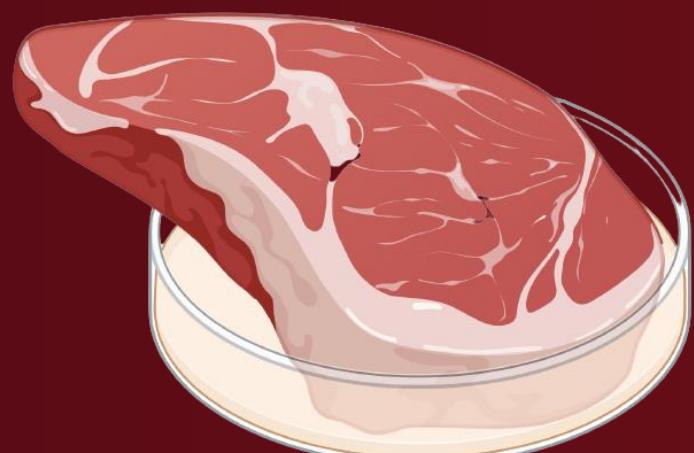

This is the **published version** of the bachelor thesis:

Pérez Mateos, Paula. A novel approach to develop cultured meat : design and simulation of a large-scale production plant. 2022. (815 Grau en Biotecnologia)

This version is available at <https://ddd.uab.cat/record/263086>

under the terms of the  license



A NOVEL APPROACH TO DEVELOP CULTURED MEAT: DESIGN AND SIMULATION OF A LARGE-SCALE PRODUCTION PLANT

Part II: Upstream and Bioreaction
Dimitri Gómez I., Pérez Mateos P., Ramírez Gómez D., Torres Domínguez L.
| Bachelor's Degree in Biotechnology (2018-2022)

INTRODUCTION

The aim of the present work is to cover a 1 % of the current market for meat substitutes in Amsterdam, so production will be 7 metric tons of cultured meat per year.

For accomplishing this goal, a large-scale process has been designed for the culture of myocytes, which comprehend two stages, proliferation and differentiation. Bioprocess is composed of:

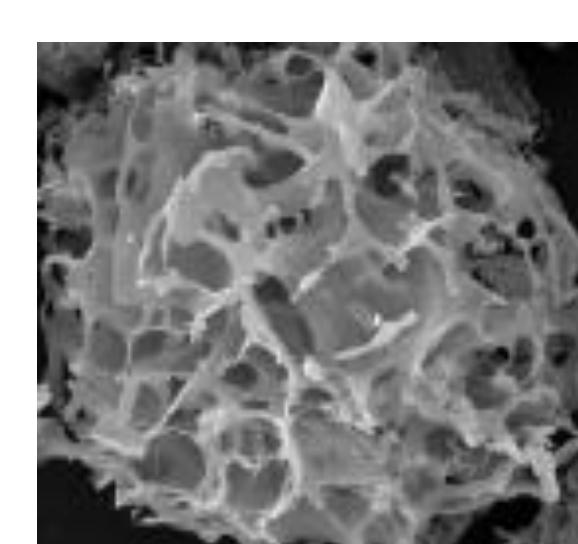
- Two 5000L industrial reactors working overlapped.
- One line semi-continuous downstream.

SCAFFOLDS

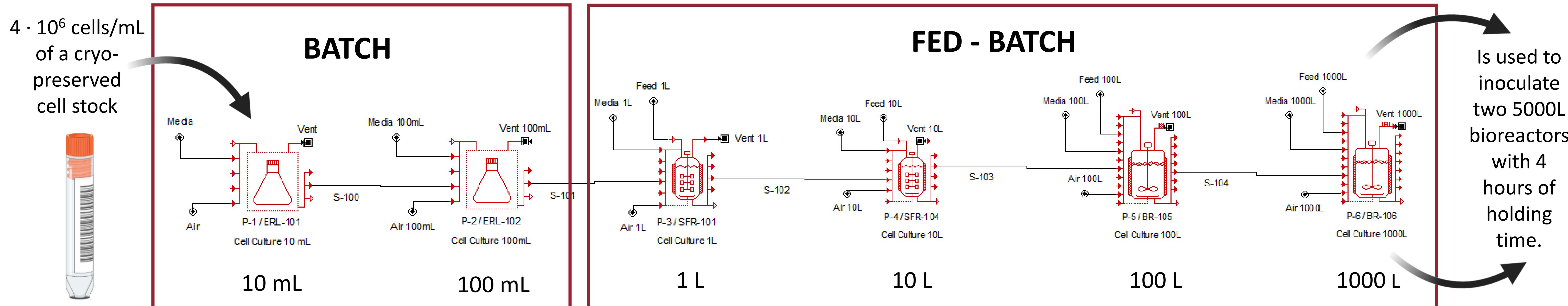
Scaffold is required because satellite stem cells need to grow under adhesion. The scaffold selected are fungal chitosan microcarriers previously described:

- Diameter 150 μm
- Porosity 98%
- Density 0.04g/ m^3
- Specific Surface Area 30 m^2/g
- Ratio Cells-Scaffold 63.3 kg/kg

**Bead-to-bead transfer
&
NO internal diffusional
limitations**



SEED TRAIN



PROLIFERATION

Initial values:

- [Glucose] \rightarrow 1 g/L
- [Glutamine] \rightarrow 0.115 g/L
- Scaffold \rightarrow 1.675 kg

Cell density $4 \cdot 10^5$ cells/mL

Proliferation Kinetics Equations:

$$\frac{dXv}{dt} = (\mu - \mu d) \cdot Xv$$

$$\frac{d[GLC]}{dt} = - \left(\frac{(\mu - \mu d)}{Y_{Xv/glc}} + m_{glc} \right) \cdot Xv + feed1$$

$$feed1 = \left(\frac{(\mu - \mu d)}{Y_{Xv/glc}} + m_{glc} \right) \cdot Xv$$

$$\frac{d[GLN]}{dt} = - \left(\frac{(\mu - \mu d)}{Y_{Xv/gln}} + m_{gln} \right) \cdot Xv - d_{gln}[GLN] + feed2$$

$$\frac{d[Lac]}{dt} = Y_{lac/glc} \left(\frac{(\mu - \mu d)}{Y_{Xv/glc}} + m_{glc} \right) \cdot Xv$$

$$\frac{d[Amm]}{dt} = Y_{amm/gln} \cdot \left(\frac{(\mu - \mu d)}{Y_{Xv/gln}} \right) \cdot Xv + d_{gln}[GLN]$$

$$feed2 = \left(\frac{(\mu - \mu d)}{Y_{Xv/gln}} + m_{gln} \right) \cdot Xv + d_{gln}[GLN]$$

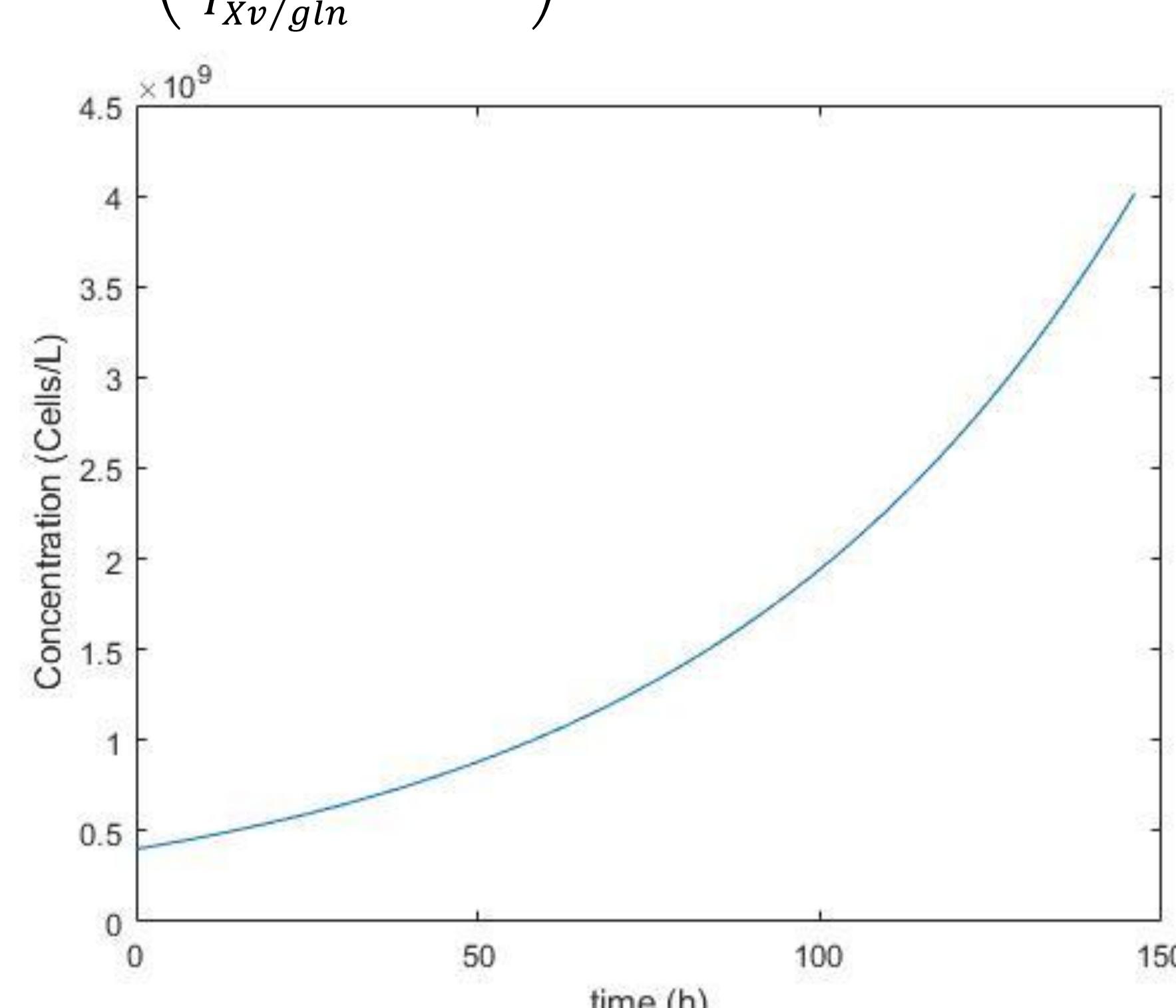


Figure 1. Representation of cell concentration in cells/L vs time in hours at the proliferation stage.

BIOREACTION

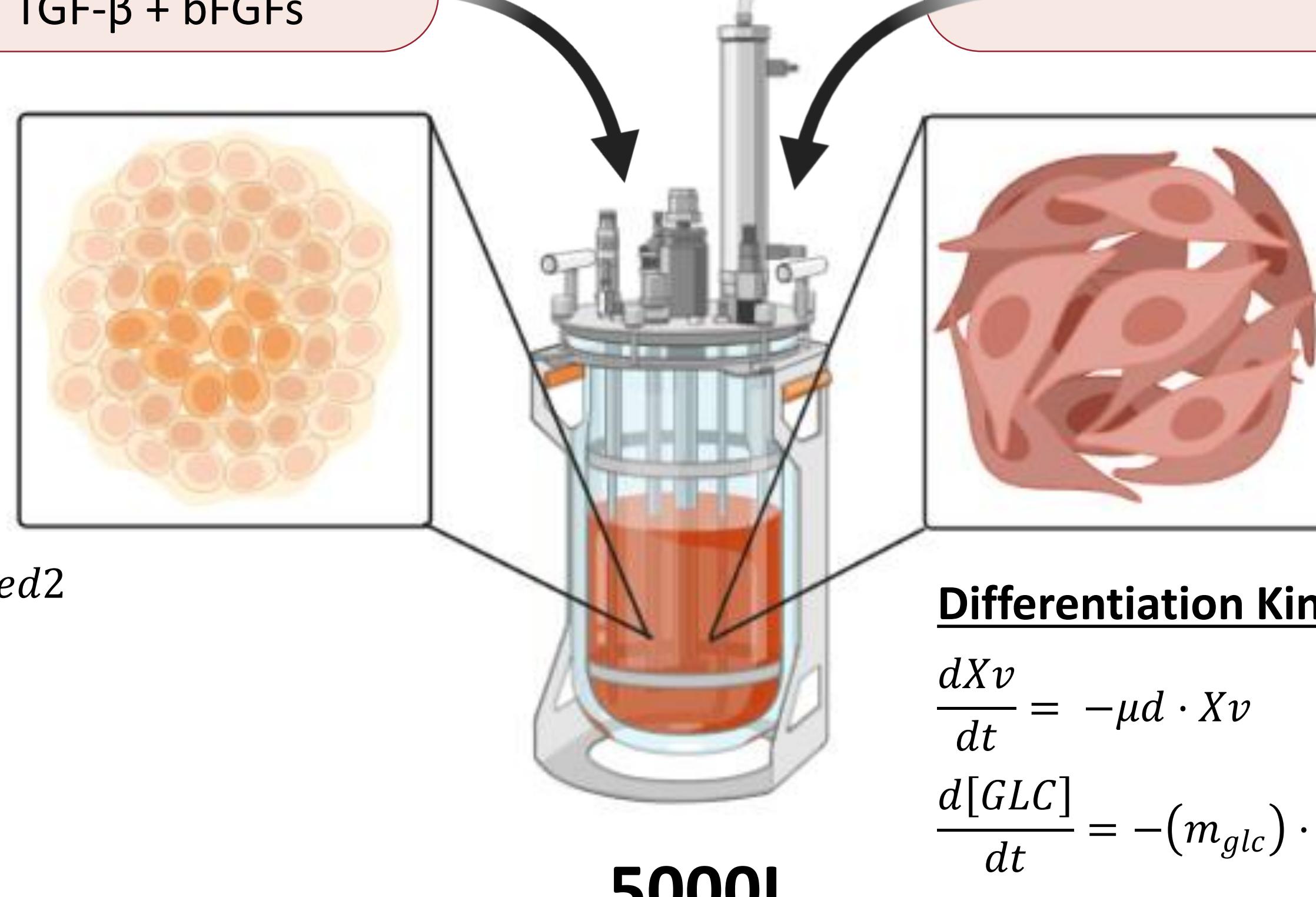
FED-BATCH

6.08 days

6.25 days

PROLIFERATION FEED
59.5 L
251.13 g/L-Glucose
25.33 g/L-Glutamine
TGF- β + bFGFs

DIFFERENTIATION FEED
72.5 L
541.12 g/L-Glucose
14.16 g/L-Glutamine



DIFFERENTIATION

Final values:

- [Glucose] \rightarrow 1 g/L
- [Glutamine] \rightarrow 0.115 g/L
- Scaffold \rightarrow 1.675 kg

By-products:

- [Lactate] \rightarrow 5.22 g/L
- [Ammonium] \rightarrow 0.048 g/L

97% viability \rightarrow $4 \cdot 10^6$ viable cells/mL

106 kg/bioreactor (x2) \rightarrow 212 kg/bioprocess' run

70 % differentiation

Differentiation Kinetics Equations:

$$\frac{dXv}{dt} = -\mu d \cdot Xv$$

$$\frac{d[GLC]}{dt} = -(m_{glc}) \cdot Xv + feed3$$

$$\frac{d[GLN]}{dt} = -(m_{gln}) \cdot Xv - d_{gln}[GLN] + feed4$$

$$\frac{d[Lac]}{dt} = Y_{lac/glc} \cdot m_{glc} \cdot Xv$$

$$\frac{d[Amm]}{dt} = d_{gln}[GLN]$$

$$feed3 = m_{glc} \cdot Xv$$

$$feed4 = m_{gln} \cdot Xv - d_{gln}[GLN]$$

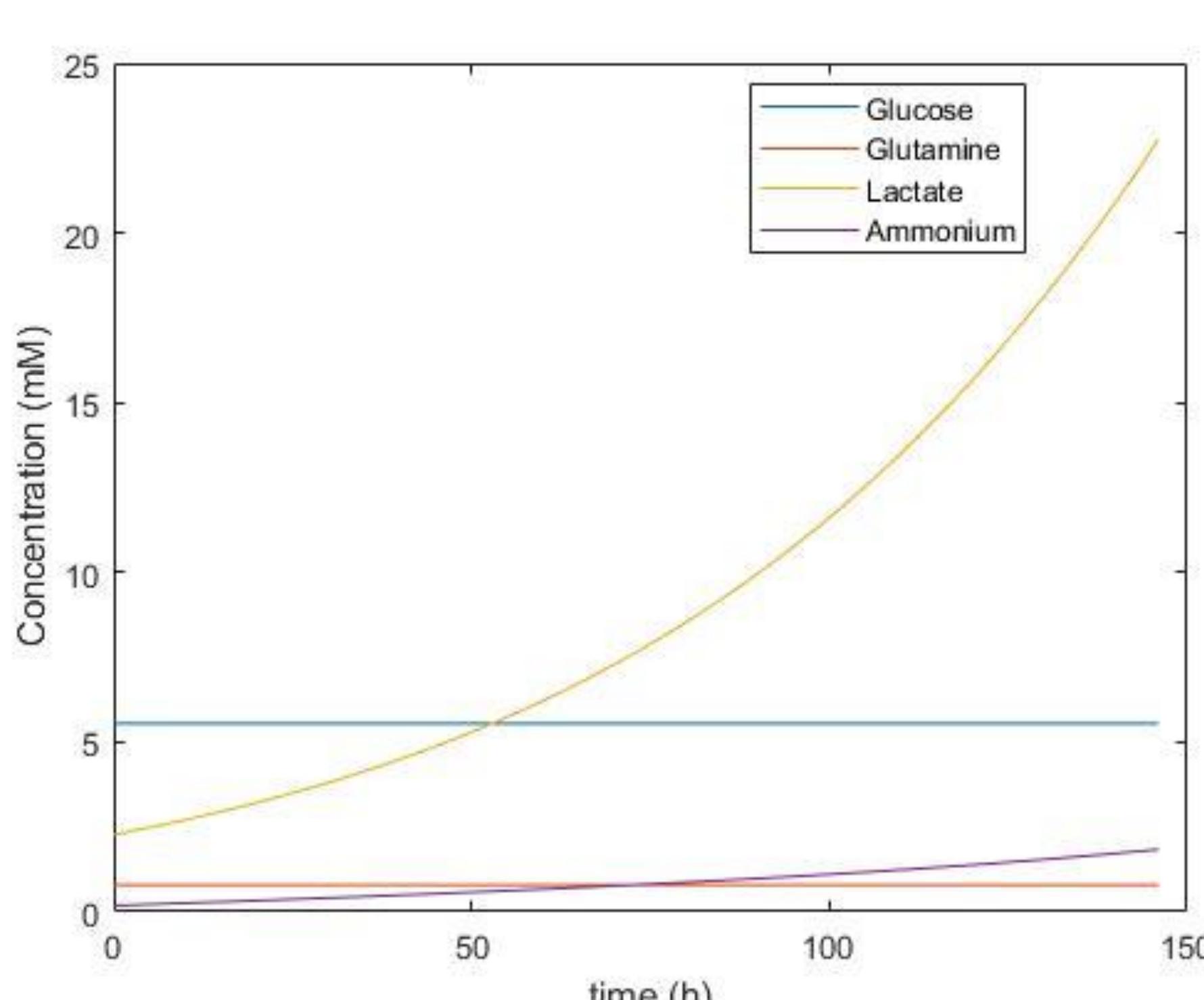


Figure 2. Evolution profile of glucose and glutamine, and of the by-products generated (lactate and ammonium) during the proliferation stage.

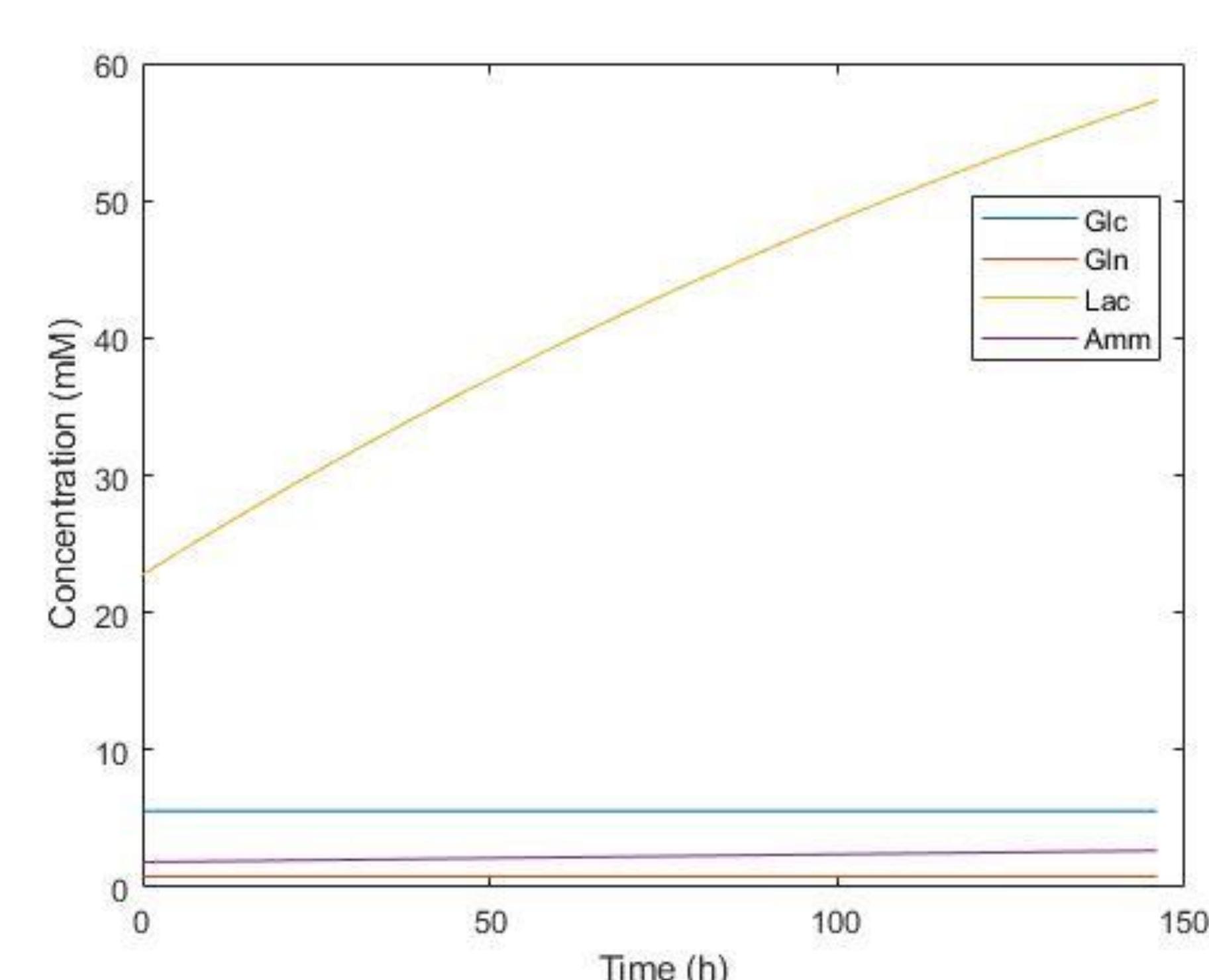


Figure 3. Evolution profile of glucose and glutamine, and of the by-products generated (lactate and ammonium) during the differentiation process.

CONCLUSIONS

In this work, we successfully carried out the simulation of a manufacturing process for the production of cell cultured meat. In summary, we started our process with a frozen vial of cells and achieved a total production of 212 kg of meat per bioprocess' run using two 5000L STR bioreactors with a process time of 12.33 days.

BIBLIOGRAPHY

1. Huang, L., Xiao, L., Jung Poudel, A., Li, J., Zhou, P., Gauthier, M., Liu, H., Wu, Z., & Yang, G. (2018). Porous chitosan microspheres as microcarriers for 3D cell culture. *Carbohydrate Polymers*, 202, 611-620.
2. Xing, Z., Bishop, N., Leister, K., & Li, Z. J. (2010). Modeling kinetics of a large-scale fed-batch CHO cell culture by Markov chain Monte Carlo method. *Biotechnology Progress*, NA.
3. Mattick, C. S., Landis, A. E., Allenby, B. R., & Genovese, N. J. (2015). Anticipatory Life Cycle Analysis of In Vitro Biomass Cultivation for Cultured Meat Production in the United States. *Environmental Science & Technology*, 49(19), 11941-11949.
4. Meyer, H.-P., Minas, W., & Schmidhalter, D. (2017). Industrial-scale fermentation. In *Industrial biotechnology: Products and processes* (pp. 1-53). Wiley, New Jersey, United States.