.08	0.02	0
High	Q8NF37	Lysophosphatidylcholine acyltransferase 1 [OS=Homo sapiens]	9.1543903	6.367040412	2	3	2	534	59.113	LPCAT1	0.4	-0.08	0.48	0
Medium	Q9EZT3_1	SRα stem-loop-interacting RNA-binding protein, mitochondrial [OS=Homo sapiens]	3.1079054	9.17431193	1	1	1	109	12.341	SLRP	0.14	-0.08	0.21	0
High	P42387	Ubiquitin-60S ribosomal protein L40 [OS=Homo sapiens]	42.0019126	3.3745	4	21	2	128	14.719	UBA52	-0.13	-0.09	-0.05	0
High	Q9BX10	GTP-binding protein 2 [OS=Homo sapiens]	15.0952725	3.32225194	2	4	2	602	65.772	GTPBP2	-0.38	-0.09	-0.3	0
Medium	Q9BZL6	Serine/threonine-protein kinase D2 [OS=Homo sapiens]	3.56290396	2.0501139	2	2	2	878	96.689	PRKD2	-0.03	-0.09	0.05	0
High	P56192	Methionine-tRNA ligase, cytoplasmic [OS=Homo sapiens]	13.1977540	38.6666667	27	43	27	900	101.052	MARS	-0.4	-0.1	-0.29	0
High	P4P0428	UPF0428 protein CXorf56 [OS=Homo sapiens]	41.0047235	28.2883883	7	17	7	1447	164.363	COIF56	0.62	-0.1	-0.37	0
High	P25241_5	Isotform 5 of Radixin [OS=Homo sapiens]	10.3753187	4.63576159	3	4	3	604	71.005	RDX	-0.56	-0.1	-0.46	0
High	Q9NS12_1	protein FAM207A [OS=Homo sapiens]	7.72545331	5.2173913	1	3	1	230	25.441	FAM207A	0.86	-0.1	0.96	0
High	Q9ZL509	tubulin beta 3 chain [OS=Homo sapiens]	51.920025	36.2222222	18	242	1	450	50.4	TUBB3	-0.18	-0.11	-0.06	2
High	P45187	40S ribosomal protein 38 [OS=Homo sapiens]	441.487307	65.8653846	22	180	22	208	24.19	RP58	-0.12	-0.11	-0.05	0
High	Q8NTIS_1	Sister chromatid cohesion protein PDS5 homolog B [OS=Homo sapiens]	176.4853809	27.4539877	30	63	30	1304	145.738	SFB1	-0.55	-0.11	-0.44	0
High	P0C7P4	PUT4028 protein CXorf56 [OS=Homo sapiens]	9.43242493	7.7058824	2	3	2	222	25.608	PHB	-1.79	-0.12	-1.68	0
Medium	Q9PFTV0	Phaf finger-like domain-containing protein SA [OS=Homo sapiens]	3.93716124	19.0909091	2	2	2	110	12.397	PHF5A	-0.1	-0.12	0.02	0
Low	Q6PJI9_1	WD repeat-containing protein 59 [OS=Homo sapiens]	1.55098468	1.43731766	1	1	1	974	109.724	WD159	0.38	-0.12	0.5	0
High	P29597	Homeobox protein Hox-A5 [OS=Homo sapiens]	39.8121191	26.6666667	6	12	5	270	29.327	HDXA5	-0.44	-0.13	-0.31	1
High	Q9Z973_1	Transportin-1 [OS=Homo sapiens]	13.8895566	6.01332603	5	6	5	898	102.289	TPN01	-0.39	-0.13	-0.27	0
High	P25232	Isotform 2 of Splicing factor, suppression of white-apricot homolog [OS=Homo sapiens]	8.24273293	3.38983051	3	3	3	1003	109.285	SFWAP	-0.29	-0.13	-0.16	0
High	Q43871	Isocitrate dehydrogenase (NAD) subunit beta, mitochondrial [OS=Homo sapiens]	5.5618637	2.85714286	1	2	1	385	42.157	IIDH3B	0.03	-0.13	0.16	0
High	Q5JTH9_1	RRP12-like protein [OS=Homo sapiens]	274.349973	38.5505012	43	129	42	1297	143.611	RRP12	0.22	-0.14	0.36	2

High	Q9P2N5	RNA-binding protein 27 [OS=Homo sapiens]	119.542771	21.8867925	21	35	20	1060	118.645	RBM27	-0.2	-0.14	-0.06	0
High	60S ribosomal protein L19 [OS=Homo sapiens]	107.7824897	34.63238776	12	48	12	196	23.451	RPL19	-0.17	-0.14	-0.04	0	
High	Q132099	T-lymphoma invasiveness and metastasis-inducing protein 1 [OS=Homo sapiens]	57.7319812	15.4619736	20	23	20	1591	177.598	TIA1	0.11	-0.14	0.24	0
High	Q8Y4W2	Ribosomal biogenesis and metabolism protein LAS1L [OS=Homo sapiens]	33.3302278	19.7547084	11	13	11	734	83.013	LSA1L	-0.21	-0.14	-0.07	0
High	P61204	ADP-ribosylation factor 3 [OS=Homo sapiens]	29.8400792	30.9392265	4	7	2	181	20.588	ARF3	1.17	-0.14	1.92	0
High	P36542-1	ATP synthase subunit gamma, mitochondrial [OS=Homo sapiens]	204.156016	52.0134228	14	65	14	298	32.975	ATPS5C1	0.03	-0.15	0.17	0
High	P62263	40S ribosomal protein S14 [OS=Homo sapiens]	188.902267	41.7218543	9	59	9	151	16.263	RP514	0.08	-0.15	0.23	0
High	Q5C924	nucleolar MifG domain-containing protein 1 [OS=Homo sapiens]	120.094109	23.1395349	19	35	19	860	96.198	NOM1	0.24	-0.15	0.38	0
High	P6734-3	Isotform 2 of Dual specificity mitogen-activated protein kinase kinase 3 [OS=Homo sapiens]	50.3336292	35.2272227	12	21	12	352	39.914	009	0.09	-0.15	0.24	0
High	P49207	60S ribosomal protein L34 [OS=Homo sapiens]	43.2084763	38.6859585	8	26	8	117	13.284	RPL34	-0.23	-0.15	-0.08	0
High	P622891	60S ribosomal protein L39 [OS=Homo sapiens]	34.2117355	25.4501961	3	67	3	51	6.403	RPL39	-0.09	-0.15	0.07	0
High	Q9BRK9	WD repeat domain-containing protein 83 [OS=Homo sapiens]	19.0760429	14.2857143	5	6	5	315	34.321	WD83	-0.7	-0.15	-0.56	0
High	P59ULX6	A-kinase anchor protein 8-like [OS=Homo sapiens]	6.51216232	4.48916409	3	3	3	646	71.604	AKAP8L	-0.18	-0.15	-0.04	0
Medium	P55658-5	Protein 5 of nuclear pore complex protein Nup214 [OS=Homo sapiens]	2.9030899	6.62111801	1	1	1	2093	215.269	NUP214	-0.83	-0.15	-0.59	0
High	P622888	60S ribosomal protein L30 [OS=Homo sapiens]	93.4056582	66.0869565	7	33	7	115	12.776	RPL30	-0.32	-0.16	-0.16	0
High	P28340	DNA polymerase delta catalytic subunit [OS=Homo sapiens]	32.917106	12.6467931	13	15	12	1107	123.553	POLD1	-0.22	-0.16	-0.06	0
High	P50772	Translational endoplasmic reticulum ATPase [OS=Homo sapiens]	10.8363175	8.21836228	3	4	3	806	83.266	VCP	-0.47	-0.16	-0.31	0
High	Q8PFQ7	Tetratricopeptide repeat protein 37 [OS=Homo sapiens]	5.77717368	1.91815857	3	4	3	1564	175.375	TTC37	-0.7	-0.16	-0.54	0
Low	O60341-2	Isotform 2 of lysine-specific histone demethylase 1A [OS=Homo sapiens]	2.15893535	0.68493151	1	1	1	876	95.096	KDM1A	-0.79	-0.16	-0.63	0
High	P62753	40S RIBOSOMAL PROTEIN 6S [OS=Homo sapiens]	646.388723	57.028124	29	308	29	249	28.663	RPS6	-0.07	-0.17	0.11	0
High	Q14498-2	Isotform 2 of RNA-binding protein 39 [OS=Homo sapiens]	531.7935074	57.6335578	26	238	23	524	58.62	RNP39	0.02	-0.17	0.2	0
High	P62841	40S ribosomal protein S15 [OS=Homo sapiens]	448.307495	67.5862069	7	27	7	145	17.029	RPS15	-0.27	-0.17	-0.1	0
High	Q9C018	pre-mRNA 3' end processing protein WDR33 [OS=Homo sapiens]	390.190579	47.6047904	56	160	56	1336	145.799	WDR33	0.08	-0.17	0.25	0
High	P12236	ADP/ATP translocase [OS=Homo sapiens]	361.24692	63.2428188	22	180	3	298	32.845	SLC25A6	-0.14	-0.17	0.03	6
High	Q15459	Splicing factor 3A subunit 1 [OS=Homo sapiens]	28.0986207	12.2320303	8	10	8	793	88.831	SF3A1	-0.66	-0.17	-0.49	0
High	P61353	60S ribosomal protein L27 [OS=Homo sapiens]	24.4089129	30.8823529	4	9	4	136	15.788	RPL27	-0.23	-0.17	-0.06	0
High	Q8NRZ9	Lymphoid-specific helicase [OS=Homo sapiens]	21.3382957	7.99532673	6	9	6	838	97.012	HELI5	-0.04	-0.17	0.13	0
High	Q9HC57	pre-mRNA splicing factor SF1 [OS=Homo sapiens]	18.4021143	8.30409557	6	6	6	855	99.946	XAB2	0.15	-0.17	0.32	0
High	Q9HC62	Sentrin-specific protease 2 [OS=Homo sapiens]	17.8764831	8.14940577	4	5	4	589	67.812	SNP2	-0.42	-0.17	-0.25	0
Medium	P00492	Hypothiamine-glutamine phosphoribosyltransferase [OS=Homo sapiens]	3.04973253	4.58715596	1	1	1	218	24.564	HPT1	-0.47	-0.17	-0.31	0
Medium	P23258	Tubulin gamma 1-chain [OS=Homo sapiens]	2.781239963	3.99113082	2	2	2	451	51.138	TUBG1	0.78	-0.17	0.96	0
Low	Q8N823-1	Zinc finger protein 611 [OS=Homo sapiens]	1.60739379	1.1347517	2	1	1	705	81.397	ZNFX1	-0.12	-0.17	0.05	0
High	Q8Y323	deoxyribonuclease triphosphate triphosphohydrodase SAMHD1 [OS=Homo sapiens]	64.1382539	21.884984	13	24	13	626	72.155	SAMHD1	-0.03	-0.18	0.14	0
High	P19755	Transmembrane emp24 domain-containing protein 10 [OS=Homo sapiens]	33.6059667	13.6986301	3	7	3	219	24.96	TMED10	-0.11	-0.18	0.07	0
High	P26304	Small nuclear ribonucleoprotein E [OS=Homo sapiens]	11.2453015	6.1956217	1	4	1	92	10.797	SNRPE	-0.26	-0.18	-0.65	0
High	O15371	Eukaryotic translation initiation factor 3 subunit D [OS=Homo sapiens]	6.14342135	3.83211679	2	3	2	548	63.932	EIF3D	-0.83	-0.18	-0.65	0
High	P78549	Endonuclease III-like protein 1 [OS=Homo sapiens]	9.95284287	14.4230769	4	4	4	312	34.368	NTHL1	-0.27	-0.19	-0.07	0
High	Q9ED072	Isotform 2 of 6S small nuclear ribonucleoprotein 40 kDa protein [OS=Homo sapiens]	1.60739379	1.1347517	2	2	2	409	44.488	SNRNP40	-0.41	-0.19	-0.22	0
High	Q9Y323	SNW domain-containing protein 1 [OS=Homo sapiens]	6.19027104	2.1884984	2	2	2	536	61.456	SNW1	-0.24	-0.19	-0.14	0
Medium	Q860X5	Up-regulated during skeletal muscle growth protein 5 [OS=Homo sapiens]	3.57384147	17.24313793	1	1	1	58	64.453	UMG5	-0.21	-0.19	-0.02	0
High	Q7Z669	E3 ubiquitin-protein ligase RBBP6 [OS=Homo sapiens]	358.6261317	36.1049107	62	170	62	1792	201.142	RBBP6	-0.81	-0.2	-0.61	0
High	P62750	60S ribosomal protein L23a [OS=Homo sapiens]	209.056546	48.0769231	12	64	12	156	17.684	RPL23A	-0.09	-0.2	0.11	0
High	Q9BQ67	Glutamateric rich WD repeat-containing protein 1 [OS=Homo sapiens]	7.500165949	7.62331939	3	3	3	446	49.388	GWMD1	-0.02	-0.2	0.17	0
Low	Q8N753	Transformation/transcription domain-associated protein 10 [OS=Homo sapiens]	1.94500414	2.03322104	1	1	1	3859	437.318	TRRAP	-0.54	-0.2	-0.34	0
High	O43592	Exportin-T [OS=Homo sapiens]	141.817427	25.2598753	20	40	20	962	109.893	XPO1	-0.15	-0.21	0.07	0
High	P12004	proliferating cell nuclear antigen [OS=Homo sapiens]	89.8809954	65.42712414	15	30	15	261	77.75	PCNA	-0.31	-0.21	0.1	0
High	P23223-3	Isotform 3 of Pyruvate-carboxylate reductase 1, mitochondrial [OS=Homo sapiens]	36.5618575	18.4971098	5	10	4	346	35.958	PCN1	0.08	-0.21	0.29	1
High	P52292	Importin subunit alpha-1 [OS=Homo sapiens]	11.7078176	4.15879017	2	4	2	529	57.826	KIN12A2	-0.56	-0.21	-0.34	0
High	P56385	ATP synthase subunit e, mitochondrial [OS=Homo sapiens]	8.27088019	15.3225805	5	5	5	69	7.928	ATP5I	0.14	-0.21	0.35	0
High	Q96571	GDP-mannose 4,6 dehydrogenase [OS=Homo sapiens]	7.75280897	2.81329923	1	2	1	391	43.923	GMDS	-0.39	-0.21	-0.18	0
High	P55209	Nucleosome assembly protein 1-like 1 [OS=Homo sapiens]	7.63204543	7.83696906	2	2	2	319	45.346	NAP1L1	-0.71	-0.21	-0.5	0
High	Q8TB37-1	Iron-sulfur protein NUP1 [OS=Homo sapiens]	2.10980261	4.80309345	1	1	1	229	71.325	NUP1	0.08	-0.21	0.28	0
Low	PE0468	Protein transport protein Sec61 subunit beta [OS=Homo sapiens]	2.02029557	11.4583333	1	1	1	96	9.968	SEC61B	-0.23	-0.21	-0.02	0
Low	Q9NVU7	Protein SDA1 homolog [OS=Homo sapiens]	1.48332097	0.87336245	1	1	1	687	79.82	SDA1	0.33	-0.21	0.54	0
High	Q8BR88	G patch domain-containing protein 1 [OS=Homo sapiens]	133.281515	26.6380236	19	36	19	931	103.282	GPATCH1	-0.81	-0.22	-0.59	0
High	Q9NV17	ATPase family AAA domain-containing protein 3A [OS=Homo sapiens]	2.36260705	12.6182965	8	12	8	634	ATAD3A	-0.22	-0.22	0	0	
High	Q9Y388	RNA-binding motif protein X-linked 2 [OS=Homo sapiens]	19.1984032	10.2484472	3	5	3	322	37.313	REMX2	0.34	-0.22	0.56	0
High	Q9UHR5-1	SAP30-binding protein [OS=Homo sapiens]	13.3812551	11.3636564	3	3	3	308	33.35	SAP30BP	-0.42	-0.22	-0.2	0
High	Q5T30-3	Isotform 3 of G patch domain-containing protein 4 [OS=Homo sapiens]	11.5113341	5.54323325	3	5	3	451	50.897	GPATCH4	-0.2	-0.22	0.01	0
High	Q6NUK1	Calcium-binding mitochondrial carrier protein ScmC-1 [OS=Homo sapiens]	5.24933983	4.19287112	2	3	2	477	53.32	SLC2AA24	-0.32	-0.22	-0.01	0
High	Q8P210	Cleavage and polyadenylation specificity factor subunit 2 [OS=Homo sapiens]	427.0005538	66.8797954	44	109	44	782	88.431	CPSF2	0.19	-0.23	0.42	0
High	Q6P2Q9	Pre-mRNA-processing-splicing factor 8 [OS=Homo sapiens]	310.220259	34.0042827	67	110	67	2335	273.427	PRPF8	-0.3	-0.23	-0.07	0
High	Q8BXF6	Rab11 family-interacting protein 5 [OS=Homo sapiens]	238.187218	58.0398162	29	72	29	653	70.372	RAB11FIP5	0.03	-0.23	0.26	0

High	Q4KMP7	TBC1 domain family member 10B [OS=Homo sapiens]	178,677476	36,2623762	24	54	24	808	87,145	TBC1D10B	-0.16	-0.23	0.07	0
High	P5786	puromycin-sensitive aminopeptidase [OS=Homo sapiens]	136,118007	55,560392	24	42	24	919	103,211	NOPPPS	-0.44	-0.23	-0.22	0
High	Q13724-1	Mannosyl-oligosaccharide glucosidase [OS=Homo sapiens]	119,110916	31,5412186	20	35	20	837	91,861	MGCS	0.07	-0.23	0.3	0
High	Q8NWK9	Box C/D snoRNA protein 1 [OS=Homo sapiens]	19,8750405	12,1276396	5	6	5	470	53,884	ZNHT6	-0.02	-0.23	0.21	0
High	Q8TE00	U3 small nucleolar RNA-associated protein 15 homolog [OS=Homo sapiens]	10,8161371	7,72200772	4	5	4	518	58,379	UTP15	-0.19	-0.23	0.04	0
High	Q9Y305-4	Isotform 4 of Acyl-coenzyme A thioesterase 9, mitochondrial [OS=Homo sapiens]	5,33854005	4,24107143	2	3	2	448	50,818	ACOT9	-0.26	-0.23	-0.03	0
High	Q6P1X5	Transcription initiation factor TFIID subunit 2 [OS=Homo sapiens]	769,268011	57,4645538	68	250	68	1199	136,883	TAF2	-0.53	-0.24	-0.29	0
High	Q9Y383	Putative RNA-binding protein Luc7-like 2 [OS=Homo sapiens]	700,1903055	62,5	40	463	32	392	46,486	LCU7L2	-0.11	-0.24	0.13	19
High	Q9H329-2	Isotform 2 of Band 4.1-like protein 4B [OS=Homo sapiens]	190,789918	43,8223938	20	67	18	518	58,529	EPB41L4B	0.25	-0.24	0.49	0
High	Q9325-2	Isotform B of Phosphate carrier protein 1, mitochondrial [OS=Homo sapiens]	162,79495	47,3684211	18	76	18	361	39,933	SLC25A3	-0.21	-0.24	-0.24	0
High	Q8NVP1	ATP-dependent RNA helicase DDX18 [OS=Homo sapiens]	101,452224	29,2537213	15	37	15	670	75,359	DDX18	0.22	-0.24	0.45	0
High	Q14974	Imidithio subunit beta-1 [OS=Homo sapiens]	51,0630732	14,4977169	11	17	11	876	97,108	KNNB1	-1.23	-0.24	-0.39	0
High	Q8WVY3	U4/6 small nuclear ribonucleoprotein Prp31 [OS=Homo sapiens]	7,20166663	8,01603206	3	3	3	499	55,421	PRPF31	-0.51	-0.24	-0.27	0
High	Q8NWV8	BRCA2 and SRSCL AC complex member 1 [OS=Homo sapiens]	6,80760345	6,07902236	2	2	2	329	36,537	BABAM1	0.43	-0.24	0.68	0
Low	Q7ZWF7-2	Isotform 2 of 39S ribosomal protein L55, mitochondrial [OS=Homo sapiens]	1,13990167	4,87804678	1	1	1	164	18,891	MRPL55	-0.27	-0.24	-0.03	0
High	Q7563-1	U5 small nuclear ribonucleoprotein 200 kDa helicase [OS=Homo sapiens]	390,229913	41,4325843	67	118	67	2136	244,353	SNRNP200	-0.18	-0.25	0.07	0
High	Q8JL4	E3 SG15-protein ligase HERC5 [OS=Homo sapiens]	349,503507	47,653507	42	92	42	1024	116,777	HERC5	-0.11	-0.25	0.14	0
High	Q14683	structural maintenance of chromosomes protein 1 alpha [OS=Homo sapiens]	117,087594	23,0332522	26	48	26	1233	143,144	SMC1A	-0.28	-0.25	-0.03	0
High	Q9HWV2-5	Bromodomain-containing protein 9 [OS=Homo sapiens]	104,472246	27,031826	13	30	13	597	66,958	BRD9	-0.41	-0.25	-0.16	0
High	Q8LGY1	Nucleolar protein 12 [OS=Homo sapiens]	50,8979685	46,093897	7	17	7	213	24,648	NOL12	-0.4	-0.25	-0.15	0
High	Q63036	Intron-binding protein aquilin [OS=Homo sapiens]	48,631451	11,13131615	13	20	12	1485	171,186	AQR	0.34	-0.25	0.59	0
High	Q8NX58	Cell growth-regulating nuclear protein [OS=Homo sapiens]	18,9874841	18,9973615	6	8	6	379	43,588	LYAR	-0.28	-0.25	-0.04	0
High	P46977	Dolichyl-diphosphooligosaccharide–protein Glycosyltransferase subunit STT3A [OS=Homo sapiens]	9,7833673	3,388794326	3	4	3	705	80,477	STT3A	-0.92	-0.25	-0.67	0
High	Q9ULX3	RNA-binding protein NOL1 [OS=Homo sapiens]	5,09520193	5,092109447	1	2	1	412	46,646	NOBL1	0.28	-0.26	0.52	0
High	P12235	ADP/ATP translocase 1 [OS=Homo sapiens]	373,23120854	68,1208054	22	167	4	298	33,043	SLC25A4	-0.22	-0.26	0.04	0
High	Q13200	26S proteasome non-ATPase regulatory subunit 2 [OS=Homo sapiens]	73,6233913	13,6563877	9	18	9	908	100,136	PSMD2	-0.6	-0.26	-0.35	0
High	QDFG8-6	Isotform 6 of Extended synaptotagmin-2 [OS=Homo sapiens]	21,5584186	9,12951168	6	8	6	942	104,643	SYNE1	0.79	-0.26	1.05	0
High	P87V18	pre-mRNA-splicing factor 1 [OS=Homo sapiens]	15,7625697	7,69230769	5	9	5	546	64,429	PRP38B	0	-0.26	0.26	0
High	Q14617-5	Isotform 5 of Arp 3 complex subunit delta-1 [OS=Homo sapiens]	6,24633261	1,97530864	2	2	2	1215	136,565	AP3D1	-0.38	-0.26	-0.12	0
Medium	P45594	Shtorbi/bifurcated chain-specific acyl-CoA dehydrogenase, mitochondrial [OS=Holo-] [OS=Holo-] [OS=Holo-] [OS=Holo-]	2,13663759	1,65185185	1	1	1	432	47,455	ACADS8	-0.22	-0.26	0.04	0
Low	Q6ZW31-1	Nucleolar GrPase-activating protein SYDEL [OS=Homo sapiens]	1,5079384	1,63265206	1	1	1	735	79,743	SYDEL	0.12	-0.26	-0.37	0
High	Q96920	60S ribosomal protein L26-like [OS=Homo sapiens]	143,334193	44,3595623	10	16	1	106	12,461	RPL36AL	-0.08	-0.27	0.19	0
High	Q16531	DNA damage-binding protein 1 [OS=Homo sapiens]	93,4904837	25,1754386	22	34	22	1140	126,887	DBP1	-0.17	-0.27	0.11	0
High	Q8N6R0-5	Methylestertransferase-like protein 13 [OS=Homo sapiens]	34,4520372	10,0143062	7	12	7	699	78,718	METTL3	0.1	-0.27	0.38	0
High	Q83597	Succinyl-CoA ligase (ADP-GDP-forming) subunit alpha, mitochondrial [OS=Holo-]	17,8339568	3,18387283	3	6	3	346	36,227	SUCNLG1	-0.22	-0.27	0.05	0
High	Q8WVA0	WD repeat-containing protein 35 [Osmo-] [Osmo-] [Osmo-] [Osmo-]	7,38863221	4,09638354	3	4	3	830	90,438	WDRT5	0.49	-0.27	0.76	0
High	Q10181	Splicing factor USAF1	217,891217	51,25	13	83	13	240	27,854	Q10724594	-0.16	-0.28	0.13	0
High	P62910	60S ribosomal protein L32-like [OS=Homo sapiens]	65,1406165	39,259293	7	16	7	135	15,85	RPL32	-0.13	-0.28	0.15	0
High	Q16531	DNA damage-binding protein 1 [OS=Homo sapiens]	93,4904837	25,1754386	3	5	3	1047	119,338	SUPT16H	-1.06	-0.28	-0.78	0
High	Q8N6R0-5	Methylestertransferase-like protein 13 [OS=Homo sapiens]	5,30319151	2,64	3	4	3	125	14,576	MEFTB4	-1.38	-0.28	-1.1	0
High	P46977	Elongation factor 1-alpha 1 [OS=Homo sapiens]	192,457137	52,8138528	19	84	19	462	50,109	EEFA1	-0.81	-0.29	-0.52	0
High	Q8Z365	Poly(RC)-binding protein 1 [OS=Homo sapiens]	115,147953	67,1328703	16	35	11	356	37,474	PBP1	-0.49	-0.29	-0.19	8
High	P35618	Cofactor subunit beta [OS=Homo sapiens]	64,7895193	14,9003148	11	21	11	953	107,074	COPB1	0.73	-0.29	0.43	0
High	P53985	Monocarboxylate transporter 1 [OS=Homo sapiens]	41,6189008	17,4	7	17	7	500	53,909	SLC16A1	-0.41	-0.29	-0.12	0
High	P61619	Protein transport protein Sec61 subunit alpha isoform 1 [OS=Homo sapiens]	32,9898583	16,3865546	5	9	5	476	52,231	SEC61A1	-0.38	-0.29	-0.09	0
High	P66561	phosphatidylserine synthase [OS=Homo sapiens]	11,1435429	7,0182424	2	8	2	473	55,491	PTBS1	0.35	-0.29	0.65	0
High	P40938	replication factor C subunit 3 [OS=Homo sapiens]	11,1502894	8,38876404	2	2	2	356	40,53	RCF3	-0.12	-0.29	0.17	0
High	Q9UQ03	Integrator complex subunit 6 [OS=Homo sapiens]	7,02103931	2,9312286	3	4	3	887	100,326	INTS6	0.12	-0.29	0.41	0
High	Q8WTT2	Nucleolar complex protein 3 homolog [OS=Homo sapiens]	201,762809	43	32	61	32	800	92,49	NOC3L	-0.19	-0.3	0.1	0
High	Q9GN5-1	F-Act complex subunit SP176 [OS=Homo sapiens]	189,997391	44,2538593	25	54	25	583	66,164	PARP2	0.21	-0.31	0.51	0
High	Q8Y384	Splicing factor 38 subunit 6 [OS=Homo sapiens]	32,6386419	27,42474792	8	10	8	299	33,276	PHB2	-1.86	-0.3	-1.56	0
High	P68104	Elongation factor 1-alpha 1 [OS=Homo sapiens]	115,147953	67,1328703	1	1	1	793	35,264	PLRKHH3	-0.03	-0.27	0.27	0
Low	Q98ZL1	ubiquitin-like protein 5 [OS=Homo sapiens]	2,14819149	12,3287671	1	1	1	73	8,541	UBL5	-0.41	-0.3	-0.12	0
High	Q8WT2	Luc7-like protein 3 [OS=Homo sapiens]	445,8131232	62,7314819	37	287	37	432	51,435	UCL7L3	-0.16	-0.31	0.14	0
High	Q9NFX1	Testis-expressed sequence 10 protein [OS=Homo sapiens]	172,319532	36,7061356	28	53	28	929	105,608	TEX10	-0.56	-0.31	0.31	0
High	Q14244	Enscs1 [OS=Homo sapiens]	90,4327392	25,361562	20	37	19	749	84,002	MAP7	-0.02	-0.31	0.28	1
Medium	P06576	pleckstrin homology domain-containing family H member 3 [OS=Homo sapiens]	3,78015361	1,51323086	12	17	12	529	56,525	ATP5B	-0.27	-0.31	0.04	0
High	Q00483	Cytochrome c oxidase subunit NDUF4A [OS=Homo sapiens]	26,9795059	45,6790123	5	10	5	81	9,364	NDUF4A	-0.1	-0.31	0.21	0
High	Q16629	serine/arginine-rich splicing factor 7 [OS=Homo sapiens]	23,1583319	27,3103944	5	11	5	238	27,35	SRSF7	0.3	-0.31	0.61	0
High	Q98805	Transmembrane 9 superfamily member 2 [OS=Homo sapiens]	16,6293592	4,3740532	3	7	3	663	17,725	TM9SF2	0.01	-0.31	0.31	0
High	Q94788-1	retinaldehyde dehydrogenase 2 [OS=Homo sapiens]	5,89073902	3,8888809	3	3	1	518	56,688	ALDH1A2	-0.33	-0.22	0.02	0
High	Q9U4K8	Protein argonaute-2 [OS=Homo sapiens]	268,491408	57,9743888	38	79	38	859	97,146	AGO2	-0.33	-0.32	0	0
High	Q15029	116 kDa U5 small nuclear ribonucleoprotein component [OS=Homo sapiens]	155,263681	37,5514403	27	44	26	972	109,366	EF3UD2	-0.24	-0.32	0.08	1
High	Q9NR8C	NAD-dependent protein deacetylase sirtuin 7 [OS=Homo sapiens]	83,551391	33,5	10	19	10	400	44,87	SIRT7	0.2	-0.32	0.51	0

High	Q9NP97	Dynein light chain rod/block-type 1 [OS=Homo sapiens]	29.3533413	33.3333333	3	7	3	96	10.915	DYNLRB1	-0.13	-0.32	0.18	0
High	P02545	Prelaminin-A/C [OS=Homo sapiens]	10.0131002	6.62650602	4	6	3	664	74.095	LMNA	-0.86	-0.32	-0.54	0
High	Q9P2V4	Zinc-finger protein 21.9 [OS=Homo sapiens]	8.37543013	30.3571429	3	4	3	722	76.53	ZNF119	-1.58	-0.32	-1.26	0
High	Q5L539	Transcription elongation factor II polypeptide 1 [OS=Homo sapiens]	8.37543013	1.33086915	0.49875512	1	1	2005	12.465	TCEB1	-0.31	-0.32	0.01	0
Low	A6NH99	Structural maintenance of chromosomes flexible hinge domain-containing protein 1 [OS=Homo sapiens]	200.16928	44.6215139	31	61	23	226.231	SMCHD1	-2.15	-0.32	-1.83	0	
High	P27448	MAP/microtubule affinity-regulating kinase 3 [OS=Homo sapiens]	13.1439335	13.618677	4	5	4	257	84.436	MARK3	-0.01	-0.33	0.32	6
High	P28294	28S ribosomal protein S15, mitochondrial [OS=Homo sapiens]	13.0293582	6.7357513	4	5	4	579	29.823	MPS5J5	-2.29	-0.33	2.62	0
High	Q9Y5A9	YTH domain-containing family protein 2 [OS=Homo sapiens]	5.50057412	2.60021668	2	2	2	923	62.296	YTHDF2	-0.2	-0.33	0.13	0
High	Q8V7T5	kinase suppressor of Ras 1 [OS=Homo sapiens]	5.50057412	5.60021668	2	2	2	102.096	0.03	KSRL	0.03	-0.33	0.36	0
High	P04121	Arginine/serine-rich coiled-coil protein 2 [OS=Homo sapiens]	5.54358178	5.76036866	3	3	3	434	50.529	RSHC2	-0.38	-0.33	-0.33	0
Medium	Q5VYK3	Proteasome-associated protein FCM29 homolog [OS=Homo sapiens]	4.24842857	1.62601626	3	3	3	1845	204.16	KIA0368	-0.9	-0.33	-0.57	0
High	P02584	40S ribosomal protein S26 [OS=Homo sapiens]	170.58864	52.173913	7	116	7	115	13.007	C101293876	-0.22	-0.34	0.12	0
High	P26641	Isotform 2 of elongation factor 1-gamma [OS=Homo sapiens]	124.75646	50.308082	20	44	20	487	56.114	EIF16	-1.16	-0.34	-0.82	0
High	Q43175	D-3-phosphoglycerate dehydrogenase [OS=Homo sapiens]	107.403916	42.2138837	16	35	16	533	56.614	PGNDH	-0.6	-0.34	-0.25	0
High	P14406	Cytochrome c oxidase subunit 7A2, mitochondrial [OS=Homo sapiens]	7.40214747	15.6626306	1	2	1	83	9.39	CODTA2	-0.24	-0.34	0.1	0
Medium	Q9N222	Midasin [OS=Homo sapiens]	3.17608859	0.37536805	2	2	2	5596	632.42	MDN1	-0.18	-0.34	0.16	0
High	P26368	Splicing factor U2AF 65 kDa subunit [OS=Homo sapiens]	441.097293	64.2105263	25	152	25	475	53.467	U2AF2	-0.19	-0.35	0.17	0
High	P25705	ATP synthase subunit alpha, mitochondrial [OS=Homo sapiens]	292.65724	59.3128391	27	73	27	553	59.714	ATPSA2	-0.29	-0.35	0.06	0
High	Q9NYV4	Cyclin-dependent kinase 12 [OS=Homo sapiens]	91.2547468	19.664495	23	40	18	1490	164.054	CDK12	-0.31	-0.35	0.03	5
High	Q9NW78	Aurora kinase A-interacting protein [OS=Homo sapiens]	27.7450546	1.05202513	3	9	3	199	22.34	AURKAIP1	-0.23	-0.35	0.11	0
High	Q3701	CCAT/enhancer-binding protein zeta [OS=Homo sapiens]	11.2888959	3.38481973	4	5	4	1054	120.598	CEBPZ	0.01	-0.35	0.36	0
High	Q5JVF3	Isotform 4 of PC1 domain-containing protein 2 [OS=Homo sapiens]	6.65700841	6.18101545	2	2	2	453	52.066	PCID2	-0.55	-0.35	-0.21	0
High	Q99459	Cell division cycle 5-like protein [OS=Homo sapiens]	155.873823	46.1346633	26	43	26	802	92.194	CDC5L	-0.24	-0.36	0.12	0
High	Q9Y80	Exportin-1 [LP] homolog [OS=Homo sapiens]	117.624636	27.3576097	24	40	24	1071	123.306	XPO1	-0.84	-0.36	-0.48	0
High	Q8BR76	Protein LP homolog [OS=Homo sapiens]	105.043339	44.9612403	6	26	6	129	15.215	LPLH	-0.44	-0.36	-0.08	0
High	P08865	40S ribosomal protein SA [OS=Homo sapiens]	87.7374086	48.1355932	10	22	10	295	32.833	RPSA	-0.23	-0.36	0.13	0
High	Q8N2M8	CK4-associating serinearginine-rich protein [OS=Homo sapiens]	36.8895205	17.5074184	7	17	7	674	77.115	CASRP	0.4	-0.36	0.76	0
High	Q9I521	Probable ATP-dependent RNA helicase DHX35 [OS=Homo sapiens]	24.90129134	13.5351305	7	8	7	703	78.86	DHH35	-0.94	-0.36	-0.58	0
High	Q95299	Isotform 2 of NADH dehydrogenase (ubiquinone) 1 alpha complex subunit 11 [OS=Homo sapiens]	19.2663036	6.60606006	3	4	3	429	48.532	NDUFA10	0.12	-0.36	0.48	0
High	P05198	Eukaryotic translation initiation factor 2 subunit 1 [OS=Homo sapiens]	6.530948039	9.84126384	3	3	3	315	36.089	EIF2S1	-0.82	-0.36	-0.46	0
High	Q8UHX1	Isotform 2 of Poly(U)-binding splicing factor PUF60 [OS=Homo sapiens]	256.262206	63.2841328	27	66	27	542	58.135	PUF60	-0.19	-0.37	0.18	0
High	P03435	Splicing factor 3B subunit 2 [OS=Homo sapiens]	70.79151731	16	16	895	100.165	SF3B2	-0.98	-0.37	-0.51	0		
High	P48047	ATP synthase subunit O, mitochondrial [OS=Homo sapiens]	51.3162719	48.3568075	8	13	8	213	23.263	ATP5O	-0.23	-0.37	0.14	0
High	Q95519	serinearginine-rich splicing factor 11 [OS=Homo sapiens]	29.8027691	12.3966942	4	13	4	484	53.51	SF3F11	-0.22	-0.37	0.15	0
High	Q9480	nuclear mitotic apparatus protein 1 [OS=Homo sapiens]	29.7308964	6.00472813	10	12	10	2115	238.115	NUMA1	-0.59	-0.37	-0.22	0
High	P07251	NAFH dehydrogenase (ubiquinone) iron-sulfur protein 7, mitochondrial [OS=Hc]	7.70295519	4.23525321	1	2	1	213	23.548	NUFUF7	-0.09	-0.37	0.28	0
High	P19474	E3 ubiquitin protein ligase TRIM21 [OS=Homo sapiens]	496.934635	66.9473684	42	175	42	475	54.135	TRIM21	-0.32	-0.38	0.06	0
High	Q95639	Cleavage and polyadenylation specificity factor subunit 4 [OS=Homo sapiens]	201.239255	61.33829	14	60	14	895	100.165	SF3B2	-0.07	-0.38	0.31	0
High	P48047	ATP synthase subunit O, mitochondrial [OS=Homo sapiens]	51.3162719	48.3568075	3	3	3	306	32.444	ATP5O	-0.23	-0.37	0.14	0
High	Q95519	serinearginine-rich splicing factor 11 [OS=Homo sapiens]	29.8027691	12.3966942	1	2	1	153	17.75	SF3F18	-0.71	-0.38	0.33	0
High	Q9480	nuclear mitotic apparatus protein 1 [OS=Homo sapiens]	29.7308964	6.00472813	1	1	1	794	89.54	UFL1	-1.18	-0.38	-1.1	0
High	P07251	NAFH dehydrogenase (ubiquinone) iron-sulfur protein 7, mitochondrial [OS=Hc]	7.70295519	4.23525321	1	2	1	594	66.487	FPLLI	-0.12	-0.39	0.28	43
High	P22806	Small nuclear ribonucleoprotein F [OS=Homo sapiens]	42.2286144	24.4186094	2	7	2	86	97.79	SNRPF	-0.07	-0.39	0.32	0
High	P12956	X-ray repair cross-complementing protein 6 [OS=Homo sapiens]	17.00634399	7.8817734	4	5	4	609	69.799	XRC6	-0.54	-0.39	-0.15	0
High	Q9Y3C1	nucleolar protein 16 [OS=Homo sapiens]	146.598239	60.6741573	13	36	13	178	21.175	NOP16	0.17	-0.4	0.57	0
High	Q96H12	Acylglycerol kinase, mitochondrial [OS=Homo sapiens]	39.8469582	9	15	9	422	47.107	AGK	-0.35	-0.4	0.05	0	
High	P04937	Replication factor C subunit 5 [OS=Homo sapiens]	18.7833108	16.4705882	5	6	5	340	38.472	RECF5	-0.35	-0.4	0.06	0
High	Q5KPK0	MitCOS complex subunit M1C13 [OS=Homo sapiens]	8.64121076	28.813593	3	3	3	118	13.079	C13orf70	-0.15	-0.4	0.25	0
Medium	P26506	Cootomer subunit beta [OS=Homo sapiens]	4.54913958	1.93867597	2	3	2	906	102.422	COPB2	-0.49	-0.4	-0.09	0
Low	Q9UBK9	RNA-binding protein SAP18 [OS=Homo sapiens]	1.06340751	2.0114925	1	1	1	348	38.767	ASMA1	-0.6	-0.4	-0.2	0
High	P04422	Histone deacetylase complex subunit SAP18 [OS=Homo sapiens]	43.0252898	4.5751634	1	2	1	660	74.996	KIF2A	-0.24	-0.41	0.17	0
Low	Q94874	E3 FM1-protein ligase 1 [OS=Homo sapiens]	17.44563967	1.25944584	1	1	1	620	71.562	FRC1	-0.35	-0.41	0.06	0
High	Q96N15	PremRNA 3'-end-processing factor Fip1 [OS=Homo sapiens]	42.2286144	19.4144444	11	21	11	684	28.143	TR58R	-0.4	-0.41	0.01	0
High	P07437	Cleavage and polyadenylation specificity factor subunit 3 [OS=Homo sapiens]	80.821521	19.4444444	11	20	16	837	96.758	TPF11	-0.7	-0.42	0.51	0
High	Q9UBK9	Tufelmin interacting protein 11 [OS=Homo sapiens]	57.3921874	22.7001195	16	61	20	134	15.19	ZNF593	-0.15	-0.42	0.28	0
High	Q9Y3C1	Protein regulation of cyclin kinase 1 [OS=Homo sapiens]	19.0754954	17.1641791	2	4	2	140	15.045	FFN1	-0.37	-0.42	0.94	0
High	Q96S44	TP53-regulating kinase [OS=Homo sapiens]	15.942627	3.174142871	4	8	4	1683	192.957	SP1PRH	-0.74	-0.42	-0.32	0
High	P07437	E3 ubiquitin-protein ligase SHPRH [OS=Homo sapiens]	9.01230886	2.7332145	4	4	4	444	49.639	TUBB	-0.37	-0.43	0.06	5
High	Q9UBK9	Tubulin beta chain [OS=Homo sapiens]	91.521076	83.3333333	31	430	5	430	57.727	UPF3B	-0.13	-0.43	0.3	0
High	Q9UBK9-1	Regulator of nonsense transcripts 3B [OS=Homo sapiens]	130.662532	38.9233954	20	41	20	483	96.372	SART1	-0.22	-0.43	0.49	0
High	P04320	U4/U6.U5 tri-snRNP-associated protein 1 [OS=Homo sapiens]	96.3730233	28.875	16	24	16	800	90.72	NDUFA11	-0.8	-0.43	-0.37	0
High	P78347	General transcription factor IIa [OS=Homo sapiens]	77.7468891	21.742487	19	30	19	998	112.346	GTF2I	-0.8	-0.43	0.88	0
High	Q9Y5B6	PAF3 and PAF7-binding protein 1 [OS=Homo sapiens]	58.3897935	20.719783	15	17	15	917	104.739	SCFC1;PAXBP	0.45	-0.43	-0.43	0
High	Q8EY39	NAFH dehydrogenase (ubiquinone) 1 alpha subcomplex subunit 11 [OS=Homo	56.8735568	57.4468085	5	13	5	141	14.843	NDUFA11	-0.28	-0.43	0.15	0

High	Q9UYK3-1	Apoptotic chromatin condensation inducer in the nucleus [OS=Homo sapiens]	35_4104819	8_12826249	9	12	9	1341	151_771	A1C1N1	-0.7	-0.43	-0.27	0
High	Q96F12	Dynein light chain 2, cytoplasmic [OS=Homo sapiens]	34_0392244	39_3258427	5	9	2	89	10_343	D1NL2	-0.52	-0.43	-0.09	3
High	Q53GQ0	Very-long-chain-3-oxacyl-CoA reductase [OS=Homo sapiens]	31_1593832	18_5897336	4	9	4	312	34_302	HSE1TB12	-0.11	-0.43	0.33	0
High	O14545	TRAF-type zinc finger domain-containing protein 1 [OS=Homo sapiens]	21_8879701	8_24742268	4	7	4	582	64_8	TRAED1	0.26	-0.43	0.69	0
High	Q63HN8-4	Isotform 2 of E3 ubiquitin-protein ligase RNF233 [OS=Homo sapiens]	11_88667257	0_91334201	4	6	4	5256	596_106	RNF213	-0.65	-0.43	-0.22	0
High	Q96K58-2	Isotform 2 of Zinc finger protein 668 [OS=Homo sapiens]	9_31586597	10_74766336	4	4	4	642	70_46	MMP6SPH10	-0.26	-0.43	0.17	0
High	O00566	U3 small nucleolar ribonucleoprotein protein Mpp10 [OS=Homo sapiens]	25_15691	8_95741557	4	7	4	681	78_816	WDrepeat60	-0.26	-0.44	0.18	0
High	Q8VVVS4	WD repeat-containing protein 60 [OS=Homo sapiens]	23_3041549	6_56660413	6	8	6	1066	122_496	WDRE60	-0.19	-0.44	0.26	0
High	P61163	Alpha-actinin-3 [OS=Homo sapiens]	6_97267817	7_18085106	3	3	3	376	42_587	ACTRIIA	-0.8	-0.44	-0.37	0
High	O95470	Sphingosine-1-phosphate lyase 1 [OS=Homo sapiens]	4_9912705	4_57164679	2	2	2	568	62_483	SGPL1	-0.62	-0.44	-0.18	0
High	P60709	Actin, cytoplasmic 1 [OS=Homo sapiens]	511_915087	77_3333333	25	205	11	375	41_71	ACTB	-0.5	-0.45	-0.05	32
High	Q16630-2	Isotform 2 of Cleavage and polyethylation specificity factor subunit 6 [OS=Homo sapiens]	21_7018869	32_9931973	19	64	19	588	63_432	CPSF6	-0.51	-0.45	-0.07	0
High	Q96K94-1	Ubiquitin carbonyl-terminal hydrolase 42 [OS=Homo sapiens]	55_2961514	13_7462236	14	19	14	1324	145_302	USP42	-0.5	-0.45	-0.05	0
High	O14004-1	Cyclin-dependent kinase 13 [OS=Homo sapiens]	39_1010993	8_53174603	10	16	5	1512	164_823	CDK13	-0.38	-0.45	0.17	0
High	Q90B54	DnaJ homolog subfamily B member 11 [OS=Homo sapiens]	4_74537036	2_23463687	1	3	1	358	40_489	DNAJ11	-0.99	-0.45	-0.53	0
High	P25490	Transcriptional repressor protein YY1 [OS=Homo sapiens]	36_5088982	21_7591304	9	14	9	414	44_685	YY1	-0.4	-0.46	0.06	0
High	P61360-1	Pleiotropic regulator 1 [OS=Homo sapiens]	23_1910149	12_6459144	5	6	5	514	57_158	PLRG1	0.1	-0.46	0.56	0
Medium	O43159	ribosomal RNA-processing protein 8 [OS=Homo sapiens]	2_8449677	4_16666667	1	1	1	456	50_683	RRP8	0.06	-0.46	0.52	0
High	P68032	Actin, alpha cardiac muscle 1 [OS=Homo sapiens]	238_953912	40_3183024	16	103	3	377	41_992	ACTC1	-0.55	-0.47	-0.08	0
High	P75746	Calcium-binding mitochondrial carrier protein Aralar 1 [OS=Homo sapiens]	42_2086702	14_0117994	9	14	4	678	74_715	SLC25A12	0.15	-0.47	0.61	0
High	P65144	G0S ribosomal protein 129 [OS=Homo sapiens]	36_668549	20_7547170	20	20	3	159	17_741	RPL29	-0.32	-0.47	0.16	0
High	Q7L014	Probable ATP-dependent RNA helicase DDX46 [OS=Homo sapiens]	366_4211	45_2958293	46	134	46	1031	117_29	DDX46	-0.44	-0.48	0.04	0
High	Q07157	Tight junction protein ZO-1 [OS=Homo sapiens]	100_963203	20_7093922	25	35	25	1748	195_34	TJP1	-0.92	-0.48	-0.44	0
High	P06265	condensin 2 complex subunit D3 [OS=Homo sapiens]	24_0667083	7_61014686	8	11	8	1498	168_783	NCAPD3	-0.17	-0.48	0.31	0
High	Q14558-2	Isotform 2 of phosphatidylinositol pyrophosphate synthase-associated protein 1 [OS=Homo sapiens]	7_77651461	7_7920779	2	4	2	385	42_44	PRPSAP1	-0.39	-0.48	0.1	0
Medium	P48444	Coatomer subunit delta [OS=Homo sapiens]	2_52607531	2_54403131	1	1	1	511	57_174	ARCN1	-1.14	-0.48	-0.66	0
High	O00541-1	Pescatillo homolog [OS=Homo sapiens]	143_010138	38_4353741	20	44	20	588	67_96	FES1	-0.35	-0.49	0.14	0
High	P60783	28S ribosomal protein S14, mitochondrial [OS=Homo sapiens]	28_1875	21_8785	2	6	2	128	15_129	MRPL54	0.04	-0.49	0.53	0
Medium	P10589-1	Thioredoxin [OS=Homo sapiens]	4_39545148	8_57142857	1	2	1	105	11_73	TXN	-0.61	-0.49	-0.12	0
Medium	Q92610	Zinc-finger protein 592 [OS=Homo sapiens]	3_26647687	1_89423836	2	2	2	1267	137_441	ZNF592	-0.34	-0.49	0.15	0
Medium	P0193MH1	Von Willebrand factor A domain-containing protein 8 [OS=Homo sapiens]	3_01686592	1_04386677	2	2	2	1905	214_689	VWA8	0.15	-0.49	0.65	0
Medium	Q9YEG4	Zinc finger protein 592 [OS=Homo sapiens]	3_68511864	2_41657077	2	2	2	869	96_559	ZNF592	-0.54	-0.49	-0.24	0
Medium	Q9Y6M4-2	Isotform 2 of Casein kinase 1 isoform gamma-3 [OS=Homo sapiens]	2_4282241	1_75824176	1	2	1	455	52_344	CSNK1G3	-0.03	-0.5	-0.47	0
High	Q9BZT3-5	Isotform 5 of Oxysterol-binding protein-related protein 6 [OS=Homo sapiens]	25_5344397	55_0573514	41	74	1	959	108_89	OBP16	-0.72	-0.51	-0.22	39
High	P059NQ2	Probable dimethyladenosine transferase [OS=Homo sapiens]	144_181397	68_6900958	21	44	21	313	23_214	DIMT1	-0.37	-0.51	0.14	0
High	Q71UUM5	4D9S ribosomal protein S27 [OS=Homo sapiens]	87_4282094	41_6666667	6	29	3	84	9_471	RPS27L	-0.32	-0.51	0.19	0
High	Q9H936	Mitochondrial glutamate carrier 1 [OS=Homo sapiens]	48_6937206	43_3436533	10	14	10	323	34_448	SLC25A22	-0.15	-0.51	0.36	0
High	P62316	Small nuclear ribonucleoprotein Sm D2 [OS=Homo sapiens]	15_1861058	33_0508475	3	6	3	118	13_518	SNRPD2	-0.75	-0.51	-0.23	0
High	Q9Y545-6	Isotform 6 of Breast cancer anti-resistance protein 1 [OS=Homo sapiens]	49_5160449	66_5938865	46	161	46	916	97_817	BCL2L1	-0.25	-0.52	-0.47	0
High	Q87VM9	Vang-like protein 1 [OS=Homo sapiens]	5_37861597	3_05343511	1	1	1	524	59_937	VANGL1	-0.74	-0.52	-0.22	0
High	P56182	Ribosomal RNA processing protein 1 homolog A [OS=Homo sapiens]	64_7183224	29_067251	15	21	15	461	52_807	RPBP1	0.01	-0.53	0.54	0
High	P18805	ADP-ribosylation factor 1 [OS=Homo sapiens]	27_3566544	26_6666667	4	8	2	180	20_498	ARE4	-0.64	-0.53	-0.11	1
Medium	O14734	Acyl-coenzyme A thioesterase 8 [OS=Homo sapiens]	2_91537489	2_923131671	1	1	1	319	35_891	ACOT8	-0.53	-0.52	-0.22	0
High	Q87VM9-2	Zinc-finger CCCH domain-containing protein 18 [OS=Homo sapiens]	12_4968416	5_35152151	5	5	5	953	106_315	ZC3H18	-0.34	-0.54	0.21	0
Low	Q9H9P8-1	L-2-hydroxyglutarate dehydrogenase, mitochondrial [OS=Homo sapiens]	1_8247782	2_59179266	1	1	1	463	50_284	L2HGDH	-0.58	-0.54	-0.04	0
High	P21AK8-3	Isotform 3 of WW domain-containing protein MUW1 [OS=Homo sapiens]	16_69655846	17_11354309	4	6	4	731	80_649	WW4	0.19	-0.55	0.74	0
High	Q9t7N4	Splicing factor, arginine/serine-rich 19 [OS=Homo sapiens]	7_23260793	1_44817073	2	3	1	1312	13_9186	SCAF1	-0.88	-0.55	-0.32	0
High	O43809	Cleavage and polyadenylation specificity factor subunit 7 [OS=Homo sapiens]	116_272624	58_1497797	10	28	10	227	26_211	NUDT21	-0.65	-0.56	-0.1	0
High	Q8V6M4-3	Isotform 3 of Cleavage and polyadenylation specificity factor subunit 7 [OS=Homo sapiens]	76_492166	31_9066148	14	24	14	514	56_341	CPSF7	-0.47	-0.56	0.09	0
High	Q96QV6	Histone H2A type 1A [OS=Homo sapiens]	55_4970157	40_5480153	5	32	1	131	142_255	HIST1H2AA	-0.07	-0.56	0.49	0
High	P51116	Fragile X mental retardation syndrome-related protein 2 [OS=Homo sapiens]	31_1383026	14_7102526	6	8	5	673	74_178	XR2	-0.53	-0.56	0.02	1
High	P23458	Tyrosine-protein kinase JAK1 [OS=Homo sapiens]	12_3717304	5_19930676	5	6	5	1154	133_191	JAK1	-0.29	-0.56	0.27	0
High	Q9UL54-1	Serine/threonine-protein kinase TAO2 [OS=Homo sapiens]	7_08689918	3_07692308	4	4	4	1235	138_166	TAOK2	0.34	-0.56	0.9	0
Medium	Q9B3G5	Probable serine carboxyproteidase CIVL [OS=Homo sapiens]	2_47716469	2_10984034	1	1	1	476	54_129	CIVL	-1.03	-0.56	-0.46	0
High	Q92522	Histone H1X [OS=Homo sapiens]	47_6860576	36_1502347	9	17	9	213	22_474	H1FX	-0.65	-0.57	-0.08	0
High	E9PR68	Uncharacterized protein C1orf98 [OS=Homo sapiens]	32_2833939	23_7704918	4	9	3	122	13_79	D2288414; C1	-0.29	-0.59	0.27	0
High	Q86V13	Ras GTPase-activating-like protein IQGAP3 [OS=Homo sapiens]	31_2649028	7_54_138565	10	12	10	1631	184_584	IQGAP3	-0.59	-0.57	-0.03	0
High	P21953	2-oxyisovalerate dehydrogenase subunit beta, mitochondrial [OS=Homo sapiens]	6_61334676	11_734639	3	3	3	392	43_095	BCKDHB	-2.56	-0.57	-1.98	0
High	Q75400	pre-mRNA-processing factor 40 homolog A [OS=Homo sapiens]	32_9370915	38_3490073	36	114	36	957	108_737	PRPF40A	-0.66	-0.58	-0.08	0
High	Q86VMS4	PremRNA-processing factor 19 [OS=Homo sapiens]	95_2623801	30_7539683	11	23	11	504	55_146	PRPF19	-0.21	-0.58	0.37	0
High	P14257-2	Isotform 2 of Reticulocalbin-2 [OS=Homo sapiens]	24_3033931	18_5074627	4	5	4	335	39_115	RCH2	-0.21	-0.58	0.38	0
High	P48643	T-complex protein 1 subunit epsilon [OS=Homo sapiens]	21_1737905	8_87245841	4	7	4	541	59_633	CCT5	-0.47	-0.58	0.12	0
High	Q6P087	RNA pseudouridylate synthase domain-containing protein 1 [OS=Homo sapiens]	14_6059261	15_0997151	5	6	5	351	38_437	RPSUD3	-0.39	-0.58	0.19	0
High	Q95696-2	Isotform 2 of Bromodomain-containing protein 1 [OS=Homo sapiens]	6_92290548	1_51387721	2	2	2	1189	133_159	RD1; LOC0083	-0.59	-0.58	-0.01	0

High	P16615	Sarcoplasmic/endoplasmic reticulum calcium ATPase 2 [OS=Homo sapiens]	150_052573	29_9424184	27	46	27	1042	114_683	ATP2A2	-0.56	-0.59	0.04	0
High	Q9B8D7	Cancer-related nucleoside-triphosphatase OS=Homo sapiens]	23_1398209	50_2633158	4	6	4	190	20_7	NTPCR	-0.48	-0.59	0.11	0
High	P17858-1	ATF-dependent 6-phosphotokinase, liver type [OS=Homo sapiens]	13_55669429	6_41025541	3	7	3	780	84_964	FFKL	-1.15	-0.56	-0.56	0
High	Q9P2R7	Succinyl-CoA ligase [ADP-forming] subunit beta, mitochondrial [OS=Homo sapiens]	10_62608243	7_99136069	4	5	4	463	50_285	SCLCA2	-0.48	-0.59	0.12	0
High	Q08123	tRNA (Cytosine-34) C(5')-methyltransferase [OS=Homo sapiens]	183_245952	55_9322034	29	56	29	767	86_416	NSUN2	-0.66	-0.6	-0.06	0
High	Q85233	76_6408812	16_4846077	16	33	16	1007	116_916	PRPF4B	-0.85	-0.6	-0.26	0	
High	Q12923-4	Isotform 4 of Tyrosine-protein phosphatase non-receptor type 13 [OS=Homo sapiens]	24_6624907	6_70582731	12	15	12	2490	277_335	PTPN13	-0.47	-0.6	0.13	0
High	Q5SY16	Polynucleotide 5'-hydroxyl kinase NOL9 [OS=Homo sapiens]	18_2149799	5_6880057	3	7	3	702	79_272	NOL9	-0.5	-0.6	0.1	0
High	P11091	eukaryotic translation initiation factor 1 subunit 3 [OS=Homo sapiens]	11_7315085	8_58864068	4	5	4	472	51_077	EF2S3	-0.9	-0.6	-0.31	0
High	P23207	F-actin-capping protein subunit alpha-1 [OS=Homo sapiens]	11_432402	8_74125874	2	4	2	286	32_927	CPZ1	-0.75	-0.6	-0.15	0
High	Q9P258	Protein RCC2 [OS=Homo sapiens]	30_976724	17_816092	9	10	9	522	56_049	RCC2	-0.66	-0.61	-0.06	0
Medium	Q86XK2-2	Isotform 2 of Condensin-2 complex subunit G2 [OS=Homo sapiens]	2_79553706	1_55709343	2	2	2	1156	132_111	NCAPG2	-0.36	-0.61	-0.35	0
Low	P78346-2	Isotform 2 of Ribonuclease P protein subunit p30 [OS=Homo sapiens]	2_09807265	1_8633504	1	1	1	322	35_542	RPP30	-0.38	-0.61	0.23	0
High	Q9H0A0	N-acetyltransferase 10 [OS=Homo sapiens]	50_5334325	65_9512195	65	151	64	1025	115_657	NAT10	-0.46	-0.62	0.16	1
High	Q9Y212	Band 4.1-like protein 3 [OS=Homo sapiens]	79_1399711	16_373345	16	24	15	1087	120_603	EPA113	-0.99	-0.62	-0.37	1
High	Q9H307	Pinin [OS=Homo sapiens]	50_919574	15_7601116	11	18	11	717	81_565	RNN	-1.32	-0.62	-0.71	0
High	P16964	WD repeat-containing protein 5 [OS=Homo sapiens]	24_6293454	26_568626	4	6	4	334	36_565	WDR5	-0.64	-0.62	-0.02	0
High	Q8N1G4	Leucine-rich repeat-containing protein 47 [OS=Homo sapiens]	8_540003848	1_88679245	1	2	1	583	63_434	LRCA7	-0.26	-0.62	0.36	0
High	P63172	Dynein light chain 1c-type 1 [OS=Homo sapiens]	7_38502501	23_0088496	2	3	2	113	12_444	DYNLT1	-0.23	-0.62	0.39	0
High	Q9BYT3-1	Serine/threonine-protein kinase 33 [OS=Homo sapiens]	91_1409464	37_037393	17	27	17	514	57_794	STK33	-0.46	-0.63	0.17	0
High	Q15344-1	E3 ubiquitin-protein ligase Midline-1 [OS=Homo sapiens]	37_380239	14_3929836	9	12	9	667	72_203	MID1	-0.63	-0.63	0.07	0
High	Q9BQG6	Ribosomal protein 63, mitochondrial [OS=Homo sapiens]	7_9597541	12_745098	1	2	1	102	12_259	RPB6; MRPL4	-0.03	-0.63	0.6	0
High	P52268	60S ribosomal protein L22 [OS=Homo sapiens]	59_4156055	58_59375	6	24	5	128	14_778	RPL22	-0.61	-0.64	0.04	0
High	P52031	Thyroid transcription factor 1-associated protein 26 [OS=Homo sapiens]	53_4916773	53_950207	7	16	7	241	28_652	CD5C9	-0.44	-0.64	0.2	0
High	P19447	TFIH basal transcription factor complex helicase XBP subunit [OS=Homo sapien	36_7031342	14_0664962	9	13	9	782	89_221	ERCC3	0.91	-0.64	1.55	0
High	P84103	Serine/threonine-protein kinase 33 [OS=Homo sapiens]	24_0111965	30_4878049	4	9	4	164	19_318	SRSF3	-1.18	-0.64	-0.54	0
High	Q96G53	FAS-associated factor 2 [OS=Homo sapiens]	5_480336455	3_37078652	2	3	2	445	52_591	FAE2	-0.36	-0.64	0.29	0
High	Q9UBX3-2	Isotform 2 of Mitochondrial dicarboxylate carrier [OS=Homo sapiens]	33_941594	37_702703	8	15	8	296	32_125	SLC25A10	-0.4	-0.65	0.26	0
High	Q9H9B4	Sidoflexin-1 [OS=Homo sapiens]	30_042322	25_4658385	7	11	7	322	35_596	SKP1N	-0.92	-0.67	-0.28	0
High	P17509	Homeobox protein Hox-B6 [OS=Homo sapiens]	12_7925099	8_92857143	2	4	1	224	25_416	HDXB6	-0.14	-0.65	0.51	0
High	Q9P522	Serine/threonine-protein kinase N3 [OS=Homo sapiens]	6_09532785	9_740157	1	3	1	889	99_358	PNK3	-0.05	-0.65	0.6	0
High	Q129866	Transcriptional regulator NF-X1 [OS=Homo sapiens]	62_9283875	17_3214286	14	20	14	120	124_312	NFK4	-0.43	-0.66	0.22	0
High	Q9HC21	Mitochondrial thiamine pyrophosphatase carrier [OS=Homo sapiens]	9_91015584	8_75	3	5	3	320	35_488	SLC25A19	-0.56	-0.66	0.1	0
High	P11177	Pyruvate dehydrogenase E1 component subunit beta, mitochondrial [OS=Homo sapien	36_8624554	25_6267409	8	15	8	359	39_208	PDH	-0.7	-0.67	-0.03	0
High	P15170-2	Isotform 2 of Galactokinase [OS=Homo sapiens]	30_0149725	14_4549763	5	9	5	422	45_329	GALK1	-0.77	-0.67	0	0
High	Q86WV1-1	Chromodomain-helicase-DNA-binding protein 1-like [OS=Homo sapiens]	16_9470668	8_80713489	6	7	6	897	100_921	CHD1L	-0.42	-0.67	0.25	0
High	P46821	microtubule-associated protein 1B [OS=Homo sapiens]	8_40318859	9_97244733	2	3	2	2468	270_468	MAP1B	-0.46	-0.67	0.22	0
Medium	Q9HA47-4	Isotform 4 of Uridine-cytidine kinase 1 [OS=Homo sapiens]	2_8680607	2_83687943	1	1	1	282	32_254	UCK1	-1.14	-0.67	-0.47	0
High	Q9BT76-8	Isotform 5 of MMS19 nucleotide excision repair protein [OS=Homo sapiens]	9_91015584	8_75	3	5	3	1051	115_631	TRM112	-0.56	-0.66	0.1	0
High	P11177	Multiple myeloma tumor-associated protein 2 [OS=Homo sapiens]	36_8624554	25_6267409	5	6	5	359	29_394	Clorf35	-0.78	-0.69	-0.09	0
High	Q5LS42	Transcription initiation factor TIF1B subunit 5 [OS=Homo sapiens]	19_8518335	9_5	6	9	6	800	86_776	TAF5	-1.21	-0.69	-0.52	0
Medium	P97740	Mitochondrial carrier homolog 1 [OS=Homo sapiens]	3_07233704	3_08483329	1	1	1	389	41_517	MTCH1	-0.55	-0.69	0.14	0
High	Q9UJ30	Nuclear pore complex protein Nap107 [OS=Homo sapiens]	2_862063718	2_70202727	2	2	2	925	106_307	NUP107	-1.23	-0.69	-0.54	0
High	Q9P015	Mutifunctional methionyltransferase subunit TM112-like protein [OS=Homo sapiens]	19_9072287	21_9594595	5	5	3	125	14_19	TRMT112	-0.87	-0.7	-0.17	0
Medium	P497W2-1	Guanine nucleotide-binding protein G1 subunit alpha isoforms Xlas [OS=Homo sapiens]	13_0734136	5_15564202	5	5	5	296	33_399	MNP15	-0.55	-0.7	0.16	0
High	P497W5-1	RNA-binding protein 25 [OS=Homo sapiens]	38_05496	48_1613286	48	125	48	1037	110_556	GNAS	-0.302	-0.7	-0.23	0
High	Q9UJS0-2	Isotform 2 of Calcium-binding mitochondrial carrier protein Aralar 2 [OS=Homo sapiens]	10_1616681	36_390525	16	25	11	676	100_124	RM25	-0.73	-0.71	-0.02	0
High	Q4680	Maternal embryonic leucine zipper kinase [OS=Homo sapiens]	13_7494516	6_75883357	4	5	4	651	74_594	MELK	-0.17	-0.71	0.54	0
High	Q9BU76-1	ATP-binding cassette sub-family F member 1 [OS=Homo sapiens]	13_78720218	6_39777469	3	5	3	719	79_678	UMC1	-0.21	-0.73	0.35	0
High	Q8NWV7	Formin-like protein 3 [OS=Homo sapiens]	57_06834391	22_01183434	12	20	12	845	95_866	ABCFL1	-0.42	-0.75	0.33	0
Medium	P497W3-2	Isotform 2 of 39S ribosomal protein L30, mitochondrial [OS=Homo sapiens]	3_07233704	2_81774704	13	1832461	2	1028	117_139	FMN1L3	-0.49	-0.75	0.25	0
High	Q9BT415	Oxysterol-binding protein-related protein 3 [OS=Homo sapiens]	427_848524	73_61890402	52	118	49	887	21_825	TUBB6	-1.7	-0.77	-0.93	0
High	Q9BEU1	Nucleolar GTP-binding protein 2 [OS=Homo sapiens]	85_0597925	24_3502052	15	27	15	731	83_603	OBBL1	-0.11	-0.76	0.54	0
High	Q8NE71-1	Guanine nucleotide-binding protein-like 3-like protein [OS=Homo sapiens]	20_495451469	11_5120275	6	13	5	582	65_532	GNL3L	-0.38	-0.76	0.38	0
High	Q14152	Euksyotic translocation initiation factor 3 subunit A [OS=Homo sapiens]	3_13161222	1_0130246	2	2	2	1382	166_168	EF5A	-0.92	-0.77	0.17	0
High	Q9BUF5	Tubulin beta-6 chain [OS=Homo sapiens]	227_558287	40_3587444	13	128	2	446	49_825	TUBB6	-1.7	-0.77	-0.93	0
High	Q8WV6	Adhesion G protein-coupled receptor A3 [OS=Homo sapiens]	122_388766	25_8894777	30	63	30	1321	146_058	PR125; ADGR	-0.64	-0.77	0.12	0
High	Q9BZ4	Nucleolar GTP-binding protein 1 [OS=Homo sapiens]	110_986421	33_2807571	20	45	20	634	73_918	GPBP4	-0.3	-0.77	0.47	0
High	Q9NMID3	heterogeneous nuclear ribonucleoprotein U-like protein 1 [OS=Homo sapiens]	40_0576761	18_0722992	11	14	11	747	85_052	HNRNPU1	-0.77	-0.77	0.36	0
High	Q9NTW7	Zinc finger protein 64 homolog, isoforms 3 and 4 [OS=Homo sapiens]	7_65230215	5_11627907	2	3	2	645	72_17	ZFP64	-1.22	-0.77	-0.45	0
Medium	Q14641	Segment polarity protein dsh1 [OS=Homo sapiens]	4_60156275	1_63043478	1	2	1	736	78_899	DVL2	-0.33	-0.77	0.44	0
High	Q13888-1	General transcription factor IIH subunit 2 [OS=Homo sapiens]	10_343519	5_82287481	2	2	2	395	44_39	F2H2; GTf2H	-0.78	-0.78	2.76	0

High	Q8NHQ9	ATP-dependent RNA helicase DDX55 [OS=Homo sapiens]	39_3723594	17_6666667	10	13	10	600	68_503	DDX55	-0.7	-0.79	0.09	0
High	Q9NPF4	Probable RNA N6-adenosine trienzylcarbamoyltransferase [OS=Homo sapiens]	6_30436626	8_95522888	2	2	2	335	36_403	OSGEP	-1.09	-0.79	-0.3	0
High	P18583-9	Isoform 1 of Protein SON [OS=Homo sapiens]	20_441342	19_14030882	32	86	32	246	267_323		-1.1	-0.8	-0.3	0
High	O75382-1	Tripartite motif-containing protein 3 [OS=Homo sapiens]	13_037723	5_37634409	3	5	3	744	80_779	TRIM3	-0.08	-0.8	0.72	0
Medium	P00558	Phosphoglycerate kinase 1 [OS=Homo sapiens]	2_69880341	2_15827938	1	2	1	417	44_586	PGK1	-8.9	-0.8	-8.11	0
High	Q9UNF1	Melanoma-associated antigen D2 [OS=Homo sapiens]	15_288748	5_61056106	4	7	4	606	64_914	MAGED2	-0.68	-0.81	0.12	0
High	Q99697-2	Isoform P2xC of Pituuary homeobox 2 [OS=Homo sapiens]	8_8013023	8_64197531	2	2	2	324	35_773	PITX2	0.19	-0.81	1	0
High	Q02040-1	A-kinase anchor protein 7A [OS=Homo sapiens]	383_63191	45_753957	42	193	42	695	80_686	AKAP17A	-0.09	-0.82	-0.08	0
High	Q9NNW5	WDR repeat-containing protein 6 [OS=Homo sapiens]	19_0450351	8_9206066	7	7	7	1121	12_647	WDR6	-0.56	-0.82	0.26	0
High	Q9Y34	Ribosomal RNA-processing protein 7 homolog A [OS=Homo sapiens]	111_706129	53_5714286	12	28	12	280	32_314	RNPL7A	-0.56	-0.83	0.27	0
High	Q9UKS8	Cyclin-L1 [OS=Homo sapiens]	8_15312335	7_03422053	3	3	3	526	59_597	CNL1	0.92	-0.83	1.75	0
High	Q9NIP1-2	Isoform 2 of Bromodomain-containing protein 7 [OS=Homo sapiens]	7_76782307	7_36196319	3	5	3	652	74_422	BRD7	-1.36	-0.83	-0.52	0
High	Q8TD1-2	Isoform 2 of ATP-dependent RNA helicase DDX54 [OS=Homo sapiens]	276_795506	62_0181406	45	83	1	882	98_605	DDX54	-0.43	-0.84	0.41	0
High	Q8TA08	Smad nuclear-interacting protein 1 [OS=Homo sapiens]	64_0363623	39_1414141	13	23	13	396	45_75	SNIP1	-0.45	-0.84	0.39	0
High	Q3A427	Peptidyl-prolyl cis-trans isomerase B [OS=Homo sapiens]	27_394432	11_8037135	8	18	8	754	88_564	PPIG	-0.76	-0.84	0.07	0
High	Q8WV3-1	F3.1 ubiquitin-protein ligase RNF138 [OS=Homo sapiens]	12_1348047	16_3265306	4	4	4	245	28_174	RNF138	-0.34	-0.84	0.5	0
High	P11021	78 kDa glucose-regulating protein 1 [OS=Homo sapiens]	330_041584	55_6714924	33	99	31	654	72_288	HSPAN4	-0.96	-0.86	-0.1	0
High	Q96C89	5-methylcytosine RNA methyltransferase NSUN4 [OS=Homo sapiens]	12_7704157	10_15625	4	6	4	384	43_061	NSUN4	-1.02	-0.86	-0.15	0
High	Q9P004-2	Isoform 2 of CXCL-type zinc finger protein 1 [OS=Homo sapiens]	6_49176468	5_3030303	3	3	3	660	76_178	CXXC1	-0.85	-0.86	0.01	0
Low	Q9Y294	Histone chaperone ASFA1 [OS=Homo sapiens]	15_4166437	3_92156663	1	1	1	204	22_954	ASFA1	-0.34	-0.86	0.52	0
High	Q9NW6	Arginine and glutamate-rich protein 1 [OS=Homo sapiens]	193_121243	46_8864469	25	118	25	273	33_197	ARGLU1	-0.81	-0.88	0.07	0
High	Q95478	Ribosome biogenesis protein NS2A homolog [OS=Homo sapiens]	17_2643474	20	6	7	6	260	30_047	NS2A	-0.83	-0.88	0.05	0
Medium	Q9HSU6-1	Zinc finger CCCH-domain-containing protein 4 [OS=Homo sapiens]	3_09750154	3_31384016	2	2	2	513	58_971	ZCCHC4	-1.02	-0.88	-0.14	0
High	Q9TP86	RNA-binding protein 26 [OS=Homo sapiens]	221_207603	31_1817279	34	69	0	1007	113_527	RMB26	-0.73	-0.89	0.16	28
High	Q7KZ7-1	Serine/threonine-protein kinase MARK2 [OS=Homo sapiens]	120_5219179	35_2791878	21	36	13	788	87_856	MARK2	-0.43	-0.89	0.46	0
High	P45880-1	Isoform 1 of Voltage-dependent anion-selective channel protein 2 [OS=Homo sapiens]	12_7750845	11_6504854	3	4	3	309	33_351	VDAC2	-0.73	-0.89	0.16	0
Medium	Q040403	Transcription initiation factor IIB [OS=Homo sapiens]	4_40557292	8_2278483	3	3	3	316	34_811	GTF2B	-0.17	-0.89	0.71	0
High	Q9H644-1	Nucleolar protein 6 [OS=Homo sapiens]	235_168713	43_8045375	35	62	35	1146	127_513	NOL6	-0.12	-0.91	-0.79	0
High	Q9BUQ8	Probable ATP-dependent RNA helicase DDX23 [OS=Homo sapiens]	36_7643383	15_3658337	11	14	11	820	95_524	DDX23	-1.26	-0.91	-0.35	0
High	Q6IAN0	Dehydrogenase/reductase SDR family member 7B [OS=Homo sapiens]	5_8410463	8_30769331	2	2	2	325	35_097	DHRS7B	-0.82	-0.91	0.09	0
High	Q9UYJ9	Probable ATP-dependent RNA hydrolase DDX41 [OS=Homo sapiens]	764_716405	88_1028839	62	268	62	627	69_793	DDX41	-0.81	-0.92	0.11	0
High	Q9RK9-1	Protein FAM130 [OS=Homo sapiens]	30_7285539	5_25101246	5	16	3	247	28_568	FAM130B	-0.48	-0.92	-0.92	2
Medium	Q75520-2	Isoform 2 of Polycomb protein EED [OS=Homo sapiens]	3_78166935	4_72103004	2	2	2	466	53_027	EED	-0.8	-0.92	0.13	0
Low	Q9UPF9-1	Ubiquitin carboxy-terminal hydrolase 22 [OS=Homo sapiens]	1_43415218	1_33333333	1	1	1	525	59_921	UPF22	-0.59	-0.92	0.33	0
High	Q9D4Y2	Tight junction protein ZO-2 [OS=Homo sapiens]	33_024375	44_2837143	47	118	47	1190	133_876	TJP2	-0.91	-0.93	0.19	0
High	Q9EME7	Zinc finger protein 542 [OS=Homo sapiens]	126_124945	42_3280423	18	35	18	567	64_641	ZNFX1	-0.81	-0.93	0.13	0
High	Q9BYW2	Histone-like N-methyltransferase SETD2 [OS=Homo sapiens]	46_0390294	8_54134165	20	20	20	2564	287_418	SETD2	-0.75	-0.93	0.18	0
High	Q9NWQ4	G patch domain-containing protein 2-like [OS=Homo sapiens]	21_0349046	12_6556017	6	8	6	482	54_226	GPATCH2L	-0.8	-0.93	0.14	0
High	Q9TSB5-1	Round spermatid basic protein 1 [OS=Homo sapiens]	14_8214431	6_61938334	5	5	5	846	94_811	RSPN1	-1.05	-0.93	-0.92	0
High	Q9Y5J1	U3 small nucleolar RNA-associated protein 1.8 homolog [OS=Homo sapiens]	38_8170592	20_0535871	7	9	7	556	61_964	UPT18	0.12	-0.94	1.06	0
High	Q12894-2	Isoform 2 of interferon-related developmental regulator 2 [OS=Homo sapiens]	5_92679718	1_484026316	1	3	1	608	65_557	IRF2	-0.75	-0.94	0.19	0
Medium	H9WNW9	Putative small nuclear ribonucleoprotein G-like protein 1 [OS=Homo sapiens]	3_80219695	17_2052632	1	2	1	76	8_538	DS3032; SNR	-0.44	-0.94	0.5	0
High	Q8TDD1	ATP-dependent RNA helicase DDX4 [OS=Homo sapiens]	270_412359	6_19750384	45	81	1	881	98_524	DDX4	-0.75	-0.95	0.2	40
High	Q48735	Isocitrate dehydrogenase [NADP], mitochondrial [OS=Homo sapiens]	22_3102924	13_4957572	5	6	5	452	50_877	IDH2	-1.23	-0.95	-0.28	0
High	P14618	Pyruvate kinase [PKM] [OS=Homo sapiens]	12_7235813	7_53295669	4	4	4	531	57_9	PKM	-2.34	-0.95	-1.39	0
High	Q9BVP2	Guanine nucleotide hydrolase domain-containing protein 1 [OS=Homo sapiens]	902_607587	6_59380692	45	376	44	549	61_954	GNL3	-1.08	-0.96	-0.12	2
High	Q9T319-1	RNA/RNP complex-1 interacting phosphatase	27_52056205	7_2454545	6	7	6	330	38_915	DPA11	-0.63	-0.96	0.33	0
High	Q8VWX5	dnaJ homolog subfamily 9 [OS=Homo sapiens]	25_2446028	15_3846154	4	8	4	260	29_891	DNAJC9	0.07	-0.96	1.03	0
Medium	Q9UDRS	Alpha-aminoacid semialdehyde synthase, mitochondrial [OS=Homo sapiens]	4_4008383	3_67170262	2	2	2	926	102_066	AASS	-0.59	-0.96	0.37	0
High	Q9TPC8-4	Isoform 3 of Proliferation initiation factor 1 (F1B) [OS=Homo sapiens]	42_850457	16_2721893	3	9	3	327_388	TAF8	-0.99	-0.97	-0.02	0	
Medium	Q75880	Protein SC01 homolog, mitochondrial [OS=Homo sapiens]	2_863227943	5_31561462	1	1	1	301	33_793	SC01	-0.39	-0.97	0.58	0
High	Q9H8G2-1	Caspase activity and apoptosis inhibitor 1 [OS=Homo sapiens]	162_422119	47_9224377	18	50	18	361	38_344	CASP1	-1.11	-0.98	-0.13	0
Medium	Q9E1U4	Alpha/beta hydrolase domain-containing protein 14B [OS=Homo sapiens]	2_46117501	6_19047619	1	1	1	210	22_332	ABHD14B	-0.87	-0.98	0.11	0
Low	Q9BXW9-1	Isoform 1 of Fanci (anemia group D2) protein [OS=Homo sapiens]	1_90329567	0_57980965	1	1	1	1471	16_356	FANCD2	-1.84	-0.98	-0.36	0
High	P15924-1	Desmoplakin [OS=Homo sapiens]	43_2111079	7_035876	20	20	20	2871	33_1569	DSP	-1.91	-0.99	-0.91	0
High	Q9HCMA	Band 4.1-like protein 5 [OS=Homo sapiens]	495_432427	72_3055935	41	190	15	733	81_805	EPB4115	-0.31	-1.01	0.7	41
High	P14340	nicotinamide phosphoribosyltransferase [OS=Homo sapiens]	10_4363201	8_14663951	4	4	4	491	55_487	NAMPT	-0.42	-0.98	0.39	0
Medium	P00966	Arabinofuranosidase synthase [OS=Homo sapiens]	2_31318505	1_69602913	1	2	1	412	46_501	ASS1	-0.68	-1.03	0.35	0
High	Q9TP86-2	Isoform 2 of RNA-binding protein 26 [OS=Homo sapiens]	42_850457	16_2721893	34	69	1	983	110_956	RM26	-0.65	-1.04	0.39	0
High	Q13393-2	Isoform PLD1B [Phospholipase D1 [OS=Homo sapiens]	80_2908956	23_1660232	19	28	3	1036	119_618	PLD1	-0.74	-1.04	0.3	0
High	Q6EP13-2	Isoform 2 of AdP-ribosylation factor-like protein 6-interacting protein 4 [OS=Homo sapiens]	50_8855477	18_6440678	8	28	3	413	44_132	ARLP4	-0.98	-1.05	0.06	10
High	Q9NY93	Probable ATP-dependent RNA helicase DDX56 [OS=Homo sapiens]	37_4426797	16_8190128	8	12	8	547	61_551	DDX56	0.01	-1.05	1.06	0
High	P07305	Histone H1.0 [OS=Homo sapiens]	74_0210513	32_472268	8	38	8	194	20_85	H1F0	-0.94	-1.06	0.12	0

High	Q9550-1	protein SCAF11 [OS=Homo sapiens]	16_12026392	5,80997949	8	8	1463	164,551	SCAF11	-1,29	-1,06	-0,24	0	
High	Q02978	Mitochondrial 2-oxoglutarate/malate carrier protein [OS=Homo sapiens]	1,953,092,919	1,938,461,54	1	1	314	34,04	SLC2A11	-1,1	-1,07	-0,93	0	
Medium	Q93034	Cullin-5 [OS=Homo sapiens]	75,101,282,6	34,848,4484	8	19	780	198	23,354	CU5	-0,33	-1,07	0,73	0
High	H095373	Immunitin-7 [OS=Homo sapiens]	36,148,115,6	11,368,015,4	8	11	1038	119,44	FCF1	-0,07	-1,08	0,38	1	
Medium	Q5VUG0	Scm1-like with four MBT domains protein 2 [OS=Homo sapiens]	3,826,136,96	1,677,852,35	2	2	894	100,498	SMBT2	-0,62	-1,08	0,46	0	
High	Q3KQJ3	MAP7 domain-containing protein 1 [OS=Homo sapiens]	24,275,592	14,86,5258	9	10	841	92,764	MAP7	-0,87	-1,09	0,22	0	
High	Q9BV16-1	U3 snRNP-associated protein 14 homolog [A [OS=Homo sapiens]]	88,07992,49	39,81841,76	24	31	771	87,924	UTP14A	-0,25	-1,1	0,85	0	
Medium	Q96A64	Leucine-rich repeat-containing protein 59 [OS=Homo sapiens]	4,384,970,14	6,840,390,988	2	3	307	34,909	LRCP59	-0,06	-1,1	1,04	0	
Medium	P4224-1	Signal transducer and activator of transcription 1-alpha/beta [OS=Homo sapiens]	2,542,22,356	1,73,333,333	1	1	750	87,28	STAT1	-0,44	-1,1	0,67	0	
High	Q9BQ39	ATP-dependent RNA helicase DDX50 [OS=Homo sapiens]	142,225,692	33,785,617,4	21	48	18	737	82,514	DDX50	-1,11	-1,11	0	0
High	P49761	Dust-specific protein kinase CLK3 [OS=Homo sapiens]	83,666,317,4	21,47,3342	13	28	638	73,469	CLK3	-0,91	-1,11	0,2	0	
High	Q9VA1	Tubulin beta-2B chain [OS=Homo sapiens]	785,56,2376	75,955,0562	29	366	1	445	49,921	TUBB2B	-1,49	-1,12	-0,37	6
High	Q94813	Slc1 homolog 2 protein [OS=Homo sapiens]	407,038,221	40,810,9876	55	155	1529	169,759	SUT2	-0,16	-1,12	0,96	0	
High	Q9BZF3	Oxysterol-binding protein-related protein 6 [OS=Homo sapiens]	263,26,2962	57,066,3812	41	76	1	934	106,239	OBPPL6	-1,61	-1,12	-0,49	0
High	P11310-2	Isotform 2 of Medium-chain specific acyl-CoA dehydrogenase, mitochondrial [OS=Homo sapiens]	30,424,60,16	23,764,7059	8	11	425	46,99	ACADM	-0,76	-1,12	0,36	0	
High	Q9VP54	Serine/threonine-protein kinase rib2 [OS=Homo sapiens]	18,539,379,94	12,368,12	5	5	552	63,243	RIOK2	-0,51	-1,12	0,62	0	
High	Q2NL82	Pre-mRNA-processing protein 7S1 homolog [OS=Homo sapiens]	301,305,036,6	52,114,4279	38	87	804	91,752	TSRL	-0,97	-1,14	0,18	0	
High	Q6ZUJ1-2	Isotform 2 of Uncharacterized protein C1orf57/1 [OS=Homo sapiens]	12,3,87,5394	39,590,4437	14	34	293	34,176	C1orf57	-1,07	-1,15	0,08	0	
High	Q96Y91-4	Isotform 4 of Kinesin-like protein KIF18B [OS=Homo sapiens]	12,606,6915	4,925,441	4	4	873	95,069	-	-0,68	-1,15	0,47	0	
High	Q9UPQ2-1	Isotform 2 of Thioesterase-repeat-containing gene 68 protein [OS=Homo sapiens]	23,357,993,1	5,397,562,39	7	10	7	1723	46,703	TRICGB	-0,92	-1,17	0,25	0
High	EPBAV3	Nascent polypeptide-associated complex subunit alpha, muscle-specific form [C [OS=Homo sapiens]]	7,077,983,38	1,299,32628	2	3	2078	205,295	NACA	-1,41	-1,17	-0,25	0	
High	Q9Y5V3-2	Isotform 2 of Melanoma-associated antigen D1 [OS=Homo sapiens]	5,586,33,162	5,635,91,61	3	3	834	91,901	MAGED1	-1,95	-1,17	-0,77	0	
High	P49781	pre-mRNA processing protein 7S1 homolog [OS=Homo sapiens]	219,46,4163	45,454,5455	36	66	847	96,499	FTS3	-0,62	-1,19	0,57	0	
High	RD4406-1	Gluceraldehyde-3-phosphate dehydrogenase [OS=Homo sapiens]	80,405,4290	56,119,0403	11	23	335	36,03	GAPDH	-2,79	-1,19	-1,6	0	
High	Q8WVA9-2	Isotform 2 of Splicing regulatory glutamyl/lysine-rich protein 1 [OS=Homo sapiens]	19,403,7965	4,647,3359	2	6	624	71,606	SREKL1	-0,82	-1,19	0,37	0	
High	Q71RC2-4	Isotform 4 of Lar-related protein 4 [OS=Homo sapiens]	10,560,5258	4,794,52055	2	4	730	81,193	LARP4	-0,36	-1,19	0,82	0	
Medium	Q72Q4Q2	HEAT repeat-containing protein 3 [OS=Homo sapiens]	2,563,649,96	1,91,764,671	1	1	680	74,355	HEATR3	0,02	-1,19	0,2	0	
Low	Q8XK2-3	Isotform 3 of Mitochondrial Rho GTPase-1 [OS=Homo sapiens]	1,182,69912	1,157,7424	1	1	691	79,496	RHO1	-1,28	-1,19	-0,09	0	
High	Q95104	Splicing factor, arginine/serine-rich 15 [OS=Homo sapiens]	97,575,176,9	22,323,381	20	45	20	1147	125,79	SCAF4	-1,19	-1,2	0,01	0
High	Q9N3E9	ThAP domain-containing protein 1A [5'-phosphodiesterase delta-3 [OS=Homo sapiens]]	440,090,079,93	70,254,626	43	113	789	89,202	PLCD3	-1,16	-1,21	0,05	0	
High	Q8TEF5-4	Isotform 4 of Rho guanosine nucleotide exchange factor 40 [OS=Homo sapiens]	50,723,074	16,11,14,988	19	27	1471	159,599	ARFGEF40	-0,34	-1,21	0,86	0	
High	Q8N9Q2	protein SREK1P1 [OS=Homo sapiens]	38,328,2055	26,451,6129	6	24	6	155	18,166	SREK1P1	-0,24	-1,21	0,96	0
High	Q8N1G0	Zinc finger protein 687 [OS=Homo sapiens]	15,61,94,627	4,042,03719	4	8	4	1237	129,446	ZNF687	-0,68	-1,21	0,53	0
Medium	Q7N9R12-1	PDZ and LIM domain protein 7 [OS=Homo sapiens]	2,750,8164	2,62,520,507	1	1	457	49,813	PDIM7	-1,25	-1,21	-0,04	0	
High	Q8Z6Z9	tRNA (uracil-5-) methyltransferase homolog A [OS=Homo sapiens]	16,497,4639	6,56	4	6	625	68,682	TRMT2A	-0,59	-1,22	0,63	0	
Medium	Q96EK4	THAP domain-containing protein 11 [OS=Homo sapiens]	4,301,4984	2,547,7707	1	2	1	314	34,433	THAP11	-0,49	-1,22	0,73	0
High	Q8TF5-4	Percentriolar material 1 protein [OS=Homo sapiens]	184,377,932	29,100,7905	40	56	40	2024	228,392	PCM1	-1,64	-1,24	-0,4	0
High	Q8N9Q2	protein SREK1P1 [OS=Homo sapiens]	31,340,966,5	15,925,9259	8	10	540	60,191	ZNF768	-0,61	-1,24	0,62	0	
High	Q7Z7H8-2	Isotform 2 of 39S ribosomal protein L10, mitochondrial [OS=Homo sapiens]	5,58,78,7559	7,01,10,7011	1	1	271	30,288	MRPL10	-1,11	-1,24	0,13	0	
Low	P49848-3	Isotform 3 of Transcription initiation factor TFIID subunit 6 [OS=Homo sapiens]	1,24,38,9663	1,12,04,4818	1	1	714	76,947	TAIF6	-1,72	-1,24	-0,47	0	
High	Q9L348-3	Isotform 3 of Treacle protein [OS=Homo sapiens]	125,50,3986	17,28,5246	25	60	1489	152,114	TFCF1	-2,87	-1,26	-1,61	0	
High	P41171-1	protein 4.1 [OS=Homo sapiens]	21,9,06,668	10,05,94,444	7	9	864	96,957	EPBP41	-1,69	-1,26	0,17	0	
High	Q8H514	Stomatin-like protein 788 [OS=Homo sapiens]	7,9,36,55,18	8,70,78,6517	2	3	356	38,51	STOML2	-2,01	-1,24	0,96	0	
Medium	P24928	DNA-directed RNA polymerase II subunit RPB1 [OS=Homo sapiens]	3,28,16,6833	1,72,58,8832	2	2	1970	217,039	POLR2A	-6,95	-1,26	-5,69	0	
High	Q96P11-2	Isotform 2 of Probable 28S rRNA (cytosine-C(5')-methyl)transferase [OS=Homo sapiens]	43,022,3974	26,6,09,4421	10	13	10	466	50,379	NSUN5	-1,09	-1,28	0,19	0
High	Q96Q625	Splitting factor 18S rRNA (guanine-N(7'))-methyltransferase [OS=Homo sapiens]	39,380,2565	20,9,47,609	7	12	6	401	44,934	RMIM17	-1,22	-1,28	0,06	2
Medium	G60701	UDP-glucose 6-dehydrogenase [OS=Homo sapiens]	4,258,4548	2,024,2915	1	1	494	54,989	UGDH	-2,24	-1,28	-0,95	0	
High	Q4981	TATA-binding protein-associated factor 172 [OS=Homo sapiens]	5,76,649,076	2,32,55,814	3	3	3	1849	206,756	BTFAF1	-0,75	-1,29	0,53	0
High	P42182	Small nuclear ribonucleoprotein Sm D1 [OS=Homo sapiens]	30,18,37,24	5,6,55,377	55	102	53	1220	139,227	DHHL2	-0,93	-1,3	0,37	0
High	Q44331	Protein FRG1 [OS=Homo sapiens]	19,20,47,13	17,44,18,05	4	5	258	29,154	FRG1	-1,15	-1,3	0,15	0	
Medium	Q44966-1	Zinc finger protein 638 [OS=Homo sapiens]	2,6,43,82,774	0,53,94,5599	1	1	1978	220,188	ZNF538	-0,73	-1,32	0,57	0	
High	Q3709-1	Splitting factor 18S rRNA (guanine-N(7'))-methyltransferase [OS=Homo sapiens]	7,6,43,26,6437	12,0,99,6441	2	2	281	31,86	WBSR22	-1,17	-1,28	-0,34	0	
High	P63208	Splice kinase-associated protein 1 [OS=Homo sapiens]	22,0,18,0564	40,49,39,75	6	6	163	18,646	SKP1	-1,99	-1,33	0,23	0	
High	P62314	Small nuclear ribonucleoprotein Sm D1 [OS=Homo sapiens]	24,16,8948	27,73,10,24	2	6	2	119	13,273	SNRPD1	-1,4	-1,34	-0,06	0
High	Q9NP564	nucleolar protein of 40 kDa [OS=Homo sapiens]	33,6,76,564	63,90,04,49	19	130	241	27,552	ZCHC17	-1,27	-1,36	0,08	0	
High	P55081	microfibrillar-associated protein 1 [OS=Homo sapiens]	128,56,9572	45,10,20,057	18	33	439	51,927	MIFAP1	-0,54	-1,36	0,82	0	
Medium	Q43172-1	U4/U6 small nuclear ribonucleoprotein Prp4 [OS=Homo sapiens]	3,0,058,13,54	2,68,19,92,34	1	1	522	58,412	PRPF4	-1,23	-1,36	0,13	0	
High	Q00268	Transcription initiation factor TFIID subunit 4 [OS=Homo sapiens]	11,19,2,78,19	3,2,25,80,45	3	3	1085	110,047	TAF4	-1,3	-1,37	0,06	0	
High	Q86U06-1	probable RNA-binding protein 23 [OS=Homo sapiens]	96,9,26,87,09	12,48,5,19,36	12	45	439	48,701	REM23	-1,06	-1,38	0,32	0	
High	Q7Z7H6-3	Isotform 3 of Centromere protein V [OS=Homo sapiens]	78,74,67,632	51,47,60,882	11	20	1	272	29,712	CENPV	-1,25	-1,39	0,14	11
High	Q9PY16-1	PhD and RING finger domain-containing protein 1 [OS=Homo sapiens]	60,81,63,015	13,46,2,047	16	29	15	1649	178,557	PHRF1	-1,54	-1,39	-0,15	1
High	P09543-1	2',3'-cyclic-nucleotide 3'-phosphodiesterase [OS=Homo sapiens]	7,14,62,61,95	8,0,76,00,95	2	2	421	47,549	CNP	-1,58	-1,4	-0,18	0	
High	U15042	U2 snRNP-associated SURP motif-containing protein [OS=Homo sapiens]	144,64,309	39,94,1691	33	60	33	1029	118,219	U2SURP	-1,62	-1,21	-0,21	0

High	O75815-1	Breast cancer anti-estrogen resistance protein 3 [OS=Homo sapiens]	139,166545	44,1212121	30	43	29	825	92,507	BCAR3	-1	-1,41	0,41	1
High	Q53G59	Isotform 2 of PhD finger protein 20-like protein 1 [OS=Homo sapiens]	18,3979617	28,0701754	7	7	1	285	32,805	PF20L1	-2,62	-1,43	-1,2	0
Low	Q4/U6.5	tr-siRNP-associated protein 2 [OS=Homo sapiens]	1,90274265	3,36283186	1	1	1	565	65,34	UPF39	-1,14	-1,43	0,03	0
High	P21741	Mickline [OS=Homo sapiens]	45,2857095	48,951049	6	14	6	143	15,575	MDK	-1,53	-1,45	-0,88	0
High	P30454	Serpin H1 [OS=Homo sapiens]	15,8923594	16,592177	4	4	4	418	46,411	SEPRINH1	-0,82	-1,45	0,63	0
High	P14773	Tripeptidyl-peptidase 1 [OS=Homo sapiens]	5,50612389	24,48867851	1	2	1	563	61,21	TPP1	-2,05	-1,47	-0,58	0
High	Q8NAV1-1	PremRNA-splicing factor 38A [OS=Homo sapiens]	27,8947465	27,5641026	7	9	7	312	37,453	PRPF38A	-0,82	-1,5	0,68	0
Medium	Q9UJ40-2	Isotform 2 of Zinc finger protein 346 [OS=Homo sapiens]	4,22776599	7,836906	2	2	2	319	35,748	ZNF346	-0,08	-1,5	1,42	0
High	Q8TA96	retinitis pigmentosa 8 protein [OS=Homo sapiens]	54,1948098	31,2217195	9	26	9	221	26,091	RPN9	-0,7	-1,51	0,81	0
High	Q2962	transcription initiation factor TFIID subunit 10 [OS=Homo sapiens]	4,8101893	6,422010378	1	1	1	218	21,698	TAF10	-1,85	-1,51	-0,34	0
High	Q9UBU9-1	nuclear RNA export factor 1 [OS=Homo sapiens]	28,5362736	10,50080738	5	8	5	619	70,139	NXF1	-1,25	-1,52	0,27	0
High	Q5T707-1	Collagen alpha-1(XIV) chain [OS=Homo sapiens]	17,1579754	5,565792873	8	8	8	1796	193,394	COL14A1	-0,61	-1,52	0,91	0
Low	Q6LE60-1	U11/U12 small nuclear ribonucleoprotein 48 kDa protein [OS=Homo sapiens]	1,0616304	3,833480826	1	1	1	339	39,941	SNRNPA18	-0,02	-1,52	1,5	0
High	Q13885	Tubulin beta 3 chain [OS=Homo sapiens]	780,378483	67,1910112	28	363	1	445	49,875	TUBB2A	-1,41	-1,54	0,13	0
Medium	Q00410-3	Isotform 3 of Importin-5 [OS=Homo sapiens]	4,20251042	2,33,183857	2	2	2	1115	125,464	IP05	-2,15	-1,54	-0,61	0
Low	Q965K2	Transmembrane protein 209 [OS=Homo sapiens]	1,3638131	2,85104991	1	1	1	561	63,881	TMEM209	-0,87	-1,57	0,7	0
High	P15397-1	Importin-8 [OS=Homo sapiens]	14,0269027	10,5852449	3	5	2	1037	119,861	IP08	-1,59	-1,58	-0,01	0
High	P30414	NK-tumor recognition protein [OS=Homo sapiens]	8,33045913	2,73597811	3	3	3	1462	165,577	NKTR	-1,74	-1,58	-0,16	0
High	P35579-1	Mysin-9 [OS=Homo sapiens]	558,932482	54,7441898	103	170	90	1960	226,392	MYH9	-2,68	-1,59	-1,09	11
High	Q9NFT7-1	F-box only protein 28 [OS=Homo sapiens]	126,800917	12,7826087	19	48	19	368	41,123	FBXO28	-0,74	-1,59	0,85	0
High	Q8NWY92	PhD finger protein 20-like protein 1 [OS=Homo sapiens]	37,7801303	16,420210516	12	12	6	1017	114,338	PHF20L1	-2,09	-1,59	-0,5	4
High	Q9H9Y2	Ribosome production factor 1 [OS=Homo sapiens]	13,2036689	9,16905444	3	4	3	349	40,086	RPF1	-1,01	-1,59	0,58	0
Medium	Q9UJ10-2	Isotform 2 of Translation initiation factor eIF-2B subunit delta [OS=Homo sapiens]	2,93903319	2,57826888	1	1	1	543	59,578	EIF2B4	-0,73	-1,59	0,86	0
High	Q6KX1	60S ribosomal protein L7/like 1 [OS=Homo sapiens]	8,69757192	15,745145	3	3	3	246	28,643	RPL7L	-1,94	-1,6	-0,34	0
High	Q8N5F7	NF-kappa-B-activating protein [OS=Homo sapiens]	69,1390875	24,8191277	9	31	9	415	47,11	NKAP	-0,86	-1,62	0,76	0
High	P50897	Palmitoyl-protein thioesterase 1 [OS=Homo sapiens]	19,6443201	7,84313725	2	4	2	306	34,171	PPT1	-1,48	-1,63	0,15	0
High	Q9NV1-3	Fancion anemia group I protein [OS=Homo sapiens]	5,2692987	2,10843373	3	3	3	1328	149,229	FANCI	-1,32	-1,65	0,33	0
High	Q6GQ7	probable ATP-dependent RNA helicase DDX27 [OS=Homo sapiens]	84,9716213	29,3696949	21	30	21	796	89,779	DDX27	-1,09	-1,66	0,57	0
High	Q6B016	Lysine-specific demethylase 4D [OS=Homo sapiens]	12,9345382	10,51625224	4	4	4	523	58,565	KDM4D	-1,39	-1,66	0,27	0
High	Q9Y34Z	Probable U3 small nucleolar RNA-associated protein 11 [OS=Homo sapiens]	37,6853831	33,9920349	10	14	10	253	30,428	JPT1L; UTP1	-1,29	-1,67	0,39	0
High	Q8Y7T6	E3 ubiquitin-protein ligase mbd1 [OS=Homo sapiens]	31,9749335	11,6302187	10	10	10	100,066	MBI1	-2,18	-1,72	-0,46	0	
High	Q32306	probable ATP-dependent RNA helicase DDX27 [OS=Homo sapiens]	9,5977935	4,11428771	3	3	3	875	100,925	DDX10	-0,09	-1,74	1,83	0
High	Q94876	Transmembrane and coiled-coil domains protein 1 [OS=Homo sapiens]	5,25178933	2,29709035	1	2	1	653	72,038	TMCC1	-0,65	-1,8	1,15	0
High	Q9UJ5	Transmembrane and coiled-coil domains protein 3 [OS=Homo sapiens]	189,445114	52,2012579	25	75	25	477	53,752	TMCC3	-1,35	-1,81	0,45	0
High	Q9B6016	Lysine-specific demethylase 4D [OS=Homo sapiens]	5,12758788	2,64084507	3	3	3	441	49,422	MY01B	-1,37	-1,81	0,91	0
High	Q9UJBM7	7-dehydrocholesterol reductase [OS=Homo sapiens]	33,5439749	12,4210526	6	11	6	475	50,454	DHCR7	-1,77	-1,83	0,06	0
High	Q9Y7L1-1	exosome complex exonuclease RRP44 [OS=Homo sapiens]	62,6392095	21,8162839	17	20	17	958	108,934	DIS3	-2,52	-1,85	-0,67	0
High	Q99599	Plakophilin-2 [OS=Homo sapiens]	5,25178933	4,11428771	8	8	8	881	97,355	PKP2	-2,17	-1,87	-0,3	0
Low	Q5P701	E3 ubiquitin-protein ligase DDX5 [OS=Homo sapiens]	2,13065102	2,0581536	1	1	1	2799	127,038	TMCC1	-0,65	-1,8	1,15	0
High	Q9EJM3	Chromosome alignment-maintaining phosphoprotein 1 [OS=Homo sapiens]	13,3404779	9,85221675	5	5	5	812	89,043	CHAMP1	-1,35	-1,95	0,19	0
High	Q9BX6	Nucleolar and spindle-associated protein 1 [OS=Homo sapiens]	99,22324854	47,3922902	19	30	3	441	131,902	NUSAP1	-1,66	-1,96	0,3	12
Medium	Q96QZ7	Membrane-associated guanylate kinase, WW and PDZ domain-containing protein 1 [OS=Homo sapiens]	33,5439749	12,4210526	2	2	2	1491	164,481	MAGI1	-1,68	-2	0,31	0
Low	Q9NVT0-1	exocyst complex component 1 [OS=Homo sapiens]	1,80937706	1,23042936	1	1	1	894	101,917	EOC1	-0,35	-2	1,95	0
High	Q6PJP8	DNA cross-link repair 1A protein [OS=Homo sapiens]	5,29503299	3,07692308	2	2	2	1040	116,226	DCREL1A	-2,37	-1,87	0,92	0
Low	Q9Y600	Choline/ethanolamine phosphotransferase 1 [OS=Homo sapiens]	1,46306298	2,64423077	1	1	1	416	46,523	CPT1	-1,17	-2,04	0,87	0
High	Q9HAZ1	Dual specificity myoinositol-phosphate transferase Click [OS=Homo sapiens]	12,8622695	7,9002079	3	3	1	481	57,455	CLK4	-0,92	-2,05	1,13	0
Low	A01T2	Mediator of RNA polymerase II transcription subunit 19 [OS=Homo sapiens]	2,01295971	4,50819672	1	1	1	244	26,257	MED19	-2,1	-2,07	-0,03	0
High	Q9Z620	pre-mRNA-splicing factor ATP-dependent RNA helicase RRP16 [OS=Homo sapiens]	5,2101111	1,629959185	2	3	1	1227	140,415	DHX38	-1,11	-2,14	1,03	0
High	Q9NZM5	Glioma tumor suppressor candidate region gene 2 protein [OS=Homo sapiens]	68,863651	24,0585774	11	20	11	478	54,356	GLTCR2	-1,81	-2,16	0,35	0
High	P63241-2	Isotform 2 of Elukaryotic translation initiation factor 5A-1 [OS=Homo sapiens]	4,68306301	19,5652174	2	2	2	184	20,157	EF5	-2,36	-2,16	0,41	0
High	Q86GD4-5	Isotform 5 of Aurora kinase B [OS=Homo sapiens]	14,3924959	12,4637681	4	4	4	345	39,442	AURKB	-1,72	-2,18	0,46	0
High	Q9E4H5	60S ribosomal protein L39-like [OS=Homo sapiens]	11,0581751	31,3737549	1	2	1	51	6,289	RPL29L	-2,12	-2,19	0,07	0
High	Q8NFW8	N-acrylneuraminate cytidylyltransferase [OS=Homo sapiens]	60,296364	35,483871	12	15	12	434	48,349	CMAS	-1,52	-2,2	0,68	0
High	Q9H0W5	Coiled-coil domain-containing protein 8 [OS=Homo sapiens]	16,4016667	10,9665428	3	6	3	538	59,339	CDC28	-0,13	-2,2	2,07	0
High	Q8YB3	Serine/threonine-protein kinase MRCK alpha [OS=Homo sapiens]	149,484163	22,0132743	16	56	16	904	102,274	SRRM1	-1,81	-2,21	0,41	0
Low	Q9BSI2-4	Isotform 3 of Gamma-tubulin complex component 2 [OS=Homo sapiens]	1,06345399	0,752668817	1	1	1	930	105,561	TUBGCP2	-0,37	-2,33	1,97	0
High	Q15334	Lethal(2) giant larvae protein homolog 1 [OS=Homo sapiens]	120,75181	34,68045151	27	33	27	1064	115,446	LGII	-2,16	-2,19	0,21	0
High	Q9HCSS	band 4.1-like protein 4A [OS=Homo sapiens]	6,23087091	3,06122449	2	2	2	686	79,01	EP414LAA	-2,46	-2,43	-0,03	0
Low	Q5V75-6	Isotform 6 of Serine/threonine-protein kinase MRCK alpha [OS=Homo sapiens]	1,373440832	0,50533408	1	1	1	1781	202,684	CDC42BPA	-0,27	-2,43	2,15	0
Low	Q60921	checkpoint protein HUS1 [OS=Homo sapiens]	1,3372525	7,14285714	1	1	1	280	31,67	HUS1	-0,92	-2,43	1,52	0
High	Q9UJ5	Vang-like protein 1 [OS=Homo sapiens]	8,83390608	6,1420355	3	3	3	521	59,677	VANG1	-1,68	-2,45	0,76	0
Low	Q86U8-1	Teleomerase-binding protein EST1A [OS=Homo sapiens]	1,70245833	0,91613813	1	1	1	1419	160,362	SMG6	-2,09	-2,45	0,36	0
High	Q9Y4F1	FERM, RhGeff and pleckstrin domain-containing protein 1 [OS=Homo sapiens]	36,5165893	14,8323559	13	13	13	1045	118,559	FARP1	-2,27	-2,51	-0,19	0
High	Q9UBC1-1	NF-kappa-B inhibitor-like protein 1 [OS=Homo sapiens]	11,852237	12,339598	4	5	4	381	43,177	NFKB1L1	-2,29	-2,53	0,24	0

High	Q6P1M3-1	Lethal(2) giant larvae protein homolog 2 [OS=Homo sapiens]	92.5043502	31.5686275	24	27	1020	113.377	LIG2	-2.05	-2.54	0.49	0		
High	Q8UVP8-1	Ankyrin repeat domain-containing protein 26 [OS=Homo sapiens]	423.049589	67.1152272	96	148	1	1709	196.202	ANKRD26	-2.52	-2.86	0.34	65	
High	O0N458	Interferon- $\gamma$ induced developmental regulator 1 [OS=Homo sapiens]	17.763305	15.1152272	5	6	5	451	50.236	IFRD1	-3.34	-2.97	-0.37	0	
High	Q8UVP0	centromeral protein of 164 kDa [OS=Homo sapiens]	10.7633195	5.47945205	6	6	6	1460	164.214	CEP164	-3.21	-2.99	-0.22	0	
High	Q9N245	CD59/iron-sulfur domain-containing protein 1 [OS=Homo sapiens]	8.0793108	26.8518319	3	3	3	108	12.191	CISD1	-2.1	-3.08	0.99	0	
High	Q9UNI2-2	Isotform 2 of Translocase-associated protein subunit gamma [OS=Homo sapiens]	6.6725462	7.0707070	1	1	1	198	22.596	SRR3	-0.26	-3.1	3.36	0	
High	Q9E7T4	serine racemase [OS=Homo sapiens]	13.9254633	17.9411765	3	4	3	340	36.543	SRR	-2.14	-3.15	1.01	0	
High	P355Z0-4	Isotform 4 of Myosin-10 [OS=Homo sapiens]	224.604036	34.1803687	57	67	44	2007	232.385	MYH10	-4.98	-3.56	-1.42	0	
Medium	P38XW7	Cateye syndrome critical region protein 5 [OS=Homo sapiens]	3.00318198	2.12765957	1	1	1	423	46.292	CECR5	-3.63	-3.67	0.03	0	
High	Q8HNP8	putative phospholipase B-like 1 [OS=Homo sapiens]	41.05339909	21.0526516	11	12	11	589	65.43	PBLB2	-4.14	-3.99	-0.16	0	
High	P02533	Keratin, type I cytoskeletal 14 [OS=Homo sapiens]	132.508456	64.6138644	29	94	9	472	51.529	KRT14	-5.77	-4.05	-1.72	9	
High	Q9H111-1	Regulator of nonsense transcripts 3A [OS=Homo sapiens]	9.96655744	6.932377311	3	4	3	476	54.663	UPF3A	-3.46	-4.51	1.05	0	
High	P41743	Protein kinase c iota type [OS=Homo sapiens]	21.6392887	11.2416107	4	4	4	596	68.218	PRKCJ	-3.94	-5.28	1.34	0	
Low	P08E01	Integrator complex subunit 3 [OS=Homo sapiens]	2.20446756	1.629931371	1	1	1	1043	117.994	INTS3	-6.99	-6.92	-0.07	0	
Low	O60716-1	Catenin delta-1 [OS=Homo sapiens]	1.2645608	1.23966842	1	1	1	968	108.103	CTNND1	-7.33	-7.26	-0.07	0	
High	P07355-2	Isotform 2 of Annexin A2 [OS=Homo sapiens]	48.4627971	32.4929972	9	12	9	357	40.386	ANXA2	-7.87	-7.32	-0.54	0	
High	Q92413	Dermogelin-1 [OS=Homo sapiens]	9.92227773	5.52907531	4	4	4	1049	113.676	DSG1	-7.58	-7.51	-0.07	0	
Medium	P23528	Cofilin-1 [OS=Homo sapiens]	2.49227902	10.2409639	1	1	1	166	18.491	CFI1	-7.96	-7.9	-0.07	0	
High	O5049-3	Isotform 3 of Tight junction protein ZO-3 [OS=Homo sapiens]	5.28898281	2.34541578	2	2	2	938	103.361	TJP3	-8.05	-7.98	-0.07	0	
Medium	Q9GD7	Zinc finger protein ZNDC [OS=Homo sapiens]	2.388336957	1.515152	1	1	1	858	89.932	ZNDC	-8.16	-8.09	-0.07	0	
Low	Q99829	Copine-1 [OS=Homo sapiens]	1.81615696	3.72439479	1	1	1	537	81.022	COPN1	-8.16	-8.09	-0.07	0	
Low	O43395	U4/U6 small nuclear ribonucleoprotein Prp3 [OS=Homo sapiens]	1.433090134	3.51390922	1	1	1	683	77.481	PRPF3	-8.28	-8.22	-0.07	0	
High	O00159-1	Unconventional myosin-ic [OS=Homo sapiens]	6.13132896	3.57478833	3	3	3	1063	121.606	MYO1C	-8.29	-8.23	-0.07	0	
Low	Q8WVY2-1	zinc finger protein 444 [OS=Homo sapiens]	1.89226882	1.892268636	1	1	1	327	35.182	ZNFX44	-8.38	-8.31	-0.07	0	
Medium	P14923	Tetrastricopeptide repeat 21B [OS=Homo sapiens]	3.10248315	2.3556231	2	3	2	1316	150.84	TTC21B	-8.4	-8.33	-0.07	0	
Medium	Q96077	Junction plakophilin [OS=Homo sapiens]	3.14138389	2.95302013	2	2	2	745	81.693	JUP	-8.41	-8.35	-0.07	0	
Medium	P51530	Zinc finger and BTB domain-containing protein 10 [OS=Homo sapiens]	3.84649001	2.98850763	1	1	1	871	94.835	ZBTB10	-8.51	-8.45	-0.07	0	
High	P0C04-1	Complement C4A [OS=Homo sapiens]	4.19889116	2.0754717	2	2	2	1060	120.337	DNAZ	-8.73	-8.67	-0.07	0	
High	P13645	Keratin, type I cytoskeletal 10 [OS=Homo sapiens]	26.7397216	6.276172329	35	221	30	584	55.792	KRT10	-1.69	-8.91	-2.78	4	
High	Q9UH99-2	Isotform 2 of SUN domain-containing protein 2 [OS=Homo sapiens]	6.04750404	4.065504065	2	3	2	738	82.452	SUN2	-8.98	-8.92	-0.07	0	
Medium	P04114	apolipoprotein B-100 [OS=Homo sapiens]	4.25156863	0.74542382	3	4	3	4563	51.285	APOB	-9.17	-9.1	-0.07	0	
High	P02751-15	Isotform 15 of fibronectin [OS=Homo sapiens]	7.15332396	1.33225676	3	3	3	2477	272.15	FBN1	-9.24	-9.17	-0.07	0	
High	Q86Y23	Homerin [OS=Homo sapiens]	75.6010623	20.7719198	17	25	17	2850	282.228	HNRN	-9.34	-9.28	-0.07	0	
Medium	P23112	U6 snRNA	1.36061313	1.35131313	1	1	1	80	91.122	LMW6	-9.36	-9.29	-0.07	0	
Medium	Q96IM7	Lethal(3)-like protein 3 [OS=Homo sapiens]	2.66054856	1.02564103	1	1	1	780	88.28	LMBT13	-9.4	-9.34	-0.07	0	
High	P09622	Dihydrofolyl dehydrogenase, mitochondrial [OS=Homo sapiens]	5.38316111	4.7151277	2	2	2	509	54.143	DLD	-9.43	-9.36	-0.07	0	
High	P06660	Mysin light polypeptide 6 [OS=Homo sapiens]	1.67302732	1.33225676	1	1	1	529	61.731	MYL6	-9.5	-9.43	-0.07	0	
Low	P24752	Acetyl-CoA acyltransferase, mitochondrial [OS=Homo sapiens]	1.67302732	1.33225676	1	1	1	427	151.57	CAT1	-9.56	-9.49	-0.07	0	
Medium	Q6R2W3	SCAN domain-containing protein 3 [OS=Homo sapiens]	2.38333984	0.52830189	1	1	1	1325	56.747	CAN3	-9.61	-9.54	-0.07	0	
Low	P30101	Protein disulfide-isomerase A2 [OS=Homo sapiens]	1.94923356	2.17821782	1	1	1	505	127.702	PDIA3	-9.63	-9.57	-0.07	0	
Low	Q8NW08	DNA-directed RNA polymerase II subunit RPB2 [OS=Homo sapiens]	1.16008222	0.61782877	1	1	1	1133	127.702	POLR3B	-9.63	-9.57	-0.07	0	
High	Q8WVY9	DEF domain-containing protein 1B [OS=Homo sapiens]	5.21632323	2.945650284	1	1	1	529	161.731	DEPDCCB	-9.67	-9.6	-0.07	0	
High	P06702	Protein S100A9 [OS=Homo sapiens]	19.7798749	3.71792882	4	6	4	114	13.234	S100A9	-9.7	-9.63	-0.07	0	
High	Q06033-1	Inter-alpha-trypsin inhibitor heavy chain H3 [OS=Homo sapiens]	5.15645579	1.46067416	1	1	1	890	99.787	ITIH3	-9.81	-9.74	-0.07	0	
High	P04695	Keratin, type I cytoskeletal 17 [OS=Homo sapiens]	54.2191556	0.9925926	15	42	4	432	48.076	KRT17	-9.89	-9.83	-0.07	0	
High	P255908	Keratin, type II cytoskeletal 2 [epidermal] [OS=Homo sapiens]	273.339083	77.4647887	50	178	37	639	65.393	KRT2	-10.64	-9.99	-0.65	6	
Medium	P29508	Serpin B3 [OS=Homo sapiens]	2.34084472	1.79487179	1	1	1	390	44.537	SERPINB3	-10.16	-10.1	-0.07	0	
High	Q5D862	Filaggrin 2 [OS=Homo sapiens]	5.90054145	0.8782936	2	3	2	2391	247.928	FLG2	-10.19	-10.13	-0.07	0	
Medium	P23435	Acetyl-CoA acyltransferase, mitochondrial [OS=Homo sapiens]	4.432654398	6.20155039	2	2	2	437.594	46.022	GMPBP	-10.16	-10.16	-0.07	0	
Medium	O75190	DnaJ homolog subfamily B member 6 [OS=Homo sapiens]	1.95550484	3.62694301	1	1	1	210.084	10.23	CBLN1	-10.19	-10.16	-0.07	0	
High	Q8WV50-2	Isotform 2 of PhD finger protein 6 [OS=Homo sapiens]	4.22148699	5.21472293	1	1	1	326	36.065	DNAJB6	-10.26	-10.19	-0.07	0	
High	Q8N5H7-1	SH2 domain-containing protein 3C [OS=Homo sapiens]	153.832909	50.9615285	14	69	1	312	35.306	FHL6	-10.33	-10.27	-0.07	0	
High	Q5T750	Skin-specific protein 32 [OS=Homo sapiens]	1.36632959	3.2	1	1	1	346	38.69	ANXA1	-10.7	-10.64	-0.07	0	
High	P04083	Anxinin A1 [OS=Homo sapiens]	1.9150953	2.60115607	1	1	1	558	62.542	RHM26	-10.72	-10.65	-0.07	0	
High	Q5T8P6-4	Isotform 4 of RNA-binding protein 26 [OS=Homo sapiens]	144.460222	42.6523297	26	42	1	352	38.603	MDH1	-10.88	-10.82	-0.07	0	
Low	Q5T925-3	Isotform 3 of Malate dehydrogenase, cytoplasmic [OS=Homo sapiens]	13.5886492	7.3863664	2	3	2	1997	226.61	OTOF	-10.98	-10.91	-0.07	0	
Low	Q5T750-1	Otoferlin [OS=Homo sapiens]	1.26533616	0.75112669	1	2	1	1	1596	174.899	SETBP1	-11.04	-10.98	-0.07	0
High	P06779	Keratin, type I cytoskeletal 16 [OS=Homo sapiens]	99.7351547	56.5965195	25	77	10	473	51.236	KRT16	-11.26	-10.3	-0.07	0	
Low	Q9NP78	ATP-binding cassette sub-family B member 9 [OS=Homo sapiens]	1.42701229	0.91383812	1	2	1	766	84.421	ABCB9	-11.39	-11.32	-0.07	0	
High	P31151	Protein S100A7 [OS=Homo sapiens]	9.90553302	22.7722772	3	4	3	101	11.464	S100A7	-11.43	-11.36	-0.07	0	
High	P02538	Keratin, type II cytoskeletal 6A [OS=Homo sapiens]	143.672933	51.503546	37	106	2	564	60.008	KRT6A	-11.46	-11.39	-0.07	1	

High	O15479	Melanoma-associated antigen B2 [OS=Homo sapiens]	9.37053898	8.15047022	3	3	3	319	35.255	MAGEB2	-1.48	-11.41	-0.07	0
High	Q7Z7K6-1	Centromere protein V [OS=Homo sapiens]	80.2317865	51.6363636	11	21	1	275	29.927	CNPV	-1.62	-11.55	-0.07	0
High	P22768-1	Serum albumin [OS=Homo sapiens]	12.2750214	6.73234811	4	8	4	609	69.321	ALB	-1.61	-11.61	-0.07	0
High	P13647	Keratin, type II cytoskeletal 5 [OS=Homo sapiens]	113.052367	43.0508475	32	95	15	590	62.34	KRT5	-1.68	-11.61	-0.07	0
High	P02765	Alpha-2-HS glycoprotein [OS=Homo sapiens]	8.07477799	5.17711172	2	2	2	367	39.3	AHG	-1.81	-11.75	-0.07	0
High	P04004	Vitronectin [OS=Homo sapiens]	4.7878124	3.13807531	1	1	1	478	54.271	VTN	-1.95	-11.89	-0.07	0
High	P01024	Complement C3 [OS=Homo sapiens]	34.3838108	5.77269994	9	15	9	1663	187.03	C3	-1.98	-11.92	-0.07	0
High	P01023	Alpha-2-macroglobulin [OS=Homo sapiens]	9.75991125	2.6458616	4	6	4	1474	163.188	A2M	-1.12	-11.93	-0.07	0
Medium	P00736	Complement C1r subcomponent [OS=Homo sapiens]	3.0838593	2.26950355	2	2	2	705	80.067	C1R	-1.12	-11.94	-0.07	0
Low	P60318	Germinal-center associated nuclear protein [OS=Homo sapiens]	1.20683847	1.21212121	1	1	1	1380	218.267	MCM3AP	-1.12	-11.94	-0.07	0
Low	Q8NNV4-2	Isotform 2 of Poly(A) RNA polymerase, mitochondrial [OS=Homo sapiens]	1.17688194	1.264044294	1	1	1	712	78.784	-	-12.02	-11.95	-0.07	0
High	P05109	Protein S100-A8 [OS=Homo sapiens]	11.7598865	52.688172	5	7	5	93	10.828	S100A8	-12.12	-12.05	-0.07	0
High	P07477	Tryptin-1 [OS=Homo sapiens]	6.60781994	11.3360324	2	5	2	247	26.541	PRSS1	-12.15	-12.09	-0.07	0
High	P04259	Keratin, type II cytoskeletal 6B [OS=Homo sapiens]	140.408359	50.7092199	37	103	2	564	60.03	KRT6B	-12.55	-12.48	-0.07	12
High	P02746	Complement C1q subcomponent subunit B [OS=Homo sapiens]	14.8911812	5.53359684	1	7	1	253	26.704	C1QB	-13.24	-13.17	-0.07	0
Low	P02042	Hemoglobin subunit delta [OS=Homo sapiens]	2.25173443	6.80272109	1	3	1	147	16.045	HBD	-13.29	-13.22	-0.07	0
High	P35527	Keratin, type I cytoskeletal 9 [OS=Homo sapiens]	264.902853	78.0096308	35	188	34	623	62.027	KRT9	-13.44	-13.37	-0.07	0
High	P04264	Keratin, type II cytoskeletal 11 [OS=Homo sapiens]	396.145244	69.5652174	55	332	46	644	65.999	KRT1	-13.71	-13.64	-0.07	5
High	P06314	Ig kappa chain V-IV region [OS=Homo sapiens]	13.1427255	17.9104478	2	10	2	134	14.956	-	-14.79	-14.73	-0.07	0
High	P01605	Ig kappa chain V-I region [OS=Homo sapiens]	14.9505445	25	2	6	2	108	11.827	-	-15.19	-15.13	-0.07	0
Low	Q7Z6K1	THAP-domain-containing protein 5 [OS=Homo sapiens]	2.08460016	1.17721519	1	10	1	395	45.388	THAP5	-15.29	-15.22	-0.07	0
High	P01859	Ig gamma-2 chain C-region [OS=Homo sapiens]	4.901446824	10.7361963	2	6	2	326	35.878	IGHG2	-15.96	-15.9	-0.07	0
Low	P14314	Glucosidase 2 subunit beta [OS=Homo sapiens]	1.41691463	4.92424242	1	1	1	528	59.388	PRKCSH	-16.61	-16.61	-0.07	0
Low	P46019	Phosphorylase b kinase regulatory subunit alpha, liver isoform [OS=Homo sapiens]	1.35261703	1.45748988	1	1	1	1235	138.32	PKHA2	-16.61	-16.61	-0.07	0

**Annex Table III: Liquid chromatography-tandem mass spectrometry (LC-MS/MS) data from SILAC (UV treatment).** The data were analyzed using SEQUEST in Proteome Discoverer 2.1 and searched in the complete human proteome database (Swiss-Prot). Peptide identification was filtered at a false discovery rate (FDR) < 1%. Coverage: percentage of the protein sequence covered by identified peptides (confident coverage threshold >1). Peptides: total number of distinct peptide sequences identified in the protein group. PSMs: total number of identified peptide spectra matched for the protein. Unique peptides: The number of peptide sequences that are unique to a protein group. MW: molecular weight. Abundance ratios are calculated based on light (for XPF-KO cells untreated), medium (for XPF-KO + XPF-Wt treated with UV 40J/m2, 1h of recovery), heavy (for XPF-KO + XPF-Wt untreated) forms of lysine and arginine. Razor peptides: peptides that can be assigned to more than one protein.

Protein FDR Confidence	Accession	Description	Sum PEP Score	Coverage	# Peptides	# PSMS	# Unique Peptides	# AAs	MW [kDa]	Gene ID	Abundance Ratio (log2); (Medium) / (Light)	Abundance Ratio (log2); (Heavy) / (Light)	# Razor Peptides
High	Q96CN7	[Isochiorismatase domain-containing protein 1 [OS=Homo sapiens]]	51,9804541	39,9328859	8	14	8	298	32,216	ISOC1	11,65	11,92	-0,27
High	Q9N256	Farnin-2 [OS=Homo sapiens]	6,7454362	0,9291525	1	3	1	1722	179,993	FMN2	11,87	11,92	-0,05
High	Q5VVV7	protein SLX4IP [OS=Homo sapiens]	13,4851316	13,9705882	4	5	4	408	45,524	SLX4IP	10,23	9,81	0,43
Medium	CB1724	TNF receptor-associated factor 4 [OS=Homo sapiens]	3,5026749	1,4893617	1	1	1	470	53,507	TRAF4	9,4	8,83	0,57
High	P51784	Ubiquitin-carboxy-terminal hydrolase 11 [OS=Homo sapiens]	8,5178222	3,1152648	3	3	3	963	109,747	USP14	8,12	7,63	0,49
High	P07922-1	DNA excision repair protein ERCC-1 [OS=Homo sapiens]	454,541232	74,4107744	29	341	29	297	32,542	ERCC1	5,31	5,4	-0,09
High	Q709F0-1	acyl-CoA dehydrogenase family member 11 [OS=Homo sapiens]	135,823536	30,3846154	24	62	24	780	87,228	ACAD11	4,65	5,15	-0,5
High	Q81Y92	Structure-specific endonuclease subunit SLX4 [OS=Homo sapiens]	659,781887	0,8015153	76	433	76	1834	199,889	SLX4	4,23	3,53	0,69
High	Q92889	DNA repair endonuclease XPF [OS=Homo sapiens]	1623,85678	75,6550224	97	250	97	916	104,42	ERCC2	4,35	3,52	0,83
High	Q9BQ83-1	Structure-specific endonuclease subunit SLX1 [OS=Homo sapiens]	17,8654641	15,636536	3	5	3	275	30,751	SLX1B;SLX1A	2,9	2,85	0,05
High	Q9HJR3	Myosin light chain kinase 2, skeletal/cardiac muscle [OS=Homo sapiens]	30,8644891	2,18120805	3	48	3	596	64,644	MLK2	2,77	2,81	-0,04
High	P08075	Protein CMSS1 [OS=Homo sapiens]	5,33953239	6,81005834	2	3	2	279	31,864	CMSS1	2,02	2,8	-0,78
High	Q96HS1-1	Serine/threonine-protein phosphatase Peam5, mitochondrial [OS=Homo sapiens]	77,0766606	39,7923875	14	46	14	289	31,985	PGAM5	3,06	2,51	0,55
High	Q02543	60S ribosomal protein L18a [OS=Homo sapiens]	6,21212243	11,9318182	2	2	2	176	20,749	RPL18A	2,48	2,5	-0,02
High	P36578	60S ribosomal protein L4 [OS=Homo sapiens]	40,9905461	15,4566745	7	17	7	427	47,667	RPL4	2,25	2,44	-0,19
High	P05387	60S acidic ribosomal protein P1 [OS=Homo sapiens]	39,1248383	60,791304	4	9	4	115	61,658	RPLP2	2,35	2,42	-0,06
High	P05386	60S acidic ribosomal protein P1 [OS=Homo sapiens]	11,4708249	14,0350877	1	3	1	114	11,507	RPLP1	1,64	2,4	-0,76
High	Q15233	Non-PoU domain-containing octamer-binding protein [OS=Homo sapiens]	8,15039186	2,33545648	1	2	1	471	54,197	NONO	1,73	2,34	-0,61
High	P11310-2	Isoform 2 of Medium-chain specific acyl-CoA dehydrogenase, mitochondrial [O	39,11152081	16,2352941	6	9	6	425	46,99	ACADM	2,42	2,32	0,1
High	Q9NPV1	ATP-dependent RNA helicase DDX18 [OS=Homo sapiens]	334,238807	62,338806	34	190	34	670	75,359	DDX18	0,72	2,18	-1,46
High	Q14204	Cytoplasmic dynein 1 heavy chain 1 [OS=Homo sapiens]	16,8504643	1,8941045	8	9	8	4645	532,072	DYNC1H1	2,04	2,06	-0,02
Low	Q96PQ7-3	Isoform 3 of Kelch-like protein 5 [OS=Homo sapiens]	1,17600441	1,55844156	1	1	1	7070	86,605	MRPL9	0,88	2,06	-1,18
High	P61313-1	60S ribosomal protein L7 [OS=Homo sapiens]	29,5569598	20,5882353	4	13	4	204	24,131	RPL15	1,8	2,01	-0,2
High	P18124	60S ribosomal protein L7 [OS=Homo sapiens]	80,6394176	49,1935484	13	30	13	248	29,207	RPL17	1,82	1,95	-0,13
High	P50914	60S ribosomal protein L14 [OS=Homo sapiens]	32,34633162	19,5348373	4	9	4	215	23,417	RPL14	1,88	1,95	-0,07
High	Q6PSPR6	60S ribosomal protein L22-like 1 [OS=Homo sapiens]	97,1588866	52,4590164	6	52	4	122	14,598	RPL22L1	1,89	1,93	-0,04
High	Q9BYD2	30S ribosomal protein 19, mitochondrial [OS=Homo sapiens]	8,91942583	3,37078652	1	3	1	267	30,224	MRPL9	0,93	1,92	-1,95
High	Q96NY9	cross-over junction endonuclease MUS81 [OS=Homo sapiens]	61,9513325	21,960726	10	33	10	551	61,135	MUS81	1,75	1,78	-0,03
High	P06782	Sh2-domain-containing protein 3A [OS=Homo sapiens]	9,99678551	6,77082333	3	7	3	576	63,054	SH2D3A	2,88	1,77	0,2
High	Q06787	fragile X mental retardation protein 1 [OS=Homo sapiens]	26,9664055	9,17772159	6	16	5	632	71,131	FMR1	1,64	1,72	-0,08
High	Q43390-2	Isoform 2 of Heterogeneous nuclear ribonucleoprotein R [OS=Homo sapiens]	5,37476803	1,88679245	1	2	1	636	71,17	HNRNPR	0,23	1,59	-2,36
High	P04029	60S ribosomal protein L13a [OS=Homo sapiens]	22,4337069	16,7487685	4	10	4	203	25,562	RPL13A	1,43	1,54	-0,11
High	Q75676	Ribosomal protein S6 kinase alpha-4 [OS=Homo sapiens]	11,4550993	7,12435233	5	8	5	772	85,552	RPS6KA4	1,41	1,52	-0,11
High	P49368-1	T-complex protein 1 subunit gamma [OS=Homo sapiens]	58,945176	18,1651376	9	22	9	545	60,495	CTC3	0,71	1,11	-0,77
High	Q07020	60S ribosomal protein L18 [OS=Homo sapiens]	40,7939192	37,2340426	6	15	6	188	21,621	RPL18	1,3	1,49	-0,19
High	Q02878	60S ribosomal protein L6 [OS=Homo sapiens]	98,0284244	44,0972222	14	46	14	288	32,708	RPL16	1,26	1,48	-0,22
High	Q15149-1	plectin [OS=Homo sapiens]	7,7510813	0,5538113	2	6	2	4645	531,467	PLEC	1,54	1,47	-0,07
High	Q9NPV8	Complex I assembly factor 1 [IMMCDC1, mitochondrial [OS=Homo sapiens]	4,68169259	7,01754386	2	2	2	285	32,158	IMMCDC1	1,07	1,47	-0,4
High	P49339	Trifunctional enzyme subunit alpha1, mitochondrial [OS=Homo sapiens]	133,48389	33,9449541	22	60	22	763	82,947	HADHA	0,84	1,45	-0,61
High	Q00567	Nucleolar protein 56 [OS=Homo sapiens]	16,2026646	6,22889623	4	6	4	594	66,009	NOP56	2,21	1,45	0,76
High	P28288	ATP-binding cassette D member 3 [OS=Homo sapiens]	4,66265337	1,82094082	1	2	1	639	75,428	AEC03	1,42	1,43	-0,01
Medium	Q9UJU6	A kinase, anchor protein 8-like [OS=Homo sapiens]	4,22371101	3,25073399	2	2	2	646	71,604	AKAP8L	0,93	1,42	-0,48
High	P18074-1	Triflin basal transcription factor complex helicase subunit 1 [OS=Homo sapiens]	20,5031099	6,9738421	6	8	6	760	86,854	ERCC2	1,23	1,41	-0,18
High	P11940-1	Polyadenylate-binding protein 1 [OS=Homo sapiens]	158,424	36,3307547	21	60	21	636	70,626	PABPC1	1,47	1,4	0,06
Medium	Q5ET7A-2	ATP-dependent zinc metalloprotease YMEL1 [OS=Homo sapiens]	2,84924356	1,16429495	1	1	1	773	86,401	YMEL1	0,25	1,4	-1,15
High	Q12996-1	Cleavage stimulation factor subunit 3 [OS=Homo sapiens]	7,97949303	2,78949028	2	3	2	717	82,869	CSTF3	1,76	1,39	0,37
High	P49959-3	Isoform 3 of Double-strand break repair protein MRPL1A [OS=Homo sapiens]	18,4176423	7,59495671	4	7	4	7111	81,02	MRE11A	1,25	1,35	-0,09
High	Q96AY2-2	Iflin basal transcription factor complex helicase subunit 1 [OS=Homo sapiens]	33,5536683	14,0651801	8	18	8	583	60,738	EME1	1,41	1,32	0,0
High	P04930-1	Nucleolar RNA helicase 2 [OS=Homo sapiens]	25,125144	5,74712644	4	11	1	783	87,29	DDX21	1,25	1,21	0,47
High	Q9NRAO-5	Formin 5 of Spingosine kinase 2 [OS=Homo sapiens]	46,4680522	8,80420499	6	15	6	761	80,154	SPHK2	1,16	1,21	-0,05
High	Q9NZ01-1	Very-long-chain enoyl-CoA reductase [OS=Homo sapiens]	37,8873051	12,6623377	5	10	5	308	36,011	TECR	0,83	1,19	-0,36
High	P33992	DNA replication licensing factor mcm3 [OS=Homo sapiens]	63,5951497	22,479564	16	28	16	734	82,233	MCM5	1,09	1,18	-0,09
High	Q92499	ATP-dependent RNA helicase DDX1 [OS=Homo sapiens]	17,1424771	10,1351351	7	9	7	740	82,388	DDX1	1,02	1,18	-0,16
High	P14654	Insulin receptor substrate 1 [OS=Homo sapiens]	112,567386	20,1272872	24	61	24	1257	133,685	IRSA	1,07	1,17	-0,11
High	Q9H521	Probable ATP-dependent RNA helicase DHX35 [OS=Homo sapiens]	33,4422228	12,802276	9	17	9	703	78,86	DHX35	0,4	1,17	-0,77
Low	P5084	Trifunctional enzyme subunit beta, mitochondrial [OS=Homo sapiens]	1,56916365	1,89873418	1	1	1	474	51,262	HADHB	0,62	1,16	-0,54
High	P11142-1	Heat shock cognate 71 kDa protein [OS=Homo sapiens]	777,753406	57,8847368	44	363	33	646	70,854	HSP48	1,3	1,15	0,15

High	P49327	Fatty acid synthases [OS=Homo sapiens]	9,243;34184	1,393;86699	3	4	3	2514	273;254	FASN	0.56	1,14	-0.58	0
Low	Q9NNV4;2	Isomerase 2 of Hept(A)RNA polymerase, mitochondrial [OS=Homo sapiens]	2,175;0953	0.983;14607	1	1	1	712	78;784		0.73	1,12	-0.39	0
High	P07900;2	Isomerase 2 of Hept(A)RNA polymerase, 90-alpha [OS=Homo sapiens]	122;169519	1.8;6921881	16	53	6	854	98;0959	HSP90AA1	0.94	1,11	-0.18	0
High	Q14119	Vesicular endothelial zinc finger 1 [OS=Homo sapiens]	4,686;21047	5.5;6621881	2	2	2	521	56;895	VEZEF1	9,37	1,1	82.6	0
Medium	P18077	60S ribosomal protein L35a [OS=Homo sapiens]	2,689;2111	18;1818182	2	2	2	110	12;53	RPL35A	0.53	1,09	-0.56	0
High	P10809	60 kDa heat shock protein, mitochondrial [OS=Homo sapiens]	67;367;612	27;2251;309	14	24	14	573	61;016	HSPD1	1,13	1,08	0.05	0
High	P19447	TFIH basal transcription factor complex helicase XBP subunit [OS=Homo sapiens]	20;743;6448	11;381;07442	8	10	8	782	83;221	ERCC3	0.69	1,03	-0.38	0
High	P52272	Heterogeneous nuclear ribonucleoprotein M [OS=Homo sapiens]	270;482;835	48;3561;644	36	204	36	730	77;464	HNRNPM	0.98	1,03	-0.05	0
High	Q92841	Probable ATP-dependent RNA helicase DDX17 [OS=Homo sapiens]	104;32;0888	19;204;896	13	52	8	729	80;222	DDX17	0.99	1,03	-0.04	0
Medium	Q9JNK9	Protein angel homolog 1 [OS=Homo sapiens]	4,115;1454	2,089;5224	1	1	1	670	75;227	ANGEL1	10,88	1,02	9.86	0
High	P00425	Insulin-like growth factor 2 mRNA-binding protein 3 [OS=Homo sapiens]	8,608;2872	3,454;23143	2	4	1	579	63;666	IGF2BP3	0.77	1,01	-0.23	0
High	P08945	FACT complex subunit SSRP1 [OS=Homo sapiens]	5,584;60918	4,956;53032	3	3	3	709	81;024	SSRP1	0.95	1,01	-0.06	0
High	Q9HDG5	Junctophilin-1 [OS=Homo sapiens]	104;47;5071	27;080;1815	19	63	19	661	71;642	JPH1	0.98	1	-0.02	0
High	P05388	60S acidic ribosomal protein P0 [OS=Homo sapiens]	45;569;769	22;397;763	6	17	6	317	34;252	RPLP0	0.7	1	-0.29	0
High	Q9Y5Y2	Cytosolic Fe-S cluster assembly factor NUBP2 [OS=Homo sapiens]	7,955;0851	3,34;209;321	1	2	1	271	28;807	NUBP2	0.64	0.97	-0.33	0
High	P50402	Emerin [OS=Homo sapiens]	48;627;5794	24;409;4488	5	17	5	254	28;976	EMD	1,2	0.96	0.24	0
High	P17844	probable ATP-dependent RNA helicase DDX5 [OS=Homo sapiens]	75;008;1662	20;032;5733	14	37	9	614	69;105	DDX5	0.99	0.95	0.04	5
High	Q9NUJ7	Probable ATP-dependent RNA helicase DDX28 [OS=Homo sapiens]	65;821;2083	24;81;48148	12	20	12	540	59;545	DDX28	0.79	0.95	-0.16	0
High	Q9UNX3	60S ribosomal protein L26-like 1 [OS=Homo sapiens]	122;41;37931	11	130	3	145	17;246	RPL26L1	0.92	0.91	-0.02	0	
High	P62244	60S ribosomal protein L7a [OS=Homo sapiens]	131;71;1446	42;857;429	15	43	15	266	42;871	RPL7A	0.87	0.9	-0.03	0
High	Q9U63;2	Isomerase 2 of ATP-binding cassette sub-family F member 2 [OS=Homo sapiens]	61;40;1092	15;615;142	9	22	9	634	72;397	ABCF2	0.84	0.86	-0.02	0
High	P043143	Pre-mRNA-splicing factor ATP-dependent RNA helicase DHX15 [OS=Homo sapiens]	192;37;662	33;333;333	25	73	25	795	90;875	DHX15	0.62	0.85	-0.23	0
High	P56270;2	Isomerase 2 of Myc-associated zinc finger protein [OS=Homo sapiens]	20;83;8963	6;683;7197	3	9	3	493	51;04	MAZ	1,26	0.85	0.41	0
High	Q8WWY3	U1/U6 small nuclear ribonucleoprotein Prp31 [OS=Homo sapiens]	4,59;961018	8;807;6123	2	2	2	499	55;421	PRP31	0.45	0.85	-0.4	0
High	Q09161	Nuclear cap-binding protein subunit 1 [OS=Homo sapiens]	21;14;4027	7;594;9671	6	9	6	790	91;781	NCBP1	0.91	0.84	0.08	0
Medium	Q9Y619	TAF6-like RNA polymerase II 1300/CBP-associated factor 65 k	2,335;56403	17;684;875	1	1	1	622	67;772	TAF6L	1,98	0.84	1.14	0
High	Q9NZ18	Insulin-like growth factor 2 mRNA-binding protein 1 [OS=Homo sapiens]	22;21;2904	9;87;868284	5	9	4	577	63;441	IGF2BP1	0.56	0.82	-0.26	1
High	Q9GUJ5;1	E2 ubiquitin-protein ligase ZFP91 [OS=Homo sapiens]	134;20;752	29;649;128	16	60	16	570	63;406	ZFP91	1,04	0.81	0.24	0
High	Q9H614;1	Receptor expression-enhancing protein 4 [OS=Homo sapiens]	89;85;4637	45;52;5918	11	34	11	257	29;376	REEP4	0.99	0.81	0.18	0
High	P38646	Serine-70 protein, mitochondrial [OS=Homo sapiens]	290;05;909	43;446;245	32	131	31	679	73;635	HSPA9	0.78	0.8	-0.02	0
High	O00139;2	Isomerase 2 of kinase-like protein KIF2A [OS=Homo sapiens]	98;06;8054	27;722;7277	19	44	19	660	74;996	KIF2A	0.64	0.8	-0.16	0
High	P7812	CTP synthase [OS=Homo sapiens]	43;32;49301	15;95;45453	9	19	9	591	66;548	CTPS1	0.47	0.8	-0.34	0
High	P38159;1	RNA-binding motif protein, X chromosome [OS=Homo sapiens]	15;82;4287	3;37;480818	1	3	1	391	42;306	RBMX	0.6	0.8	-0.2	0
High	Q96134;1	Mab/microtubule-associated protein ligase ZFP91 [OS=Homo sapiens]	133;45;6021	37;769;574	24	64	22	752	82;468	MARK4	0.84	0.79	0.04	0
High	P53007	Tricarboxylate transport protein, mitochondrial [OS=Homo sapiens]	124;15;0655	41;80;6431	12	64	12	311	33;991	SIC25A1	0.85	0.79	0.06	0
High	Q16629	serine/arginine-rich splicing factor 7 [OS=Homo sapiens]	32;50;19365	27;73;0924	5	10	5	238	27;35	SCF7	1.91	1.12	0	0
Low	Q15392	Delta(24)-sterol reductase [OS=Homo sapiens]	1,175;8856	1;744;18605	1	1	1	516	60;062	DHCR24	0.16	0.78	-0.63	0
High	Q67622	Dolichol phosphate mannosyltransferase subunit 1 [OS=Homo sapiens]	122;43;6018	65;769;308	14	45	14	260	29;616	DPM1	2,67	0.77	1.9	0
High	P62987	Ubiquitin-E63 ribosomal protein L40 [OS=Homo sapiens]	49;13;57029	37;5	4	48	1	128	14;719	UBA52	0.55	0.77	-0.22	0
High	P54886	delta-1-pyrroline-5-carboxylate synthase [OS=Homo sapiens]	11;73;9122	5;40;80503	4	5	4	795	18;248	ALDH18A1	1,62	0.77	0.85	0
High	P04350	Tubulin beta-1A chain [OS=Homo sapiens]	689;96;9925	75;90;09099	28	430	2	444	49;554	TUBB4A	0.73	0.75	-0.02	0
High	Q9BQ33	Tubulin alpha-1C chain [OS=Homo sapiens]	544;74;5632	63;91;98218	29	269	4	449	49;863	TUBA1C	0.27	0.74	-0.47	0
High	P13010	X-ray repair cross-complementing protein 5 [OS=Homo sapiens]	76;29;65189	20;90;16393	15	27	15	732	82;652	XRC5	0.54	0.74	-0.2	0
High	P49593	Protein phosphatase 1F [OS=Homo sapiens]	16;37;17181	2;64;31;887	1	1	1	454	9;48	PPM1F	0.55	0.74	-0.19	0
High	Q05823;2	2-SF-dependent ribonuclease [OS=Homo sapiens]	9,05;87;1193	3;91;362;023	3	4	3	741	83;481	RNASE1	0.79	0.74	0.05	0
High	Q49A26;1	Putative oxidoreductase GLYR1 [OS=Homo sapiens]	23;11;8431	14;285;7143	6	11	6	553	60;518	GLYR1	1.09	0.73	0.37	0
High	P39023	60S ribosomal protein L3 [OS=Homo sapiens]	48;55;2685	20;09;9256	9	23	9	403	46;08	RPL3	0.48	0.72	-0.24	0
High	P19338	Nucleolin [OS=Homo sapiens]	46;02;7625	13;94;3662	10	20	10	710	76;568	NCL	0.2	0.72	-0.52	0
Low	Q9Y619	mitochondrial ornithine transporter 1 [OS=Homo sapiens]	1,27;31;096	2;25;55814	1	1	1	301	32;715	SLC25A15	0.66	0.71	-0.06	0
High	P53621;2	Coatomer subunit alpha [OS=Homo sapiens]	26;43;2456	5;7;5831;36	7	11	7	1233	139;23	COPA	-0.28	0.69	-0.97	0
High	P04843	Dolichyl-diphospholigosaccharide-protein glycosyltransferase subunit 1 [OS=Homo sapiens]	121;01;5786	4;28;336079	3	4	3	607	68;527	RPN1	0.83	0.69	0.14	0
High	P39023	Protein SDA1 homolog [OS=Homo sapiens]	74;70;41691	14;70;16012	11	38	11	687	79;82	SDAD1	0.53	0.68	-0.15	0
High	P19338	Metal-response element-binding transcription factor 2 [OS=Homo sapiens]	25;47;21072	15;682;968	7	11	7	593	67;063	MTT2	0.2	0.72	-0.52	0
High	Q9Y619;4	Spingomyelin phosphodiesterase 4 [OS=Homo sapiens]	6;70;64;1199	3;20;73;31;04	3	3	3	873	98;086	HST1HIE	0.8	0.67	0.12	15
High	P104112	Histone H4 [OS=Homo sapiens]	173;56;553	39;72;60274	13	174	3	219	21;852	CEP27	0.54	0.67	-0.13	6
Medium	P26767	40S ribosomal protein S27 [OS=Homo sapiens]	121;01;5786	4;28;571429	6	46	2	84	9;455	RPS27	0.54	0.67	0.54	0
High	Q57545	Splitting factor 45 [OS=Homo sapiens]	4;15;29335	4;23;94;405	2	2	1	401	44;934	RBM17	0.21	0.67	0.54	0
High	Q75746	Calcium-binding mitochondrial carrier protein Atrial 1 [OS=Homo sapiens]	71;91;428	17;984;1003	11	28	6	678	74;715	SLC25A12	0.86	0.66	0.2	0
High	Q6884	Dihydrofolate reductase [OS=Homo sapiens]	13;90;66;736	5;0;97;08738	2	6	2	412	45;717	DNAJA2	0.63	0.66	-0.03	0
Low	Q51TW2;2	Centrosomal protein of 78 kDa [OS=Homo sapiens]	1,753;4769	0;83;102;493	1	1	1	722	80;023	CEP78	0.99	0.66	0.33	0
High	Q9eA33;1	Collel-coil domain-containing protein 47 [OS=Homo sapiens]	28;18;1;8193	12;62;9596	6	12	6	483	55;838	CCDC47	0.54	0.65	-0.11	0

High	Q92878-2	Isoform 2 of DNA repair protein RAD50 [OS=Homo sapiens]	19,821,017,1	5,007,587,25	6	8	6	1318	154,491,1	0,5	0,65	-0,15	0	
High	Q75419-3	Isoform 3 of Cell division control protein 45 homolog [OS=Homo sapiens]	9,677,776,26	6,02,006,689,0	3	4	3	598	68,724	CDC45	0,81	0,65	0,15	0
High	Q966CM3	RNA pseudouridyl synthase domain-containing protein 4 [OS=Homo sapiens]	7,426,515,72	6,89,006,689,0	3	4	3	377	42,178	RPSU4	0,4	0,65	-0,25	0
High	Q07955-1	Serine/arginine-rich splicing factor 1 [OS=Homo sapiens]	12,737,206,1	20,96,774,19	4	6	4	248	27,728	SRSE1	0,32	0,64	-0,31	0
High	Q9Y678	Catomer subunit gamma-1 [OS=Homo sapiens]	8,125,212,7	2,288,32,952	2	3	2	874	97,655	COPG1	0,54	0,63	-0,1	0
High	Q95349-1	Structural maintenance of chromosomes protein 2 [OS=Homo sapiens]	6,182,405,636	2,339,181,29	3	3	3	1197	135,572	SMC2	0,1	0,63	-0,53	0
High	Q8WKL1	Mitochondrial Rhod GTPase 2 [OS=Homo sapiens]	15,725,556,3	7,285,515,4	4	8	4	618	68,075	RHOT2	0,79	0,62	0,18	0
High	Q8UF8	Bifunctional lysine-specific demethylase and histidine-hydroxylase MINA [OS=Hg]	8,564,312,304	2,365,591,4	2	3	2	465	52,767	MINA	1,17	0,62	0,54	0
High	P33993-1	DNA replication licensing factor MCM7 [OS=Homo sapiens]	336,658,735	58,831,710,7	37	137	37	719	81,257	MCM7	0,54	0,61	-0,06	0
High	P22695	Cytochrome b-c1 complex subunit 2, mitochondrial [OS=Homo sapiens]	53,875,017,8	19,426,044,86	9	19	9	453	48,413	UQCRC2	0,69	0,6	0,09	0
High	Q9EY1-1	DnaJ homolog subfamily A member 3, mitochondrial [OS=Homo sapiens]	34,225,762,9	21,875,1	8	12	8	480	52,456	DNAJA3	1,02	0,6	0,42	0
High	Q75306-1	NADH dehydrogenase (ubiquinone) iron-sulfur protein 2, mitochondrial [OS=Hg]	4,855,984,89	1,727,851,77	1	2	1	463	52,512	NDUF52	0,14	0,59	-0,05	0
High	P16403	Histone H1.2 [OS=Homo sapiens]	174,045,804	37,558,854	12	180	2	213	21,352	HIST1H1C	0,55	0,58	-0,03	0
High	Q9UBM7	7-dehydrocholesterol reductase [OS=Homo sapiens]	24,805,726,5	9,263,157,89	5	10	5	475	50,454	DHCR7	0,44	0,58	0,15	0
High	Q96544	TF55-regulating kinase [OS=Homo sapiens]	6,244,905,7	10,67,139,68	2	3	2	253	28,143	TPB3RK	0,23	0,58	-0,34	0
High	Q9BZT-1	Regulator of nonsense transcripts 3B [OS=Homo sapiens]	149,31,457,1	6,58,385,09	22	74	22	483	57,723	UPF3B	0,58	0,57	0,02	0
High	Q9NT3-1	Structural maintenance of chromosomes protein 4 [OS=Homo sapiens]	27,675,684,6	6,832,298,14	9	13	9	1288	147,091	SMC4	0,64	0,57	0,07	0
Medium	Q9UH62	Armadillo repeat-containing X-linked protein 3 [OS=Homo sapiens]	3,824,07,297	4,749,340,937	2	2	2	379	42,474	ARMCK3	0,63	0,56	0,08	0
High	O0571	ATP-dependent RNA helicase DDX3X [OS=Homo sapiens]	422,365,694	59,667,657,37	40	191	40	662	73,198	DDX3X	0,38	0,55	-0,17	0
High	P04911	Elongation factor 1, mitochondrial [OS=Homo sapiens]	317,45,722	47,12,399,38	19	90	19	453	49,51	TUFM	0,44	0,55	-0,12	0
High	Q13724-1	Mannosyl-oligosaccharide glucosidase [OS=Homo sapiens]	99,086,285	23,297,491	16	44	16	837	91,861	MOGS	0,64	0,55	0,09	0
High	P62906	60S ribosomal protein L10A [OS=Homo sapiens]	48,15,553	44,239,53,13	9	20	9	217	24,816	RPL10A	0,79	0,55	C24	0
High	P51116	Fragile X mental retardation syndrome-related protein 2 [OS=Homo sapiens]	14,906,010,1	4,98,341,753	3	8	2	673	74,178	FXR2	0,43	0,55	-0,11	0
Low	Q9UH58	Cyclin-L1 [OS=Homo sapiens]	1,53,466,441	1,14,066,226,3	1	1	1	526	59,597	CCNL1	-0,15	0,55	-0,7	0
High	P17987	T-complex protein 1 subunit alpha [OS=Homo sapiens]	193,045,162,6	45,503,971	23	70	23	556	60,306	TCP1	0,63	0,54	0,09	0
High	Q9NW7-2	Isoform 2 of ATPase family AAA domain-containing protein 3A [OS=Homo sapiens]	43,476,755,2	21,84,300,34	15	23	15	586	66,177	ATAD3A	0,46	0,54	-0,08	0
High	Q8N2M8	CK4-associated serine/arginine rich protein [OS=Homo sapiens]	46,561,420,3	13,946,875	8	23	8	674	77,115	CLASRP	0,28	0,53	-0,25	0
High	Q9Y4R8	Telomere length regulation protein TE12 homolog [OS=Homo sapiens]	5,66,596,045	3,2,258,012,8	3	3	3	837	91,689	TEL02	-0,04	0,52	-0,56	0
High	Q14880	Microsomal glutathione reductase 2-transferrase 3 [OS=Homo sapiens]	21,826,057	8,52,263,158	1	5	1	152	16,506	MEST3	1,01	0,51	0,5	0
High	Q14739	Lamin-B receptor [OS=Homo sapiens]	20,822,942,6	7,64,22,762	4	5	4	615	70,658	LBR	0,34	0,51	-0,18	0
Medium	P63167	Dynein light chain 1, cytoplasmic [OS=Homo sapiens]	3,57,013,233	13,483,1,461	1	1	1	89	10,359	DYNLL1	-9,93	0,51	-0,44	0
Low	Q8XK8-1	DNA helicase family AAA domain-containing protein 3A [OS=Homo sapiens]	1,847,871,166	1,26,105,405	1	1	1	793	91,021	DNAH10	0,48	0,5	-0,02	0
High	P78527	DNA-dependent protein kinase catalytic subunit [OS=Homo sapiens]	338,17,202,21	20,37,306,2	77	214	77	4128	468,788	PRKDC	0,47	0,48	0	0
High	Q9BQ7-0	Transcription factor 25 [OS=Homo sapiens]	4,889,581,115	3,63,922,485	2	2	2	676	76,619	TCF25	1,29	0,48	0,81	0
Medium	Q15070-1	Mitochondrial inner membrane protein OXA1L [OS=Homo sapiens]	3,509,165,38	2,29,883,057	1	2	1	438	48,516	OXA1L	0,16	0,48	-0,32	0
High	P05141	Alp/Alp translocase [OS=Homo sapiens]	59,15,146,8	1,07,38,255	26	352	8	298	31,831	SIC25A5	0,47	0,47	0,07	38
High	Q9Y3Z3	Deoxyynucleotide triphosphate triphosphohydrolase 5/AMHD1 [OS=Homo sapiens]	54,35,920,49	16,61,34,185	10	20	10	26	72,155	SANMD1	0,43	0,47	-0,03	0
High	P25490	Transcriptional repressor protein YY1 [OS=Homo sapiens]	28,382,912,3	8,2,12,560,39	5	15	5	414	44,685	YY1	0,27	0,47	-0,2	0
High	P12956	X-ray repair cross-complementing protein 6 [OS=Homo sapiens]	16,20,502,3	10,01,64,204	5	7	5	609	69,799	XRC6	0,36	0,47	-0,11	0
Low	Q7Z736	Pleckstrin homology domain-containing family H member 3 [OS=Homo sapiens]	2,25,44,148	1,1,34,9,604	1	1	1	793	83,264	PLEKH3	-0,45	0,47	-0,32	0
High	P52597	Heterogeneous nuclear ribonucleoprotein F [OS=Homo sapiens]	14,33,31,48	7,95,18,073	2	4	1	415	45,46,73	HNRNPF	0,66	0,46	0,46	0
Medium	Q95816	BAF family molecular chaperone regulator 2 [OS=Homo sapiens]	3,34,996,248	2,1,32,70,14	1	1	1	211	23,757	BAG2	11,8	0,46	11,34	0
High	P25495-1	SPB30-binding protein [OS=Homo sapiens]	12,90,905,2	9,09,90,909	3	6	3	308	33,85	SAP30BP	-0,26	0,45	0,72	0
High	Q9Y2X3	Nucleolar protein 58 [OS=Homo sapiens]	57,18,561,3	10,95,2,74,1	10	21	10	529	59,541	NOP58	0,77	0,44	0,83	0
Medium	Q96CM1	Arp-2 complex subunit mu [OS=Homo sapiens]	3,67,78,299,21	2,3,67,81,609	2	3	2	435	49,62,3	AP2M1	0,46	0,44	0,02	0
Medium	Q99805	Transmembrane 9 superfamily member 2 [OS=Homo sapiens]	3,38,33,39,69	1,2,06,63,35	1	2	1	663	75,725	TM9SF2	0,12	0,44	-0,32	0
High	Q9UKV8	Protein argonaute-2 [OS=Homo sapiens]	384,36,47,34	51,57,15,949	40	198	40	859	97,146	AGO2; EIF2C2	0,29	0,42	-0,13	0
High	Q94W2	Ribosomal biogenesis protein LAS1L [OS=Homo sapiens]	67,90,71,73	34,05,99,95	16	31	16	734	83,013	WHSC1	0,54	0,41	0,13	0
High	Q9NWQ4	G patch domain-containing protein 2-like [OS=Homo sapiens]	40,01,04,93	15,14,52,382	7	13	7	482	54,226	GPATCH2L	0,83	0,42	C41	0,41
High	Q5SS5-1	Heterochromatin protein 1-binding protein 3 [OS=Homo sapiens]	35,88,048,19	16,27,486,44	9	15	9	533	61,169	HPLBP3	0,72	0,42	0,3	0
High	Q9NWK9	Box C/D snoRNA protein 1 [OS=Homo sapiens]	6,65,61,18,18	4,6,60,85,106	2	4	2	470	53,884	ZNHIT6	0,67	0,42	C25	0
Medium	Q60934	Nibrin [OS=Homo sapiens]	4,42,15,82,69	1,06,100,79,6	1	2	1	754	84,906	NBN	0,85	0,42	0,43	0
High	Q96028-1	Histone-lysine N-methyltransferase NSD2 [OS=Homo sapiens]	6,01,95,23,93	6,65,93,9,66	1	3	1	1365	15,2,16	WHSC1	0,54	0,41	0,13	0
Low	Q14592	Zinc finger protein 460 [OS=Homo sapiens]	1,3,60,15,26	1,4,2,34,8,754	1	1	1	562	63,624	ZNF460	0,2	0,41	-0,22	0
High	Q16891	MC1C5 complex subunit Mc160 [OS=Homo sapiens]	34,90,95,55	10,68,60,158	8	16	8	758	83,626	IMMT	0,49	0,4	0,09	0
Medium	P78371-1	T-complex protein 1 subunit mu [OS=Homo sapiens]	2,57,94,914	2,24,29,906	1	1	1	535	57,452	CT12	0,36	0,4	-0,05	0
High	Q8V1F7-1	Nuclear pore complex protein Nup93 [OS=Homo sapiens]	17,66,84,74	4,1,26,94,13	31	89	31	819	93,43	NUP93	0,5	0,39	0,11	0
High	P50911	T-complex protein 1 subunit delta [OS=Homo sapiens]	31,53,44,44	12,05,93,692	6	13	6	539	57,888	CT14	0,95	0,39	0,56	0
Low	Q9P0J6	35S ribosomal protein 136, mitochondrial [OS=Homo sapiens]	1,38,57,3571	5,8,52,92,72	1	2	1	103	11,777	MRP136	0,24	0,39	-0,16	0
High	Q08211	Atp-dependent rna helicase a [OS=Homo sapiens]	13,12,70,505	4,1,17,32,835	5	7	5	1270	140,86,9	DHX9	0,3	0,38	-0,08	0
High	Q96EV4	Translation machinery-associated protein 16 [OS=Homo sapiens]	139,80,057,6	46,79,80,26	13	73	13	203	23,849	TMA16	0,63	0,37	0,27	0

Low	Q14257-2	Isoform 2 of Reticulocalbin-2 [OS=Homo sapiens]	1,72955409	2,3880557	1	1	1	335	39,115	RCN2	-0.3	0.37	-0.67	0	
Medium	Q9H6F5	Coiled-coil domain-containing protein 86 [OS=Homo sapiens]	3,95130966	5,2777778	2	2	2	360	40,211	CCDC86	0.62	0.36	0.26	0	
Low	Q05BV3-5	Isoform 5 of Echinoderm microtubule-associated protein-like 5 [OS=Homo sapiens]	1,2037269	0,40465668	1	1	1	1977	220,126	EMIL5	-0.16	0.36	-0.52	0	
High	Q9UJ50-2	Isoform 2 of Calcium-binding mitochondrial carrier protein Atrial natriuretic peptide 1 [OS=Homo sapiens]	1,65145568	20	41,221,893	20	65	15	676	74,257	SLC25A13	0.27	0.35	-0.08	0
High	Q96Z51-1	DNA-directed RNA polymerase I subunit RPA49 [OS=Homo sapiens]	15,8356123	6,02910603	3	7	3	481	53,928	POLR1E	0.69	0.35	0.34	0	
High	Q14949	Cytochrome b-c1 complex subunit 8 [OS=Homo sapiens]	6,6453899	15,8538585	1	2	1	82	9,9	UQCRCQ	0.63	0.35	0.27	0	
High	P27448-5	MAP/microtubule affinity-regulating kinase 3 [OS=Homo sapiens]	304,627752	55,7768924	36	128	2	753	84,436	MARK3	0.42	0.34	0.08	29	
High	Q71354	Zinc finger protein 771 [OS=Homo sapiens]	157,639962	50,157787	20	65	20	317	35,679	ZNF771	0.26	0.34	-0.08	0	
High	O43660-1	Phleiotropic regulator 1 [OS=Homo sapiens]	20,3624524	19,0661479	9	10	9	514	57,158	PLRG1	0.32	0.34	-0.02	0	
High	Q8TCI2	Dolichyl-diphospholigosaccharide-protein glycosyltransferase subunit STT3B	8,44665769	5,08474576	3	3	3	826	93,614	STT3B	0.27	0.34	-0.07	0	
High	Q75533-1	Splicing factor 3B subunit 1 [OS=Homo sapiens]	43,2838375	10,429479	11	18	11	1304	145,738	SF3B1	-0.14	0.33	-0.47	0	
High	Q9HOU3	Magnesium transporter protein 1 [OS=Homo sapiens]	7,57155398	8,05970149	3	3	3	359	38,011	MAGT1	0.37	0.33	0.04	0	
High	Q13247	Serine/arginine-rich splicing factor 6 [OS=Homo sapiens]	11,6631668	6,68604651	3	4	3	344	39,563	SRSF6	0.24	0.32	-0.08	0	
High	Q9HZ9	Zinc finger protein Ets [OS=Homo sapiens]	21,0041293	14,8717949	7	8	7	585	64,065	IKZF4	0.78	0.31	0.47	0	
Medium	Q75530-2	Isoform 2 of Polycomb protein EED [OS=Homo sapiens]	3,15249698	5,79389914	2	2	2	466	53,027	EED	0.58	0.31	0.28	0	
High	Q9UHX1-2	Isoform 2 of Poly(U)-binding-splicing factor PUF60 [OS=Homo sapiens]	311,059489	68,0811808	26	136	26	542	58,135	PUF60	0.43	0.29	0.14	0	
High	Q9NSD4	Zinc finger protein 275 [OS=Homo sapiens]	16,7628187	10,955711	5	9	5	429	48,412	ZNF275	0.6	0.29	0.31	0	
High	Q9Y5Q9-1	YTH domain-containing family protein 2 [OS=Homo sapiens]	7,82461158	5,87219344	3	4	3	579	62,296	YTHDF2	0.35	0.29	0.07	0	
High	Q14680	Maternal embryonic leucine zipper kinase [OS=Homo sapiens]	11,2073142	5,98078341	4	5	3	651	74,594	MELK	0.48	0.28	0.21	1	
High	Q46411	Segment polarity dishevelled-related gene DV12 [OS=Homo sapiens]	5,24663503	16,0442447	1	2	1	736	78,899	DV12	0.04	0.27	-0.22	0	
High	Q6UN15-3	Isoform 3 of Pre-mRNA 3'-end-processing factor Fhl1 [OS=Homo sapiens]	532,4251759	54,8076923	31	417	1	520	58,341	FIP1L1	0.4	0.26	0.15	0	
High	P61513	60S ribosomal protein L37a [OS=Homo sapiens]	86,18308	52,173913	6	44	6	92	10,268	RPL37A	0.14	0.26	-0.12	0	
High	Q9H936	Mitochondrial glutamate carrier 1 [OS=Homo sapiens]	4,92860748	4,98356037	2	2	2	323	34,448	SLC25A22	-0.13	0.26	-0.38	0	
High	P9N2X0	30S ribosomal protein S21 [OS=Homo sapiens]	8,25565381	8,36565386	2	2	2	251	28,431	MRPL16	0.22	0.25	-0.03	0	
Medium	P42695	condensin 2 complex subunit D3 [OS=Homo sapiens]	4,64634636	1,8024032	2	2	2	1498	168,783	NCAPD3	-0.26	0.25	-0.52	0	
High	Q2TA8-1	PWWP domain-containing protein NuM1 [OS=Homo sapiens]	38,2265844	14,0845457	10	16	10	710	78,587	MUM1	0.59	0.24	0.35	0	
High	Q9YT9	Nucleolar complex protein 2 homolog [OS=Homo sapiens]	10,8241446	2,67022697	2	3	2	749	84,866	NOC2L	0.72	0.24	0.48	0	
High	P62805	histone H4 [OS=Homo sapiens]	127,070315	52,42717845	7	44	7	103	11,36	ST114H/HST1	0.22	0.23	-0.01	0	
High	Q14545	TRAF4-type zinc finger domain-containing protein 1 [OS=Homo sapiens]	5,649640913	3,26460481	2	2	2	582	64,8	TRAID1	0.5	0.23	0.27	0	
High	P07814	Bifunctional glutamate/proline-tRNA ligase [OS=Homo sapiens]	4,96421477	0,9592351	1	3	1	1512	170,483	EPRS	-0.16	0.23	-0.39	0	
High	Q9BX5-2	Isoform 2 of AP-1 complex subunit mu-1 [OS=Homo sapiens]	4,71980771	1,6091954	1	3	1	435	49,809	AP1M1	0.01	0.23	-0.22	0	
High	P08238	HeLa shock protein HSP-90-beta [OS=Homo sapiens]	155,490121	33,010497	22	65	11	724	83,212	HSP90AA1	-0.06	0.22	-0.27	12	
High	P067233-1	alpha-enolase [OS=Homo sapiens]	40,6701762	14,0552995	7	16	7	344	47,139	ENO1	-0.16	0.22	-0.37	0	
High	P68431	Histone H3.1 [OS=Homo sapiens]	15,5323695	16,9117647	3	16	3	136	15,394	ST113H/HST1	0.15	0.22	-0.07	0	
High	P15531-2	Isoform 2 of Nucleoside diiphosphate kinase A [OS=Homo sapiens]	4,82131076	6,77966102	1	1	1	1777	19,641	NME1	0.37	0.22	0.15	0	
High	Q9RHZ2	Probable ATP-dependent RNA helicase DDX3 [OS=Homo sapiens]	3,40775558	2	2	2	851	94,029	DDX31	0.32	0.22	0.1	0		
High	Q9URK6	Cleavage and polyadenylation specificity factor subunit 3 [OS=Homo sapiens]	53,7017994	22,8070175	13	30	13	68	77,436	CPSF3	0.43	0.21	0.22	0	
High	P22087	tRNA 2'-O-methyltransferase fibrillarin [OS=Homo sapiens]	5,82600955	4,36137072	2	2	2	321	33,763	FBL	0.8	0.21	0.6	0	
Low	Q99728	BRCA1-associated RING domain protein 1 [OS=Homo sapiens]	1,47404866	1,54440154	1	1	1	777	86,593	BARD1	0.19	0.21	-0.03	0	
High	P62273-1	40S ribosomal protein S20 [OS=Homo sapiens]	12,8553868	26,7857143	2	25	2	56	6,672	RPS29	0.1	0.2	-0.1	0	
High	Q8AN57	Solute carrier family 35 member 6 [OS=Homo sapiens]	5,05090759	5,1212938	2	2	2	371	40,188	RCC2	0.29	0.19	0.51	0	
High	P62913	60S ribosomal protein L11 [OS=Homo sapiens]	177,663272	66,8339326	13	189	13	178	80,24	RPL11	-0.06	0.19	-0.25	0	
High	P32969	60S ribosomal protein L9 [OS=Homo sapiens]	114,201463	49,479167	6	29	6	192	21,85	RPL9	0.01	0.19	-0.18	0	
High	P62861	40S ribosomal protein S30 [OS=Homo sapiens]	66,5719454	33,8897051	6	59	6	59	6,644	FAU	0.1	0.19	-0.09	0	
High	Q15235	28S ribosomal protein S12, mitochondrial [OS=Homo sapiens]	63,7979486	44,8253968	7	26	5	138	15,163	MRPS12	0.13	0.19	-0.06	0	
High	Q9P258	Protein RCC2 [OS=Homo sapiens]	21,5626035	17,4329502	8	11	8	522	56,049	RCC2	0.29	0.19	0.09	0	
Medium	Q9H6F2	Sentrin-specific protease 2 [OS=Homo sapiens]	3,70961871	52,5466383	3	15	3	86	86	SEN1P2	0.22	0.18	0.04	0	
High	P00549-1	Pescadillo homolog [OS=Homo sapiens]	229,91752	45,7482993	17	124	17	361	39,933	SLC25A3	0.15	0.17	-0.02	0	
High	P62277	40S ribosomal protein S13 [OS=Homo sapiens]	227,086541	51,6556291	13	155	13	151	10,602	GTPBP2	0.14	0.17	-0.04	0	
High	Q8N257	Histone H2B type 3-B [OS=Homo sapiens]	115,02113	46,8253968	7	64	2	126	20,79	TRIM3	0.79	0.17	0.62	0	
High	P04637	Cellular tumor antigen p53 [OS=Homo sapiens]	36,988621	25,9541985	10	15	10	393	43,625	TIF53	-0.26	0.18	-0.43	0	
High	P49458	Signal recognition particle 9 kDa protein [OS=Homo sapiens]	33,2719809	32,5583395	3	15	3	86	10,105	SRP9/SRP91	0.22	0.18	0.04	0	
High	Q00325-2	Isoform B of Phosphate carrier protein, mitochondrial [OS=Homo sapiens]	300,738215	45,7063712	17	124	17	361	39,933	SLC25A3	0.15	0.17	-0.02	0	
High	Q9BX10	GTP-binding protein 2 [OS=Homo sapiens]	39,579444	18,1063123	10	13	10	602	65,727	GTPBP2	0.14	0.17	-0.04	0	
High	Q75328-1	Tripartite motif-containing protein 3 [OS=Homo sapiens]	23,098127	14,6505376	8	9	8	744	80,779	TRIM3	0.79	0.17	0.62	0	
High	Q9H5V9-1	UnRpd428 protein Corf56 [OS=Homo sapiens]	95,947348	43,2432432	9	35	9	222	25,608	CXorf56	0.28	0.16	0.12	0	
High	P14678-3	Isoform S1-B of small nuclear ribonucleoprotein-associated proteins D and B	5,70530729	7,6124567	3	4	3	289	30,012	-0.07	0.16	-0.23	0		
Medium	Q00743-3	Isoform 3 of Serine/threonine-protein phosphatase catalytic subunit [OS=Hor	4,550382	5,84795322	2	2	2	342	38,921	PPBP6C	0.25	0.16	0.09	0	
High	Q9BGN5-1	Polypeptidyl-prolyl isomerase 2 [OS=Homo sapiens]	243,220267	49,2281304	30	109	30	583	66,164	PARP2	-0.03	0.15	-0.17	0	
High	P32322-3	Isoform 3 of Pyrrolidine-5-carboxylate reductase 1, mitochondrial [OS=Homo sapiens]	35,3052422	12,4277457	4	11	3	346	35,958	PYCR1	0.02	0.15	-0.13	1	
High	P13639	Elongation factor 2 [OS=Homo sapiens]	6,61817663	4,07925408	3	3	3	838	95,277	EEF2	0.14	0.15	-0.01	0	

High	Q9UET6	Putative tRNA cytidine(32)/guanosine(34)-2'-O-methyltransferase [OS=Homo sapiens]	5.9344437	8.20668693	2	2	329	36.056	FTS11	-1.43	0.15	-1.58	0	
High	P18621-3	Isoform 3 of 60S ribosomal protein L17 [OS=Homo sapiens]	641.552293	56.5788474	22	898	21	228	26.356	PL17-C18QR3	0.02	0.14	-0.13	2
High	P62895-2	Isoform 2 of 60S ribosomal protein L31 [OS=Homo sapiens]	103.921716	40.3688092	9	64	9	128	14.623	RPI31	0.08	0.13	-0.07	0
High	Q9H8M2-5	Bromodomain-containing protein 9 [OS=Homo sapiens]	109.618858	24.74353897	17	62	17	597	66.958	BRD9	-0.12	0.13	-0.24	0
High	P17838-1	ATP-dependent 6-phosphofructokinase, liver-type [OS=Homo sapiens]	54.5860376	24.74353897	16	26	12	780	84.964	PKL	-0.46	0.13	-0.59	3
High	Q9NG29	Plastid RNA-binding protein 9 [OS=Homo sapiens]	427.2276402	49.8652291	30	313	21	371	43.701	JUC7L	0.13	0.12	0	0
High	Q9BXF6	Rab11 family-interacting protein 5 [OS=Homo sapiens]	254.053484	53.1395568	28	105	28	653	70.372	RAB11FIP5	0.29	0.11	0.18	0
High	Q8IWA0	WD repeat-containing protein 75 [OS=Homo sapiens]	9.85332563	5.54216867	4	4	4	830	94.438	WDR75	0.3	0.11	0.19	0
Medium	O92769	Histone deacetylase 2 [OS=Homo sapiens]	3.30636977	2.45901639	1	2	1	488	55.329	HDAC2	0.43	0.11	0.32	0
High	Q16630-2	Isoform 2 of Cleavage and polyadenylation specificity factor subunit 6 [OS=Hon	241.75803	32.1428571	20	84	20	588	63.432	CPSF6	0.07	0.1	-0.03	0
High	Q8WWN0	Dimethyladenosine transferase 1, mitochondrial [OS=Homo sapiens]	172.144392	76.5895954	21	79	21	346	39.518	TFB1M	-0.1	0.1	-0.2	0
Low	Q8UKW9-1	RNA-binding protein Raly [OS=Homo sapiens]	1.3621018	2.9411747	1	1	1	306	32.444	RALY	-0.11	0.1	-0.22	0
High	Q96K58-2	Isoform 2 of Zinc finger protein 668 [OS=Homo sapiens]	26.97673819	17.2897196	8	14	8	642	70.46	0.07	0.09	-0.01	0	
High	Q9NVW8	Guanine nucleotide-binding protein-like 3-like protein [OS=Homo sapiens]	12.0623254	4.63917526	3	14	2	582	65.532	GNL3L	0.17	0.09	0.07	0
High	P27766	60S ribosomal protein L35 [OS=Homo sapiens]	285.227524	49.5936959	13	201	13	123	14.543	RPL35	0.03	0.08	-0.05	0
High	P27708	CDP protein [OS=Homo sapiens]	119.613577	15.0561798	28	54	28	225	242.829	CAD	-0.11	0.08	-0.19	0
Medium	Q9B067	Glutamine-rich WD repeat-containing protein 1 [OS=Homo sapiens]	4.41920567	1.79372197	1	2	1	446	49.388	GRWD1	-0.07	0.08	-0.15	0
High	Q9XKF1	Testis-expressed sequence 10 protein [OS=Homo sapiens]	229.625453	38.2131324	30	93	30	929	105.608	TEX10	0.3	0.07	0.23	0
High	P61927	60S ribosomal protein L37 [OS=Homo sapiens]	21.3795605	20.261856	6	49	6	97	11.071	RPL37	0.08	0.07	0.01	0
High	P62995	Transformer 2 protein homolog beta [OS=Homo sapiens]	27.4201129	17.7082333	5	13	5	288	33.646	TRA2B	0.07	0.06	0.06	0
High	Q5T310-3	Isoform 3 of G patch domain-containing protein 4 [OS=Homo sapiens]	24.1962883	7.7605215	4	10	4	451	50.897	GPATCH4	0.61	0.06	0.55	0
High	P365242-1	ATP synthase subunit gamma, mitochondrial [OS=Homo sapiens]	181.969865	5.74624161	17	75	17	298	32.975	ATP5C1	0.11	0.05	0.06	0
High	Q9V285	Phenylalanine-tRNA ligase alpha subunit [OS=Homo sapiens]	153.326948	15.976378	16	62	16	508	57.528	FARSA	0.59	0.05	0.54	0
High	P39019	40S ribosomal protein S19 [OS=Homo sapiens]	99.4494982	50.3449276	10	40	10	145	16.051	RPS19	0	0.05	-0.04	0
High	Q7LY03	Mitochondrial ribonuclease P protein 1 [OS=Homo sapiens]	13.0287609	7.94044665	3	4	3	403	47.317	TRMT10C	-0.05	0.05	-0.1	0
High	Q9H70-3	Isoform 3 of Methyltransferase-like protein 17, mitochondrial [OS=Homo sapiie	10.4728544	5.85774059	3	5	3	478	53.017	METTL17	0	0.05	-0.06	0
High	P13674-2	Isoform 2 of Prolyl 4-hydroxylase substrate alpha-1 [OS=Homo sapiens]	8.18482947	6.92883895	4	5	4	534	60.929	P4HA1	1	0.05	0.95	0
High	P23025	DNA repair domain-containing XP-A cells	8.4052915	7.69320769	1	2	1	273	31.348	XPA	10.27	0.05	10.22	0
Medium	Q966221	U3 small nucleolar ribonucleoprotein complementing XP-A cells	2.412127683	4.81099656	1	1	1	291	33.736	IMPA	10.86	0.05	10.81	0
Low	P09543-1	2'-cyclic nucleotide 3'-phosphodiesterase [OS=Homo sapiens]	1.45173345	2.6128266	1	1	1	421	47.549	CNP	-0.5	0.05	-0.35	0
High	P62829	60S ribosomal protein L23 [OS=Homo sapiens]	665.60464	77.1428571	17	678	17	140	14.856	RPL23	0.01	0.04	-0.03	0
High	P62269	40S ribosomal protein S18 [OS=Homo sapiens]	398.226245	59.86842111	25	350	25	152	17.078	RPS18	0.03	0.04	0.03	0
High	Q01813	ATP-dependent 6-phosphofructokinase, platelet type [OS=Homo sapiens]	25.01605	9.94887959	8	14	4	784	85.542	PFKP	0.07	0.04	0.03	0
High	Q9BIX9	WD repeat domain-containing protein 83 [OS=Homo sapiens]	6.58452767	5.3968254	2	4	2	315	34.321	WDR83	-0.58	0.04	-0.63	0
High	P46783	40S ribosomal protein S10 [OS=Homo sapiens]	80.2044269	53.9395939	11	37	11	165	18.886	RPS10	0.09	0.03	0.07	0
High	P61682	Ribosomal RNA processing protein L10 [OS=Homo sapiens]	52.5151891	18.8720174	9	27	9	461	52.807	RRP1	0.09	0.03	0.06	0
High	P61353	60S ribosomal protein L27 [OS=Homo sapiens]	48.820781	54.4117647	9	24	9	136	15.788	RPL27	0.6	0.03	0.57	0
High	Q96HAI-1	Nuclear envelope pore membrane protein POM121 [OS=Homo sapiens]	5.11616388	0.88070456	1	2	1	1249	127.642	POM121	0.26	0.03	0.23	0
High	P62280	40S ribosomal protein S11 [OS=Homo sapiens]	291.471926	59.4936709	18	395	18	158	18.419	RPS11	-0.11	0.02	-0.13	0
High	DE0783	28S ribosomal protein 54, mitochondrial [OS=Homo sapiens]	42.314125	3.02128105	3	11	3	128	15.129	MRPS14	0.16	0.02	0.14	0
High	H49755	Transmembrane protein 24 domain-containing protein 10 [OS=Homo sapiens]	50.00049434	5.02283105	1	1	1	219	24.96	TMED10	0.45	0.02	0.43	0
Low	Q13573	SNW domain-containing protein 1 [OS=Homo sapiens]	1.723716804	1.86567164	1	1	1	536	61.456	SNW1	0.32	0.02	0.31	0
High	P27635	60S ribosomal protein S7 [OS=Homo sapiens]	556.166375	64.0186916	19	732	19	214	24.588	RPL10	-0.04	0.01	0.05	0
High	P62081	40S ribosomal protein S7 [OS=Homo sapiens]	411.385447	22.668041	20	316	20	194	22.113	RPS7	0.12	0.01	0.11	0
High	Q8NHQ9	ATP-dependent RNA helicase DDX55 [OS=Homo sapiens]	87.267184	23.5	13	32	13	600	68.503	DDX55	0	0.01	0.02	0
High	Q86779	Probable peptidyl-tRNA hydrolase [OS=Homo sapiens]	4.95442894	11.682243	1	1	1	214	22.922	PTRH1	-0.27	0.01	-0.28	0
High	P62249	40S ribosomal protein S16 [OS=Homo sapiens]	138.905241	56.5212391	6	71	6	69	7.836	RPS16	-0.04	0.01	0.09	0
High	Q8NB0-5	Methyltransferase-like protein 13 [OS=Homo sapiens]	98.557531	28.8984263	18	36	18	699	78.718	METTL13	0.2	0.01	0.21	0
High	Q8Y37	Probable ATP-dependent RNA helicase DHX37 [OS=Homo sapiens]	24.232647	6.65514261	8	11	8	1157	129.464	DHX37	-0.03	0.01	-0.03	0
High	P48047	ATP synthase subunit O, mitochondrial [OS=Homo sapiens]	18.377571	25.5521127	5	5	5	213	23.263	ATPSO	0.33	-0.01	0.34	0
High	Q9BTG0	Death-inducer obligator 1 [OS=Homo sapiens]	8.335216	1.66178571	4	5	4	2240	243.723	DIDO1	0.29	0.01	0.3	0
High	P12236	ADP/ATP translocase 3 [OS=Homo sapiens]	390.362187	67.496644	24	233	4	298	32.946	SIC25A6	-0.05	-0.02	-0.03	0
High	P83731	60S ribosomal protein 124 [OS=Homo sapiens]	154.070702	48.4076433	15	239	15	157	17.768	RPL24	0.08	-0.02	0.1	0
High	O43653-1	Protein regulator of cyclinésis 1 [OS=Homo sapiens]	48.3420281	19.6774194	11	23	10	620	71.562	PRCL	0.3	-0.02	0.33	0
High	P62244	40S ribosomal protein S15a [OS=Homo sapiens]	269.883761	75.3846154	13	141	13	143	14.83	RPS15A	-0.15	-0.03	-0.12	0
High	P63173	60S ribosomal protein 38 [OS=Homo sapiens]	84.9443821	52.8571429	6	66	6	70	8.213	RPL38	0.02	-0.03	0.05	0

High	O95470	sphingosine-1-phosphate lyase 1 [OS=Homo sapiens]	7393956362	4,40140845	3	4	3	568	63,483	SGPL1	-0.1	-0.03	-0.07	0
High	P82266	40S ribosomal protein S23 [OS=Homo sapiens]	371,017231	72,027922	17	1188	17	143	12,433	RPS23	-0.08	-0.04	-0.04	0
High	P83881	60S ribosomal protein L36a [OS=Homo sapiens]	160,856691	73,735269	13	191	4	106	12,433	RPL36A	0.03	-0.04	-0.04	0.07
High	Q9Y6A4	Cilia- and flagella-associated protein 20 [OS=Homo sapiens]	26,1343312	23,3160622	5	9	5	193	22,76	160rF80; CFA P	0.27	-0.04	-0.04	0.31
High	P46777	60S ribosomal protein L5 [OS=Homo sapiens]	5,22047176	5,3820539	2	2	2	297	34,341	RPL5	-0.38	-0.04	-0.04	-0.34
High	P41252	Isoleucine-tRNA ligase, cytoplasmic [OS=Homo sapiens]	4,98886525	2,13946117	3	3	3	1262	144,406	IARS	-0.27	-0.04	-0.04	-0.23
Low	P49321-3	Isoform 3 of Nuclear autoantigenic sperm protein [OS=Homo sapiens]	1,243656833	0,88407595	1	1	1	790	86,216	NASP	0.03	-0.04	-0.04	0.07
High	P60866-2	Isoform 2 of 40S ribosomal protein S20 [OS=Homo sapiens]	109,22762	36,6197183	7	49	7	142	15,995	RPS20	-0.11	-0.05	-0.06	0.15
High	Q14683	structural maintenance of chromosomes protein 1a [OS=Homo sapiens]	66,7068715	14,2741281	17	28	17	1233	143,144	SMC1A	0.1	-0.05	-0.05	0.15
High	P08670	Vimentin [OS=Homo sapiens]	32,45533228	12,4463519	6	17	5	466	53,619	VIM	-0.14	-0.05	-0.05	-0.09
High	Q9NQ22	Something about silencing protein 10 [OS=Homo sapiens]	12,9257267	9,39457293	4	7	4	479	55,525	UTP3	0.17	-0.05	-0.05	0.22
Low	Q9BWQ9	Colleled-coil domain-containing protein 106 [OS=Homo sapiens]	2,05128271	2,85714286	1	1	1	280	32,012	CCDC106	0.65	-0.05	-0.05	0.71
High	P07437	tubulin beta chain [OS=Homo sapiens]	932,87256	84,069009	33	542	5	444	49,639	TUBB	-0.05	-0.06	-0.01	2
High	P46782	40S ribosomal protein S5 [OS=Homo sapiens]	386,784851	72,0588235	17	477	17	204	22,862	RPS5	-0.07	-0.06	-0.02	0
High	Q8W50-1	Ph finger protein 6 [OS=Homo sapiens]	234,9945348	45,4794521	20	112	20	365	41,264	PHF6	-0.31	-0.06	-0.06	-0.25
High	Q5P519	serine/arginine-rich splicing factor 11 [OS=Homo sapiens]	18,8808565	13,4297521	4	8	4	484	53,51	SRSF11	0.46	-0.06	-0.06	0.52
High	P11021	78 kDa Glucosidase-regulated protein [OS=Homo sapiens]	456,642055	57,1865443	40	190	37	654	72,288	HSPAS	-0.56	-0.07	-0.07	-0.49
High	Q536Q0	Very-long-chain 3-oxacyl-CoA reductase [OS=Homo sapiens]	82,08143	34,9358974	9	28	9	312	34,302	HSDB17B12	0.01	-0.07	-0.07	0.08
High	Q9Y5J1	Ub3 small nucleolar RNA-associated protein 18 homolog [OS=Homo sapiens]	65,9566942	24,4604317	10	20	10	556	61,964	UPTB18	0.14	-0.07	-0.07	0.2
High	O43175	D-3-phosphoglycerate dehydrogenase [OS=Homo sapiens]	31,3666842	12,0075477	7	15	7	533	56,614	PGDH	0.2	-0.07	-0.07	0.27
High	Q14488-2	Isoform 2 of RNA-binding protein 39 [OS=Homo sapiens]	622,175138	60,1145038	32	342	29	524	58,62	RBM39	0	-0.08	-0.08	0.08
High	P62910	60S ribosomal protein L32 [OS=Homo sapiens]	119,1616358	50,3702704	8	51	8	135	15,85	RPL22	0	-0.08	-0.08	0.08
High	P07910-1	Heterogeneous nuclear ribonucleoproteins C1/C2 [OS=Homo sapiens]	94,4708834	39,2156863	15	42	15	306	33,65	HRNPBC	-0.37	-0.08	-0.08	0.29
High	P08708	40S ribosomal protein S17 [OS=Homo sapiens]	572,5335962	73,3333333	17	474	17	135	15,54	RPS17L	-0.16	-0.09	-0.09	-0.27
High	P62847-4	Isoform 4 of 40S ribosomal protein S24 [OS=Homo sapiens]	282,5052277	21,1072664	8	190	8	289	32,41	RPS24	0.16	-0.09	-0.09	0.26
High	P46779	60S ribosomal protein L28 [OS=Homo sapiens]	65,0565305	38,6861314	7	47	7	137	15,738	RPL28	-0.03	-0.09	-0.09	0.06
High	Q9P014-2	Isoform 2 of CXCL-type zinc finger protein 1 [OS=Homo sapiens]	14,7961915	10,1515152	6	9	6	660	76,178	CXCL1	-0.53	-0.09	-0.09	0.45
Low	Q9Z797-1	Sympiekin [OS=Homo sapiens]	1,55083207	0,8642229	1	1	1	1274	141,05	SYMPK	-0.06	-0.09	-0.09	0.03
High	Q6UN15-1	Pre-mRNA 3'-end-processing factor, FIP1 [OS=Homo sapiens]	634,5870207	68,6868687	40	482	10	594	66,487	FIP1L1	0.17	-0.1	-0.1	0.27
High	Q5QNWW6-2	Isoform 2 of Histone H2B type 2-E [OS=Homo sapiens]	134,6200149	49,2537313	8	82	1	134	14,832	HIST2H2BF	0.02	-0.1	-0.1	0.11
High	Q8N1684-3	Isoform 3 of Cleavage and polyadenylation specificity factor subunit 7 [OS=Hon	13,73738409	13,618677	6	17	6	514	56,341	CPSF7	-0.04	-0.1	-0.06	0
High	Q8WVWC4	Uncharacterized protein C20f47, mitochondrial [OS=Homo sapiens]	13,1969215	10,1515152	3	5	3	291	32,523	C2orf47	0.07	-0.1	-0.1	0.17
High	A0FG188-6	Isoform 6 of Extended synaptosomal protein 2 [OS=Homo sapiens]	12,813407	37,1549894	4	6	4	942	104,643	-	-0.15	-0.1	-0.05	
High	P43307	Translocon-associated protein subunit alpha [OS=Homo sapiens]	7,23864742	2,7972038	1	3	1	286	32,215	SSR1	-0.35	-0.1	-0.25	0.27
High	P84098	60S ribosomal protein L19 [OS=Homo sapiens]	136,170823	33,1632653	12	75	12	196	23,451	RPL19	-0.05	-0.11	-0.06	0
High	Q9NNM5	Glioma tumor suppressor candidate region gene 2 protein [OS=Homo sapiens]	13,1969215	43,1465444	19	62	19	478	41,782	GLTSCR2	0.05	-0.11	-0.11	0.15
High	P20700	Lamin-B1 [OS=Homo sapiens]	68,7695292	22,0136519	13	26	12	586	66,368	LMNB1	0.37	-0.11	-0.11	0.47
High	P46776	60S ribosomal protein L27a [OS=Homo sapiens]	98,3929002	34,4594595	6	144	6	148	16,551	RP127A	-0.14	-0.12	-0.12	0.02
High	P16104	Histone H2AX [OS=Homo sapiens]	62,3567585	37,7622338	6	59	1	143	15,135	H2AFX	-0.27	-0.12	-0.12	0.15
High	Q9NP11-2	Isoform 2 of Bromodomain-containing protein 7 [OS=Homo sapiens]	59,8559707	23,1595092	13	29	13	652	16,782	BRD7	-0.46	-0.12	-0.12	-0.33
High	P31689-1	DnaI homolog subfamily A member 1 [OS=Homo sapiens]	50,3265002	24,1813602	7	15	7	397	40,839	DNAI1	0	-0.11	-0.11	0.12
Medium	Q9P035	Very-long-chain (3R)-3-hydroxyacyl-CoA dehydratase 3 [OS=Homo sapiens]	2,379753881	1,93370166	1	1	1	362	43,132	THLAD1; HACD	0.29	-0.13	-0.13	0.42
High	P61247	40S ribosomal protein L33a [OS=Homo sapiens]	526,6212121	30	306	30	264	22,926	RPS2A	-0.14	-0.14	-0.14	0.01	
High	P010570	Cleavage and polyadenylation specificity factor subunit 1 [OS=Homo sapiens]	525,6143451	62	243	62	1443	16,078	CPSF1	-0.09	-0.14	-0.14	0.05	
High	P04908	Histone H2A type 1-B [OS=Homo sapiens]	104,964422	8	70	2	130	14,127	H2AB; H1H1	-0.03	-0.14	-0.14	0.11	
High	P37108	Signal recognition particle 14 kDa protein [OS=Homo sapiens]	57,444773	41,9117647	7	25	7	136	14,561	SRP14	0.02	-0.14	-0.14	0.16
High	Q14897-2	Isoform 2 of Chromodomain-helicase-DNA-binding protein 4 [OS=Homo sapiens]	5,62457206	2,21649485	4	4	4	1940	22,709	CHD4	0.09	-0.14	-0.14	0.23
Medium	P22061-2	Isoform 2 of Protein-L-isoparpartate-D-aspartate O-methyltransferase [OS=Homo sapiens]	4,21745572	6,57894737	1	1	1	228	24,664	PCMT1	-0.23	-0.14	-0.14	0.08
Medium	Q43772	mitochondrial carnitine/acylcarnitine carrier protein [OS=Homo sapiens]	34,79003322	1	1	1	301	32,922	SLC25A20	-0.19	-0.14	-0.14	0.05	
High	P0C08	Histone H2A type 1 [OS=Homo sapiens]	151,483093	36,1538462	8	90	2	130	14,083	M: HIST1H2A;	-0.07	-0.15	-0.15	0.08
High	Q9P2NS	RNA-binding protein 27 [OS=Homo sapiens]	53,7666246	16,1320755	14	31	13	1060	118,645	RBM27	-0.8	-0.15	-0.15	-0.66
High	Q7KZ7-1	Serine/threonine-protein kinase MARK2 [OS=Homo sapiens]	17,025147	37,9441624	23	72	2	788	87,856	MARK2	0.09	-0.16	-0.16	0.25
High	Q9Y3C1	nucleolar protein 16 [OS=Homo sapiens]	173,225736	56,741573	13	61	13	178	21,175	NOP16	-0.31	-0.17	-0.17	0.15
Low	Q9BQG6	Ribosomal protein 63, mitochondrial [OS=Homo sapiens]	1,67060212	12,745088	1	1	1	102	12,259	VRP63; MRPL5	-0.07	-0.17	-0.17	0.1
High	P0DMV8	Heat shock 70 kDa protein 1A [OS=Homo sapiens]	622,866515	68,7458752	45	301	36	641	70,009	SPB18; LSP1A1	0.21	-0.18	-0.18	0.38
High	O95232-1	Lip7-like protein 3 [OS=Homo sapiens]	406,918784	43,9814815	28	453	28	432	51,435	LUC7L3	0.1	-0.18	-0.18	0.28
High	O94761	ATP-dependent DNA helicase Q4 [OS=Homo sapiens]	53,682556	10,7615894	11	22	11	1208	13,799	RECQL	-0.02	-0.18	-0.18	0.16
High	Q9HOW5	Colleled-coil domain-containing protein 8 [OS=Homo sapiens]	18,3573614	11,33829	4	8	4	538	59,339	CCDC8	0.11	-0.18	-0.18	0.29
High	Q9BQG62	Isoform 2 of Myb-binding protein 1A [OS=Homo sapiens]	16,3042154	2,7777778	4	8	4	1332	149,274	MYBBP1A	-0.02	-0.18	-0.18	0.16
Medium	O95391	Pre-mRNA-splicing factor SLU7 [OS=Homo sapiens]	2,65432331	1,525383618	1	1	1	586	68,344	SLU7	0.36	-0.18	-0.18	0.54
High	P62854	40S ribosomal protein S26 [OS=Homo sapiens]	171,275376	52,173913	6	171	6	115	13,007	JCI01929876;	-0.22	-0.19	-0.19	0

High	Q9Y3U8	60S ribosomal protein L36 [OS=Homo sapiens]	51.2706185	30.4761905	4	23	4	105	12.246	RPI236	-0.07	-0.19	0.12	0
High	P53885	Monocarboxylate transporter 1 [OS=Homo sapiens]	26.1101033	6.6	3	5	3	500	53.909	SIC16A1	-0.31	-0.19	-0.12	0
High	P12235	ATP translocase 1 [OS=Homo sapiens]	395.76739	68.1208054	23	257	4	298	33.043	SIC25A4	-0.01	-0.2	0.19	0
High	Q9UY51	Nucleolar protein 11.2 [OS=Homo sapiens]	127.458048	49.497652582	11	40	11	213	24.648	NOL12	-0.24	-0.2	-0.03	0
High	Q9ESCA4-1	Nucleolar protein 10 [OS=Homo sapiens]	19.1795598	6.83139535	5	7	5	688	80.251	NOL10	-0.7	-0.2	-0.5	0
High	P23396-1	40S ribosomal protein S3 [OS=Homo sapiens]	419.003623	83.9506173	26	285	26	243	26.671	RPS3	-0.13	-0.21	0.08	0
High	P49207	60S ribosomal protein L34 [OS=Homo sapiens]	56.4054084	38.4615385	8	49	8	117	13.284	RPL34	-0.5	-0.21	-0.29	0
Medium	Q6KCM7-3	Isform 3 of Calcium-binding mitochondrial carrier protein ScamC-2 [OS=Homo sapiens]	2.50003816	2.52427184	1	1	1	515	56.846	SIC25A5	-0.53	-0.21	-0.32	0
High	P62753	40S RIBOSOMAL PROTEIN S6 [OS=Homo sapiens]	636.656637	49.3975904	21	534	21	249	28.663	RPS6	-0.06	-0.23	0.17	0
High	Q6P2Q9	Pre-mRNA-processing-splicing factor 8 [OS=Homo sapiens]	92.5078058	7.06638116	16	39	16	2335	273.427	RPRF8	-0.38	-0.23	-0.15	0
High	Q8V15H7-1	SH2 domain-containing protein 33 [OS=Homo sapiens]	63.8335089	16.7441586	13	35	12	860	94.352	SNRBC3	0.4	-0.23	0.63	0
High	P62218	Small nuclear ribonucleoprotein snRNPd31 [OS=Homo sapiens]	10.0313709	7.14285714	1	4	1	126	13.907	SNRNPd3	-0.35	-0.23	0.13	0
Low	Q9RL1L	BICA1-A complex subunit RAP80 [OS=Homo sapiens]	1.28382965	1.25173853	1	1	1	719	79.678	UIMCL	-0.12	-0.23	0.1	0
High	P62701	40S ribosomal protein S4 X-isomeric [OS=Homo sapiens]	553.2288587	73.7642536	26	451	26	263	29.579	RPS4X	-0.27	-0.24	-0.03	0
High	Q8UE6	Histone H2A type Z-B [OS=Homo sapiens]	29.5546623	35.3846154	4	33	1	130	13.987	HIST2H2AB	0.03	-0.24	-0.27	0
High	P60891	Ribose-rich repeat byrophosphokinase 1 [OS=Homo sapiens]	18.0460297	14.4654088	4	5	4	318	34.812	RPRP51	-0.33	-0.25	-0.08	0
High	Q8N1G4	Leucine-rich repeat-containing protein 47 [OS=Homo sapiens]	16.2253881	8.9193825	4	6	4	583	63.434	LRRK47	0.07	-0.25	0.32	0
High	Q9Y283	Putative RNA-binding protein Luc7-like 2 [OS=Homo sapiens]	781.30089	64.2857143	40	641	31	392	46.486	LUC7L2	0.42	-0.26	0.68	20
High	P46781	40S ribosomal protein S9 [OS=Homo sapiens]	305.111856	59.2783505	23	306	23	194	22.578	RPS9	-0.28	-0.26	-0.01	0
High	P61254	60S ribosomal protein L26 [OS=Homo sapiens]	160.012	53.1034985	13	179	5	15	17.248	RPL26	-0.21	-0.26	0.05	16
High	Q9Y2R4	Probable ATP-dependent RNA helicase DDX52 [OS=Homo sapiens]	21.7962365	7.01168614	4	11	3	599	67.456	DDX52	0.36	-0.26	0.63	0
High	P06493	Cyclin-dependent kinase 1 [OS=Homo sapiens]	17.4457571	11.1111111	3	5	2	297	34.074	CDK1	0.25	-0.26	0.51	0
High	P00483	Cytochrome c oxidase subunit NDUF4A [OS=Homo sapiens]	15.5458917	46.9135802	4	7	4	81	9.364	NDUF4A	0.37	-0.26	0.63	0
High	P622917	60S ribosomal protein L8 [OS=Homo sapiens]	300.7042802	67.7042802	20	140	20	257	28.007	RPL8	-0.18	-0.27	0.1	0
High	P62888	60S ribosomal protein L30 [OS=Homo sapiens]	93.561983	66.0869565	7	33	7	115	12.776	RPL30	-0.39	-0.27	-0.11	0
High	Q9ZB21	Nuclear pore complex protein Nup205 [OS=Homo sapiens]	16.6131351	2.48580946	5	8	5	2012	227.776	NUP205	-0.46	-0.27	-0.19	0
High	P31947-1	14-3-3 protein sigma [OS=Homo sapiens]	9.3428917	6.4516129	2	4	1	248	27.757	SFN	-0.19	-0.27	0.08	1
High	P62750	60S ribosomal protein L23a [OS=Homo sapiens]	302.215925	63.076921	14	223	14	156	17.684	RP123A	-0.13	-0.28	0.14	0
High	P29372-2	Isform 2 of DNA 3'-methyladenine glycosylase [OS=Homo sapiens]	92.6985759	47.4402273	11	37	11	293	32.16	MPG	-0.11	-0.28	0.17	0
High	P61964	WD repeat-containing protein 5 [OS=Homo sapiens]	22.183194	15.26946111	4	8	4	334	36.565	WDR5	-0.08	-0.28	0.2	0
High	P25705-1	ATP synthase subunit alpha, mitochondrial [OS=Homo sapiens]	148.154238	49.5479204	26	59	26	553	59.714	ATP5A1	-0.31	-0.29	-0.02	0
High	Q969Q0	60S ribosomal protein S4a [OS=Homo sapiens]	142.371867	50.5439562	11	174	2	106	12.461	RPL36AL	-0.05	-0.29	0.25	0
High	P82914	28S ribosomal protein S15, mitochondrial [OS=Homo sapiens]	23.7470749	24.1245136	7	14	7	257	29.823	MRP515	1.33	-0.29	1.61	0
High	Q8P1X5	Transcription initiation factor TIF1 subunit 2 [OS=Homo sapiens]	912.301186	63.2195495	79	574	79	1199	136.3883	TAF2	-0.43	-0.3	-0.13	0
High	Q5TH9-1	RPL12-like protein [OS=Homo sapiens]	249.507338	38.387047	38	115	38	1297	143.611	RPL12	-0.31	-0.32	0.01	0
High	P62263	40S ribosomal protein S4a [OS=Homo sapiens]	41.7218543	9.5478543	9	114	9	151	12.163	RPS14	-0.26	-0.32	0.06	0
High	P12004	proliferating cell nuclear antigen [OS=Homo sapiens]	86.7085997	54.0229885	13	41	13	261	28.75	PCNA	-0.2	-0.32	0.11	1
High	Q9JIBX-2	Isform 2 of Mitochondrial dicarboxylate carrier [OS=Homo sapiens]	81.5951815	43.9188189	11	27	11	296	32.125	SIC25A10	-0.34	-0.32	-0.02	0
High	Q9HS4	Zinc finger protein 768 [OS=Homo sapiens]	30.981625	21.1111111	10	14	10	540	60.191	ZNF768	-0.26	-0.32	0.07	0
High	P51571	transcytosome-associated protein subunit delta [OS=Homo sapiens]	30.7161098	18.4971098	4	9	4	173	18.987	SSR4	-0.07	-0.32	0.24	0
High	P0C7P4	Putative cytochrome bc1 complex subunit Rieske-like protein 1 [OS=Homo sapiens]	47.4457924	4.90469947	1	1	1	283	30.796	UQCRC5I1	-0.19	-0.32	0.13	0
High	Q13309	tubulin beta-3 chain [OS=Homo sapiens]	47.4482268	38.	17	284	1	450	50.4	TUBB3	-0.43	-0.33	-0.11	1
High	Q96ME7	Zinc finger protein 512 [OS=Homo sapiens]	67.9333388	35.8024691	13	37	13	567	60.641	ZNF641	-0.26	-0.33	0.07	0
High	P12906-7	Isform 7 of Interleukin enhancer-binding factor 3 [OS=Homo sapiens]	4.636472477	3.63647439	3	3	3	898	95.748	ILF3	-0.43	-0.33	-0.1	0
High	P27448-6	Isform 5 of MAP/microtubule affinity-regulating kinase 3 [OS=Homo sapiens]	278.367019	35.66119916	35	125	1	713	79.886	MARK3	0.11	-0.34	0.45	0
High	Q5SY16	Polynucleotide 5'-hydroxyl-kinase NL09 [OS=Homo sapiens]	72.6521207	24.2165242	12	31	12	702	29.722	NOL9	0.34	-0.34	0.67	0
Medium	O95163	Elongator complex protein 1 [OS=Homo sapiens]	8.11010631	4.90469947	1	1	1	1332	150.159	IKBKBAP	0.27	-0.34	0.6	0
High	P30050-1	60S ribosomal protein S23 [OS=Homo sapiens]	29.6793036	33.333333	4	105	8	125	13.734	RPS25	-0.04	-0.39	0.35	0
High	Q96MU7-1	YTH domain-containing protein 1 [OS=Homo sapiens]	83.9705018	9.88146067	16	34	16	2136	244.353	SNRNP200	-0.71	-0.39	-0.32	0
High	Q95639	Cleavage and polyadenylation specific factor subunit 4 [OS=Homo sapiens]	13.7570787	25.8675079	8	7	4	727	84.649	YTHDC1	-0.51	-0.35	-0.16	0
High	Q9JUO7	Structural maintenance of chromosomes protein 3 [OS=Homo sapiens]	25.174746	75.464684	18	108	2	269	30.235	CPSF4	-0.25	-0.36	0.12	24
High	Q9JUW1	Thyroid hormone receptor-associated protein 3 [OS=Homo sapiens]	95.8083383	20.8370953	20	48	18	955	108.601	THRAP3	-0.3	-0.37	0.08	0
High	Q14684	Ribosomal RNA processing protein 1 homolog B [OS=Homo sapiens]	15.3396462	0.67567568	1	7	4	758	84.375	RRP1B	0.13	-0.38	0.6	0
High	P62251	Elongator complex protein 1 [OS=Homo sapiens]	99.8898079	39.92	8	105	8	125	13.734	RPL12	-0.04	-0.39	0.35	0
High	Q7P3X3	Ub small nuclear ribonucleoprotein 200 kDa helicase [OS=Homo sapiens]	83.9705018	9.88146067	16	34	16	2136	244.353	SNRNP200	-0.71	-0.39	-0.32	0
High	P63244	Guanine nucleotide-binding protein subunit beta2-lik1 [OS=Homo sapiens]	22.67439	25.8675079	8	7	4	727	84.649	SNB2L1;RACK	-0.69	-0.39	-0.31	0
High	P0UQ07	Structural maintenance of chromosomes protein 3 [OS=Homo sapiens]	22.5254284	5.42317173	7	12	7	1217	141.454	SMC3	0.03	-0.39	0.42	0
High	P46743-3	Isform 2 of Dual specificity mitogen-activated protein kinase 3 [OS=Homo sapiens]	14.7805645	5	5	32	32	13.734	-0.41	-0.39	-0.01	-0.01	0	
High	Q6P2X3	Teratocopeptide repeat protein 27 [OS=Homo sapiens]	6.677682	4.15183867	3	3	3	843	96.571	TTC27	1.38	-0.39	1.77	0
High	Q9HCIM-2	Isform 2 of Band 4.1-lik1 protein 5 [OS=Homo sapiens]	394.583991	72.8712871	30	339	6	505	57.849	EPB4115	-0.24	-0.4	0.16	0
High	Q00839	Heterogeneous nuclear ribonucleoprotein U [OS=Homo sapiens]	106.512561	23.5151515	17	45	17	825	90.528	HNRNPU	1.68	-0.4	2.08	0
High	P55081	microfibrillar-associated protein 1 [OS=Homo sapiens]	102.550418	48.5195622	19	46	19	439	51.927	MFAP1	-0.47	-0.07	-0.07	0

High	G92522	Histone H4X [OS=Homo sapiens]	100,9458138	37,0892019	10	41	10	213	22,474	H4FX	-0.22	-0.4	0.18	0	
High	P06748	Nucleophosmin [OS=Homo sapiens]	14,5917647	16,66666667	3	4	3	29	475	32,555	NPM1	-0.49	-0.4	-0.1	0
High	P26368	Splicing factor UFAF55 ribosomal subunit [OS=Homo sapiens]	554,175764	67,076921	29	384	29	475	53,467	UAF2	-0.24	-0.41	-0.17	0	
High	O9UIK0	Ribosome biogenesis protein TSR3 homolog [OS=Homo sapiens]	123,403015	48,0769231	11	45	11	312	33,575	TSR3	-0.25	-0.41	0.16	0	
High	O53H12	Acylglycerol kinase, mitochondrial [OS=Homo sapiens]	14,0766899	11,6115744	4	5	4	422	47,107	AGK	-0.56	-0.41	-0.15	0	
High	Q14974	Importin subunit beta-1 [OS=Homo sapiens]	9,59774362	5,59360731	5	6	5	876	97,108	KPNB1	-0.3	-0.41	0.1	0	
High	P62841	40S ribosomal protein S15 [OS=Homo sapiens]	685,822563	73,1034483	9	487	9	145	17,029	RPS15	-0.41	-0.42	0.05	0	
High	Q9BYT3_1	Serine/threonine-protein kinase 33 [OS=Homo sapiens]	39,9578826	24,3190615	11	22	11	514	57,794	STK33	0.23	-0.42	0.65	0	
High	Q9UNF1	Melanoma-associated antigen 12 [OS=Homo sapiens]	14,607003	10,5610561	5	7	5	606	64,914	MAGED2	-0.21	-0.42	0.21	0	
High	P15880	40S ribosomal protein S2 [OS=Homo sapiens]	282,905051	59,3856655	18	136	18	293	31,305	RPS2	-0.66	-0.43	-0.23	0	
High	Q9UNQ2	Probable dimethyladenosine transferase [OS=Homo sapiens]	160,6515479	73,8019169	21	94	21	313	35,214	DIMT1	-0.71	-0.43	-0.28	0	
High	Q86Y39_1	NADH dehydrogenase (ubiquinone) 1 alpha subcomplex subunit 11 [OS=Homo sapiens]	35,8847479	42,2269520	4	11	4	141	14,843	NDUFA11	-0.03	-0.43	0.4	0	
High	O14776_1	Transcription elongation regulator 1 [OS=Homo sapiens]	20,5210734	4,1894534	4	8	4	1098	123,823	TCERG1	-0.7	-0.43	-0.27	0	
Low	Q96GY0	Zinc finger C2H2 domain-containing protein 1A [OS=Homo sapiens]	1,95566345	1,84615385	1	1	1	325	35,07	ZC2H2C1A	-0.3	-0.44	0.14	0	
High	P62891	60S ribosomal protein L39 [OS=Homo sapiens]	37,8380653	37,4901961	3	85	3	51	6,403	RP13P	-0.41	-0.45	0.04	0	
High	P62241	40S ribosomal protein S8 [OS=Homo sapiens]	444,552606	62,9807692	21	282	21	208	24,19	RPS8	-0.49	-0.48	0.01	0	
High	P62826	GTP-binding nuclear protein RAN [OS=Homo sapiens]	63,327485	39,8148148	8	23	8	216	24,408	RAN	-0.62	-0.48	-0.14	0	
Medium	Q15054	DNA polymerase delta subunit 3 [OS=Homo sapiens]	4,31680061	2,14592275	1	2	1	466	51,368	POLD3	0.2	-0.49	0.7	0	
Low	P53997	Succinyl-CoA ligase (ADP/GDP-forming) subunit alpha, mitochondrial [OS=Homo sapiens]	1,460915607	1	1	1	1	346	36,227	SUCNL1	-0.86	-0.49	-0.37	0	
High	Q9JMS4	Pre-mRNA-processing factor 19 [OS=Homo sapiens]	33,246565	17,8971429	7	15	7	504	55,146	DRP19	-0.76	-0.5	-0.27	0	
High	Q9BYG3	MKL67/FHA domain-interacting nucleolar phosphoprotein [OS=Homo sapiens]	266,00165	57,337884	17	93	17	293	34,201	MKL67P_NIK	-0.36	-0.52	0.17	0	
High	P56134	ATP synthase subunit f, mitochondrial [OS=Homo sapiens]	25,7915018	25,5319149	2	7	2	94	10,911	ATPS12	-0.44	-0.52	0.07	0	
Medium	P9V5Y3_2	Isoform 2 of Melanoma-associated antigen D1 [OS=Homo sapiens]	2,3821301	1,31894484	1	2	1	834	91,901	MAGED1	-0.61	-0.52	-0.1	0	
High	P26233_1	60S ribosomal protein L13 [OS=Homo sapiens]	219,650139	56,6066351	17	176	17	211	24,247	RPL13	-0.39	-0.54	0.15	0	
High	Q9BFE4	Nucleolar GTP-binding protein 1 [OS=Homo sapiens]	278,225058	60,5678233	36	137	36	634	73,918	GTPBP4	-0.33	-0.55	0.23	0	
Low	Q43852_3	Isoform 3 of Calumenin [OS=Homo sapiens]	2,20873052	4,0247678	1	2	1	323	38,027	CALU	-0.83	-0.55	-0.28	0	
High	Q5T280	Plastid methyletranferase Cof114 [OS=Homo sapiens]	86,6327083	36,1702128	10	28	10	376	41,982	Cof114	-0.36	-0.56	0.2	0	
High	Q01081	Splicing factor, 55 kDa subunit [OS=Homo sapiens]	288,880568	56,6666567	12	169	2	207	27,854	DCU02724594	-0.42	-0.57	0.15	18	
High	P31943	Heterogeneous nuclear ribonucleoprotein H [OS=Homo sapiens]	19,501598	13,1403118	4	7	3	449	49,198	HNRNPH1	0.14	-0.57	0.71	1	
High	Q9UB54	Dinal homolog subfamily B member 11 [OS=Homo sapiens]	5,8921264	4,71860335	2	3	2	358	40,489	DNAI11	-0.57	-0.57	-0.12	0	
High	P62304	Small nuclear ribonucleoprotein E [OS=Homo sapiens]	5,76081681	25	2	2	2	92	10,797	SNRPE	-0.5	-0.57	-0.06	0	
Medium	Q15369_2	Isoform 2 of Band 4.1-like protein 1 [OS=Homo sapiens]	2,390533945	9,82144857	1	1	1	12	12,465	TCFB1	-0.96	-0.57	-0.39	0	
High	E9PRG8	Uncharacterized protein C11orf98 [OS=Homo sapiens]	21,102788	12,192788	12	27	12	305	33,691	Q2288414_C11	-0.3	-0.58	0.28	0	
High	P68363	Tubulin alpha-1B chain [OS=Homo sapiens]	619,13432	63,6363636	29	296	4	451	50,12	TUBA1B	-0.39	-0.59	0.19	37	
Low	P08047	Transcription factor Sp1 [OS=Homo sapiens]	1,534631471	0,76433121	1	1	1	785	80,644	SP1	0.75	-0.59	1.34	0	
High	Q9J329_2	Isoform 2 of Band 4.1-like protein 4 [OS=Homo sapiens]	34,2129135	36,4332724	12	101	15	518	58,529	EPBP1L4B	-0.57	-0.6	-0.03	0	
High	Q9NY93	Probable ATP-dependent RNA helicase DDX56 [OS=Homo sapiens]	68,9150338	29,4332724	12	27	12	547	61,551	DDX56	0.04	-0.6	0.64	0	
High	Q14232	Translation initiation factor eIF-2B subunit alpha [OS=Homo sapiens]	23,5358082	13,1147541	4	11	4	305	33,691	EIF2B1	-0.65	-0.6	-0.05	0	
High	P61619	Protein transport protein Sec61 subunit alpha isoform 1 [OS=Homo sapiens]	19,4059852	8,61344538	3	6	3	476	52,231	SEC61A1	-0.75	-0.6	-0.15	0	
High	P41250	Glycine-tRNA ligase [tRNA ligase]	8,04424338	3,4645391	3	4	3	739	83,113	GARS	-0.11	-0.61	0.5	0	
High	Q15365	Poly(RC) binding protein 1 [OS=Homo sapiens]	137,429135	64,8876404	14	52	9	356	37,470	PCBP1	-0.73	-0.62	-0.11	0	
High	Q87TC3_2	Isoform 2 of 30S ribosomal protein 130, mitochondrial [OS=Homo sapiens]	6,97784759	5,7591623	1	2	1	191	21,825	-	-0.39	-0.62	0.23	0	
High	Q9N93	Apoptosis-inducing factor 1 [OS=Homo sapiens]	112,101084	30,1939058	11	56	11	361	38,344	CAAP1	-0.58	-0.63	0.05	0	
High	Q14232	T-lymphoma invasion and metastasis-inducing protein 1 [OS=Homo sapiens]	14,287318	12,9541691	4	6	4	1591	177,398	TIAM1	-0.57	-0.63	0.06	0	
High	P77815_1	Breast cancer anti-estrogen resistance protein 3 [OS=Homo sapiens]	159,3748665	43,8787297	28	74	27	825	92,507	BCAR3	-0.61	-0.64	0.03	1	
High	P62979	Ubiquitin-t40S ribosomal protein 52alpha [OS=Homo sapiens]	280,03675	58,3333333	10	161	7	156	17,953	RP52TA	-0.53	-0.65	0.13	3	
High	Q87TC3	NAD-dependent protein deacetylase sirtuin-7 [OS=Homo sapiens]	101,810576	39,75	13	29	13	400	44,87	SIRT7	-0.34	-0.65	0.32	0	
High	Q9NB81_1	Apoptosis-inducing factor 1, mitochondrial [OS=Homo sapiens]	6,97784759	5,7591623	1	17,7814029	9	16	613	86,859	AIFM1	0.76	-0.65	0.05	0
High	Q37CC8_2	Isocapne 2 of Mitochondrial import membrane translocase subunit Tim50	16,8049135	10,9649123	4	7	4	456	50,433	TIMM50	-0.6	-0.6	0.06	0	
High	Q92616	translational activator GCN1 [OS=Homo sapiens]	9,37412495	1,906938723	4	6	4	2671	292,572	GCN11; GCN1	-1.04	-0.66	-0.38	0	
High	Q9UQ35	serine/arginine-repetitive matrix protein 2 [OS=Homo sapiens]	64,3474235	5,33790698	12	43	12	2752	299,438	SRM2	-0.76	-0.67	-0.09	0	
High	Q9BY54	Serine/threonine-protein kinase rho2 [OS=Homo sapiens]	18,9778468	12,6811594	5	9	5	552	63,243	RIOK2	-0.45	-0.67	0.22	0	
High	Q9HB4	Sideroflexin-1 [OS=Homo sapiens]	27,3738122	18,0124224	5	8	5	322	35,596	SFYN1	-0.44	-0.69	0.25	0	
High	P35268	60S ribosomal protein L22 [OS=Homo sapiens]	46,5785545	57,8125	6	28	4	128	14,778	RPL22	-0.77	-0.7	-0.07	0	
High	P07305	Histone H4.0 [OS=Homo sapiens]	90,8534368	28,3505155	6	60	6	194	20,85	H1FO	-0.72	-0.72	0	0	
High	P47914	60S ribosomal protein L29 [OS=Homo sapiens]	46,7375078	24,5283019	4	44	4	159	17,741	RPL29	-0.31	-0.72	0.41	0	
High	P31040	Succinate dehydrogenase (ubiquinol) flavoprotein subunit, mitochondrial [OS=Homo sapiens]	47,986214	17,7814029	4	4	4	664	72,7084	SDHA	0.29	-0.72	0.01	0	
High	Q8Y174	Isocapne 4 of Neuropathy target esterase [OS=Homo sapiens]	5,18633985	1,16363636	1	2	1	1375	150,859	PNPLA6	0.92	-0.72	1.41	0	
High	Q15366_2	Isocapne 2 of Poly(C) binding protein 2 [OS=Homo sapiens]	5,19059976	1,1202186	6	26	1	366	38,627	PCBP2	-1.32	-0.73	-0.59	0	
High	Q75683	Surfactant locus protein 6 [OS=Homo sapiens]	5,98828022	6,09481283	2	2	2	361	41,426	SURE6	-0.59	-0.73	0.14	0	
High	Q8N3E9	1-phosphatidylinositol 4,5-bisphosphate phosphodiesterase delta 3 [OS=Homo sapiens]	600,078554	72,1166033	43	272	43	789	89,202	PLCD3	-0.75	-0.74	0	0	

High	P49756-1	RNA-binding protein 25 [OS=Homo sapiens]	372,825,518	46,263,452	44	191	44	843	100,124	RBM25	-0.72	-0.74	0.02	0
High	Q9HC55	band 4-like protein 4A [OS=Homo sapiens]	26,502,639	19	10	12	10	686	79,01	EPB414A	-0.51	-0.75	0.25	0
High	P96P11-2	Isoform 2 of Probable 2.8S rRNA (cytosine-C5')-methyltransferase [OS=Homo sapiens]	9,839,1289	6,437,1389	3	6	3	466	50,379	NSUN5	-0.64	-0.75	0.12	0
Medium	P48444	Ceafamer subunit delta [OS=Homo sapiens]	2,812,261,38	2,152,641,88	1	1	1	511	57,174	ARCN1	-0.15	-0.75	0.6	0
High	O43809	Cleavage and polyadenylation specificity factor subunit 5 [OS=Homo sapiens]	83,653,8558	58,590,5084	10	31	10	227	26,211	NUDT11	-0.71	-0.76	0.05	0
High	Q95748	Ribosome biogenesis protein NSA2 homolog [OS=Homo sapiens]	60,254,2194	34,230,7692	10	30	10	260	30,047	NSA2	-0.53	-0.76	0.23	0
High	Q8NP45-1	Zinc finger protein 64 homolog, isoforms 1 and 2 [OS=Homo sapiens]	9,834,9925	5,139,50073	3	5	1	681	74,596	ZFP64	-0.75	-0.76	0.02	2
High	Q9BU67-1	Multiple myeloma tumor-associated protein 2 [OS=Homo sapiens]	51,3,275,33	63,878,327	29	548	29	263	29,394	C1orf35	-1.11	-0.77	0.34	0
High	Q96CB9	5-methylcytosine RNA methyltransferase NSUN4 [OS=Homo sapiens]	15,421,0302	10,156,252	4	6	4	384	43,061	NSUN4	-0.98	-0.77	-0.21	0
Medium	Q53G10-1	pleckstrin homology domain-containing family O member 1 [OS=Homo sapiens]	2,731,9027	3,667,481,66	1	1	1	409	46,209	PLEKHO1	-0.83	-0.77	-0.06	0
Medium	Q9UH99	Signal recognition particle subunit SRP16 [OS=Homo sapiens]	3,163,83163	2,551,834,13	2	2	2	627	59,688	SRP88	-1.58	-0.78	-0.8	0
High	Q9YB9	FATCT complex subunit SPT16 [OS=Homo sapiens]	5,743,9714	1,146,131,81	1	2	1	104	70,167	SUPT16H	-0.11	-0.79	0.67	0
High	Q9YAA4	Ribosomal RNA-processing protein 7 homolog A [OS=Homo sapiens]	150,758,105	60,357,14,29	13	73	13	280	32,314	RBP7A	-0.77	-0.81	0.04	0
High	P49759-3	Isoform 3 of Dual specificity protein kinase CLK1 [OS=Homo sapiens]	90,982,0311	24,714,8289	13	37	12	526	61,718	CLK1	-0.67	-0.81	0.14	1
High	Q02978	Mitochondrial 2-oxoglutarate/malate carrier protein 1 [OS=Homo sapiens]	69,108,218	61,700,803	19	79	19	314	34,04	SLC25A11	-0.77	-0.82	0.05	0
High	Q8YI93	Serine/arginine repetitive matrix protein 1 [OS=Homo sapiens]	150,124,778	19,91,150,44	18	93	18	904	102,274	SRRM1	-1.2	-0.83	0.37	0
High	Q9H941-4	Ubiquitin carboxy-terminal hydrolase 42 [OS=Homo sapiens]	20,543,26	5,891,12,867	6	10	6	1324	145,302	USP42	-0.54	-0.83	0.29	0
High	P56955-6	Isoform 6 of Breast cancer anti-estrogen resistance protein 1 [OS=Homo sapiens]	603,224,124	61,026,029	45	324	45	916	97,817	BCAR1	-0.61	-0.84	0.24	0
High	P51570-2	Isoform 2 of Galactokinase [OS=Homo sapiens]	28,755,221	13,381,0427	6	11	6	422	45,329	GALK1	-0.9	-0.84	-0.07	0
High	Q9YV1	Tubulin beta 28 chain [OS=Homo sapiens]	749,14,862	61,707,752	26	45	1	45	49,921	TUBB2B	-1.1	-0.85	-0.25	0
High	Q6DK11	60S ribosomal protein L7/L17.1 [OS=Homo sapiens]	46,04,19228	33,739,83,74	7	15	7	246	28,643	RPL7L1	-0.53	-0.85	0.32	0
High	P21980	Protein-glutamine gamma-glutamyltransferase 2 [OS=Homo sapiens]	41,537,074	17,61,260,93	10	16	10	687	77,78	TGM2	-0.92	-0.85	-0.07	0
High	Q9ZX6-2	Isoform 2 of Nucleolar and spindle-associated protein 1 [OS=Homo sapiens]	124,747,7792	56,81,88,18	24	71	3	440	49,294	NUSAP1	-0.76	-0.86	0.1	15
High	Q7Z6E9	E3 ubiquitin-protein ligase RBBP6 [OS=Homo sapiens]	77,845,229	13,504,643	24	71	24	1792	201,442	RBBP6	-1.03	-0.87	-0.16	0
High	Q8NYV4-1	Cyclin-dependent kinase 12 [OS=Homo sapiens]	26,63,659	6,91,275,168	8	14	7	1490	164,054	CDK12	-0.76	-0.87	0.11	1
High	P45954	Short/branched chain specific acyl-CoA dehydrogenase, mitochondrial [OS=Homo sapiens]	61,088,8813	4,16,666,667	2	2	2	432	47,455	ACADS	-0.75	-0.87	0.12	0
Medium	P63220	40S ribosomal protein S21 [OS=Homo sapiens]	2,779,8837	8,43,373,494	1	2	1	83	9,106	RPS21	-0.19	-0.87	0.68	0
High	Q8IZ69	RNA (uracil-5')-methyltransferase homolog A [OS=Homo sapiens]	23,418,3551	11,36	7	11	7	626	68,682	TRMT2A	-0.42	-0.88	0.46	0
High	Q9YX6-2	Multifunctional methytransferase subunit TRM112-like protein [OS=Homo sapiens]	14,69,430,32	31,2	3	4	3	125	14,19	TRMT112	-1.83	-0.88	-0.95	0
High	P41091	eukaryotic translation initiation factor 2 subunit 3 [OS=Homo sapiens]	7,169,9,99	4,44,915,254	2	2	2	472	51,077	EIF2S3	-0.74	-0.89	0.15	0
Medium	Q15293	Reticulocalbin-1 [OS=Homo sapiens]	24,898,5499	3,9,274,9245	1	1	1	331	38,866	RCN1	-0.81	-0.89	0.09	0
High	Q9PQ31	Thyroid transcription factor 1-associated protein 26 [OS=Homo sapiens]	55,527,4884	37,43,59,833	9	31	9	241	28,652	CCDC59	-0.68	-0.92	0.25	0
High	P08865	40S ribosomal protein SA [OS=Homo sapiens]	66,95,740,56	38,98,30,508	9	24	9	295	32,833	RPSA	-1	-0.93	-0.07	0
High	P68371	Tubulin beta-4B chain [OS=Homo sapiens]	906,88,866	83,82,02,47	33	540	1	445	49,799	TUBB4B	-0.89	-0.94	0.05	71
High	Q9Y3A2	Probable U13 small nucleolar RNA-associated protein 11 [OS=Homo sapiens]	53,708,7915	37,549,040,71	11	23	11	253	30,428	UTP11L; UTP14	-1.16	-0.94	-0.22	0
High	Q8WQU7-2	Isoform 2 of Cactin [OS=Homo sapiens]	37,87,9671	23,006,633	17	33	33	935	10,781	TOP1	-0.97	-0.96	-0.01	0
High	P16085	ADP-ribosylation factor 4 [OS=Homo sapiens]	35,70,582,9	28,33,33,33	4	13	3	180	20,498	ARF4	-0.81	-0.96	0.14	0
Medium	Q9H972	Ribosome production factor 1 [OS=Homo sapiens]	3,8,40,3281	3,4,38,39,542	1	1	1	349	40,086	RPF1	-1.67	-0.96	-0.71	0
High	Q9NW78	Aurora kinase A-interacting protein [OS=Homo sapiens]	22,07,9635	7,035,175,88	2	9	2	199	22,34	AURKAIP1	-1.37	-0.97	-0.39	0
High	P0B339	ATP-dependent RNA helicase DDX30 [OS=Homo sapiens]	187,63,744	51,17,63,744	31	99	28	737	52,514	DDX50	-0.71	-0.98	0.28	2
High	P11387	DNA topoisomerase 1 [OS=Homo sapiens]	78,38,696	23,006,633	17	33	17	765	90,669	TOP1	-1.39	-0.98	-0.42	0
High	P16615	Sarcoplasmic/endoplasmic reticulum calcium ATPase 2 [OS=Homo sapiens]	72,90,76,3	16,98,65,643	16	34	16	1042	114,683	ATP2A2	-1.02	-0.98	-0.04	0
High	Q8TER5	Guanine nucleotide exchange factor 40 [OS=Homo sapiens]	18,20,1181	15,13,49,721	7	9	7	1519	164,555	ARHGFE40	-1.28	-0.98	0.3	0
High	Q9P848-3	Isoform 3 of Transcription initiation factor TFIID subunit 6 [OS=Homo sapiens]	2,6,61,06,443	2	2	2	714	76,947	TAIF6	-0.59	-0.98	0.39	0	
High	Q9UJ55	Transmembrane and coiled-coil domains protein 3 [OS=Homo sapiens]	191,01,7373	48,008,357	26	83	25	477	53,752	TMCC3	-0.8	-0.99	0.19	0
High	P47756-2	Isoform 2 of F-actin-capping protein subunit beta 1 [OS=Homo sapiens]	4,760,2018	6,25	2	3	2	272	30,609	CAPZB	-0.39	-0.99	0.6	0
Low	P17026	Cell division cycle 5-like protein [OS=Homo sapiens]	14,20,0213	6,69,642,857	1	1	1	224	25,899	ZNF22	-0.39	-0.99	0.6	0
High	Q8NFM7	Lysophosphatidylcholine acyltransferase 1 [OS=Homo sapiens]	6,64,04,6897	2,4,34,65,693	1	4	1	534	59,113	LPCAT1	-0.34	-0.98	-0.02	0
High	Q8P087	Rho guanine nucleotide exchange factor 40 [OS=Homo sapiens]	11,56,60,75	10,56,61,93	4	5	4	351	38,437	RPUS3	-1.05	-1.01	0.73	0
High	Q6GP33-2	Isoform 2 of ADP-ribosylation factor-like protein 6-interacting protein 4 [OS=Homo sapiens]	36,32,9146	15,25,42,33	6	16	6	413	44,132	ARL6IP4	-1.72	-1.04	-0.68	0
High	Q8IWK6	Adhesion G protein-coupled receptor A3 [OS=Homo sapiens]	31,28,019,5	7,41,86,2,26	9	19	9	1221	146,058	PN125; ADG124	-1.28	-1.04	-0.24	0
High	Q99459	Cell division cycle 5-like protein [OS=Homo sapiens]	123,68,1495	27,43,142,14	18	52	18	802	87,145	CDC5L	-1.01	-1.05	0.04	0
High	Q4KMP7	TET1 domain family member 10B [OS=Homo sapiens]	122,77,6005	25	18	64	18	808	59,113	TBC1D10B	-1.07	-1.05	0.02	0
High	Q8WXA9-2	Isoform 2 of Splicing regulatory glutamine/lysine-rich protein 1 [OS=Homo sapiens]	19,11,7126,9	3,8,46,15,385	2	11	2	624	71,606	SREK1	-0.34	-1.07	0.73	0
High	P58557	Putative ribonuclease [OS=Homo sapiens]	16,59,8751	16,16,76,647	2	4	2	167	19,286	YBEY	-0.8	-1.07	0.27	0
High	Q13823	Nucleolar GTP-binding protein 2 [OS=Homo sapiens]	106,17,8821	33,37,89,33	20	49	20	731	83,603	GNL2	-1.39	-1.08	-0.31	0
High	P20719	Hmboob protein Hox-A5 [OS=Homo sapiens]	51,72,65,64	27,77,77,77	6	12	6	270	25,327	H0X5	-1.25	-1.08	-0.17	0
High	P05023	Sodium/potassium transporting ATPase subunit alpha-1 [OS=Homo sapiens]	147,08,925	29,32,51,32	28	76	25	1023	112,824	ATP1A1	-1.1	-1.12	0.03	3
High	Q95299	Na/DH <sub>+</sub> dehydrogenase (ubiquinone) 1 alpha subcomplex subunit 0, mitochondrial [OS=Homo sapiens]	13,03,17,91	14,92,55,75	4	5	4	355	40,725	NDUFA10	0.06	-1.12	1.18	0
High	Q9H300	Presentins-associated rhomboid-like protein, mitochondrial [OS=Homo sapiens]	10,98,736,6	10,29,02,375	3	4	3	379	42,163	PARL	-1.39	-1.12	-0.27	0
Low	P49792	E3 SUMO-protein ligase RanBP2 [OS=Homo sapiens]	1,53,65,5497	0,27,91,533	1	1	1	3224	357,974	RANBP2	-0.33	-1.12	0.79	0

High	Q8N4G0	zinc finger protein 687 [OS=Homo sapiens]	32,4269297	7,59302991	8	16	8	1237	129,446	-0.63	-1.13	0.5	0	
High	Q94876	Transmembrane and coiled-coil domains protein 1 [OS=Homo sapiens]	22,149746	8,72889334	6	12	5	653	72,038	-0.49	-1.13	0.64	0	
High	O00488	Zinc finger protein 553 [OS=Homo sapiens]	37,2916799	55,225806	5	10	5	103	15,19	-0.82	-1.14	0.31	0	
High	O75644	ATP synthase subunit 8, mitochondrial [OS=Homo sapiens]	19,81829827	37,864077	3	5	3	103	11,421	ATP5I	-0.81	-1.16	0.34	0
High	Q13523	Serine/threonine-protein kinase PRK4 homolog [OS=Homo sapiens]	65,2307608	13,6047666	13	32	13	1007	116,916	PRPF4B	-0.94	-1.18	0.24	0
High	Q9UDY2-7	Isomeric 7 of Tight junction protein 2α-2 [OS=Homo sapiens]	196,913853	34,8075348	38	93	38	1221	137,258	TJP2	-1.1	-1.19	0.09	0
High	P26641-2	Isomeric 2 of Elongation factor 1-gamma [OS=Homo sapiens]	25,6579682	13,963039	7	9	7	487	56,114	EEF1G	-1.07	-1.19	0.12	0
High	Q9NWU5	35S ribosomal protein 122, mitochondrial [OS=Homo sapiens]	5,13917532	10,1941748	2	2	2	206	23,626	MRPL22	0.59	-1.19	1.78	0
High	Q13428-3	Isomeric 3 of Treacle protein [OS=Homo sapiens]	105,488563	17,7300201	26	81	26	1489	152,114	TCOF1	-2.82	-1.2	-1.62	0
High	O75367-1	Core histone macro-H2A.1 [OS=Homo sapiens]	29,2906933	17,7419355	5	8	5	372	39,592	H2AFY	-0.44	-1.21	0.76	0
Medium	P11177	Pyruvate dehydrogenase E1 component subunit beta, mitochondrial [OS=Homo sapiens]	2,30129065	2,22841262	1	1	1	359	20,208	PDHIB	-2.4	-1.22	-1.19	0
High	P11171-1	protein 4.1 [OS=Homo sapiens]	109,2912732	27,5662763	20	46	17	864	96,957	EPBF1	-1.36	-1.24	-0.12	0
High	P63208	sphase kinase-associated protein 1 [OS=Homo sapiens]	48,783619	50,3067485	7	14	7	163	18,646	SKP1	-0.61	-1.24	0.63	0
High	Q9P210	Cleavage and polyadenylation specificity factor subunit 2 [OS=Homo sapiens]	267,518448	51,6624041	31	110	31	782	88,431	CPSE2	-0.88	-1.25	0.36	0
High	Q9Y244	RNA-processing protein FCF1 homolog [OS=Homo sapiens]	112,674876	38,8888889	10	37	10	198	23,354	FCF1	-1.02	-1.28	0.26	0
High	Q71014	probable ATP-dependent RNA helicase DDX16 [OS=Homo sapiens]	104,408263	20,659538	20	67	20	1031	11,729	DDX16	-0.94	-1.28	0.34	0
High	Q9BX6-4	Isomeric 4 of Nucleolar and spindle-associated protein 1 [OS=Homo sapiens]	107,842872	25,2941176	22	64	1	425	47,541	NUSAP1	-1.03	-1.29	0.26	0
High	Q8WWD3-1	E2 ubiquitin-protein ligase RNF138 [OS=Homo sapiens]	11,7150511	25,7142857	5	5	5	245	28,174	RNF138	0.79	-1.29	0.51	0
High	Q9NWV6	Arginine and glutamatergic-rich protein 1 [OS=Homo sapiens]	202,126562	47,222473	28	290	28	273	33,197	ARG1U1	-1.25	-1.34	0.09	0
High	P50416	Carnitine O-palmitoyltransferase 1, liver isoform [OS=Homo sapiens]	13,676151	8,92626132	6	7	6	773	88,311	CPT1A	-1.16	-1.34	0.19	0
High	P19474	E3 ubiquitin-protein ligase TRIM21 [OS=Homo sapiens]	532,752206	67,1578947	37	260	37	475	54,135	TRIM21	-0.75	-1.35	0.6	0
Medium	P04792	Heat shock protein beta-1 [OS=Homo sapiens]	3,13299144	6,34146341	1	1	1	205	22,768	HSPB1	-1.91	-1.35	-0.57	0
High	Q8E6U0-1	probable RNA-binding protein 23 [OS=Homo sapiens]	59,5766925	17,0842825	7	47	4	439	48,701	RBM23	-1.45	-1.38	-0.07	0
Low	Q8N0172-1	zinc finger protein 444 [OS=Homo sapiens]	1,82554019	2,14067278	1	1	1	327	35,182	ZNF444	0.18	-1.38	1.57	0
High	Q5CZ24	nucleolar Mif4G domain-containing protein 1 [OS=Homo sapiens]	112,6473033	20,1167791	16	45	16	860	96,198	NOM1	-2.15	-1.4	-0.74	0
High	Q14244	Enrichsin [OS=Homo sapiens]	109,41114	27,9038718	24	54	23	749	84,002	MATP7	-1.17	-1.4	0.23	1
High	Q96009	G-protein coupled receptor-associated sorting protein 2 [OS=Homo sapiens]	6,97054575	2,2673031	2	5	2	838	93,715	P2_ARMCX5_G	-1.43	-1.4	-0.02	0
High	Q99590-1	protein SCAF11 [OS=Homo sapiens]	7,3972122	3,3923445	3	3	3	1463	35,551	SCAF11	-1.27	-1.42	0.15	0
High	P21127-1	Cyclin-dependent kinase 11B [OS=Homo sapiens]	190,31212	39,7484277	34	114	34	795	92,65	CDK11B	-1.75	-1.43	-0.32	0
Medium	P78549	Endonuclease III-like protein 1 [OS=Homo sapiens]	3,0075045	3,2051821	1	2	1	312	34,368	NTHL1	-0.5	-1.44	0.94	0
Low	Q5QMY1-1	Nucleolar protein 7 [OS=Homo sapiens]	1,67019548	3,89105058	1	1	1	257	29,409	NOL7	-1.39	-1.45	0.06	0
High	Q77C8-4	Isomeric 3 of Transcription initiation factor TFIID subunit 2 [OS=Homo sapiens]	6,97054575	24,2672513	5	15	5	338	37,388	TAF1	-1.7	-1.46	-0.24	0
High	Q9C018	pre-mRNA 3' end processing protein WDR33 [OS=Homo sapiens]	192,481051	26,2724551	34	34	34	1336	145,799	WDR33	-1.48	-1.47	-0.02	0
High	Q8IWCV1	MAP7 domain-containing protein 3 [OS=Homo sapiens]	20,9192056	7,53424658	7	9	7	876	98,368	MAP7D3	-1.1	-1.47	0.38	0
High	Q75400	pre-mRNA-processing factor 40 homolog A [OS=Homo sapiens]	314,244181	33,4378265	32	150	32	957	108,737	PRPF40A	-1.7	-1.48	-0.23	0
Medium	P50897	Palmitoyl-protein thioesterase 1 [OS=Homo sapiens]	14,9853893	7,841315983	2	2	2	306	34,171	PTPRT	-1.41	-1.48	0.06	0
High	Q8TAD8	Smad nuclear-interacting protein 1 [OS=Homo sapiens]	18,0181549	9,84848485	3	8	3	396	45,75	SNIP1	-1.45	-1.51	0.06	0
High	Q9BV2	Glutamine nucleotide-binding protein-like 3 [OS=Homo sapiens]	1199,55305	62,8415301	53	682	52	549	61,954	GNL3	-1.59	-1.52	-0.06	2
High	Q8NAV1-1	Pre-mRNA-splicing factor 3BA [OS=Homo sapiens]	7,8874982	8,97435897	3	3	3	312	37,453	PRPF38A	-1.79	-1.52	-0.26	0
High	P42704	Leucine-rich PRP motif-containing protein, mitochondrial [OS=Homo sapiens]	31,812574	7,96269727	10	19	10	1394	157,805	LRRPRC	-2.1	-1.55	-0.54	0
High	H17509	Homeobox protein Hox-B6 [OS=Homo sapiens]	19,8872059	5,6	5	24	24	25,370	108,706	HoxB6	-0.92	-1.57	0.65	0
High	Q9HFM4	Band 4.1-like protein 5 [OS=Homo sapiens]	485,7606842	66,9849932	37	389	2	733	81,805	EPB4115	-1.56	-1.6	0.04	42
High	Q13427	Peptidyl-prolyl cis-trans-isomerase E [OS=Homo sapiens]	55,738904	15,1193634	11	24	11	754	88,564	PIP1	-1.5	-1.6	0.1	0
High	Q95104	Splicing factor, arginine/serine-rich 15 [OS=Homo sapiens]	38,5142534	7,7537228	9	24	9	147	125,79	SCAF4	-1.46	-1.62	0.16	0
High	Q6BD16	Lysine-specific demethylase 4D [OS=Homo sapiens]	21,6584822	11,663479	7	9	7	523	58,565	KDM4D	-0.94	-1.63	0.69	0
High	A8MVN32	PhD finger protein 20-like protein 1 [OS=Homo sapiens]	21,607765	10,1278269	8	14	8	1017	114,938	PHB20L1	-2.33	-1.63	-0.7	0
High	Q9H477	Protein SREBP1 [OS=Homo sapiens]	7,5945477	0,93786635	1	3	1	853	95,848	MCM3	-1.63	-1.63	0.04	0
High	Q8NFW8	N-acetylneuraminate cytidylyltransferase [OS=Homo sapiens]	38,7544113	23,0414747	9	18	9	434	48,349	CNAAS	-1.51	-1.64	0.13	0
Low	Q6ZNG9	KRAB-A-domain-containing protein 2 [OS=Homo sapiens]	1,80574625	1,2195122	1	1	1	492	56,169	KRB2	-1.65	-1.65	0.07	0
High	Q77K6-1	Centromere protein V [OS=Homo sapiens]	110,493775	49,0908091	11	40	2	275	29,927	CENPV	-1.94	-1.69	-0.24	8
High	Q9NVF7-1	F-box only protein 28 [OS=Homo sapiens]	108,069983	38,8586957	15	41	15	388	41,123	FBXO28	-0.88	-1.72	0.84	0
High	Q8NQ2	protein SREBP1 [OS=Homo sapiens]	32,972146	21,290226	5	22	5	155	18,166	SREK1P1	-0.92	-1.75	0.83	0
High	Q8WTT2	Nucleolar complex protein 3 homolog [OS=Homo sapiens]	153,772152	38	27	67	27	800	92,49	NOC3L	-1.06	-1.78	0.72	0
High	Q8UKV3-1	Apoptotic chromatin condensation inducer in the nucleus [OS=Homo sapiens]	14,062847	2,31170768	3	4	3	1341	151,771	ACIN1	-1.79	-1.79	0	0
High	Q8Q8123	RNA (Cytosine-34-C(5)-methyl)transferase [OS=Homo sapiens]	90,6923432	29,8565841	19	55	19	767	86,416	NSUN2	-1.41	-1.8	0.39	0
Medium	Q8NB90-1	Spermatogenesis-associated protein 5 [OS=Homo sapiens]	3,70816163	2,75955207	2	2	2	893	97,843	SPATA5	5,29	-1.8	7.09	0
Medium	Q92974	Rho GTPase nucleotide exchange factor 2 [OS=Homo sapiens]	2,58185348	0,912779	1	3	1	986	11,473	ARHGEF2	7,29	-1.8	0.98	0
High	Q9BR16	Protein LP homolog [OS=Homo sapiens]	61,6713522	34,8837209	5	26	5	129	15,245	LPH	-1.09	-1.81	0.73	0
High	Q9H445	Oxysterol-binding protein-related protein 3 [OS=Homo sapiens]	37,955504	59,7519729	41	162	5	887	101,16	OSBP13	-1.27	-1.83	0.56	37
High	P63012-2	Isomeric 2 of AP-2 complex subunit beta [OS=Homo sapiens]	26,3193666	7,78128286	7	11	7	951	105,625	AP2B1	-1.85	-1.85	0	0
High	Q12986-3	Isomeric 3 of Transcriptional repressor NF-X1 [OS=Homo sapiens]	45,6886031	15,4861945	10	26	10	833	92,618	NFX1	-1.18	-1.87	0.69	0

High	Q9Y4X4-1	Krueppel-like factor 12 [OS=Homo sapiens]	10,056,6381	6,965,17413	3	5	3	402	44,212	KLF12	-1.63	-1.89	0.26	0
High	P18533-9	Isoform of Protein SON [OS=Homo sapiens]	42,138,1604	6,606,88457	11	50	11	1666	267,923	TIP1	-1.8	-1.9	-0.08	0
High	Q7157-2	Isoform Short of Tight junction protein ZO-1 [OS=Homo sapiens]	31,575,5932	8,393,28537	10	12	10	1668	186,852	PKN3	-1.82	-1.94	0.12	0
High	Q6P572	Serine/threonine-protein kinase N3 [OS=Homo sapiens]	8,629,51363	2,249,71212	2	4	2	889	99,358	DUSP1	-1.55	-1.96	0.42	0
High	O75319-1	RNA/RNP complex-1-interacting phosphatase [OS=Homo sapiens]	46,057,336	41,21,21212	10	20	10	330	38,915	BClAF1	-2.1	-1.97	-0.13	2
High	Q9NYF8-1	Bcl-2-associated transcription factor 1 [OS=Homo sapiens]	88,308,3774	24,130,4348	23	52	21	920	106,059	FAM133B	-2,24	-1.98	-0.26	4
High	P05K19-1	Protein FAM133B [OS=Homo sapiens]	40,896,6643	35,62,75304	6	23	3	247	28,368	CAPTA1	-1,24	-1.98	0.74	0
High	P52907	F-actin-capping protein subunit alpha-1 [OS=Homo sapiens]	11,742,8203	17,83,21678	4	4	4	286	32,902	PHRF1	-1,67	-1.99	0.32	0
High	Q9R1V6-1	PhD and RING finger domain-containing protein 1 [OS=Homo sapiens]	40,712,688	7,39,842329	10	22	10	1649	178,557	TSR1	-2,07	-2.03	-0.04	0
High	Q2NI82	Pre-mRNA-processing protein TSR1 homolog [OS=Homo sapiens]	302,50614	40,67,16418	35	163	35	804	91,752	RBM26	-2,21	-2.05	-0.15	1
High	Q8TBP6	RNA-binding protein 26 [OS=Homo sapiens]	106,488132	18,768,5197	19	55	18	1007	113,527	RP9	-1,68	-2.05	0.37	0
High	Q8TA86	Retinitis pigmentosa 9 protein [OS=Homo sapiens]	80,111,7903	35,746,6053	11	57	11	221	26,091	SF3B3	-1,95	-2.06	0.11	0
High	Q15333-1	Splicing factor 3b subunit 3 [OS=Homo sapiens]	23,461,7632	6,16,69515	8	11	8	1217	135,492	SIC25A19	-0.84	-2.09	1.25	0
High	Q9HC21	Mitochondrial thiamine pyrophosphate carrier [OS=Homo sapiens]	14,681,3634	14,375,	4	5	4	320	35,488	TRIM28	-2,34	-2.12	-0.22	0
High	Q13263	Transcription intermediary factor 1-beta [OS=Homo sapiens]	24,520,9709	14,59,28859	5	7	5	835	88,493	PRKCI	-1.86	-2.13	0.27	0
High	P17143	Protein kinase C iota type [OS=Homo sapiens]	19,314,6227	7,55,608028	5	9	5	847	94,565	MATR3	-1.8	-2.13	0.34	0
High	P43243	Matrin-3 [OS=Homo sapiens]	13,19,7664	4,9,19,7861	5	7	5	935	101,495	MTHFD1	-2,44	-2.14	0.3	0
High	P11586	C-1-tetrahydrofolate synthase, cytoplasmic [OS=Homo sapiens]	19,988,706	6,44,831116	6	8	6	977	107,478	AIF2A1	-1.8	-2.15	0.35	0
High	P05782	Arf-2 complex subunit alpha-1 [OS=Homo sapiens]	19,092,049	42,76,29809	14	47	14	462	50,109	EF1A1	-2,51	-2.16	-0.35	0
High	Q02040-1	A-kinase anchor protein 17A [OS=Homo sapiens]	479,44,3032	47,76,97842	47	348	47	695	80,686	AKAP17A	-2,25	-2.17	-0.08	0
High	Q9BR88	G patch domain-containing protein 1 [OS=Homo sapiens]	55,524,5784	16,32,65306	11	20	11	931	103,282	GPATCH1	-2,92	-2.17	-0.75	0
Medium	Q15459	Splicing factor 3A subunit 1 [OS=Homo sapiens]	2,479,8348	0,88,272383	1	1	1	793	88,831	SF3A1	-2,3	-2.21	-0.09	0
High	Q8NN57	NF-kappa-B-activating protein [OS=Homo sapiens]	48,441,526	24,78,29816	9	29	9	415	47,111	NKAP	-2,03	-2.22	0.2	0
High	Q8IWX8	Calcium homeostasis endoplasmic reticulum protein [OS=Homo sapiens]	78,359,9887	18,44,97817	15	30	15	916	103,637	CHERP	-2,66	-2.24	-0.42	0
High	P09874	Poly [ADP-ribose] polymerase 1 [OS=Homo sapiens]	18,056,1311	5,02,95888	5	8	5	1014	113,012	PARP1	-2	-2.24	0.24	0
High	Q13435	Splicing factor 3b subunit 2 [OS=Homo sapiens]	14,17,6119	7,03,910615	6	8	6	895	100,165	SF3B2	-2,21	-2.25	0.04	0
High	P33991	DNA replication licensing factor CMCM4 [OS=Homo sapiens]	28,855,615	1,01,52,843	1	2	1	8	96,498	MCMV4	-2,03	-2.26	0.23	0
Low	Q4KWH8-1	1-phosphatidylinositol 4,5-diphosphate phosphodiesterase eta-1 [OS=Homo sapiens]	2,187,8713	4,41,346722	1	1	1	1693	189,104	PLCH1	-1,31	-2.26	0.95	0
High	Q8NH18	Putative phospholipase B-like 2 [OS=Homo sapiens]	59,154,9376	24,78,77759	13	25	13	589	65,43	PBLD2	-2,62	-2.27	-0.35	0
High	Q00458	Interferon-related developmental regulator 1 [OS=Homo sapiens]	6,799,8421	7,09,534,368	2	2	2	451	50,236	JFRD1	-2,32	-2.27	-0.05	0
High	P68032	Actin, alpha cardiac muscle 1 [OS=Homo sapiens]	171,87,102738	40,53,0504	15	172	2	377	41,893	XPTB	-1.96	-2.28	0.32	0
High	P60709	Actin, cytoplasmic 1 [OS=Homo sapiens]	13,00,74373	4,43,895,172	5	5	5	1657	189,134	ACTC1	-1.63	-2.28	0.65	0
Medium	Q6ZN55-2	Isoform 2 of Zinc finger protein 574 [OS=Homo sapiens]	4,102,5418	1,01,52,843	1	2	1	985	108,504	-1,03	-2.28	1.25	0	
High	Q96137-1	Putative RNA-binding protein 15 [OS=Homo sapiens]	19,137,5171	6,03,88458	5	7	5	977	107,124	RBM15	-2,06	-2.3	0.24	0
High	Q35922	Exportin-1 [OS=Homo sapiens]	78,516,947	12,88,981,29	11	27	11	962	109,893	XPOT	-2,1	-2.31	0.21	0
High	P46940	Actin, cytoplasmic 1 [OS=Homo sapiens]	326,82,3363	70,93,3333	24	240	11	375	41,71	ACTC1	-2,36	-2.33	-0.03	21
High	Q9NPF64	nucleolar protein of 40 kDa [OS=Homo sapiens]	268,63,1356	45,22,8158	16	210	16	241	27,552	ZCHC17	-2,52	-2.33	-0.18	0
High	Q3KQU3	MAP7 domain-containing protein 1 [OS=Homo sapiens]	21,58,33152	9,75,029727	8	14	7	841	92,764	MAP7D1	-1,99	-2,35	0.36	0
High	P29597	Non-receptor tyrosine-protein kinase TK2 [OS=Homo sapiens]	7,51,779871	0,92,95938	1	3	1	1187	133,565	TYK2	-2,68	-2.38	-0.3	0
High	Q8UJ49	E3ISG15-protein ligase HERC5 [OS=Homo sapiens]	384,51,6762	45,996,0938	38	148	38	1024	116,777	HERC5	-2,1	-2.39	0.29	0
High	Q75147	Oisourin-1-like protein 1 [OS=Homo sapiens]	30,27,096	4,43,037975	8	12	8	1896	206,817	OB5L1	-1,62	-2.39	0.77	0
High	P55786	purimycin-sensitive aminopeptidase [OS=Homo sapiens]	22,90,397	5,76,713819	5	9	5	919	103,211	NPEPPS	-2,08	-2.39	0.31	0
High	P5189-1	Tufelin-interacting protein 11 [OS=Homo sapiens]	17,65,15062	7,16,845,878	6	7	6	837	96,758	TRP11	-2,64	-2.44	-0.2	0
High	Q14241	Transcription elongation factor B polypeptide 3 [OS=Homo sapiens]	17,75,4689	6,76,691,29	5	13	7	798	89,835	TCEB3	-2,2	-2.45	0.25	0
High	P55060-1	Exportin-2 [OS=Homo sapiens]	11,65,10409	2,26,570546	3	5	3	971	110,346	CSE1L	-2,81	-2.47	-0.34	0
Medium	Q43395	U4/U6 small nuclear ribonucleoprotein Prp31 [OS=Homo sapiens]	3,357,9389	3,80,673499	2	2	2	683	77,481	PRPF3	-1,49	-2.47	0.98	0
Medium	Q8XK2-2	Isoform 2 of Condensin complex subunit G2 [OS=Homo sapiens]	7,328,73791	2,85,006196	1	1	1	1156	132,111	NCAPG2	-1,94	-2.49	0.55	0
Low	Q9ULD4	Bromodomain and PH finger-containing protein 3 [OS=Homo sapiens]	1,493,63028	5,86,981,86	1	1	1	1205	135,66	BRPF3	-2,33	-2.5	0.17	0
High	Q9J388	RNA-binding motif protein, X-linked 2 [OS=Homo sapiens]	43,17,556	25,4658,385	7	13	7	322	37,313	RBMX2	-2,34	-2.52	0.18	0
Medium	Q15542	Transcription initiation factor Cl1orf57 [OS=Homo sapiens]	146,77,1392	40,61,4345	15	71	15	293	34,176	C11orf57	-2,3	-2.53	0.23	0
High	Q966Q7	Probable ATP-dependent RNA helicase DDX27 [OS=Homo sapiens]	91,255,7244	29,648,4412	23	58	23	796	89,779	DDX27	-1,62	-2.53	0.91	0
High	Q9HCB6	Spondin-1 [OS=Homo sapiens]	2,738,46244	0,77,854,671	5	2	2	807	90,914	SPON1	-2,24	-2.53	0.3	0
High	P56192	Methionine-tRNA ligase, cytoplasmic [OS=Homo sapiens]	39,72,7155	13,22,22222	11	19	11	900	101,052	MARS	-2,28	-2.54	0.26	0
High	Q94906-1	Pre-mRNA-processing factor 6 [OS=Homo sapiens]	20,28,3445	7,01,38159	6	8	6	941	106,358	PRPF6	-2,7	-2.55	-0.15	0
Medium	Q6ZUT1-2	Isoform 2 of Uncharacterized protein Cl1orf57 [OS=Homo sapiens]	3,774,69072	1,375,	1	1	1	800	86,776	TAf5	-3,55	-2.55	-1	0
High	Q8PIP8	DNA cross-link repair 1A protein [OS=Homo sapiens]	17,12,605,58	4,90,384615	5	5	5	1040	116,326	DCLRE1A	-2,36	-2.58	0.23	0
High	Q99599	Plakophilin-2 [OS=Homo sapiens]	25,73,4605	13,393876	9	12	9	881	97,355	PKP2	-1,81	-2.59	0.78	0
High	Q14M03	Heterogeneous nuclear ribonucleoprotein U-like protein 1 [OS=Homo sapiens]	40,18,65804	17,53,68139	11	21	11	747	85,052	HNRNPU2	-2,23	-2.62	0.39	0
High	Q15029	116 kDa US small nuclear ribonucleoprotein component [OS=Homo sapiens]	61,23,388766	20,47,3251	16	25	16	972	109,366	ETUD2	-2,61	-2.64	0.03	0
High	P49761	Dual specificity protein kinase CLK3 [OS=Homo sapiens]	78,167,1703	15,0470219	8	24	8	638	73,469	CLK3	-2,67	-2.65	-0.02	0

High	Q14562	ATP-dependent RNA helicase dhh8 [OS=Homo sapiens]	179,872895	36,3934426	35	83	35	1220	139,227	DHH8	-2.73	-2.68	-0.05	0
High	P53618	Ceatorner subunit beta [OS=Homo sapiens]	22,092,392	8,91,920252	7	9	7	953	-	-	-2.69	-	-0.04	0
Medium	Q96776-8	Isoform 5 of MMS2/9 nucleotide excision repair protein homolog [OS=Homo sa	2,157,2185	0,85,632731	1	1	1	1051	115,63,61	COPB1	-2.6	-2.7	0.1	0
High	Q14980	Exoprotein-1 [OS=Homo sapiens]	47,814,6828	12,605,042	12	19	12	1071	123,306	XPO1	-2.82	-2.72	-0.11	0
High	P43246-1	DNA mismatch repair protein MSH2 [OS=Homo sapiens]	14,983,9768	4,28,655,25	4	6	4	934	104,67,77	MSH2	-1.65	-2.74	-1.09	0
High	Q8NIE71-1	ATP-binding cassette sub-family F member 1 [OS=Homo sapiens]	13,541,9358	4,14,201,183	3	5	3	845	95,86,6	ABCF1	-2.67	-2.74	0.07	0
High	Q9NNR29	Lymphoid-specific helicase [OS=Homo sapiens]	5,430,7832	3,63,982401	3	3	3	838	97,01,2	HELLS	-2.31	-2.75	0.44	0
High	Q9Y2I2	Band 4.1-like protein 3 [OS=Homo sapiens]	88,048,802	19,963,2015	17	31	14	1087	120,50,3	EPR4113	-2.41	-2.76	0.35	0
High	Q9IBZ3-5	Isoform 5 of Oxyesterol-binding protein-related protein 6 [OS=Homo sapiens]	143,684,751	30,969,602	24	64	24	959	108,89	OSBPL6	-2.25	-2.77	0.53	0
High	Q8BYF7	Formin-like protein 3 [OS=Homo sapiens]	16,809,594	3,2,01,01673	4	6	2	1028	117,13,9	FMN13	-2.67	-2.77	0.1	0
High	P16331	DNA damage-binding protein 1 [OS=Homo sapiens]	49,648,4258	11,92,98246	12	18	12	1140	126,88,7	DDI1	-2.96	-2.78	-0.18	0
High	Q9H307	Pinin [OS=Homo sapiens]	22,355,4048	7,51,38075	6	11	6	717	81,56,5	PNN	-2.69	-2.78	0.09	0
High	Q14331	Protein FRG1 [OS=Homo sapiens]	33,176,7188	21,705,2464	6	13	6	258	29,15,4	FRG1	-2.65	-2.79	0.15	0
High	Q9PP015	36S ribosomal protein L15, mitochondrial [OS=Homo sapiens]	9,613,07735	14,864,8649	4	5	4	296	33,39,9	MRPL15	-3.12	-2.83	-0.29	0
High	Q9YBB6	PAX3- and PAX7-binding protein [OS=Homo sapiens]	21,780,7579	7,30,64,402	5	6	5	917	104,73,9	GCF1; PAXBP1	-2.36	-2.85	0.49	0
Low	P54198	Protein HIRA [OS=Homo sapiens]	1,403,9297	1,17,9941	1	1	1	1017	111,76,4	HIRA	-2.16	-2.86	0.7	0
High	Q727K6-3	Isoform 3 of Centromere protein V [OS=Homo sapiens]	105,760617	48,16,17647	10	40	1	272	29,71,2	CENPV	-1.91	-2.87	0.95	0
High	Q9UJV9	Probable ATP-dependent RNA helicase DDX41 [OS=Homo sapiens]	1077,8304	91,96,14148	74	675	74	622	69,79,3	DDX41	-3.57	-2.92	-0.64	0
High	Q9EPV5-3	Isoform 2 of Formin-like protein 2 [OS=Homo sapiens]	8,413,07337	3,06,12,821	4	4	2	1092	124,02,9	FMLN2	-2.73	-2.93	0.2	1
High	Q99679-2	Isoform Pm22 of pituitary homeobox 2 [OS=Homo sapiens]	11,728,9751	3	3	3	324	35,77,5	PTX2	-1.67	-2.95	-0.27	0	
High	Q9H1J1-1	Regulator of nonsense transcripts 3A1 [OS=Homo sapiens]	11,816,7297	10,294,1176	5	9	5	476	54,66,3	UPF3A	-3.3	-2.98	-0.32	0
High	Q9BUQ8	Probable ATP-dependent RNA helicase DDX23 [OS=Homo sapiens]	13,981,9576	7,07,31,073	5	5	5	820	95,52,4	DDX23	-2.69	-3	0.3	0
High	Q9HOAO	N-acetyltransferase 10 [OS=Homo sapiens]	533,956463	58,92,68293	58	227	58	1025	115,65,7	NAT10	-2.88	-3.01	0.13	0
High	Q43290	U4/U6/U5 tri-snRNP-associated protein 1 [OS=Homo sapiens]	64,631,3822	12,0,25	12	25	12	800	90,2	SART1	-3.1	-3.07	-0.03	0
High	P00338-3	Isoform 3 of L-lactate dehydrogenase A chain [OS=Homo sapiens]	10,113,3893	8,58,725,762	3	3	2	361	39,81,2	LDHA	-3.31	-3.09	-0.22	1
Low	Q9I8W3-3	Isoform 3 of Protein ECT2 [OS=Homo sapiens]	1,591,42087	1,35,69,374	1	1	1	958	108,46,1	ECT2	-2.53	-3.12	0.58	0
Low	P23458	Tyrosine-protein kinase Jak1 [OS=Homo sapiens]	1,661,54351	0,69,32,469	1	1	1	1154	133,19,1	JAK1	-2.46	-3.14	0.68	0
High	Q13393-2	Isoform P1D127B of phospholipase D1 [OS=Homo sapiens]	114,51,1277	22,68,3917	20	44	20	1036	119,61,8	PLD1	-2.75	-3.22	0.47	0
High	Q9H6R4-1	Nucleolar protein 6 [OS=Homo sapiens]	218,46,3919	30,279,321	29	88	29	1146	127,51,3	NOL6	-3.12	-3.23	0.11	0
High	Q9ULK5	VenG-like protein 2 [OS=Homo sapiens]	6,220,3029	4,4,145,733	2	2	2	521	59,67,7	VANGL2	-2.54	-3.29	0.75	0
High	Q14697-2	Isoform 2 of Neutral alpha-glucosidase AB [OS=Homo sapiens]	4,637,25639	1,96,668,371	2	2	2	966	109,36,9	GANAB	-2.74	-3.29	0.55	0
High	P78347	General transcription factor II-I [OS=Homo sapiens]	47,447,1521	16,93,86,77	15	19	15	998	112,34,6	GTF2I	-3.07	-3.33	0.26	0
High	Q9RVW6-1	U3 small nucleolar RNA-associated protein 14 homolog A [OS=Homo sapiens]	2,377,8082	1,31,33,203	18	30	18	717	87,94,6	UTP14A	-2.7	-3.35	0.65	0
High	Q14566	DNA replication licensing factor CMCM6 [OS=Homo sapiens]	5,493,6446	3,04,50,6399	2	2	2	821	92,83,1	MCM6	-3.4	-3.4	0.01	0
High	Q13206	probable ATP-dependent RNA helicase DDX10 [OS=Homo sapiens]	6,611,53906	4,2,28,57,143	3	3	3	875	100,82,5	DDX10	-2.58	-3.43	0.85	0
High	P78316	Nucleolar protein 14 [OS=Homo sapiens]	20,318,531	11,31,85,31	9	9	9	857	97,18,9	NDP40	-2.62	-3.48	0.86	0
Medium	Q8WV54	WD repeat-containing protein 60 [OS=Homo sapiens]	3,377,8082	1,31,33,203	1	2	1	1066	122,49,6	WDR60	-2.09	-3.5	1.4	0
High	Q95686-2	Isoform 2 of Bromodomain-containing protein 1 [OS=Homo sapiens]	5,893,59415	1,51,13,87,721	2	2	2	1189	133,15,9	BRD1; LOC10833	-2.04	-3.61	1.56	0
High	P21741	Midline 1 [OS=Homo sapiens]	55,315,584	60,13,96,01	9	22	9	143	15,57,5	MDK	-3.9	-3.64	-0.26	0
High	Q8TDD1	ATP-dependent RNA helicase DDX54 [OS=Homo sapiens]	213,72,647	53,27,58,24	38	92	1	881	98,53,4	DDX54	-3.15	-3.74	0.59	29
High	Q94813	Slt1 homolog 2 protein [OS=Homo sapiens]	228,66,429	26,35,98,03	47	112	47	1529	169,7,9	SLT2	-3.84	-3.87	0.03	0
Medium	Q9UDR5	Alpha-aminoacidic semialdehyde synthase, mitochondrial [OS=Homo sapiens]	2,807,15,388	1,07,99,13,61	1	1	1	926	102,06,6	AASS	-2.99	-3.88	0.89	0
High	Q9NNW5	WD repeat-containing protein 61 [OS=Homo sapiens]	12,10,64,491	3,36,98,05,1	4	5	4	1121	121,64,7	WDR6	-3.39	-3.93	0.53	0
High	Q9EDT7	Zinc finger and BTB domain-containing protein 10 [OS=Homo sapiens]	1,752,81,774	0,9,18,84,05	1	1	1	871	94,83,5	ZBTB10	-2.85	-3.94	1.1	0
High	Q86Y76	E2 ubiquitin-protein ligase mbl1 [OS=Homo sapiens]	22,29,50,693	10,57,58,481	6	6	6	1006	110,81,6	MBI1	-4.86	-4.09	-0.77	0
High	Q90566	U3 small nucleolar ribonuclease mbl1 [OS=Homo sapiens]	17,19,36,03	10,57,28,872	6	8	6	681	78,81,6	MPHOSPH10	-4.14	-4.13	0	0
High	P13647	Keratin, type II cytoskeletal 5 [OS=Homo sapiens]	89,678,2869	29,83,05,85	23	128	9	590	62,34	KRT5	-4.55	-4.4	-0.16	2
Low	P15924-1	Desmoplakin [OS=Homo sapiens]	1,677,07,8071	0,95,32,885	1	1	1	1049	113,67,6	DSG1	-5,24	-5,12	-0.12	0
High	Q9UP8-3	Isoform 3 of Ankyrin repeat domain-containing protein 26 [OS=Homo sapiens]	288,74,8634	40,76,02,339	65	123	65	2871	331,56,9	DSP	-4,59	-4,53	0.38	0
Low	Q96D77	Zinc finger and BTB domain-containing protein 20 [OS=Homo sapiens]	20,79,73,373	48,60,82,306	32	95	32	847	96,74,9	FTS3	-4,15	-4,53	-0.25	0
High	Q94fH1-2	Isoform 2 of FERM, RhGEF and pleckstrin domain-containing protein 1 [OS=Homo sapiens]	20,66,491,3	6,8,77,33,42	7	8	7	1076	122,03,7	-	-3.81	-4.68	0.87	0
High	Q15154-1	Pericentriolar material 1 protein [OS=Homo sapiens]	96,27,20,5	14,77,22,73	25	38	24	2024	228,39,2	PCM1	-4,06	-4,78	0.71	1
High	P35579-1	Miosin-9 [OS=Homo sapiens]	886,6,125	65,15,30,612	143	375	123	1060	226,39,2	MYH9	-5,55	-4,93	-0.62	14
Low	Q02413	Desmoglein-1 [OS=Homo sapiens]	1,384,22,124	0,20,89,642	1	1	1	1049	113,67,6	DSG1	-5,24	-5,12	-0.12	0
High	Q9UP8-3	Isoform 3 of Ankyrin repeat domain-containing protein 26 [OS=Homo sapiens]	288,74,8634	40,76,02,339	65	123	65	1710	196,28,9	ANKRD26	-5,04	-5,25	0.21	0
High	Q9HC84	Mucin-5B [OS=Homo sapiens]	11,20,83,978	2,0,99,95,29	4	5	4	5762	59,5,96	MUC5B	-5,37	-5,25	-0.12	0
High	Q9PfN3-1	Lethal(2) giant larvae protein homolog 2 [OS=Homo sapiens]	79,86,24,86	25,68,62,745	19	31	19	1020	113,37,7	LGII2	-5,34	-5,36	0.02	0
High	Q9UP8-1	Upf3B-associated SdRP motif-containing protein [OS=Homo sapiens]	19,90,74,64	6,80,72,719	7	10	6	1029	118,21,9	U2SURP	-5,86	-6,03	0.17	0
High	Q15334	Lethal(2) giant larvae protein homolog 1 [OS=Homo sapiens]	104,05,22,38	30,73,08,27	25	43	25	1064	115,34,6	LGII1	-6,3	-6,21	-0.09	0
Medium	Q00178	GTP-binding protein 1 [OS=Homo sapiens]	4,12,35,176	2,39,16,293	1	2	1	659	72,40,8	GTPBP1	1,31	-6,31	7,62	0
Low	Q9EfI-5-2	Isoform 2 of GTPase MAP family member 5 [OS=Homo sapiens]	1,31,31,4835	5,76,53,68,876	1	2	1	347	39,55,9	GIMAP5	-6,46	-6,41	-0.06	0
Medium	Q9fBE1-1	P02, AT hook-, and zinc finger-containing protein 1 [OS=Homo sapiens]	3,34,26,3419	3,34,88,937	2	2	2	687	74,01,3	PATZ1	-6,83	-6,65	-0.17	0

Low	Q13123	Protein Red [OS=Homo sapiens]	1.53555779	1.61579892	1	1	537	65.5562	IK	-6.82	-6.65	-0.17	0	
Medium	Q5T749	Keratinocyte proline-rich protein [OS=Homo sapiens]	3.38033881	3.45423143	2	3	579	64.0933	KRPB	-7.15	-7.02	-0.12	0	
Medium	O15344-1	E3 ubiquitin-protein ligase Millidine-1 [OS=Homo sapiens]	4;49972043	5.09124127	2	2	667	75.203	MID1	-7.21	-7.07	-0.13	0	
Medium	P06702	Protein S100-A9 [OS=Homo sapiens]	3.60240575	13.1578947	1	1	114	13.234	S100A9	-7.48	-7.35	-0.13	0	
Medium	Q131617-2	Isoform 2 of Cullin-2 [OS=Homo sapiens]	2.77667553	2.8758115	2	2	764	83.434	CUL2	-7.54	-7.37	-0.17	0	
Low	Q8WV15-1	Extracellular sulfatase Sulf-2 [OS=Homo sapiens]	1.41240127	1.14942529	1	2	870	100.39	SULF2	-7.52	-7.39	-0.13	0	
Medium	P35637-1	RNA-binding protein FUS [OS=Homo sapiens]	4;49596018	5.89535612	2	2	526	53.394	FUS	-7.77	-7.64	-0.13	0	
Medium	P52292	Importin alpha-1 [OS=Homo sapiens]	2.46205504	3.78071834	1	1	529	57.826	KPNAA2	-7.82	-7.68	-0.13	0	
High	P02545	Prefamin-A/C [OS=Homo sapiens]	6;3280034	4.81277111	3	3	664	74.095	LMNA	-7.82	-7.69	-0.13	0	
Low	P14923	Junction plakoglobin [OS=Homo sapiens]	1.96337311	1.61073826	1	1	745	81.693	JUP	-8.14	-7.97	-0.17	0	
High	Q9P2Y4	Zinc finger protein 219 [OS=Homo sapiens]	12.7969393	9.41828255	5	5	722	76.83	ZNF219	-8.27	-8.1	-0.17	0	
High	Q9UGM3-4	Isoform 4 of Deleted in malignant brain tumors 1 protein [OS=Homo sapiens]	10.4710515	6.8762279	3	4	1527	166.395	DNMT3B	-8.26	-8.14	-0.12	0	
Medium	Q5SRE5	Nucleoporin NUP188 homolog [OS=Homo sapiens]	2.860403573	9.91480846	1	1	1749	195.917	NUP188	-8.37	-8.17	-0.2	0	
High	Q86723	Homerin [OS=Homo sapiens]	29.5309549	9.75438596	9	28	2850	282.228	HRNR	-8.37	-8.23	-0.14	0	
Medium	Q55985	DNA topoisomerase 3-beta-1 [OS=Homo sapiens]	3.53549913	1.85614849	2	2	862	96.599	TOPBP1	-0.03	-8.27	8.24	0	
High	Q9H089	Large subunit GTPase 1 homolog [OS=Homo sapiens]	5.14549848	4.7112462	2	3	658	75.178	LSG1	-8.68	-8.5	-0.17	0	
Low	P55201-2	Isoform 2 of Peregian [OS=Homo sapiens]	1.30644891	0.491180328	1	1	1220	138.099	BRPF1	-8.75	-8.55	-0.2	0	
High	P02533	Keratin, type I cytoskeletal 14 [OS=Homo sapiens]	59.71040561	26.9067797	15	146	472	51.529	KRT14	-8.7	-8.57	-0.13	0	
High	P43824	Plutelate GTP-binding protein 6 [OS=Homo sapiens]	21.310887	9.46612403	4	6	516	56.848	GTPBP6	0	-8.58	8.58	0	
High	Q9NW1-3	Fanci anemia group 1 protein [OS=Homo sapiens]	4;0705031	21.0842373	3	5	1238	149.229	FANCI	-8.79	-8.58	-0.2	0	
Medium	Q8IXM3	35S ribosomal protein L41, mitochondrial [OS=Homo sapiens]	2.74328225	7.25992707	1	1	137	15.373	MRP141	-8.81	-8.75	-0.06	0	
High	Q14617-5	Isoform 5 of AP-2 complex subunit delta-1 [OS=Homo sapiens]	6.36967915	1.06995855	1	2	1215	136.565	AP3D1	9.04	-8.83	-0.2	0	
High	P61204	ADP-ribosylation factor 3 [OS=Homo sapiens]	22.4366442	28.72928818	3	6	181	20.588	ARF3	-9.07	-9.01	-0.06	0	
Low	Q8UWM00-1	Transmembrane and coiled-coil domain-containing protein 1 [OS=Homo sapien	21.919061295	7.9787244	1	1	188	21.161	TMC01	-9.13	-9.07	-0.06	0	
High	P62316	Small nuclear ribonucleoprotein Sm D2 [OS=Homo sapiens]	9.26388775	24.5762712	2	2	18	13.518	SNRDP2	-9.16	-9.1	-0.06	0	
Low	P42345	Serine/threonine-protein kinase mTOR [OS=Homo sapiens]	1.66655597	0.4315178	1	1	2439	288.707	MTOR	-9.32	-9.15	-0.17	0	
Low	Q53HL2	Borealin [OS=Homo sapiens]	1.61403643	3.57142857	1	1	280	31.304	CDC48	-9.44	-9.28	-0.16	0	
High	Q9NTW7	Zinc finger protein 64 homolog, isoforms 3 and 4 [OS=Homo sapiens]	9.42200071	6.666666567	3	5	645	72.17	ZIPF64	-9.52	-9.33	-0.19	0	
Medium	Q9NSB2-1	protein FAM207A [OS=Homo sapiens]	3.314957521	5.21273913	1	1	230	25.441	FAM207A	-9.53	-9.46	-0.07	0	
Medium	P14618	Pyruvate kinase PMK [OS=Homo sapiens]	2.35704112	2.44821092	1	1	531	57.9	PKM	-9.64	-9.51	-0.13	0	
Low	Q1240-2	Isoform 2 of Eukaryotic initiation factor 4A-II [OS=Homo sapiens]	1.84771166	1.96078431	1	1	408	46.46	EIF4A2	-9.73	-9.54	-0.19	0	
Medium	P62937	Peptidyl-prolyl cis-trans isomerase A [OS=Homo sapiens]	3.12833887	8.48384188	1	1	165	18.001	PIPA	-9.62	-9.56	-0.06	0	
High	P01605	Ig kappa chain V-I region lay [OS=Homo sapiens]	21.79040992	25	2	19	1	108	11.827	SNRNP27	-9.67	-9.62	-0.06	0
Medium	P00747	Phosphoglycerate kinase [OS=Homo sapiens]	3.3482815	0.98765432	1	4	810	90.51	PLG	-9.82	-9.65	-0.17	0	
High	Q9QH99-2	Isoform 2 of SUN domain-containing protein 2 [OS=Homo sapiens]	12.9579532	5.6395664	5	6	738	82.452	SUN2	-9.83	-9.66	-0.17	0	
High	P10268-1	Serum albumin [OS=Homo sapiens]	15.5860401	9.638801314	6	21	609	69.321	ALB	-9.89	-9.73	-0.16	0	
Low	Q96TB8	Mxs2-interacting protein [OS=Homo sapiens]	1.91384017	0.272792576	1	6	1	3664	402.004	SPEN	-9.92	-9.78	-0.13	0
Medium	Q96NB2	Sideroflexin-2 [OS=Homo sapiens]	2.76170293	3.727670807	1	1	322	36.208	SFXN2	-9.87	-9.8	-0.07	0	
Medium	P07336	Complement C1r subcomponent [OS=Homo sapiens]	3.3575814	1.27659574	1	1	705	80.067	C1R	-9.95	-9.88	-0.07	0	
High	P07355-2	Isoform 2 of Annexin A2 [OS=Homo sapiens]	55.26388992	42.8571492	12	43	12	357	40.386	ANXA2	-9.95	-9.88	-0.07	0
High	P1368-1	Nascent polypeptide-associated complex subunit alpha, muscle-specific form [Q9T744]	15.5860401	9.638801314	6	21	609	82.452	SUN2	-9.83	-9.66	-0.16	0	
Low	P01764	Ig heavy chain V-III region 23 [OS=Homo sapiens]	7.40072724	16.2395162	1	2	117	12.574	IGHV3-23	-10.13	-9.94	-0.19	0	
High	Q9ZT4	Serine racemase [OS=Homo sapiens]	5.7961264	8.23529412	2	2	340	36.543	SRR	-10.23	-10.23	-0.07	0	
Medium	Q95Z73	Importin-7 [OS=Homo sapiens]	3.65332984	2.697495318	2	2	1038	119.44	IP07	-10.12	-10.11	-0.14	0	
High	P0C04-1	Complement C4-A [OS=Homo sapiens]	18.7308884	1.97574884	5	16	5	1744	192.664	C4A	-10.34	-10.18	-0.16	0
High	E9PAV3	Nascent polypeptide-associated protein subunit gamma [OS=Homo sapiens]	313.36054996	66.609589	3	3	2078	205.295	NACA	-9.96	-9.9	-0.06	0	
High	P02768-1	Histone H2A.Z [OS=Homo sapiens]	27.2140846	31.25	4	26	2	128	13.545	H2AFZ	0.37	-9.92	10.29	0
High	P01764	Ig heavy chain V-III region 23 [OS=Homo sapiens]	7.40072724	16.2395162	1	1	1042	117.729	SKIV2L2	-10.21	-10.21	-0.16	0	
High	Q9HCS7	Pre-mRNA-splicing factor SFY1 [OS=Homo sapiens]	8.14359667	3.27488538	3	3	855	99.946	XAB2	-10.26	-10.25	0	0	
High	P02746	Complement C1q subcomponent subunit B [OS=Homo sapiens]	15.0241623	5.53359684	1	15	1	253	26.704	C1QB	-10.4	-10.26	-0.14	0
Low	Q9UNI2-2	Isoform 2 of Transoocan-associated protein subunit gamma [OS=Homo sapiens]	1.60153915	2.80963303	5	1	1	198	22.596	SSR3	-10.4	-10.34	-0.06	0
Medium	P17482	Keratin, type I cytoskeletal 10 [OS=Homo sapiens]	3.22320749	5.2	1	1	250	28.041	H0XB9	-10.49	-10.43	-0.07	0	
High	P07477	Proteinase-3 [OS=Homo sapiens]	6.812322138	11.3360324	2	15	247	26.541	PRSS1	-10.69	-10.56	-0.13	0	
Low	P42285	Suppressor of cytokine signaling 1 [OS=Homo sapiens]	6.161475132	0.671795593	1	3	1997	226.611	OTOF	-10.79	-10.59	-0.2	0	
High	Q9HC10	Otoferlin [OS=Homo sapiens]	1.33451282	0.75112659	1	1	1997	231.226	MYH10	-10.87	-10.67	-0.2	0	
High	P35580-3	Isoform 3 of Myosin-10 [OS=Homo sapiens]	458.2245604	49.9248873	97	78	1997	62.027	KRT19	-10.81	-10.7	-0.11	0	
Medium	P35527	Keratin, type I cytoskeletal 9 [OS=Homo sapiens]	280.6622681	68.057749	38	494	37	623	44.767	C10orf54	-10.81	-10.75	-0.11	0
High	P06XAT0	Uncharacterized protein C12orf54 [OS=Homo sapiens]	2.29946934	13.3858258	1	1	127	14.476	C12orf54	-10.81	-10.75	-0.11	0	
High	P01859	Ig gamma-2 chain C region [OS=Homo sapiens]	7.046133134	10.7361963	2	15	326	35.878	IGHG2	-10.92	-10.77	-0.15	0	
High	O60264	SWI/SNF-related matrix-associated actin-dependent regulator of chromatin subunit 1 [OS=Homo sapiens]	4.99566813	2.18631179	2	2	1052	121.828	SMARCAS5	-10.88	-10.87	0	0	
High	Q8TF09	Dynein light chain rod/block-type 2 [OS=Homo sapiens]	4;94652156	12.5	1	1	96	10.848	DYNLRB2	-10.94	-10.88	-0.06	0	

High	P027245	Complement C1c subcomponent subunit A [OS=Homo sapiens]	7,07746598	4,483975592	1	8	1	245	26	C1Q0A	-13,11	-10,92	-0,19	0
Low	P23435	Cerebellin-1 [OS=Homo sapiens]	1,856046888	3,62694301	1	12	1	193	21,084	CBLN1	-11,17	-11,01	-0,16	0
High	P06753-2	Isoform 2 of Trypotryptase alpha-3 chain [OS=Homo sapiens]	19,4267759	26,6129032	6	7	4	248	29,015	TPN3	-13,15	-15,09	-0,07	0
High	P35908	Keratin, type II cytoskeletal 2, epidemal [OS=Homo sapiens]	314,418826	78,4037559	57	465	42	639	65,393	KRT2	-11,32	-11,18	-0,14	0
High	Q8N9E0	Protein FAM13A [OS=Homo sapiens]	26,4183991	18,1451613	5	26	2	248	28,923	FAM13A	-11,42	-11,29	-0,12	0
High	P06310	Ig kappa chain V-ii region RPMI 6410 [OS=Homo sapiens]	6,21360354	9,77443609	1	9	1	133	14,697		-11,44	-11,38	-0,06	0
High	Q94832	Unconventional myosin-1d [OS=Homo sapiens]	32,5981977	10,7353865	11	11	11	1006	116,129	MYO1D	-11,44	-11,44	0	0
Low	Q8VY11	Copine-9 [OS=Homo sapiens]	1,97224228	5,06329114	1	4	1	553	61,825	CPNE9	-11,69	-11,57	-0,11	0
High	O14950	Myosin regulatory light chain 12B [OS=Homo sapiens]	10,835137	18,0232558	3	4	3	172	19,767	MYL2B	-11,7	-11,63	-0,06	0
Medium	P044334	Ig kappa chain V-iii region VH [OS=Homo sapiens]	4,18002199	12,9310345	2	5	1	116	12,749	IGKV3-7	-11,76	-11,7	-0,07	1
Low	Q9H019	Poly [ADP-ribose] polymerase 12 [OS=Homo sapiens]	1,22380749	2,2845364	1	1	1	701	79,013	PARP12	-11,89	-11,7	-0,19	0
Low	Q5U67-1	Uncharacterized protein C9orf117 [OS=Homo sapiens]	1,93330145	2,11538462	1	1	1	520	60,496	C9orf117-CEA121	-11,77	-11,71	-0,06	0
High	O43795	Unconventional myosin-1b [OS=Homo sapiens]	27,2572044	11,7077465	9	9	9	1136	131,902	MYO1B	-11,79	-11,78	0	0
High	P06314	Ig kappa chain V-iv region B17 [OS=Homo sapiens]	10,7214567	23,1343284	3	33	3	134	14,956		-12,04	-11,88	-0,17	0
High	Q8ZY91-4	Isoform 4 of kinase-like protein KIF18B [OS=Homo sapiens]	7,70165384	2,29095074	2	3	2	873	95,069		-11,93	-11,93	0	0
High	P04264	Keratin, type II cytoskeletal 1 [OS=Homo sapiens]	404,0633232	63,5093168	53	806	44	644	65,999	KRT1	-12,14	-12,03	-0,11	1
Medium	P62314	Small nuclear ribonucleoprotein Sm D1 [OS=Homo sapiens]	4,39340369	10,9243697	1	1	1	119	13,273	SNRP1	-12,09	-12,03	-0,06	0
High	P01876	Ig alpha-1 chain C-region [OS=Homo sapiens]	16,5063344	17,84702355	5	9	5	353	37,631	IGHA1	-12,23	-12,23	0	0
Medium	P01023	Alpha-2-macroglobulin [OS=Homo sapiens]	3,82179886	1,01763908	2	6	2	1474	163,188	A2M	-12,39	-12,33	-0,06	0
High	P06060	Myosin light polypeptide 6 [OS=Homo sapiens]	36,266485	47,0198675	7	14	7	151	16,919	MYL6	-12,71	-12,65	-0,06	0
High	P01024	Complement C3 [OS=Homo sapiens]	33,2572719	4,98098016	6	18	6	1663	187,03	C3	-12,8	-12,71	-0,09	0
High	Q001159-1	Unconventional myosin-1c [OS=Homo sapiens]	55,8664137	18,2502352	20	24	20	1063	121,606	MYO1C	-12,73	-12,72	0	0
Low	Q3W672-1	Girdin [OS=Homo sapiens]	14,3711262	0,64136825	1	2	1	1871	215,909	CCDC88A	-13,19	-13,14	-0,06	0
High	P01765	Ig heavy chain V-iii region T1L [OS=Homo sapiens]	5,1094044	7,8268696	1	4	1	115	12,348		-13,43	-13,24	-0,19	0
Low	Q15247	Chloride intracellular channel protein 2 [OS=Homo sapiens]	1,60380065	6,07287449	1	5	1	247	28,338	CLIC2	-14,74	-14,55	-0,19	0

**Annex Table IV: Liquid chromatography-tandem mass spectrometry (LC-MS/MS) data from SILAC (MMC treatment).** The data were analyzed using SEQUEST in Proteome Discoverer 2.1 and searched in the complete human proteome database (Swiss-Prot). Peptide identification was filtered at a false discovery rate (FDR) < 1%. Coverage: percentage of the protein sequence covered by identified peptides (confident coverage threshold >1). Peptides: total number of distinct peptide sequences identified in the protein group. PSMs: total number of identified peptide spectra matched for the protein. Unique peptides: The number of peptide sequences that are unique to a protein group. MW: molecular weight. Abundance ratios are calculated based on light (for XPF-KO cells untreated), medium (for XPF-KO + XPF-Wt treated with MMC 0.5mg/mL during 21h), heavy (for XPF-KO + XPF-Wt untreated) forms of lysine and arginine. Razor peptides: peptides that can be assigned to more than one protein.