Research proposal: Can heavy metals protect Brassica napus from pathogens and pests?

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Background

Some plants show unusually high levels of heavy metals accumulation in their tissues (Brooks et al. 1977). Over 500 plant species and 0,2 per cent of all angiosperms have been described as metal hyperaccumulators (Sarma, 2011). Many hypotheses have been raised to explain such phenomenon. One of these suggests that heavy metal accumulation can protect plants from pathogens and pests (Boyd and Martens, 1992).

Several studies have focused on determining whether metals can defend plants from biotic stress (e.g., Fones et al. 2010; Poschenrieder et al. 2011). They reached the conclusion that, sometimes, it is actually possible.

Plant defense by metals is rellevant on many fields such as phytorremediation, phytomining or metallomics (Boyd, 2007), which may benefit from further research. Also, new insights on specificity and cross talk in plant responses to environmental stresses can be provided (Poschenieder et al. 2006).

Objectives and hypotheses

The objectives of the research project are to:

- 1) Obtain new information about the biology and ecology of Brassica napus
- Provide additional insights on the role of heavy metals in plant-pathogen and plant-herbivore interactions
- 3 Become a data source for further Brassica napus studies or applied uses

The inital hypotheses are the following:

- Heavy metals accumulation can protect Brassica napus from pathogens
 and pests
- (2) Different metals offer different grades of protection



Why Brassica napus?

It is cultivated worldwide
Many uses

Vegetal oil production Phytorremediation Biofuel production Animal fodder

High biomass production Well known pathogens and pests Defense by metals not studied yet

Which metals will be tested?

Cd and Zn

Its accumulation in *B. napus* leaves has been proven and quantified

Using two metals allows to test whether they offer different protection

concentrate along the food chain



Materials and methods

Experiment 1: Metal concentration in plant

Objective: Determine which concentration of Cd and Zn should plants be exposed to for an optimum accumulation and quantify metal incorporation into leaves.

Procedure: Different hydroponic solutions with a variable metal concentration will be prepared. Plants will be visually examined. The higher metal concentration showing no apparent symptoms of toxicity in plants will be used in the following experiments. Metal concentration in tissue will be quantified by inductively coupled plasma spectrometry.

[CdsO,] or [ZnsO,]



Experiment 2: Biotic factors toxicity evaluation

Objective: Determine whether pathogens and herbivores are susceptible to Cd and Zn.

Procedure: Pathogens tolerance will be estimated by inoculating the fungi on to MEA plates and mesuring resultant mycelum diameter.

Herbivores will be placed on metal treated and non treated plants and their fresh weight gain will be measured.



Experiment 3: Pathogen infection

Objective: Verify whether accumulation of Cd or Zn in B. napus protects it from *Alternaria brassicicola* and *Peronospora parasitica* infection.

Procedure: Inoculation of pathogens will be carried out by spraying plants with a spore suspension. 4 days after inoculations, the number of lesions per leaf will be counted as a measure of plant infection degree

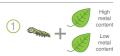




Experiment 4: Herbivore choice feeding

Objective: Verify whether accumulation of Cd or Zn in B. napus protects it from *Pieris rapae* and *Helix aspersa* herbivory.

Procedure: One herbivore and two pieces of leaf will be placed on a recipient, one containing high metal concentration and one containing low metal concentration. Leaves fresh weight will be measured before and after the experiment. Feeding will be quantified by measuring fresh weight loss.

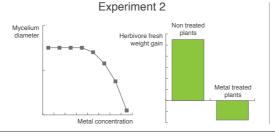


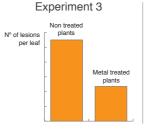


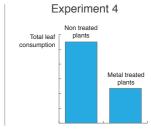
Expected results

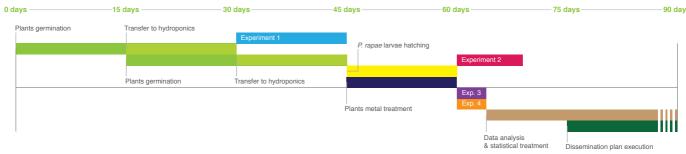
How expected results would look like

Working plan









BARCELG J and POSCHENHIEDER C. (2003) Phytomenoidation: principles and perspectives. Contributions to Science 2 (6): 332-8070 PS and MARTENS SN. (1992) The stand or diet for trend in plyenizacionalisation by given in: The Vegetiation of Unbrandic, Ceperative Solin, p. 277-856 E. Edister A. BOYD, R. (2007) The defense hypothesis of elemental hyperaccomutation: status, challenges and rew deriction. Plant Sol 2013-159.

BROOKS RR, LEE J, REEVES RD and JAWRÉT. (1973) The defense hypothesis of elemental hyperaccomutation: status, challenges and rew derictions. Plant Sol 2013-159.

BROOKS RR, LEE J, REEVES RD and JAWRÉT. (1973) Contribution demental analyses by plasma entesion specificationy. FASSEL W. (1978) Quantitative elemental analyses by plasma entesion specificationy. Science 2021: 159.

FONESH, DAVIS GAR, RICO A, FRANC F, SMITH AC, et al. (2010) Media Hyperaccomutation rhormer Plants against Disease. PLOS STRIPA 6009; et 001